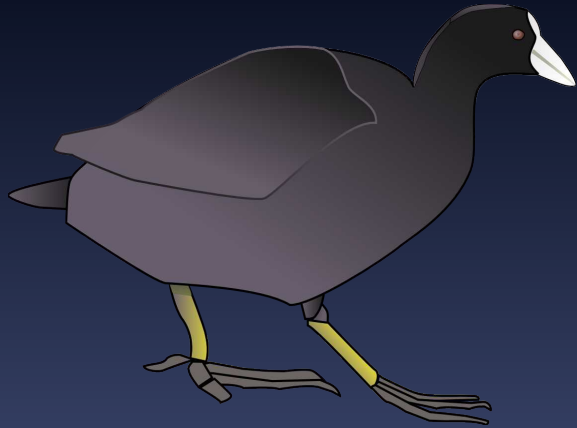


December 2011 Okinawa



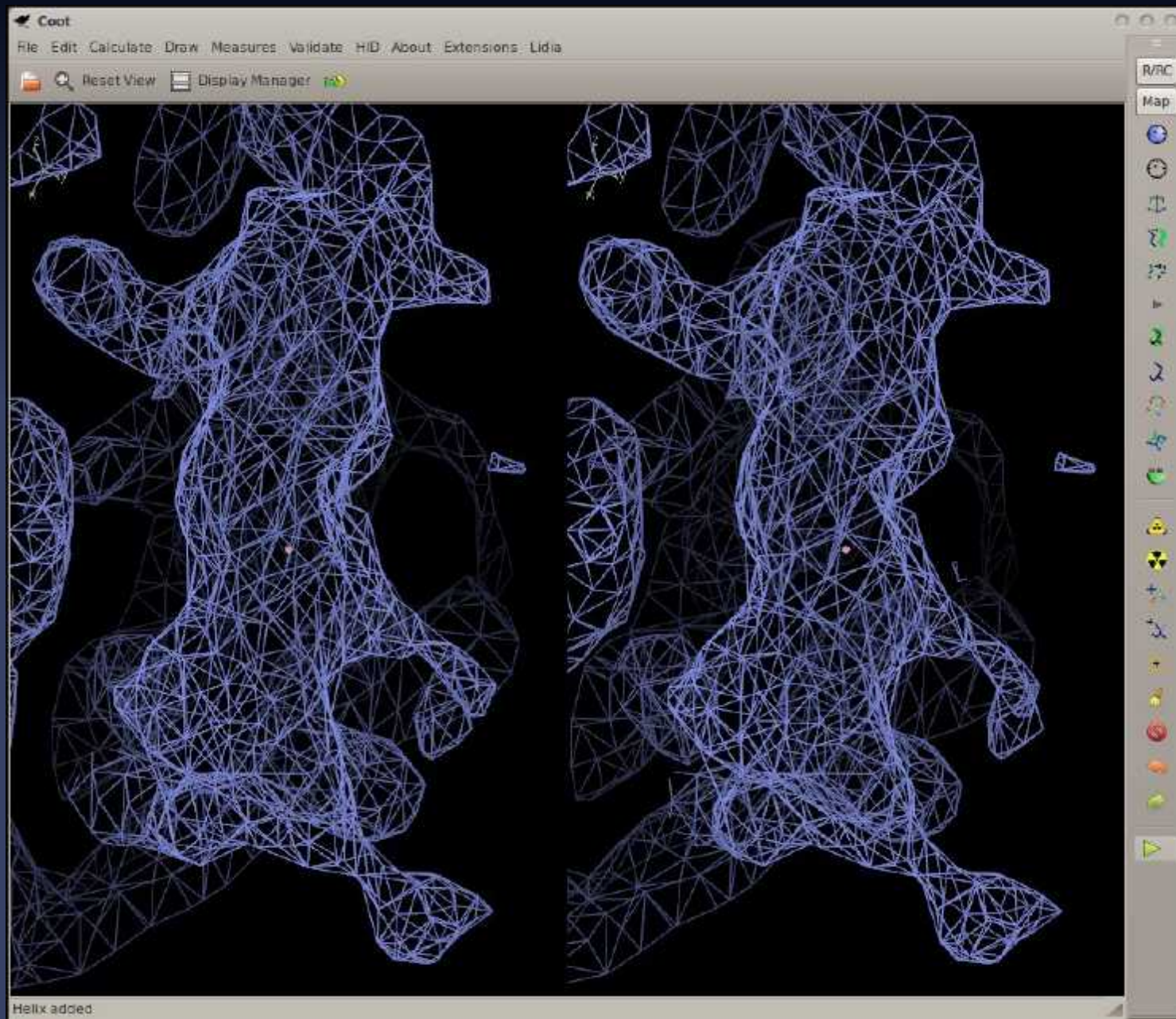
Model-Building with Coot

Secondary structure tools

Bernhard Lohkamp

Karolinska Institutet

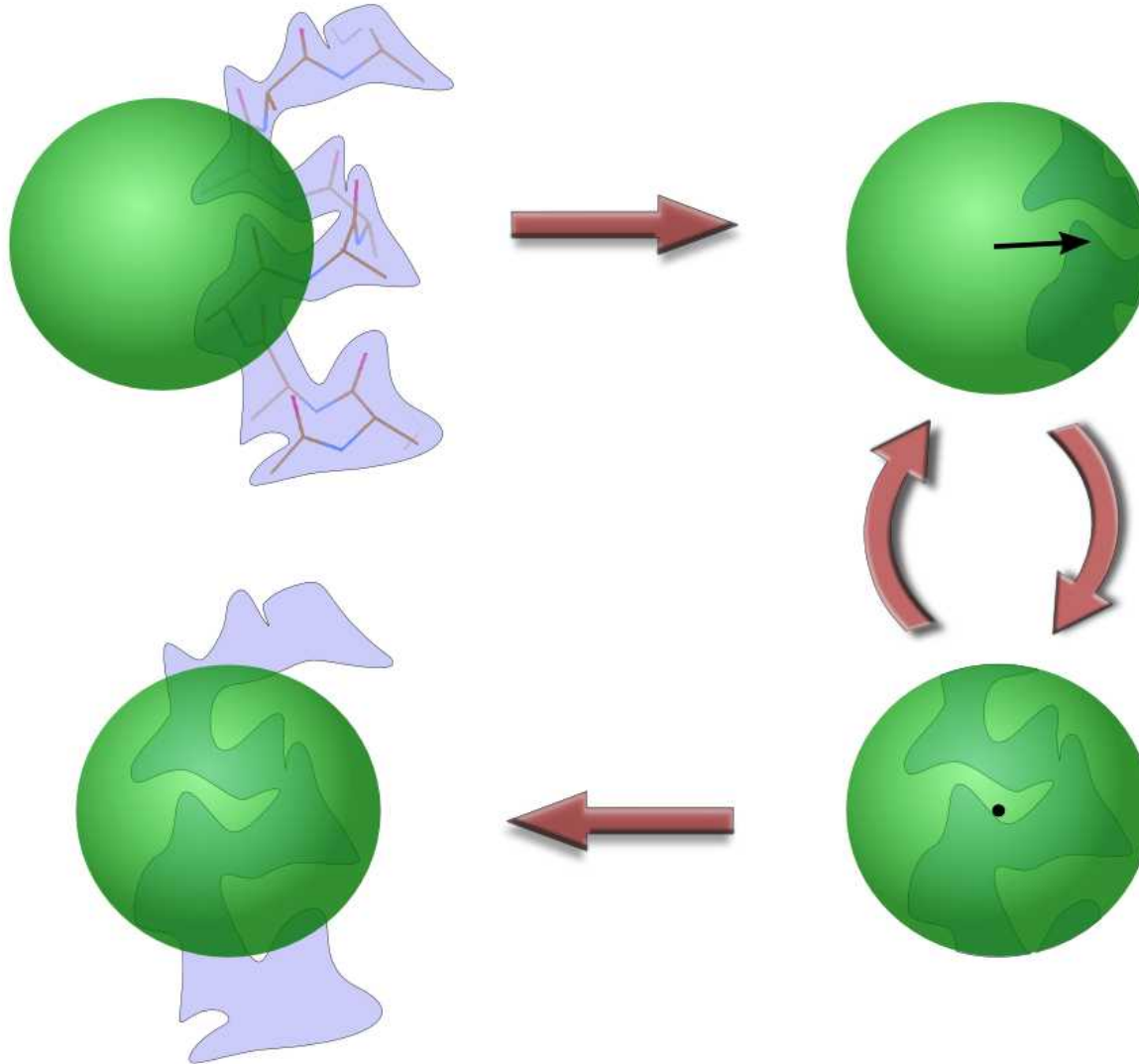
Helix-Building



Alpha Helix Placement

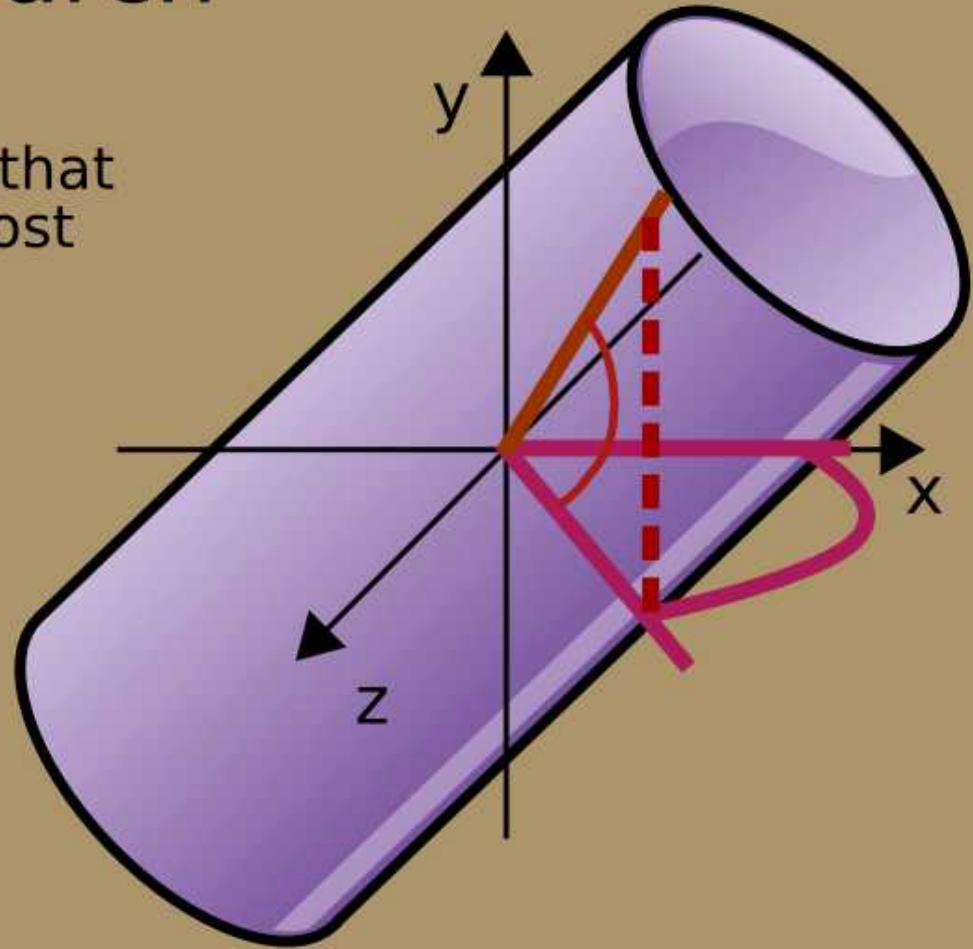
- Scenario: Looking at a new map, not built with automatic tools:
 - “I can see that there’s a helix here - build it for me!”
- From a given point:
 - Move to local averaged maximum
 - Do a 2D MR-style orientation search on a cylinder of electron density
 - Build a helix (both directions)
 - 1D Rotation search to find best fit
 - Score based on density at CB positions
 - Trim ‘n Grow

Centering the Rotation point

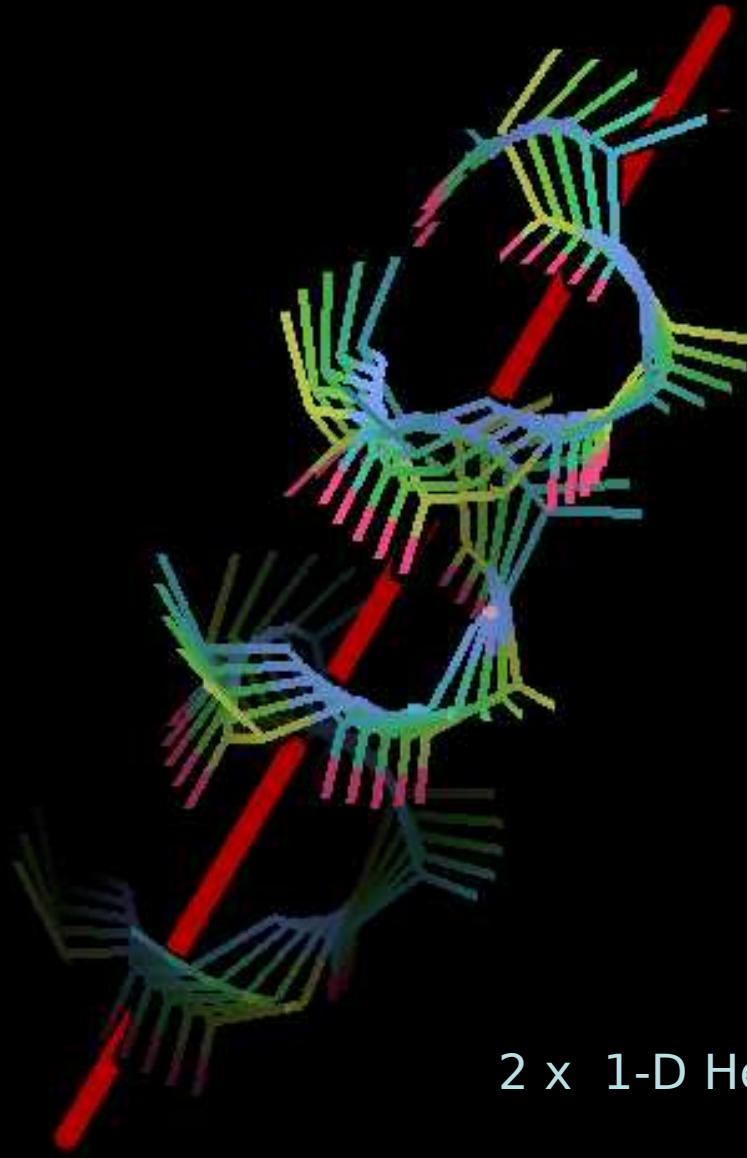


Cylinder Search

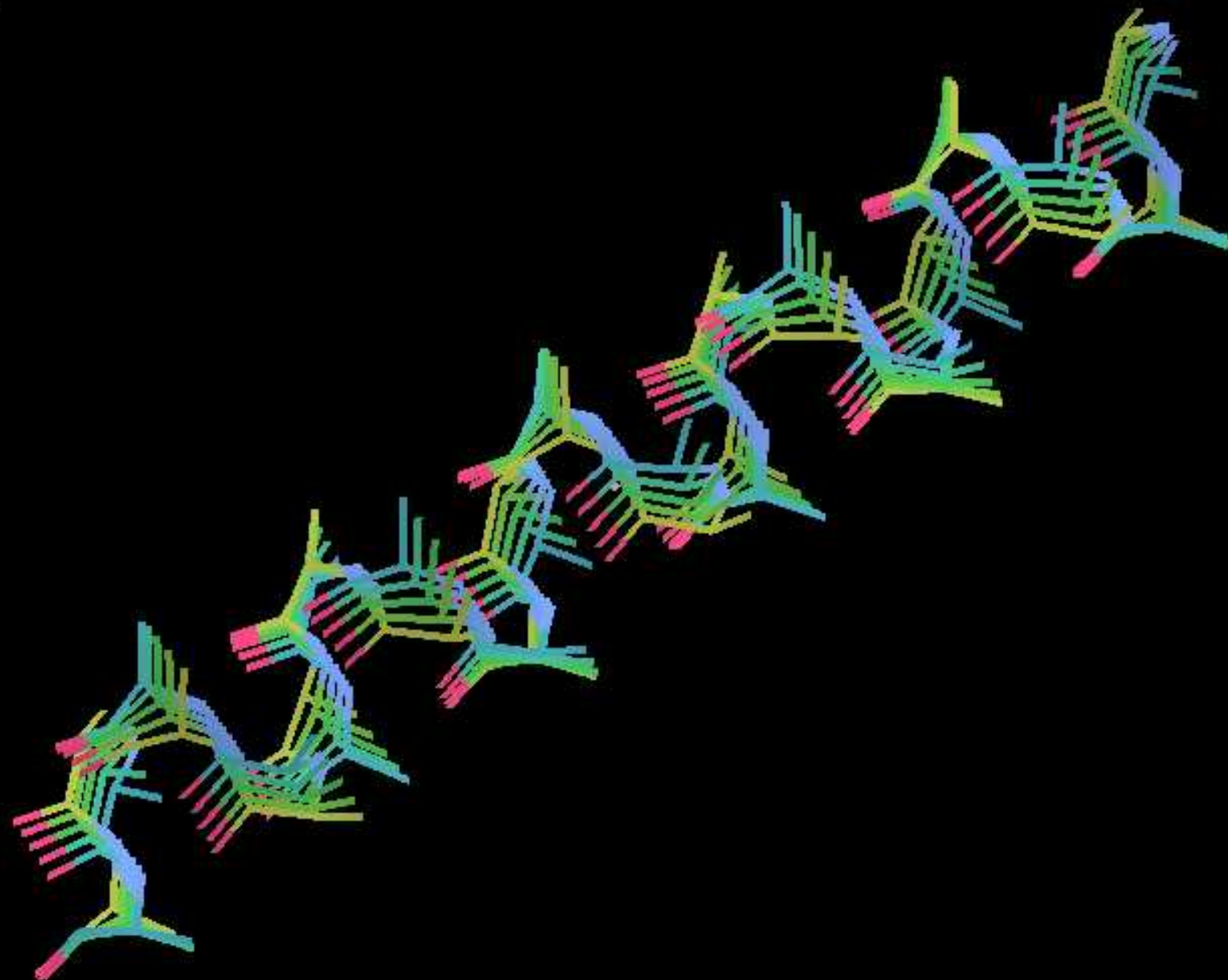
Pick the orientation that encapsulates the most electron density

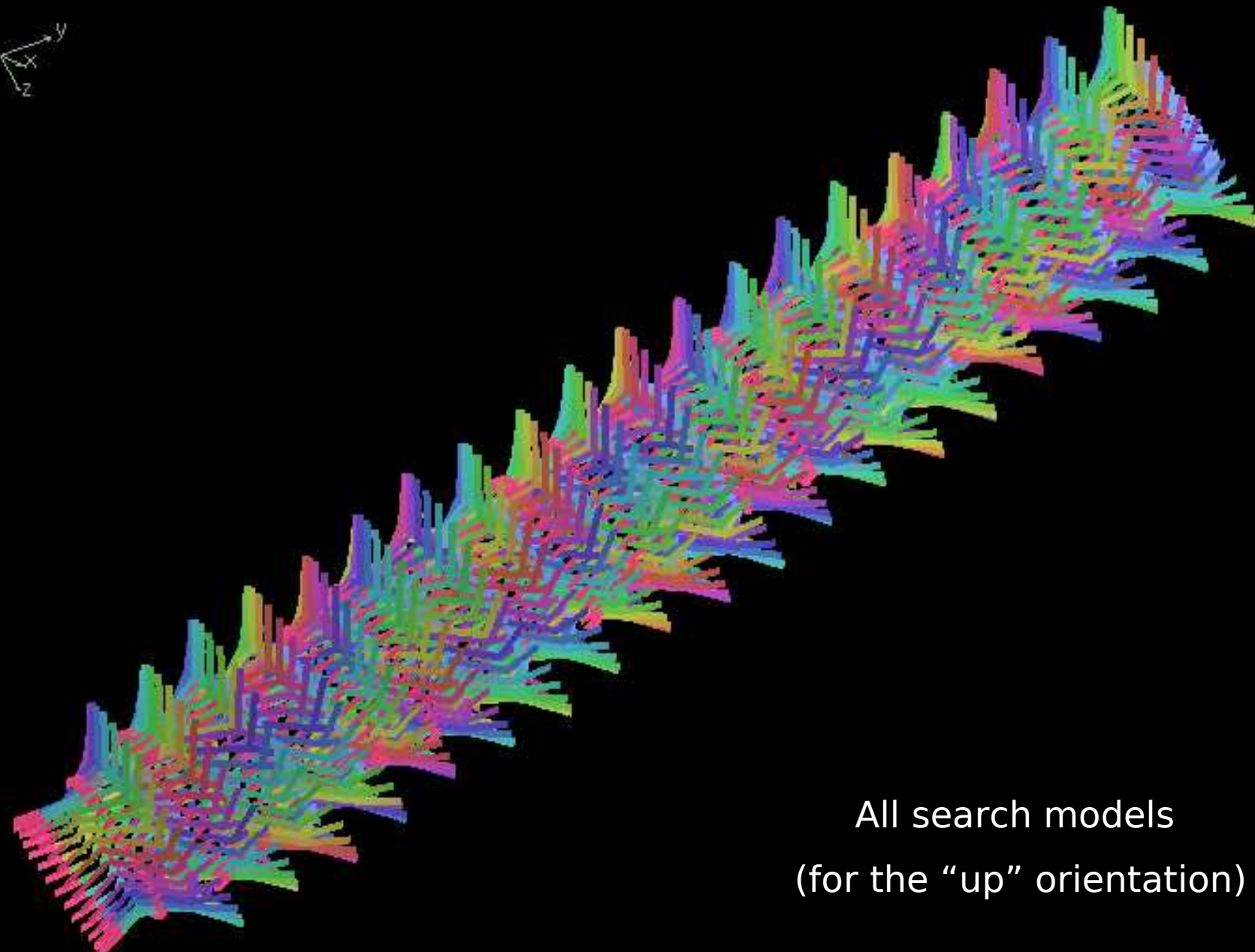


2 orientation axes

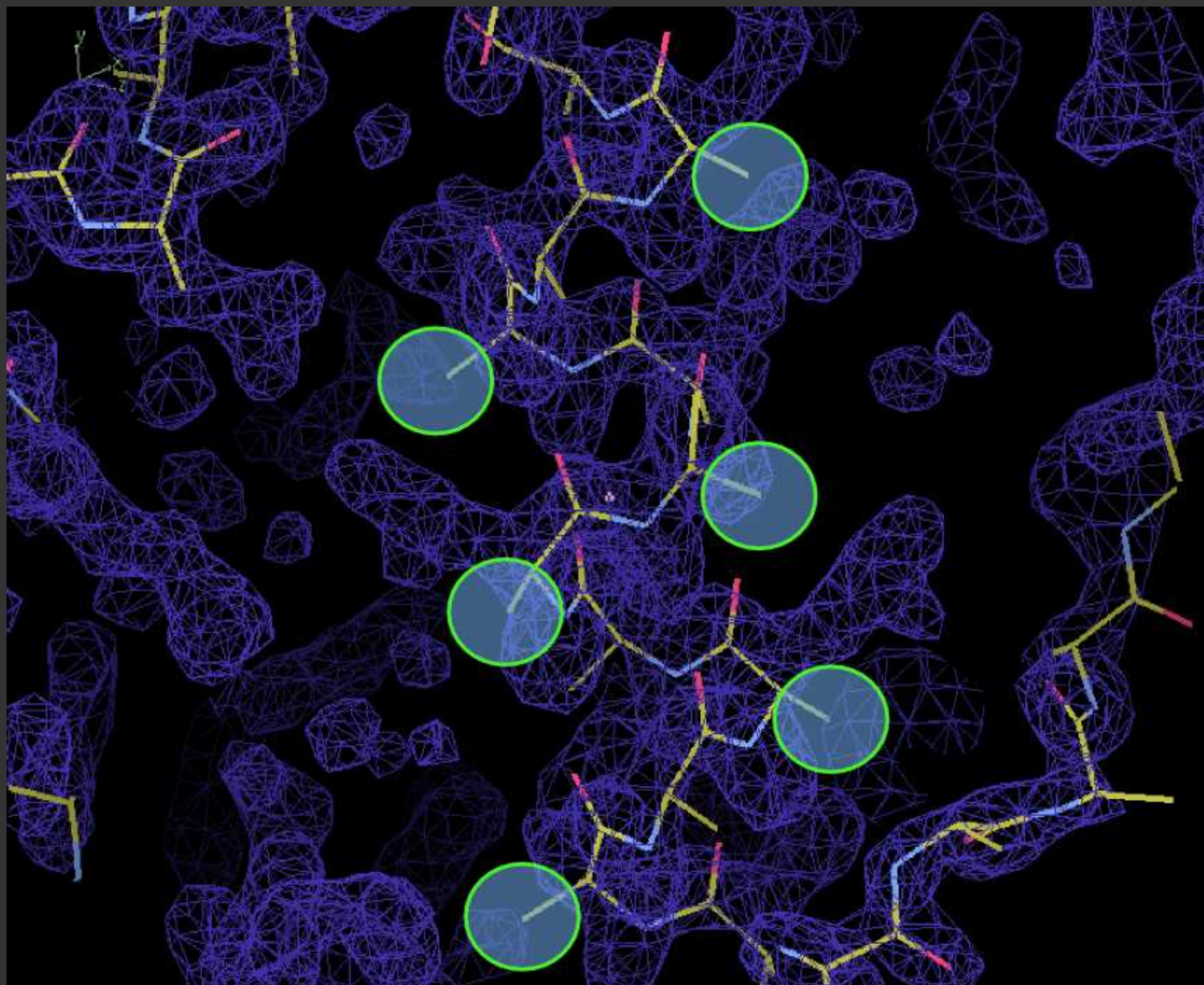


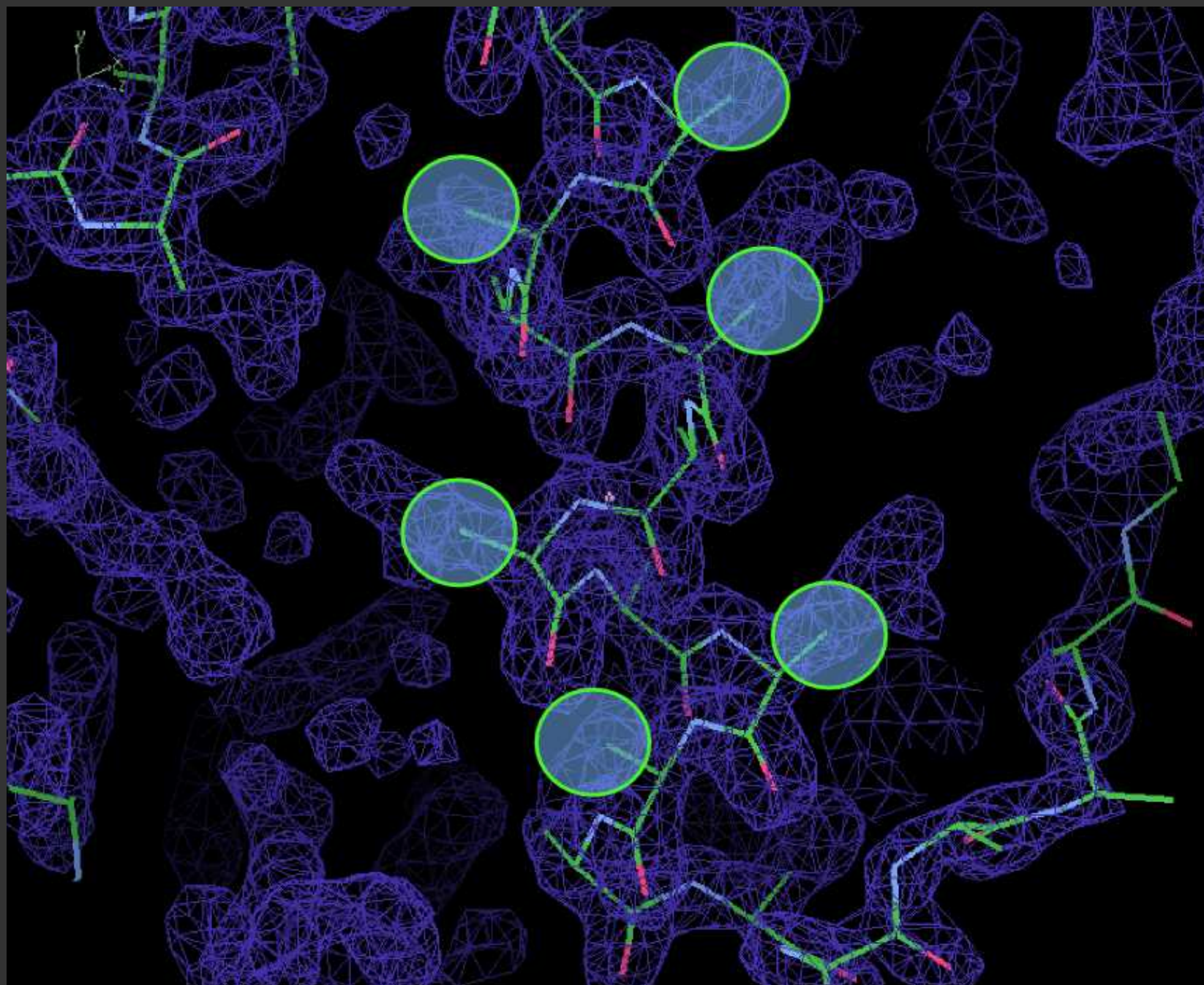
2 x 1-D Helix orientation searches





All search models
(for the “up” orientation)





Fitting Strands

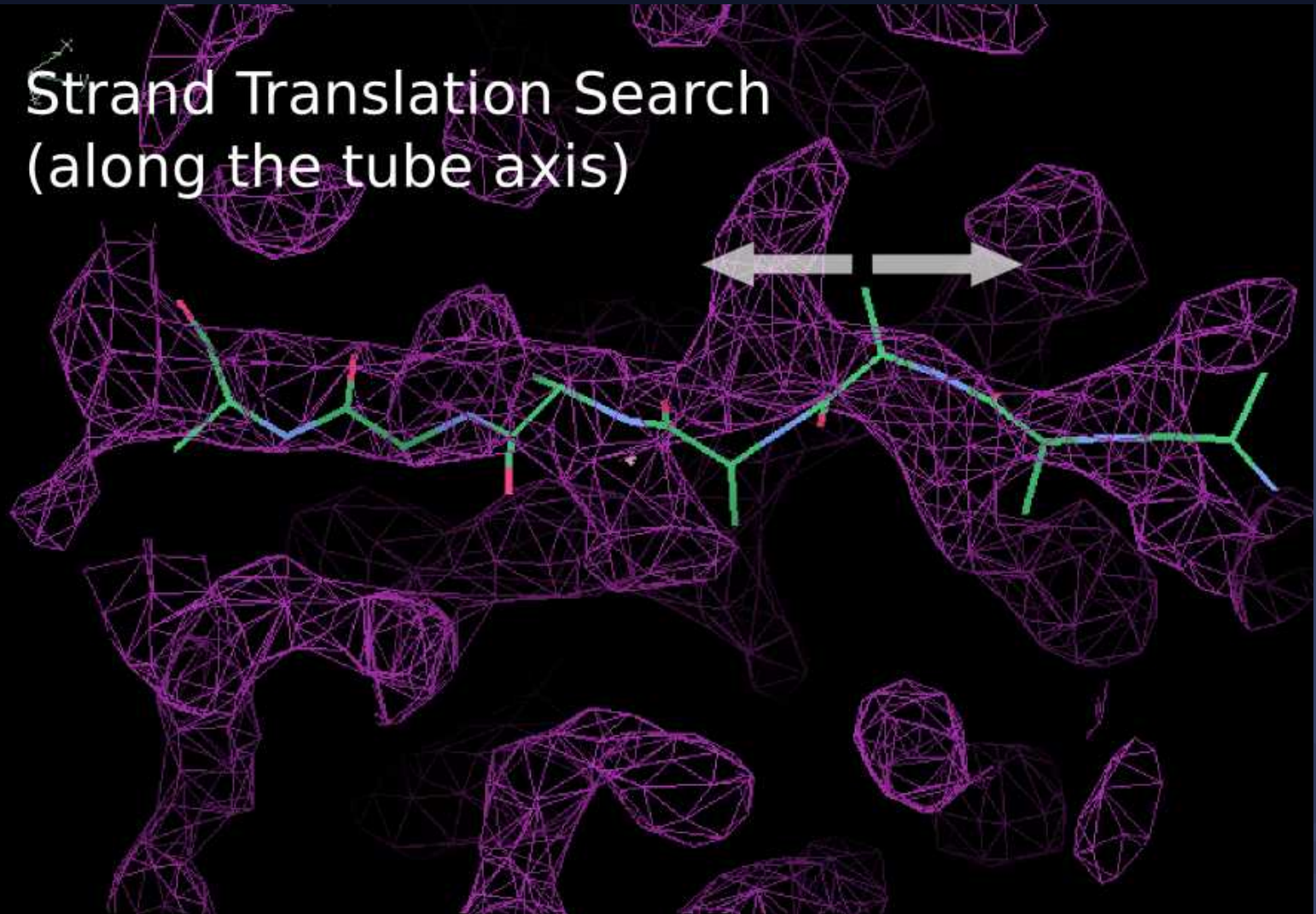
Placing Strands

- Unlike Helices, Strands have to be treated as non-idealized
 - ◆ Repeating a single phi/psi value doesn't make a structure that fits “real-world” density
- Curvature of strands should be taken into account
 - ◆ Use selections from a “database” of good structures

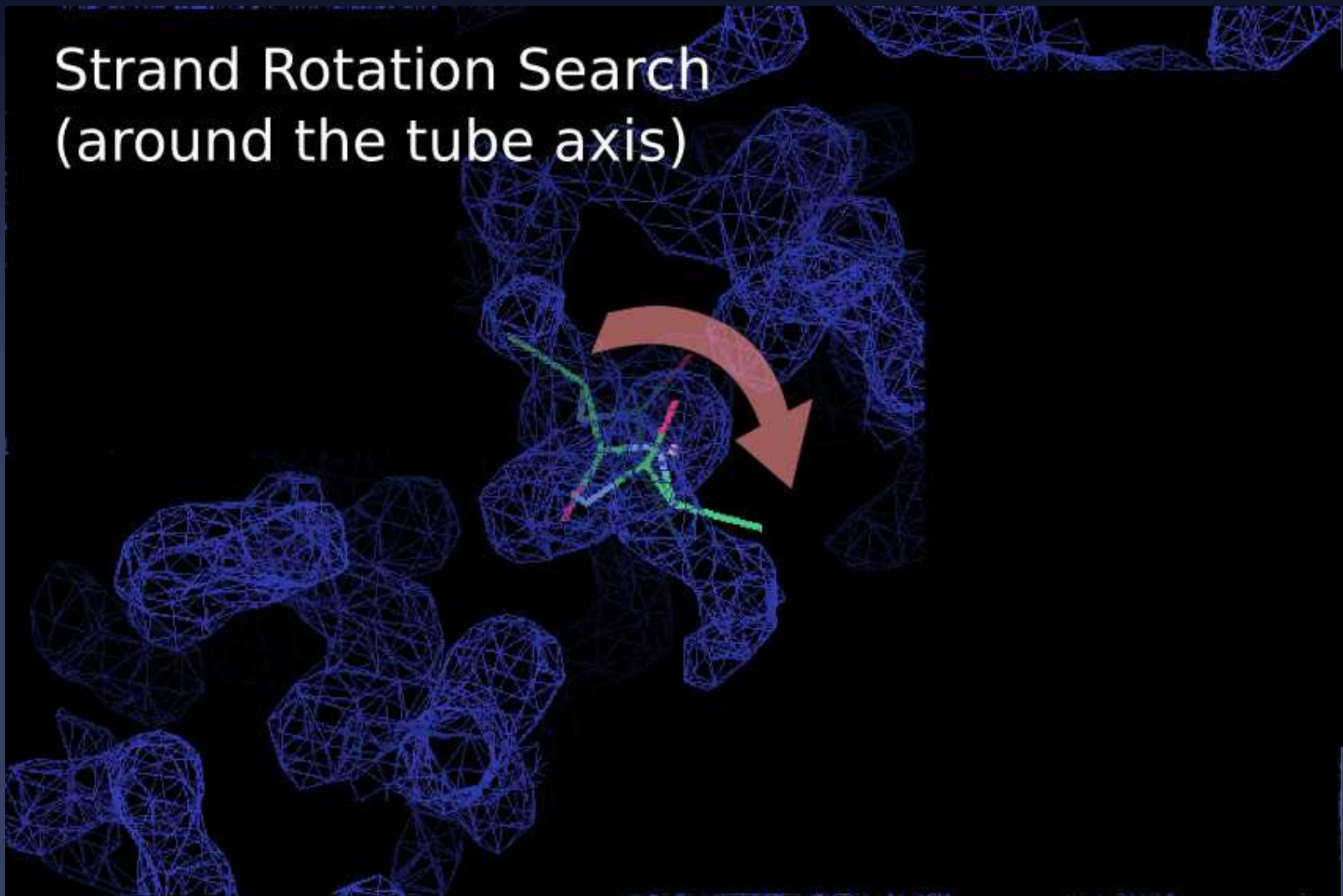
Strand fitting algorithm

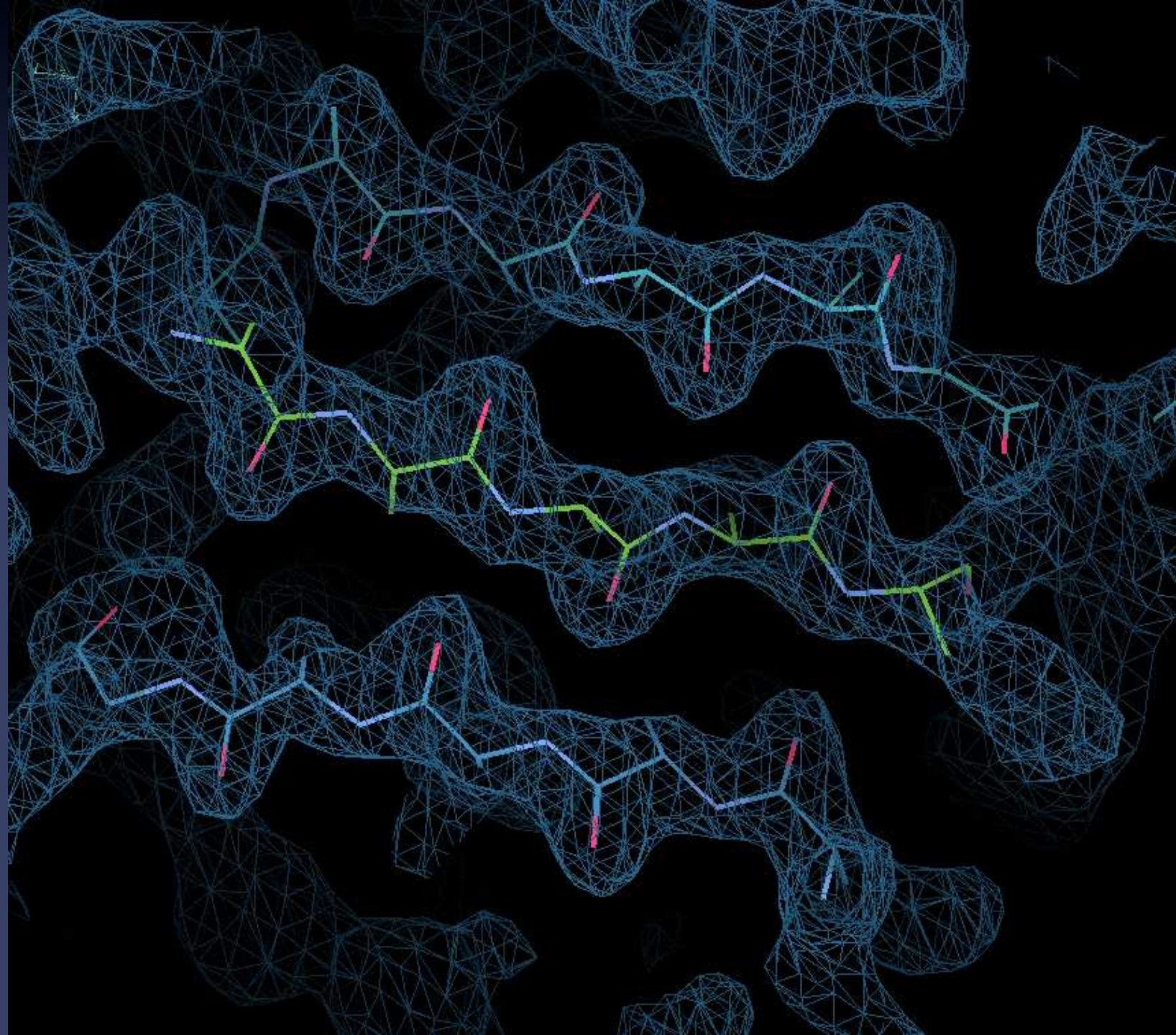
- Cylinder search
- Get N fragments of length l from database
 - ◆ 1-D Translation search along the tube
 - 1-D Rotation search around the tube
 - Direction flip search
- Rigid body refine best solutions
- Real-space refine best solution

Strand Translation Search (along the tube axis)

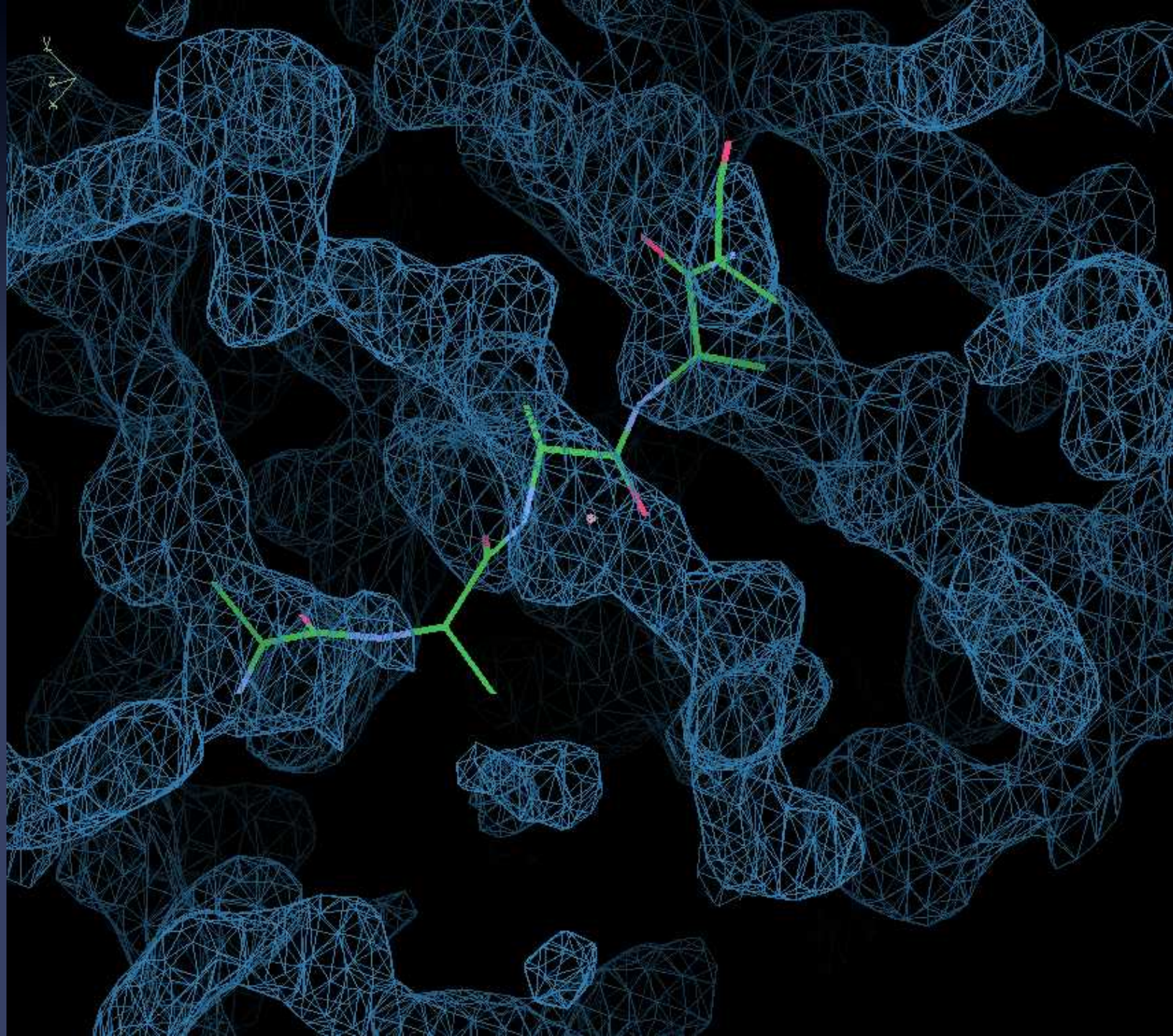


Strand Rotation Search (around the tube axis)





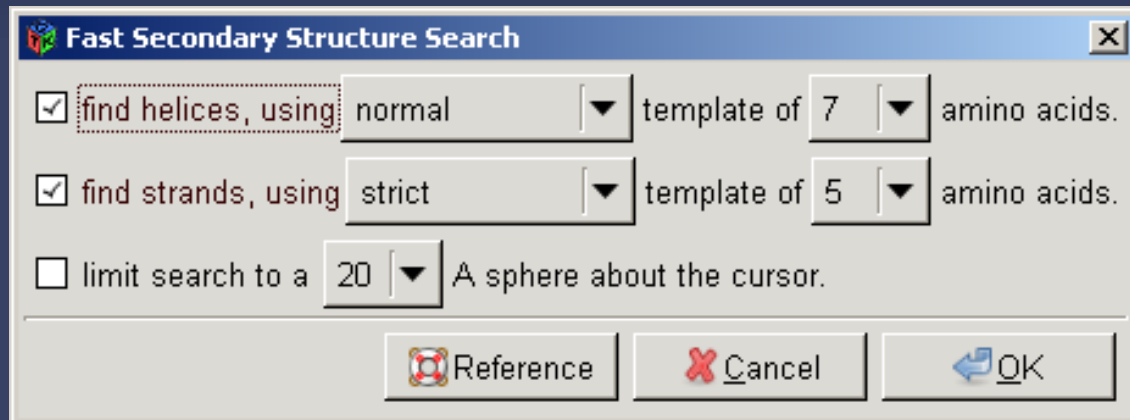
Not all is rosy...



Fitting Strands caveat

- In the case of strand-fitting, the initial translation search centring the cylinder is not performed (the search cylinder is too thin)
- The user is responsible for centring the search point “in the middle of the tube”
- Not at a C-alpha position

Automated Fast Secondary Structure Search

A screenshot of a software dialog box titled "Fast Secondary Structure Search". The dialog box has a blue title bar with a close button (X) in the top right corner. It contains three rows of settings, each with a checkbox, a label, a dropdown menu, and a text field. The first row is "find helices, using" with a checked checkbox, a dropdown menu set to "normal", a "template of" label, a text field with "7", and the text "amino acids.". The second row is "find strands, using" with a checked checkbox, a dropdown menu set to "strict", a "template of" label, a text field with "5", and the text "amino acids.". The third row is "limit search to a" with an unchecked checkbox, a text field with "20", and the text "A sphere about the cursor.". At the bottom of the dialog box are three buttons: "Reference" with a red cube icon, "Cancel" with a red X icon, and "OK" with a blue arrow icon.

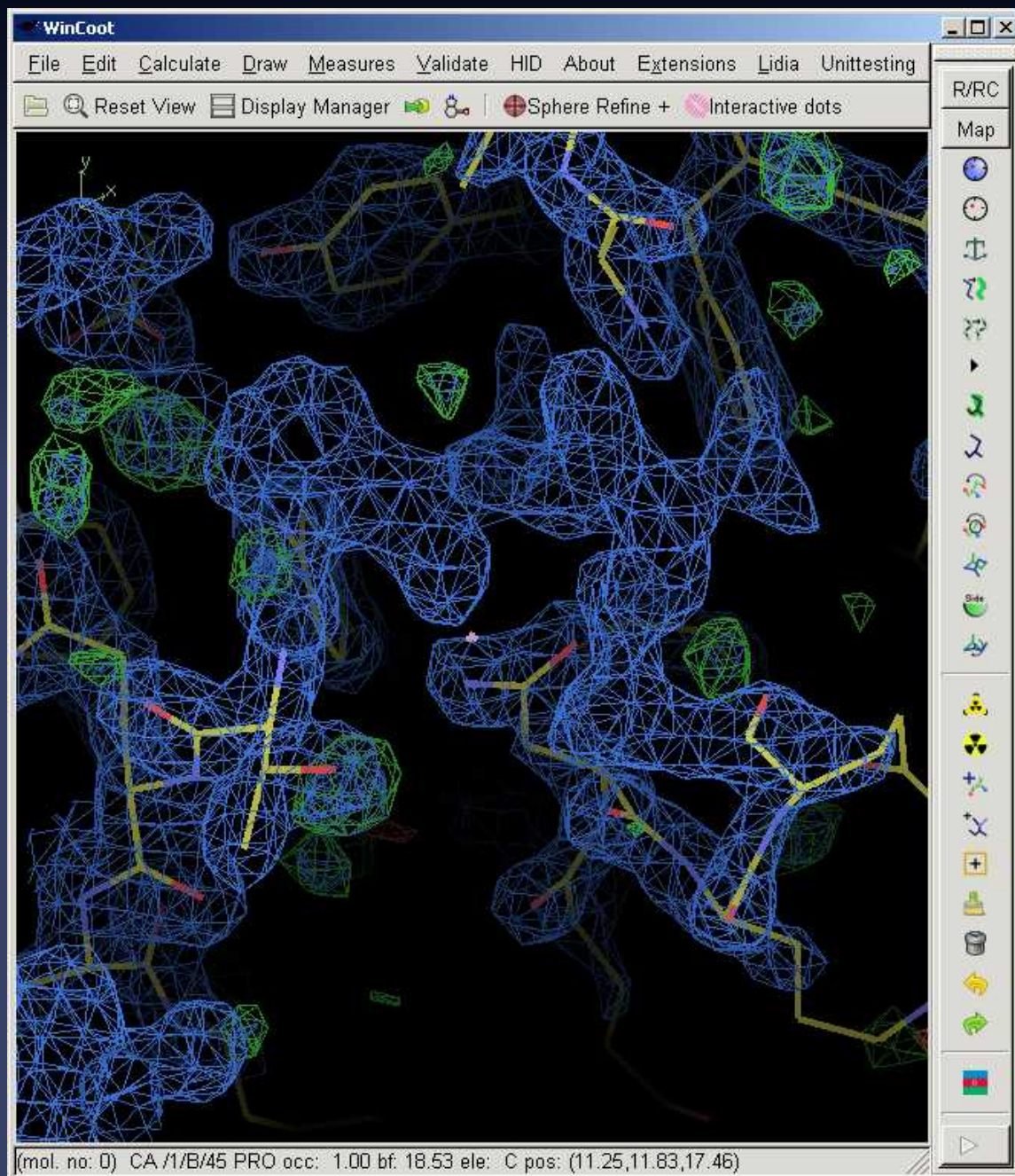
Fast Secondary Structure Search

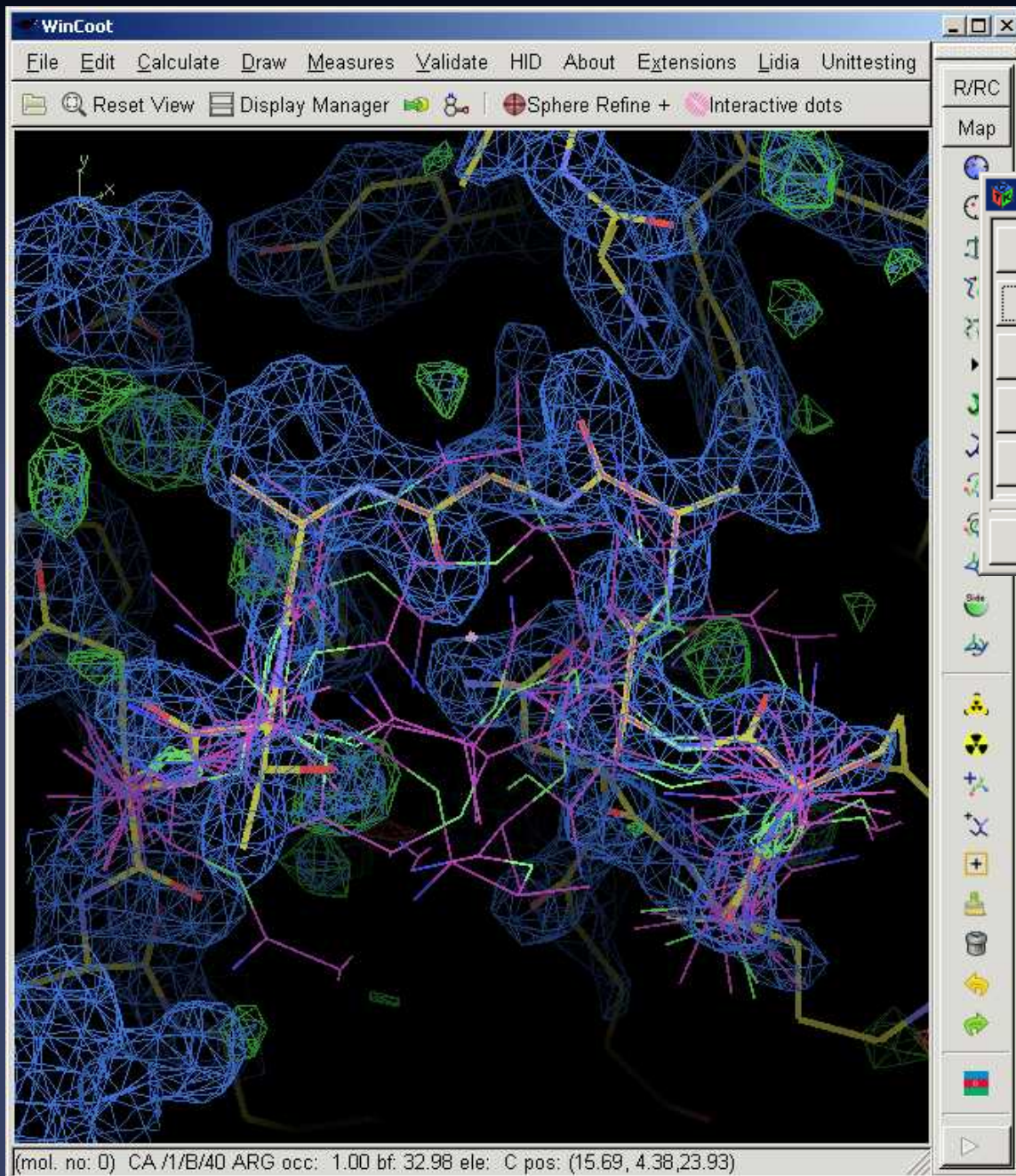
☒ find helices, using normal template of 7 amino acids.

☒ find strands, using strict template of 5 amino acids.

☐ limit search to a 20 A sphere about the cursor.

 Reference  Cancel  OK





Fitting Ligands

Ligands in Coot

- Fitting ligand
- Ligand validation
- Ligand representation
- Ligand builder

Ligand Type

Ligand Site

Known

Unknown

Known



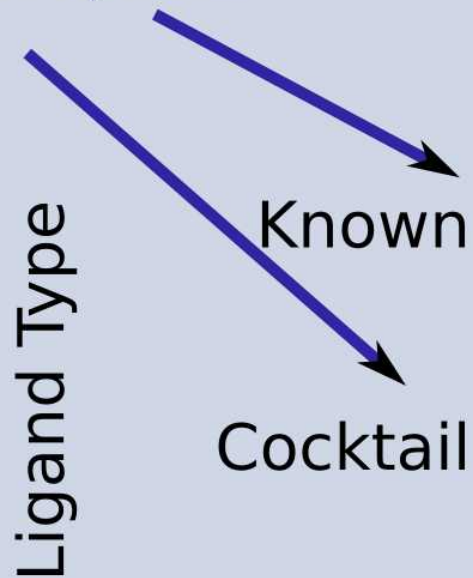
Cocktail



Unknown



Optimize with "ligand expert" options



Unknown

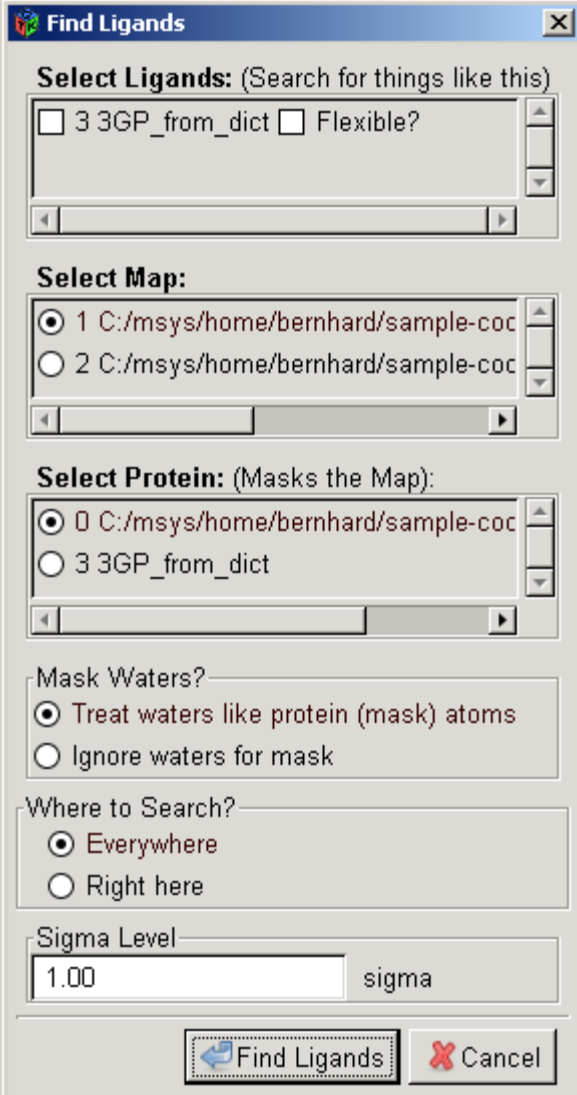
Ligand Site

Known

Unknown

Ligand search in Coot



Find Ligands

Select Ligands: (Search for things like this)

☐ 3 3GP_from_dict ☐ Flexible?

Select Map:

☒ 1 C:/msys/home/bernhard/sample-coc
☐ 2 C:/msys/home/bernhard/sample-coc

Select Protein: (Masks the Map):

☒ 0 C:/msys/home/bernhard/sample-coc
☐ 3 3GP_from_dict

Mask Waters?

☒ Treat waters like protein (mask) atoms
☐ Ignore waters for mask

Where to Search?

☒ Everywhere
☐ Right here

Sigma Level

1.00 sigma

Ligand Fitting

- *c.f.* Oldfield (2001) *Acta Cryst. D* X-LIGAND
- Somewhat different torsion search algorithm
- Build in crystal-space

REFMAC Monomer Library

chem_comp_bond

```
loop_  
_chem_comp_bond.comp_id  
_chem_comp_bond.atom_id_1  
_chem_comp_bond.atom_id_2  
_chem_comp_bond.type  
_chem_comp_bond.value_dist  
_chem_comp_bond.value_dist_esd
```

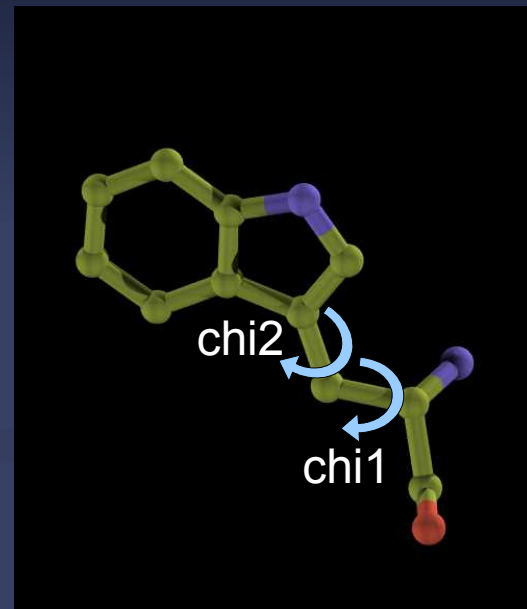
ALA	N	H	single	0.860	0.020
ALA	N	CA	single	1.458	0.019
ALA	CA	HA	single	0.980	0.020
ALA	CA	CB	single	1.521	0.033
ALA	CB	HB1	single	0.960	0.020
ALA	CB	HB2	single	0.960	0.020
ALA	CB	HB3	single	0.960	0.020
ALA	CA	C	single	1.525	0.021
ALA	C	O	double	1.231	0.020

REFMAC Monomer Library

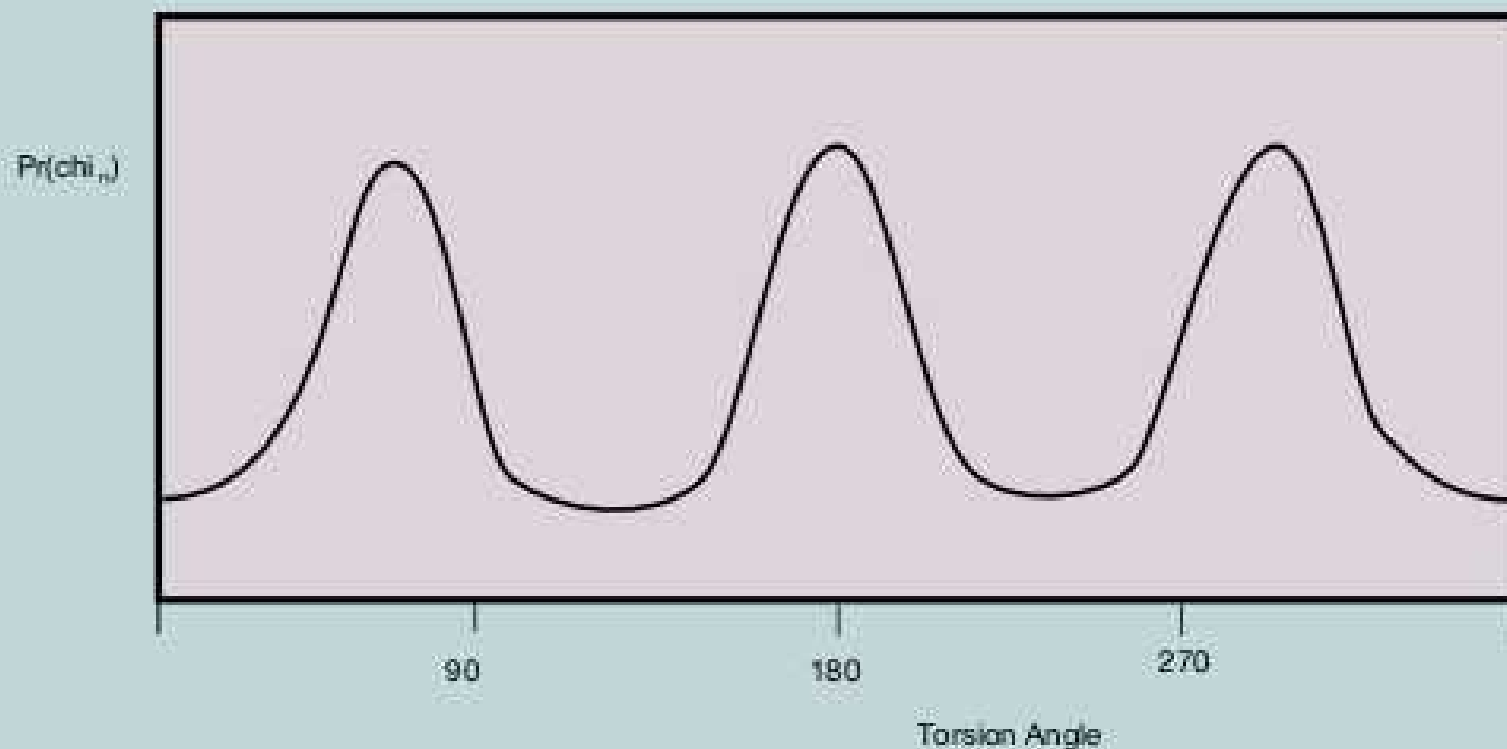
chem_comp_tor

```
loop_  
_chem_comp_tor.comp_id  
_chem_comp_tor.id  
_chem_comp_tor.atom_id_1  
_chem_comp_tor.atom_id_2  
_chem_comp_tor.atom_id_3  
_chem_comp_tor.atom_id_4  
_chem_comp_tor.value_angle  
_chem_comp_tor.value_angle_esd  
_chem_comp_tor.period
```

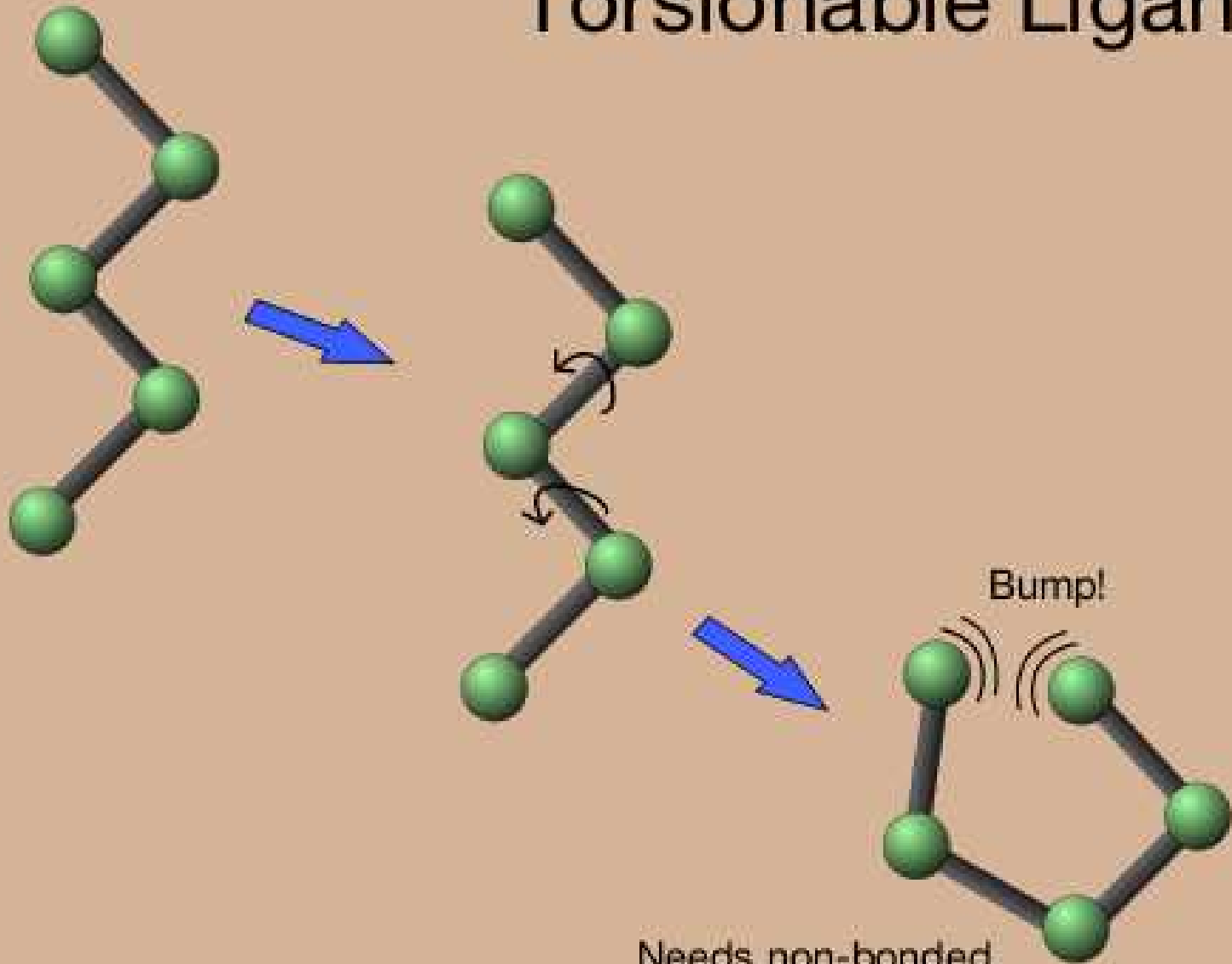
TRP	chi1	N	CA	CB	CG	180.000	15.000	3
TRP	chi2	CA	CB	CG	CD1	90.000	20.000	2



Ligand Torsionable Angle Probability from CIF file



Torsionable Ligands



Needs non-bonded
contact idealization

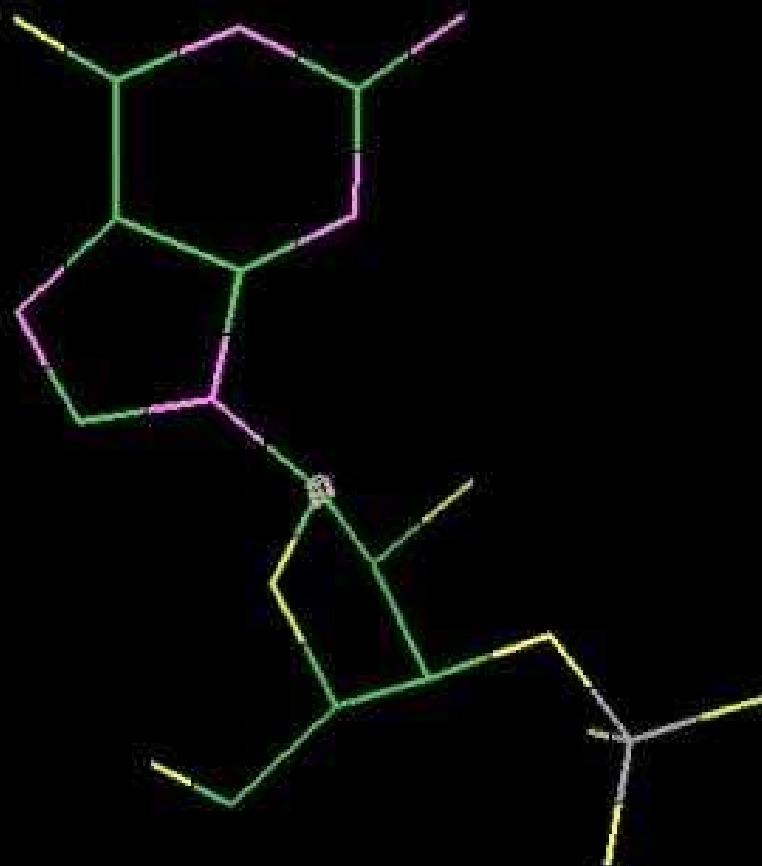
Crystal Space

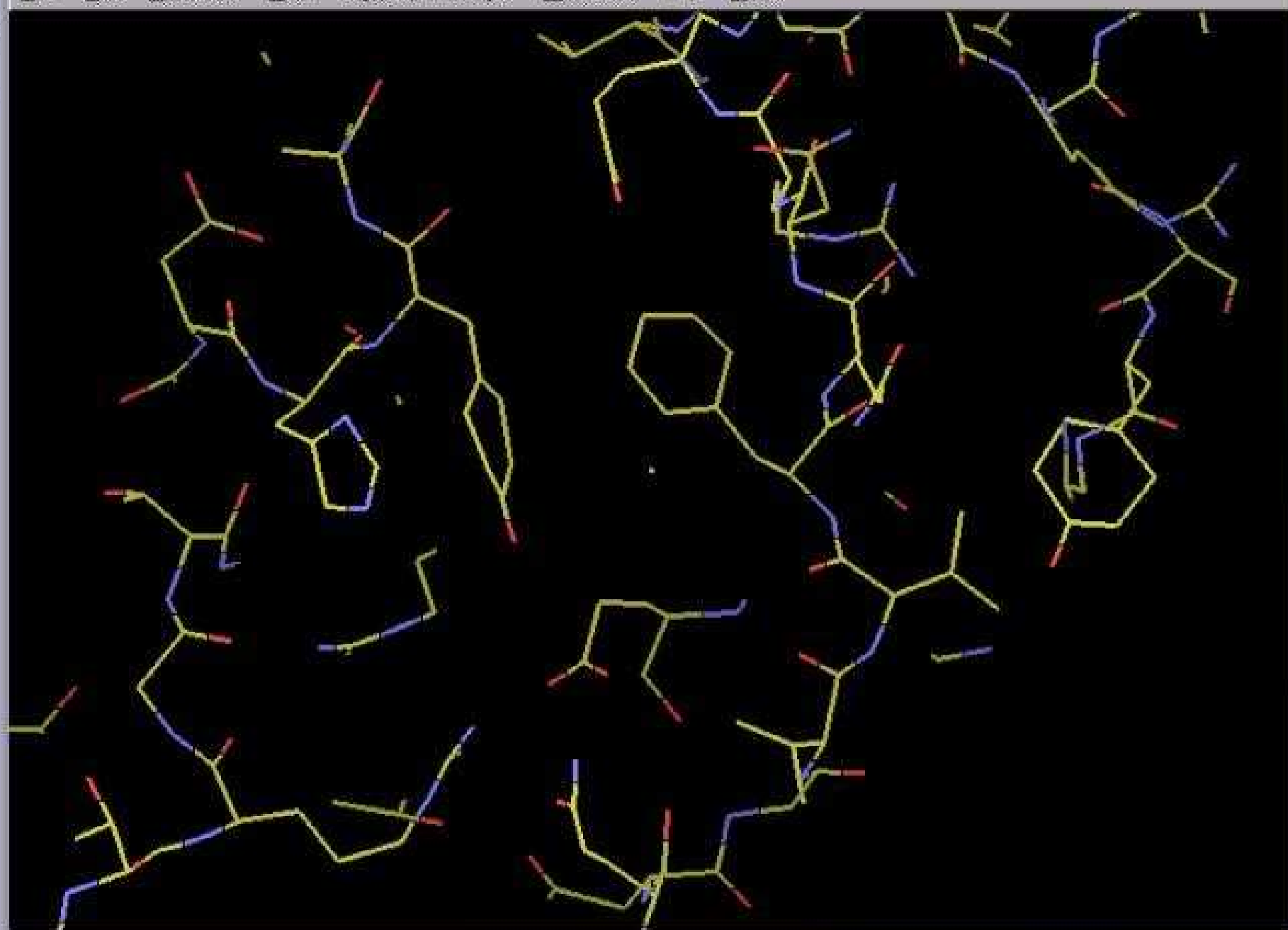
- Build in “crystal space”
 - Like real-space, but wrapped by crystal symmetry
 - Like “Asteroids”
 - Assures only one real-space representation of map features
 - Build everything only once
 - No symmetry clashing
 - However, more difficult to calculate real space geometries
 - ...such as bonds, torsions

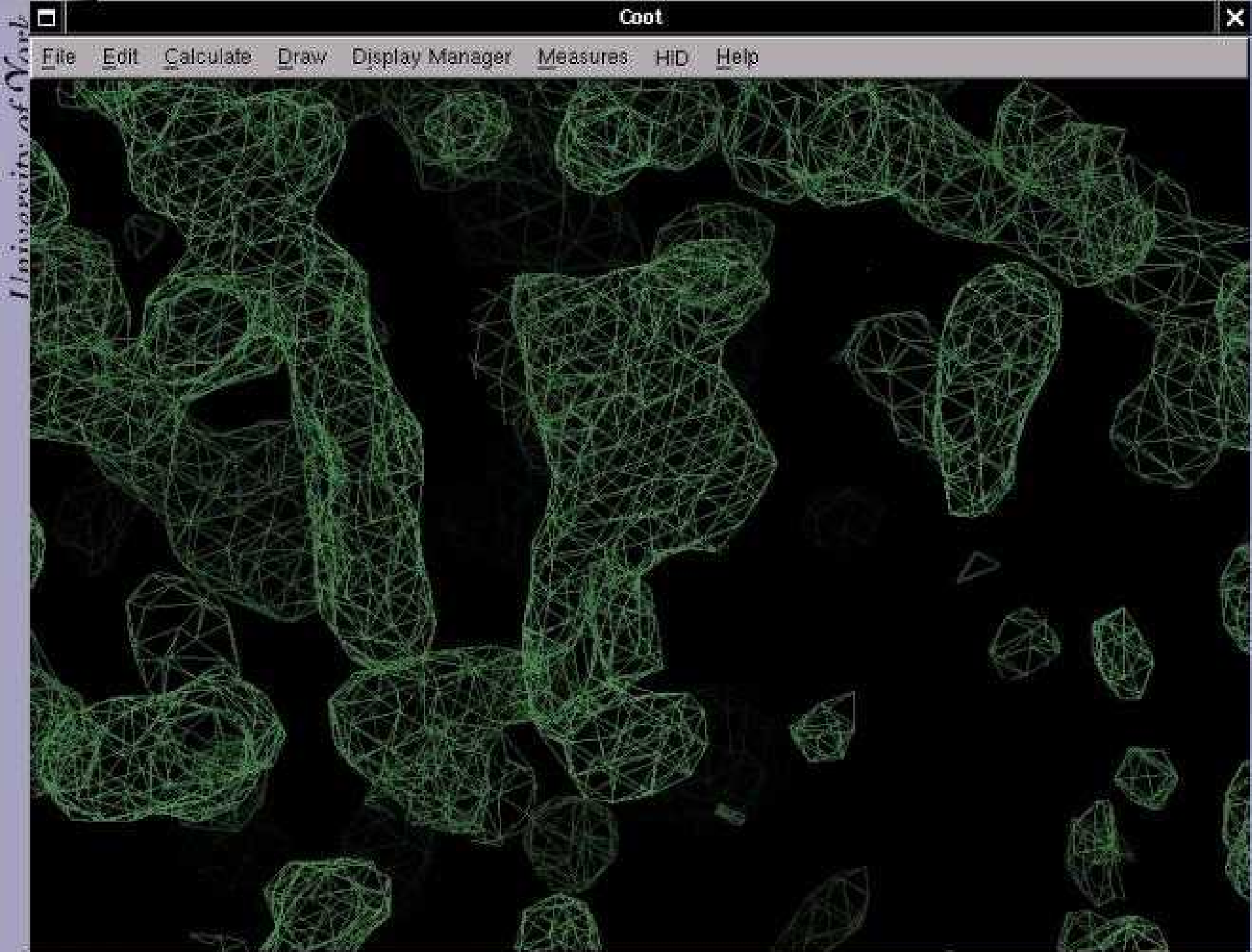


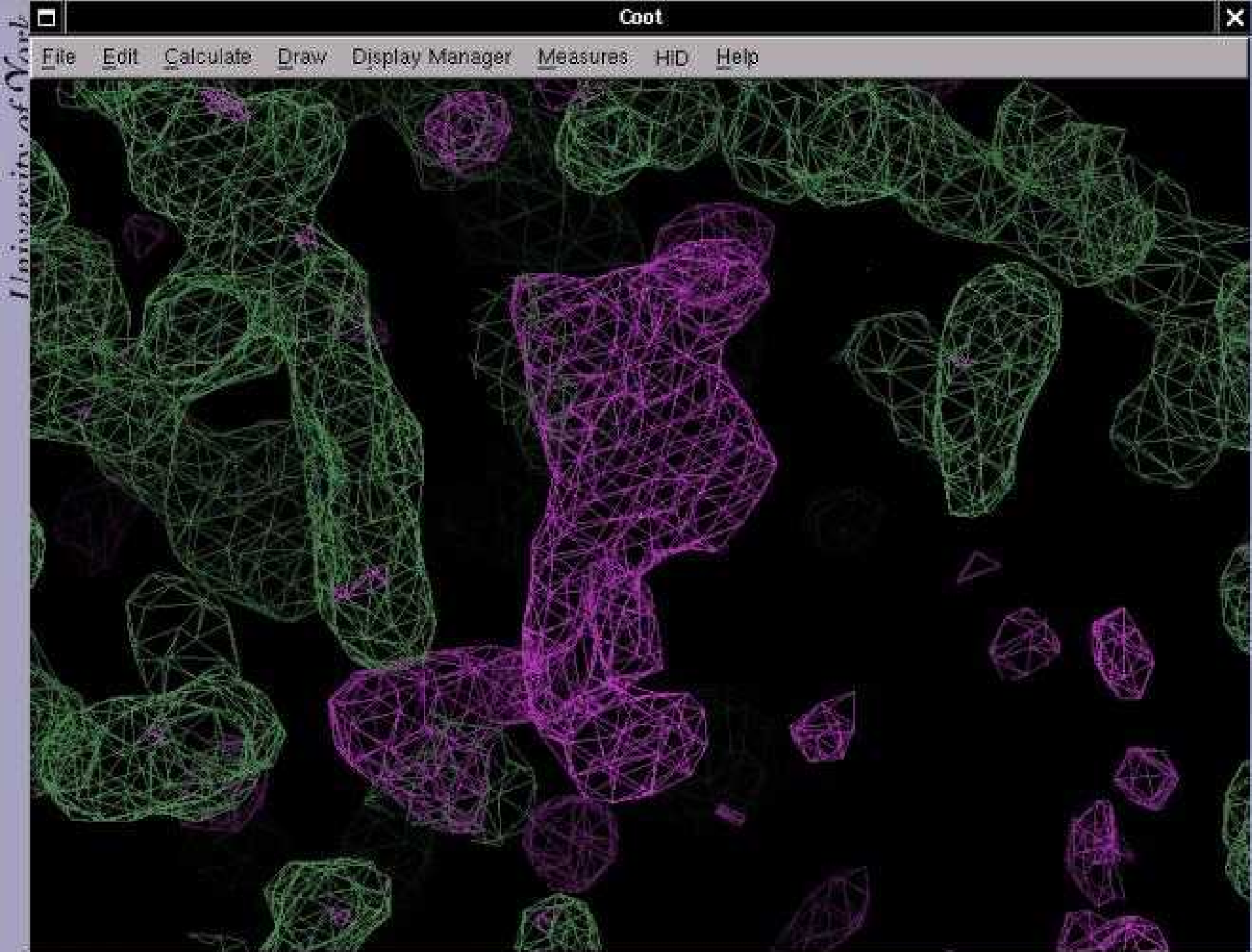
Crystal space

- Building in crystal space is good:
 - We don't need to define where the protein is and create an extended map that surrounds it
 - We don't have to worry about the relative position of the ligand and the protein
 - Unknown "BORDER" parameter
 - We find (and fit) each site exactly once
 - No symmetry problems



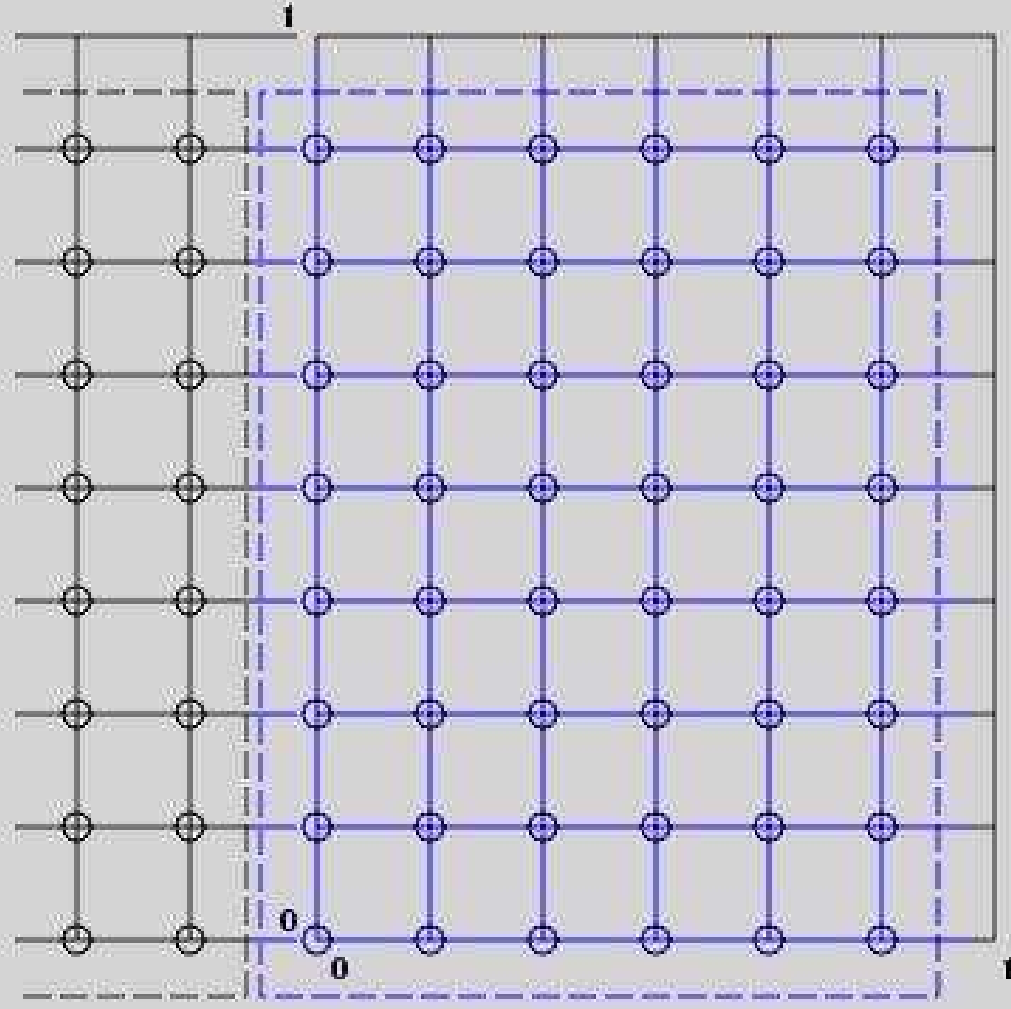


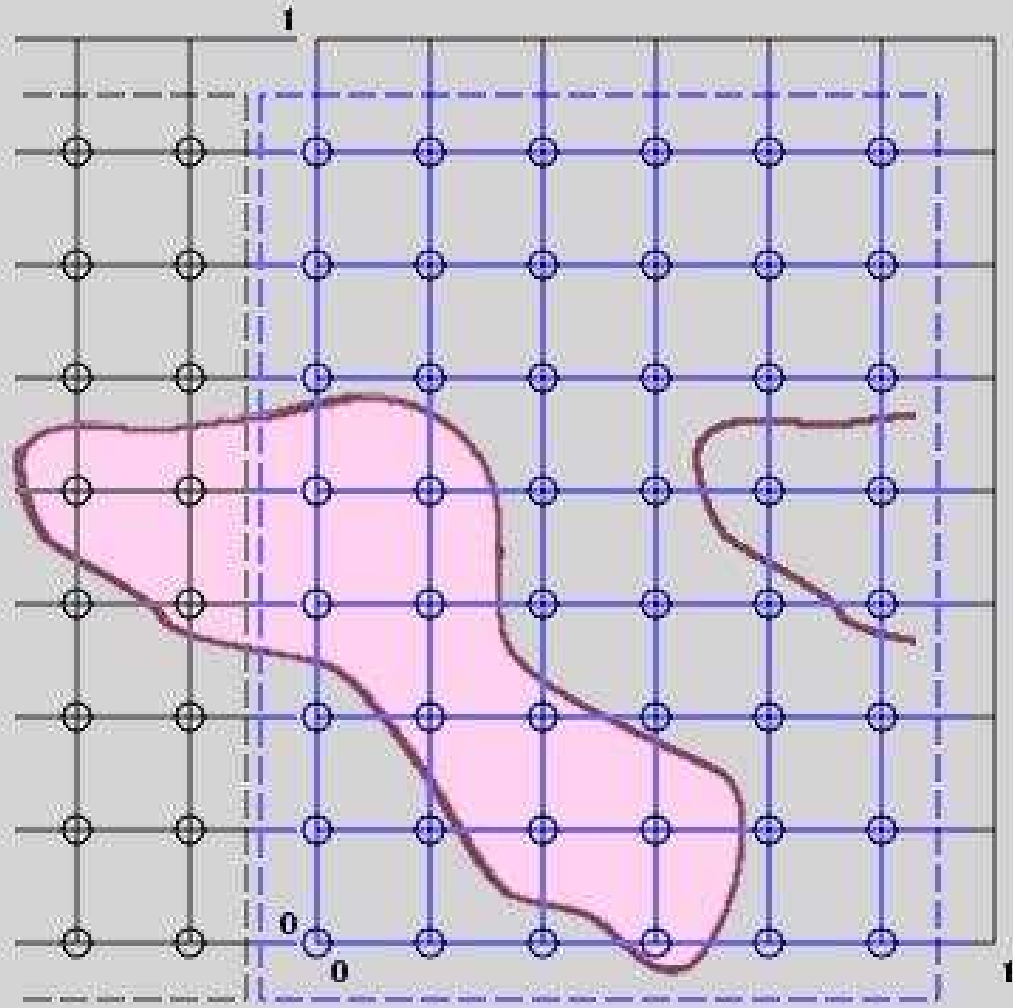


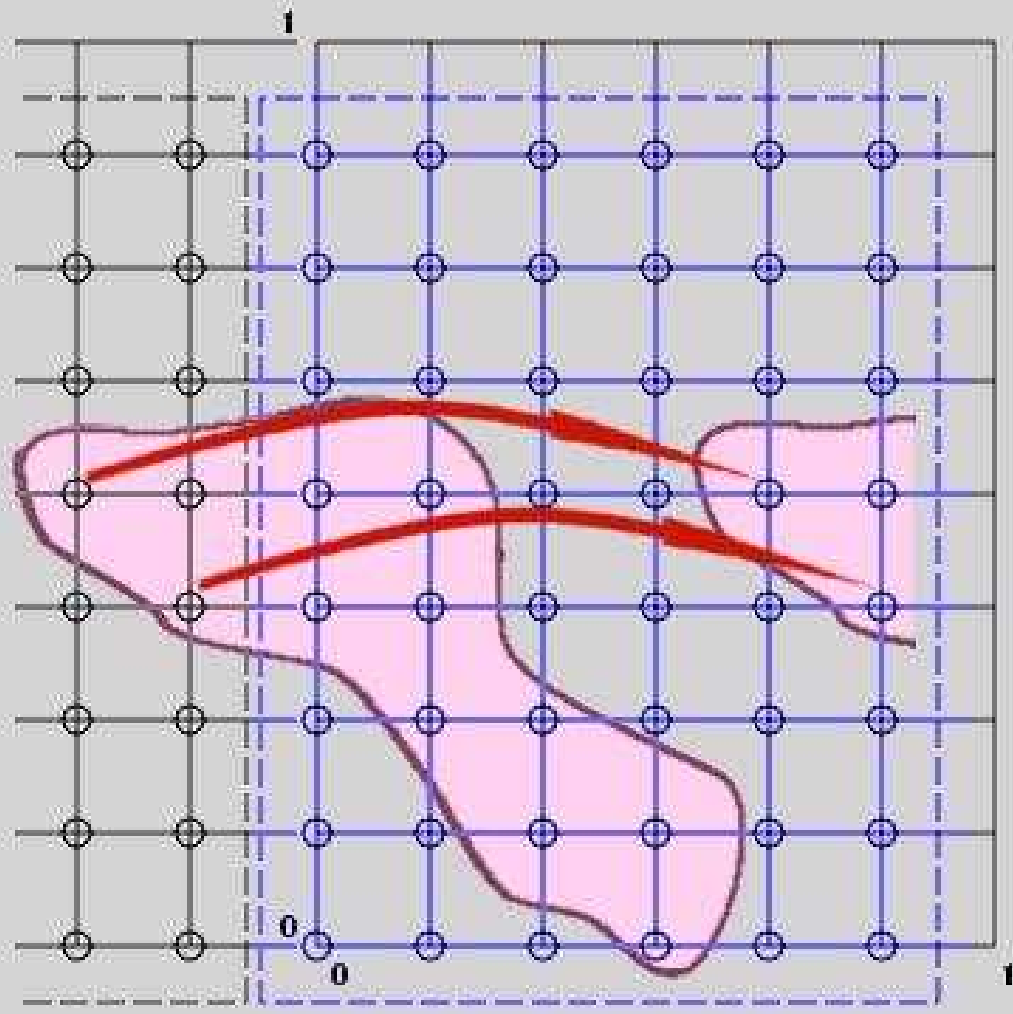


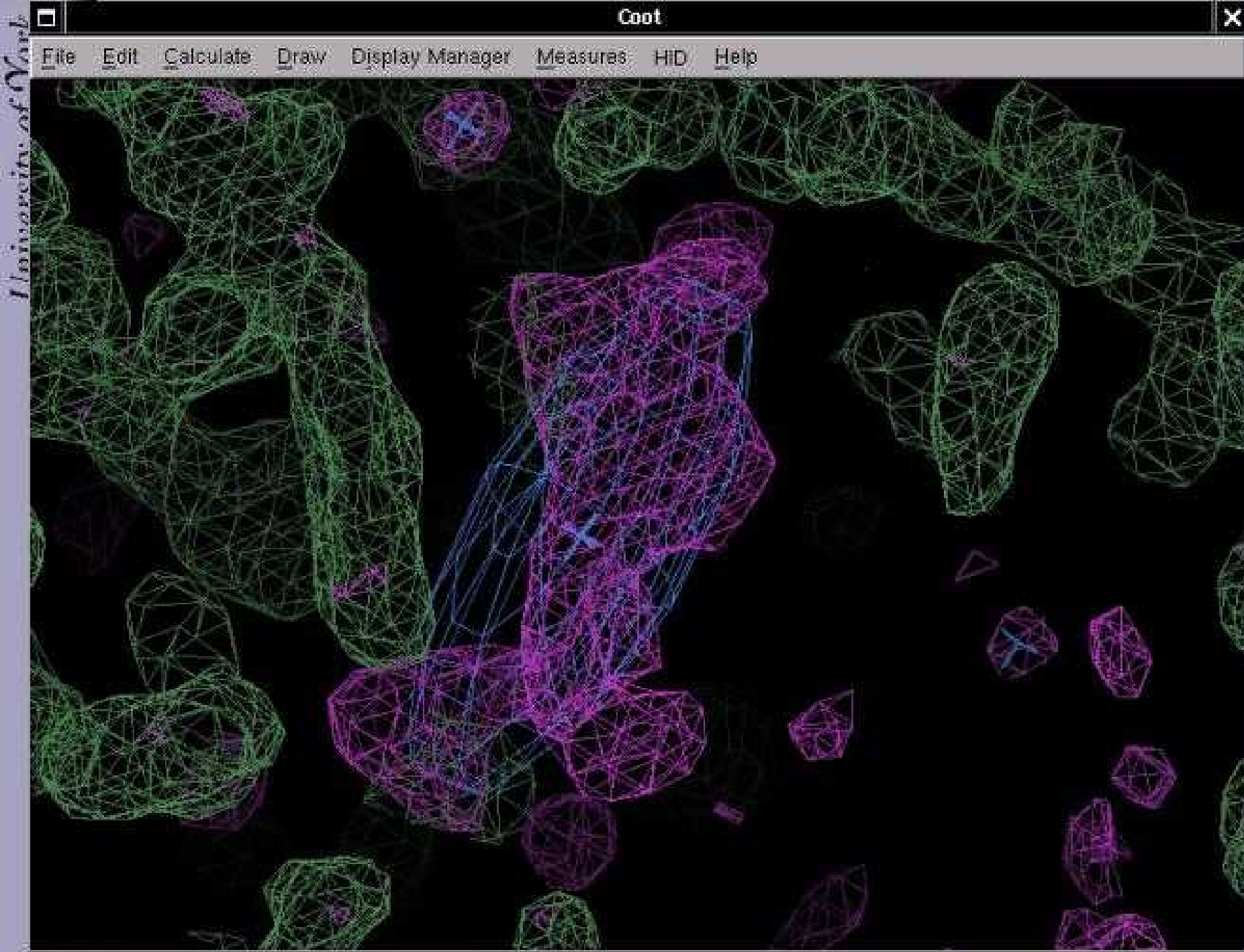
Clipper Map Mapping

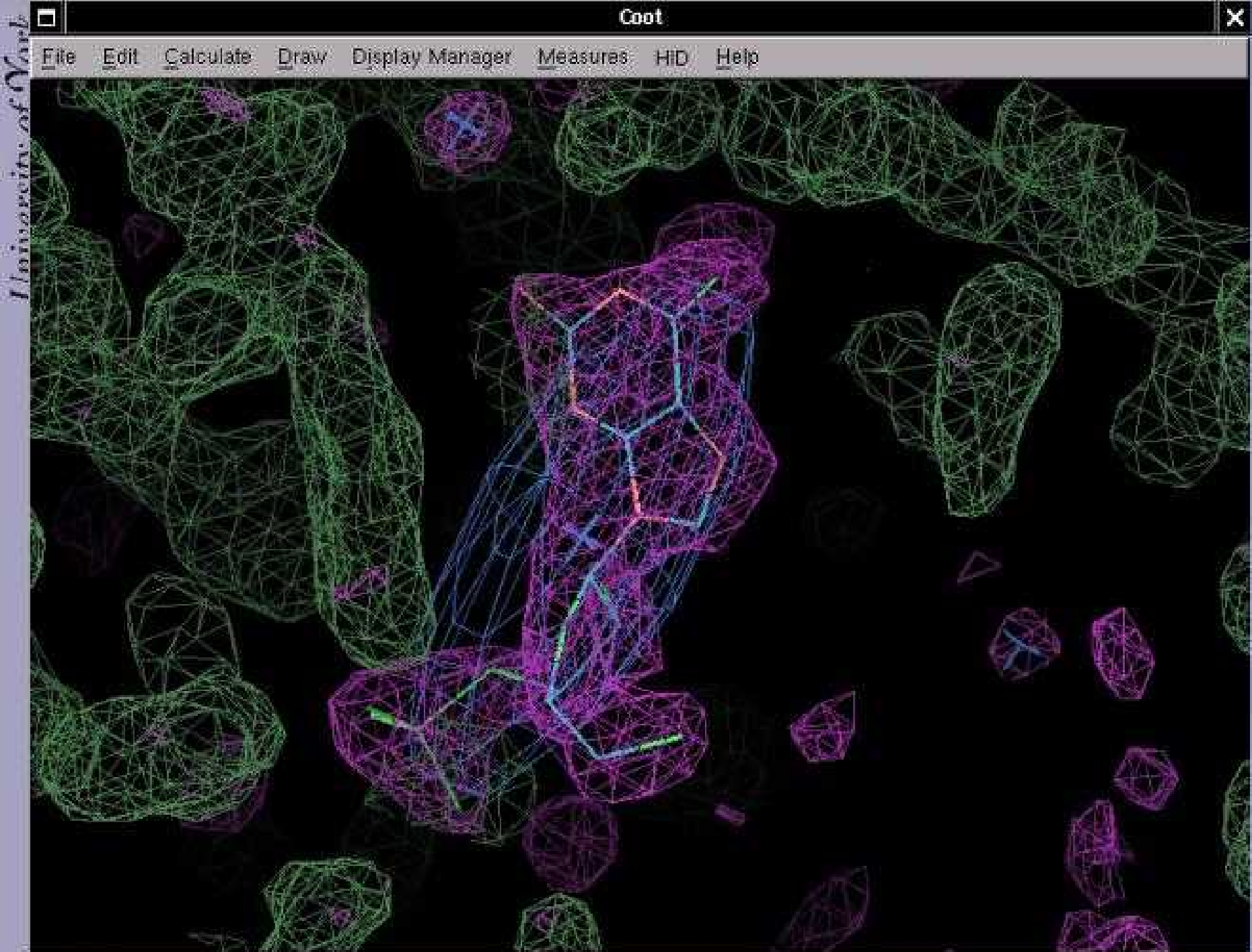
- Clipper maps
 - Appear to be “infinite”
 - Density value can be queried anywhere in space

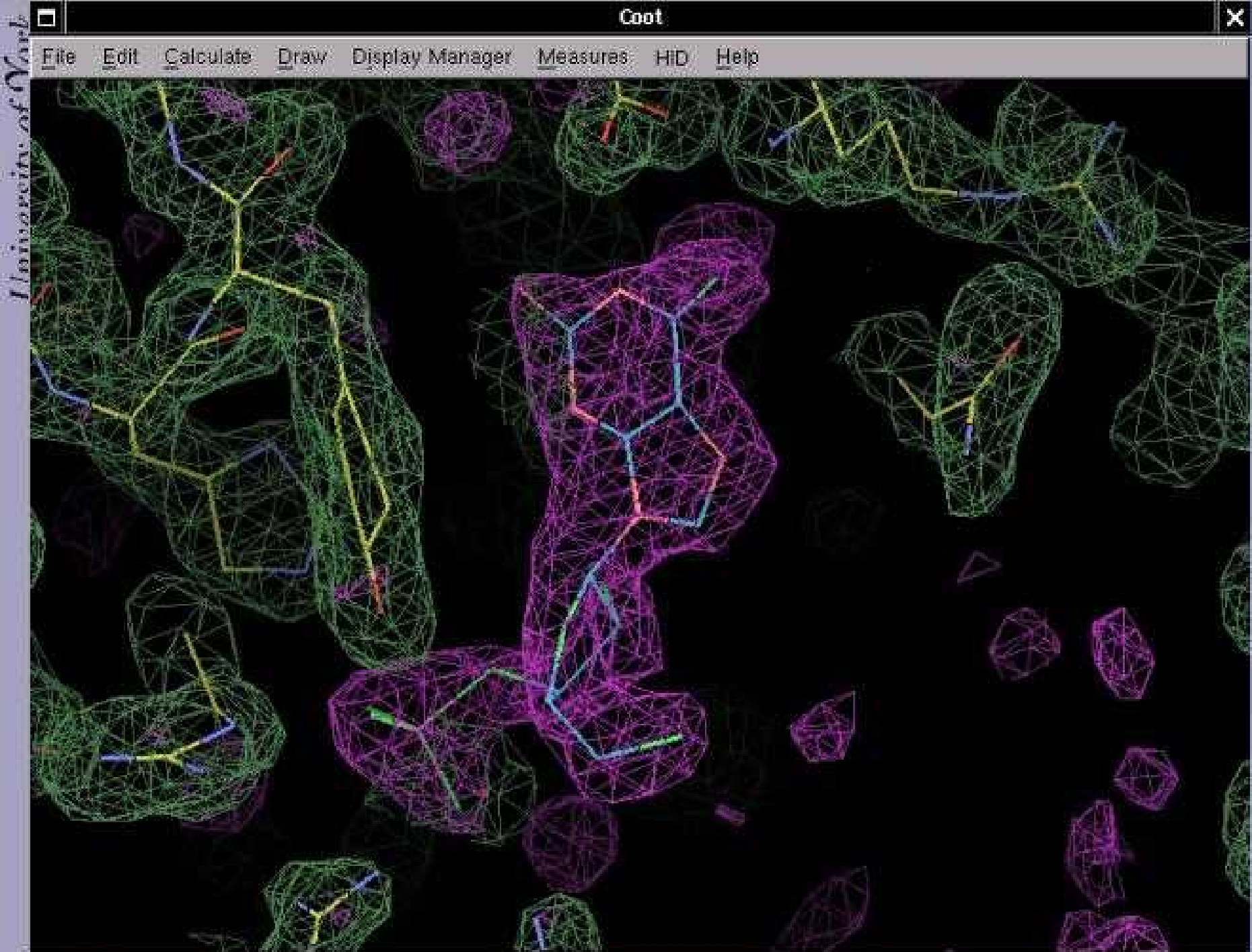












Conformation Idealization

- Each conformer is passed through the “Regularization” function of Coot
 - Non-bonded terms included
- Better to have hydrogen atoms on the model
- Slows things down a good deal...
 - May not be the best method to explore conformational variability for many rotatable bonds

Scripting

- Python or scheme
- 100s of functions are scriptable
- Accessed via:
 - the command line: **--script**
 - the GUI: Calculate -> Run Script...
 - Interactive: Calculate -> Scripting
- Use **--no-graphics** for “batch mode”

Delete residues by Scripting

- (delete-residue-range *imol chain_id resno_start end_resno*)
- e.g. in scheme
 - (delete-residue-range 0 "A" 10 20)
- e.g. in Python
 - delete_residue_range(0, "A", 10, 20)
- General command:
 - Scheme: (scheme-command arg1 arg2 ...)
 - Python: python_command(arg1, arg2, ...)

More on Scripting

- If something is boring, stop it
 - Write a script
 - Or get someone to do it for you
 - Paul, me?
- Scripting available in Python or Scheme (lisp)
- Scripting example available on the mailing list
 - and the Coot Wiki

Some key bindings

- Any function can be bound to a key
 - Allows for personalization/customization
- Here's how you do it:
 - `(add-key-binding "Refine residue" "x" (lambda () (refine-active-residue)))`
 - `add_key_binding("Refine residue", "x", lambda: refine_active_residue())`
- Makes Coot easy to use
 - (but harder to learn)
- See crib sheet...

Acknowledgements

- Paul Emsley
- Kevin Cowtan
- Eleanor Dodson
- Keith Wilson

<http://www.biop.ox.ac.uk/coot/>

or

Google: Coot

or for WinCoot

<http://www.ysbl.ac.uk/~lohkamp/coot>

- Libraries, dictionaries
 - Alexei Vagin, Eugene Krissinel, Stuart McNicholas
 - Dunbrack, Richardsons
- Coot Builders and Testers
 - William Scott, Ezra Peisach
 - York YSBL, Dundee, Glasgow (early adopters)
 - Coot Mailing List subscribers