

Macromolecular Complexes in Crystals and Solutions

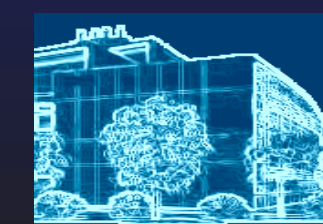
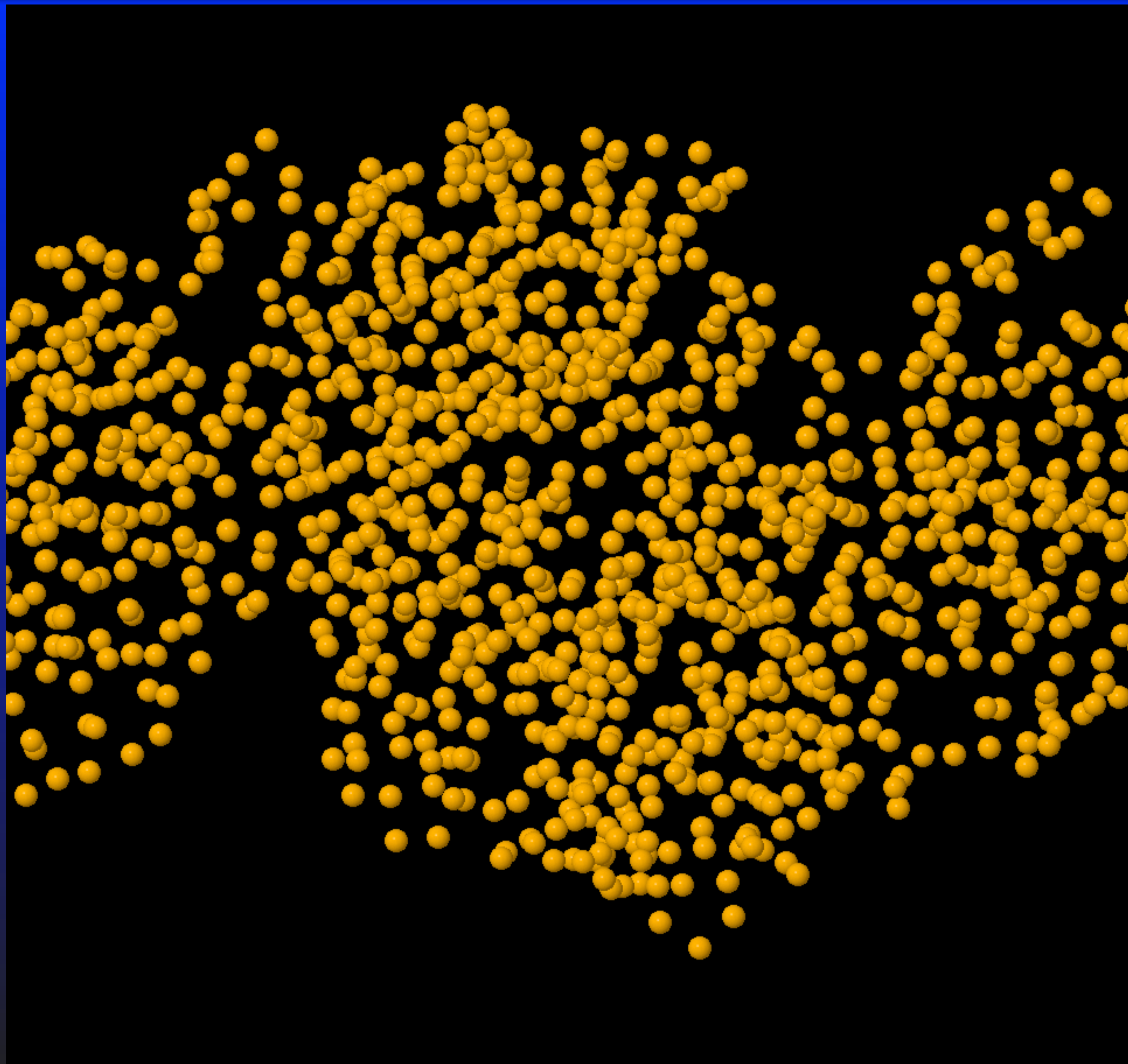
Eugene Krissinel

CCP4, STFC Research Complex at Harwell
Didcot, United Kingdom

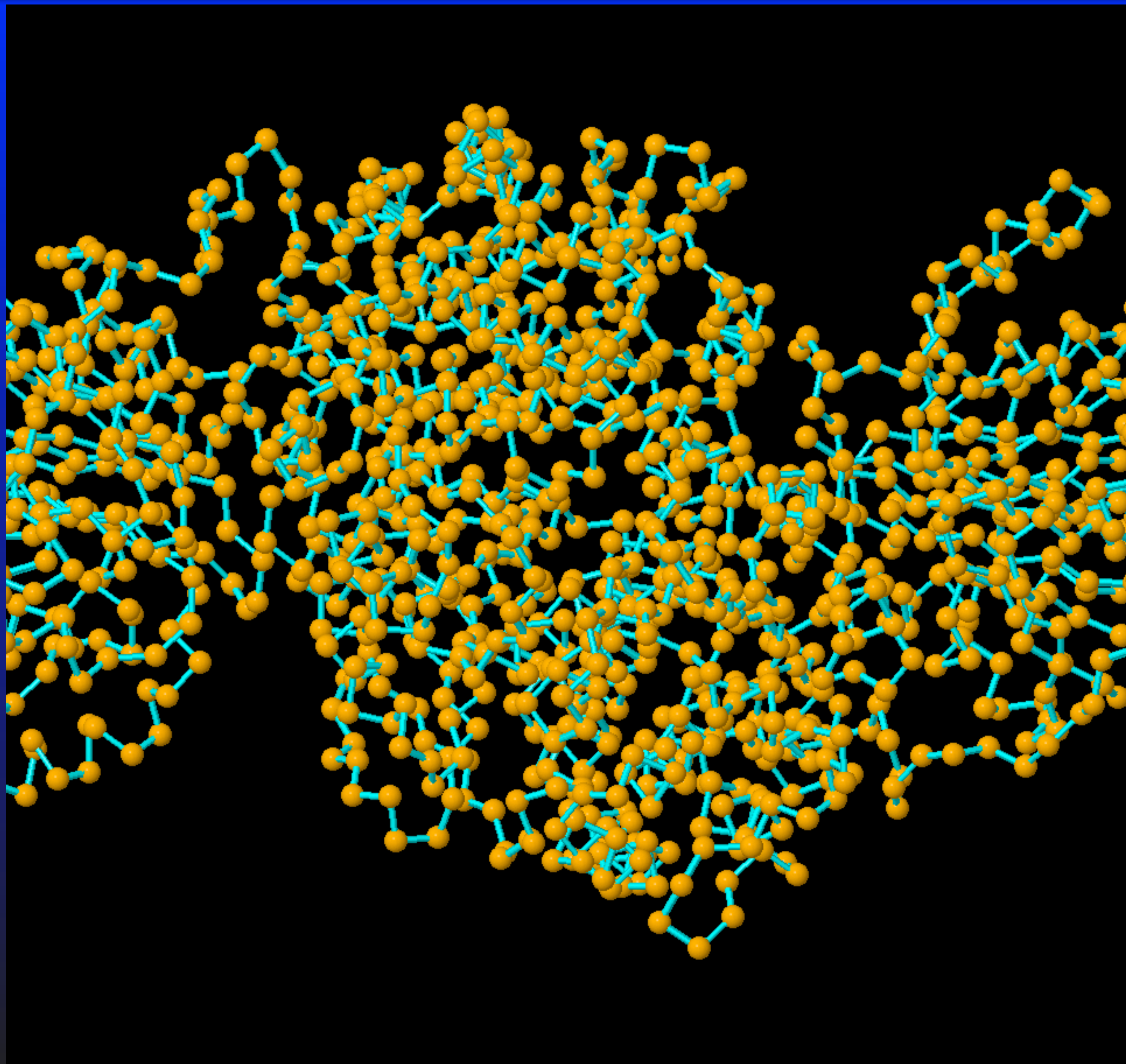
eugene.krissinel@stfc.ac.uk

E. Krissinel and K. Henrick (2007) J. Mol. Biol. 372, 774-797

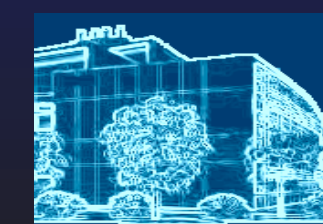
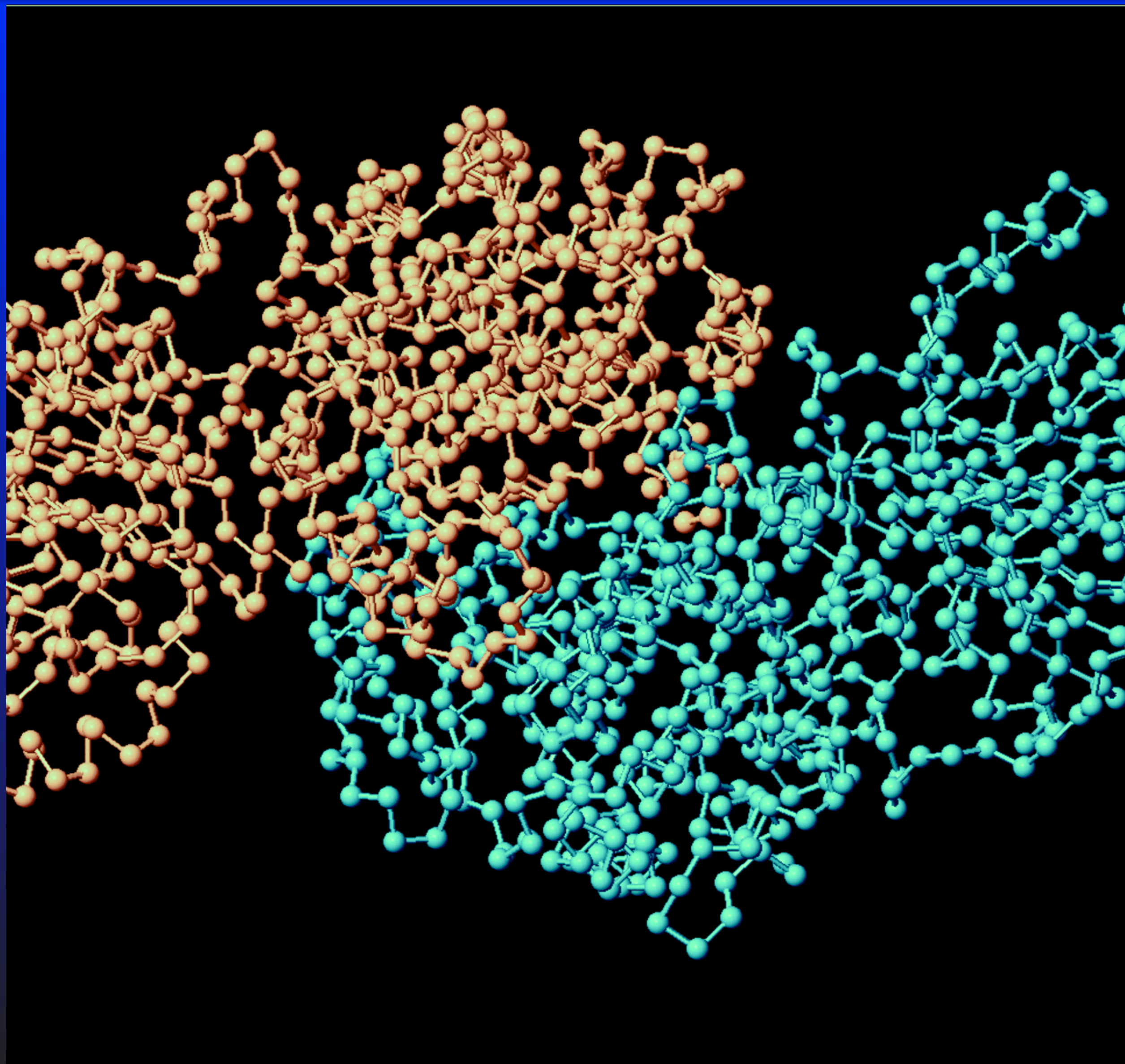
E. Krissinel (2010) J. Comp. Chem. 31, 133-143



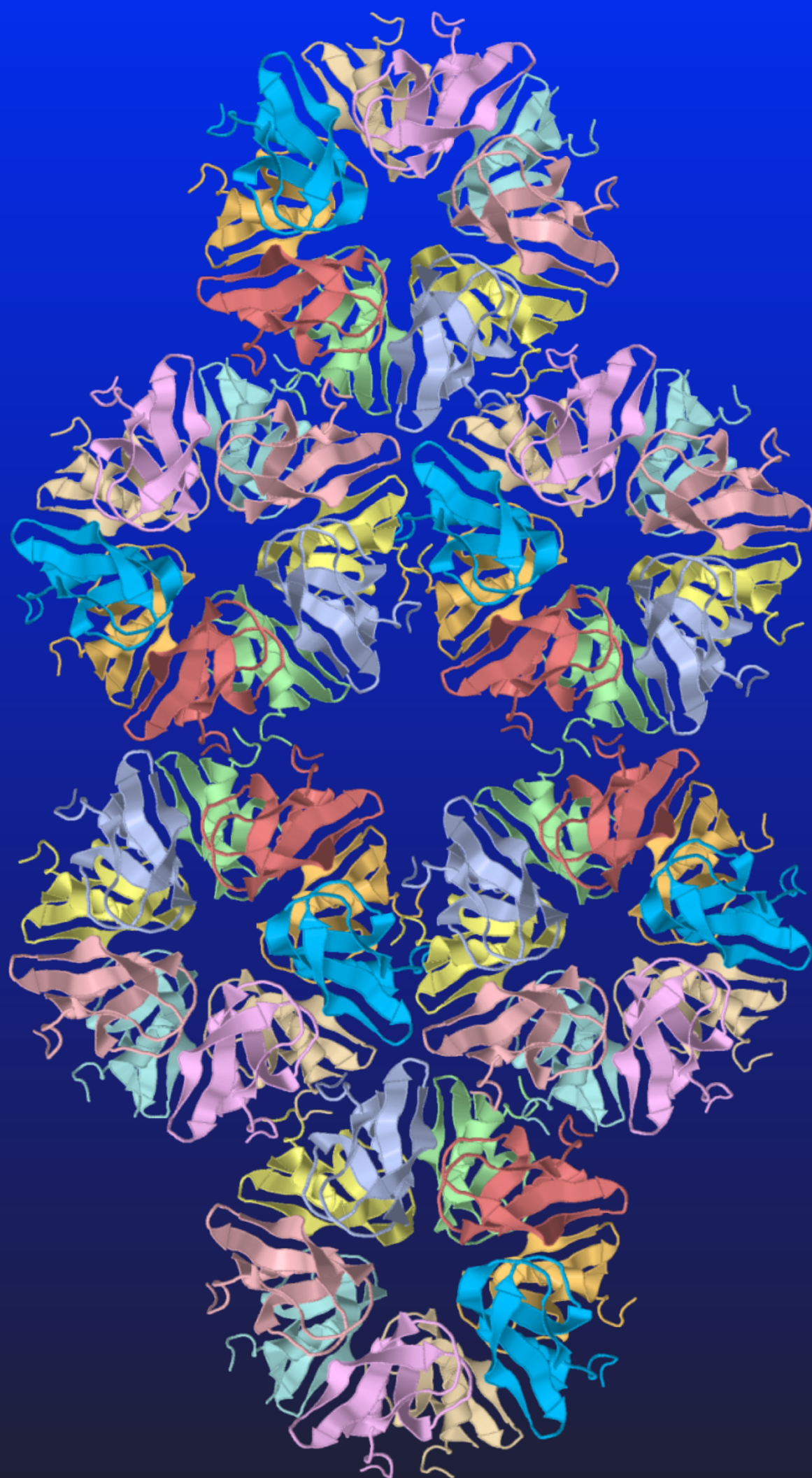
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★ Why would we want to know structure of a macromolecule?

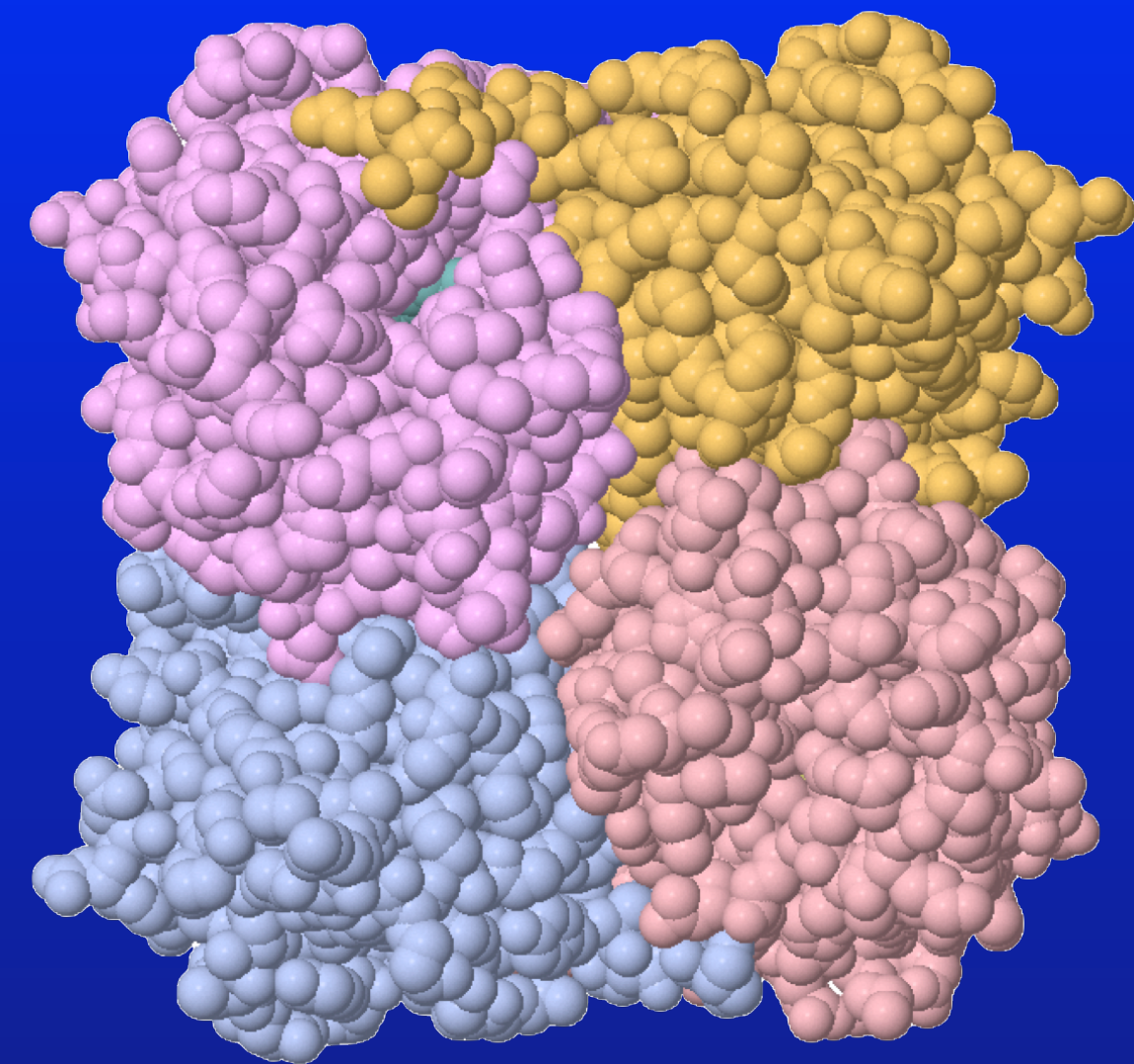
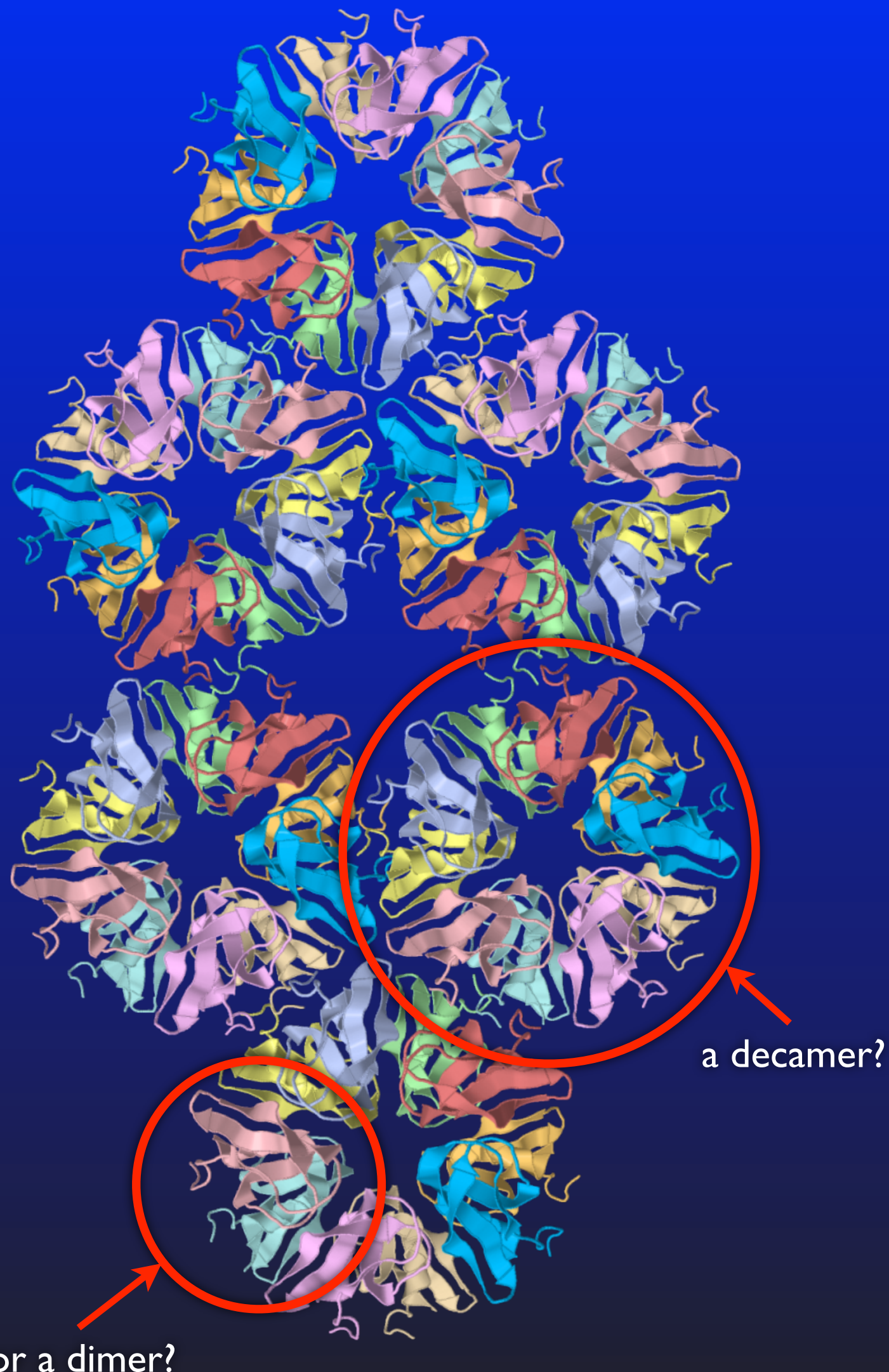
➔ for many reasons, but probably firstly for finding out how it interacts with other molecules

★ Macromolecular crystals present us with models of biological structures and interactions between them

➔ “if you want to know how A interacts with B - crystallize them together!” (crystallographer’s sweet dream)



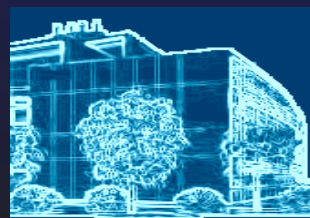
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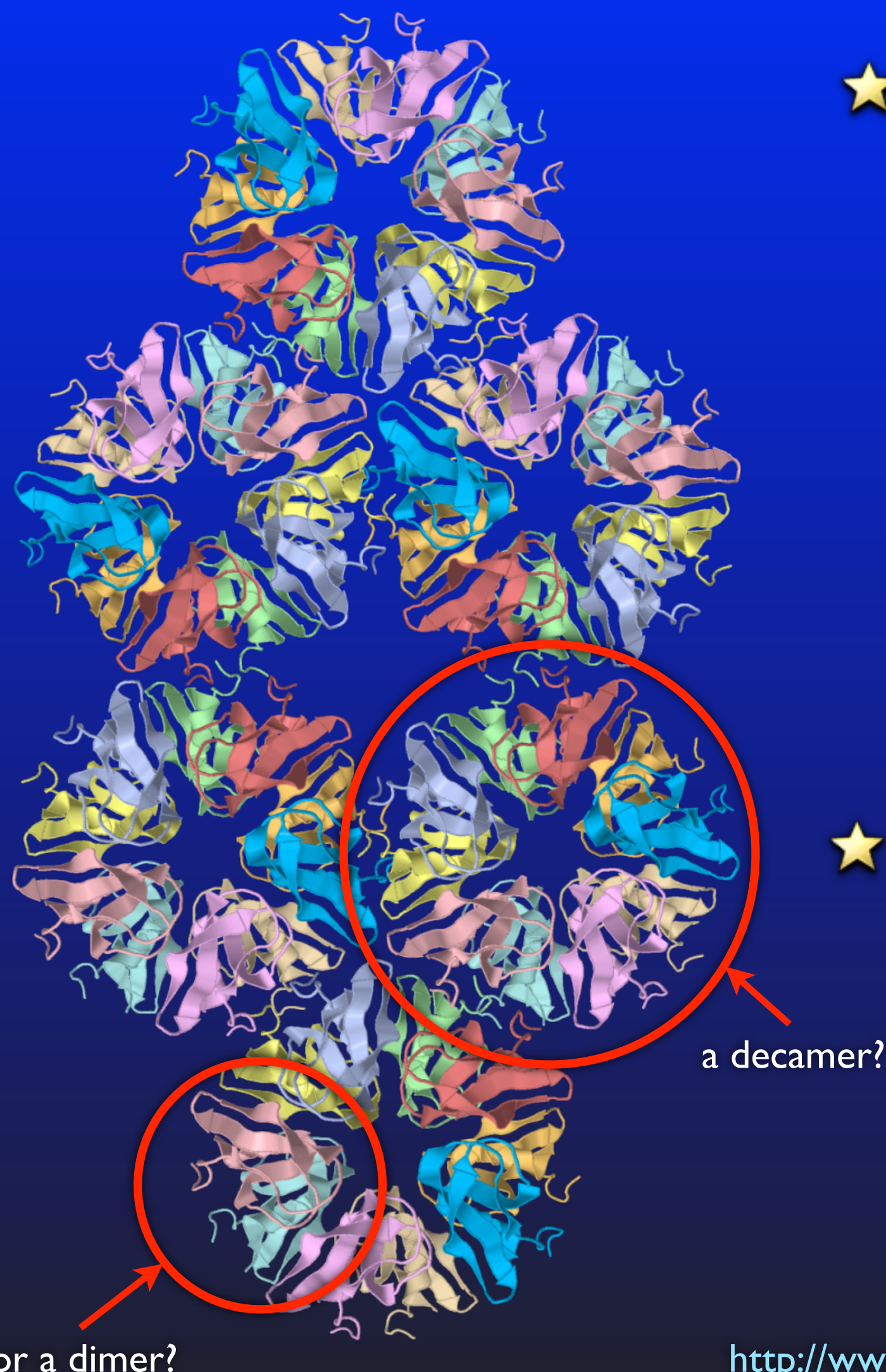


3bxc

“Monomer Association—According to gel filtration data, mKate exists in solution in the monomeric state at concentration as high as 10 mg/ml (13). However, in the crystalline state, which corresponds to a much higher protein concentration, mKate adopts at all pH values tetrameric arrangement with 222 symmetry, typically seen in GFP-like proteins.”

Pletnev et.al. (2008), J. Biol. Chem. 283, 28980-7





★ Crystals present us with both real and artifactual interactions, which may be difficult to differentiate. Frequently used techniques:

Theoretical: sharp eye and scientific authority

Rules of thumb: e.g. manifestation in different crystal forms

Experimental: complementing studies (MS, EM, NMR, scattering)

Bioinformatical: homology and interface similarity analysis

Computational: energy estimates and modelling

★ PISA software infers significant interactions and macromolecular assemblies from crystals by evaluating their free Gibbs energy:

$$\Delta G_0 = -\Delta G_{\text{int}} - T\Delta S > 0$$

http://www.ebi.ac.uk/msd-srv/prot_int/pistart.html



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$$\Delta G_0 = -\Delta G_{\text{int}} - T\Delta S > 0$$





Protein interfaces

http://www.ebi.ac.uk/msd-srv/prot_int/cgi-bin/piserver

Home RAL Mail Google Mail Yahoo! Mail Staff Central Google Maps Wikipedia Apple YouTube

EMBL-EBI

EB-eye Search

All Databases

Databases Tools EBI Groups Training

Submission Form for

Protein structure to be analysed:

- ☒ Coordinate file ☐ PDB entry

1e94.pdb uploaded

Wait for page to update after you change

6 aminoacid chains and 2 ligands

Cell parameters:

A: 172.022 Alpha: 90
B: 172.022 Beta: 90
C: 276.569 Gamma: 120

You may change the values of the cell
Your input will super

Process ligands: ☒ ANP

Processing mode: Auto

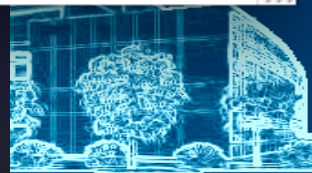
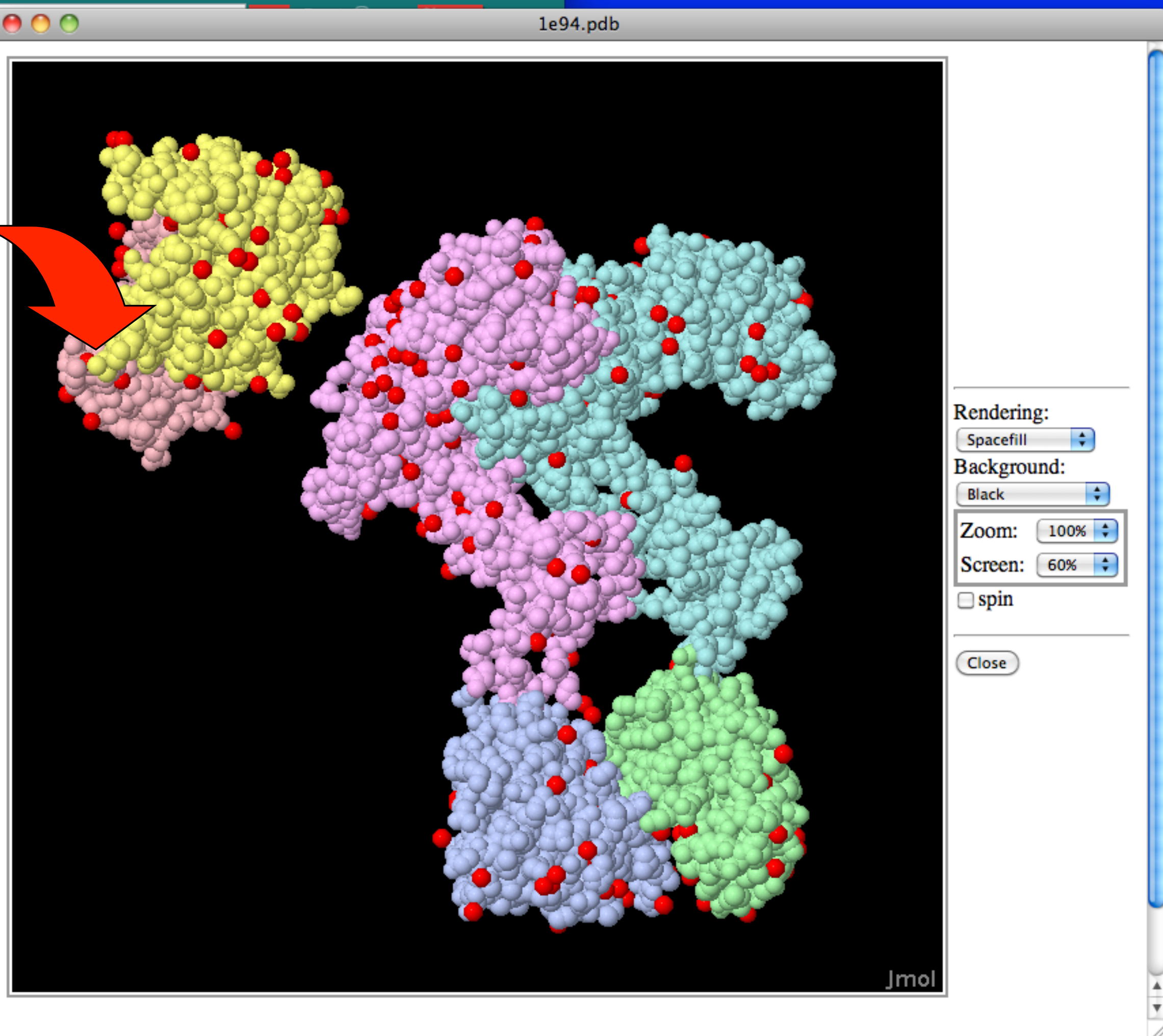
interfaces

PISA v1.18 << 30/03/2009 >>

developer: Dr. Eugene Krissinel
last modified: 30/03/2009

the Collaborative Computational Project Number 4 in Protein Crystallography
of the Biotechnology and Biological Sciences Research Council

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Protein interfaces

http://www.ebi.ac.uk/msd-srv/prot_int/cgi-bin/piserver

Home RAL Mail Google Mail Yahoo! Mail Staff Central Google Mail

EMBL-EBI EB-eye Search All Databases Enter Text Here

Databases Tools EBI Groups Training Industry About Us

Home > Databases > PDB > Services > PISA

Session 766-54-O99 map

query **1e94.pdb** → interfaces : → interface search results
 monomers :
 assemblies :

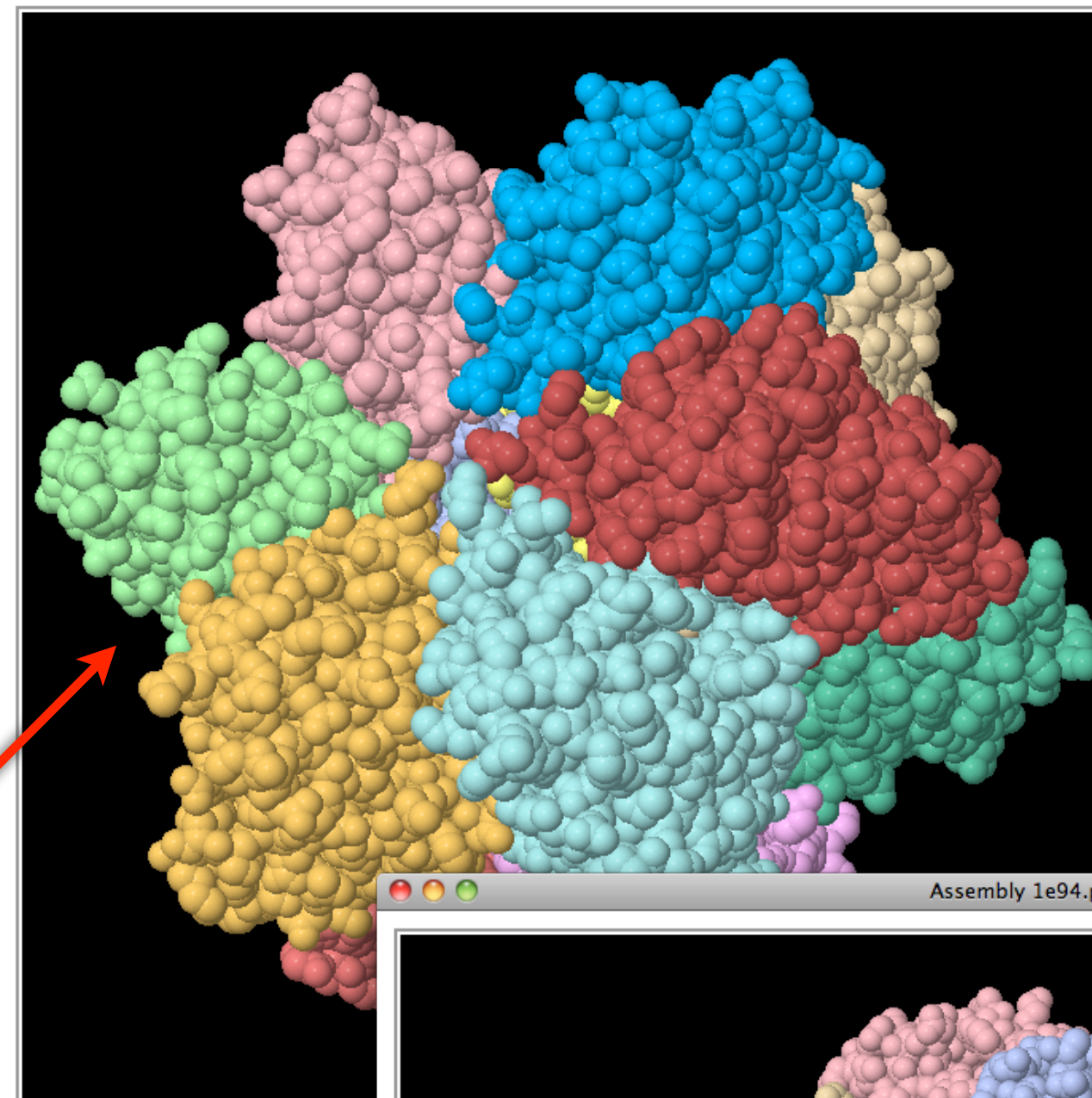
PQS sets 1 to 10 of total 10

Analysis of complex represented As Is by uploaded file is found

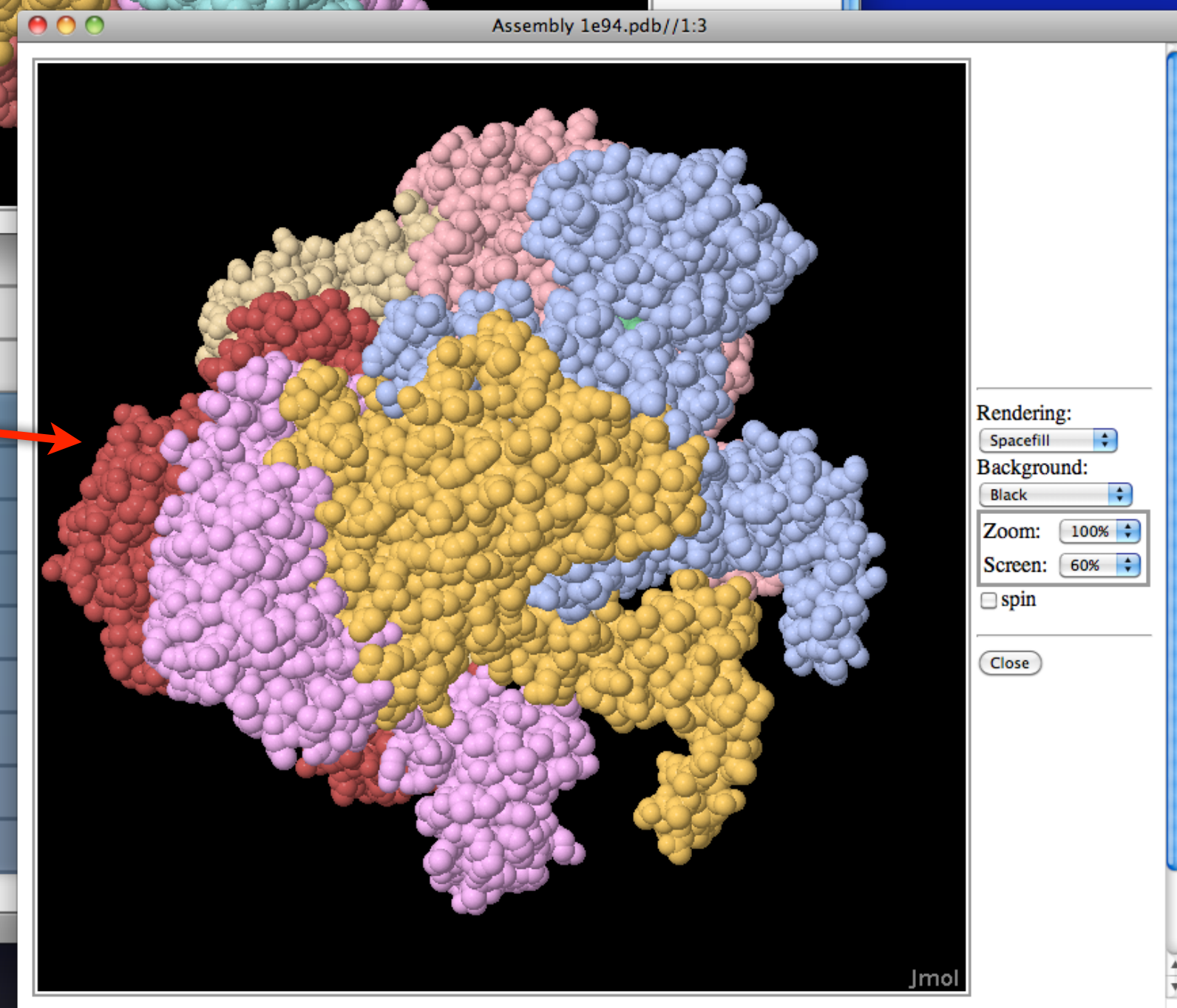
NOTE: In one or more solutions (PQS sets), not all chains appear in their original order to the set number.

Analysis of protein interfaces suggests that the following qu

PQS set		mm Size	Formula	Composition	Id	Biomol R350	Stable		
NN	«»								
1	☒	12	A ₁₂	<u>A₆B₆</u>	<u>1</u>	1	yes	77220	25620
	☐	12	A ₁₂	<u>C₆D₆</u>	<u>1</u>	2	yes	77200	25140
	☐	6	A ₆ a ₆	<u>E₃F₃[ANP]₆</u>	<u>2</u>	3	yes	104500	32220
2	☐	12	A ₁₂	<u>A₆B₆</u>	<u>1</u>	1	yes	77220	25620
	☐	12	A ₁₂	<u>C₆D₆</u>	<u>1</u>	2	yes	77200	25140
	☐	1	Aa	<u>E[ANP]</u>	<u>3</u>	—	yes	21850	850
	☐	1	Aa	<u>F[ANP]</u>	<u>3</u>	—	yes	22030	840
3	☐	6	A ₆ a ₆	<u>E₃F₃[ANP]₆</u>	<u>2</u>	3	yes	104500	32220
	☐	1	A	<u>B</u>	<u>4</u>	—	yes	8560	0
	☐	1	A	<u>C</u>	<u>4</u>	—	yes	8460	0
	☐	1	A	<u>D</u>	<u>4</u>	—	yes	8600	0
	☐	1	A	<u>A</u>	<u>4</u>	—	yes	8580	0



Rendering:
 Spacefill
 Background:
 Black
 Zoom: 100%
 Screen: 60%
☐ spin
 Close



Rendering:
 Spacefill
 Background:
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 Zoom: 100%
 Screen: 60%
☐ spin
 Close

```
Terminal — bash — 88x11
rcccp4mw001:setup Eugene$ ./pisa 1e94 -analyse 1e94.pdb pi

... session '1e94' created
... structure read and understood
... structural equivalence analysed
... molecular surfaces analysed
... 30 potential contacts identified, now processing:
|||||
... total 24 interfaces calculated
... assembly analysis: done
... results written
```

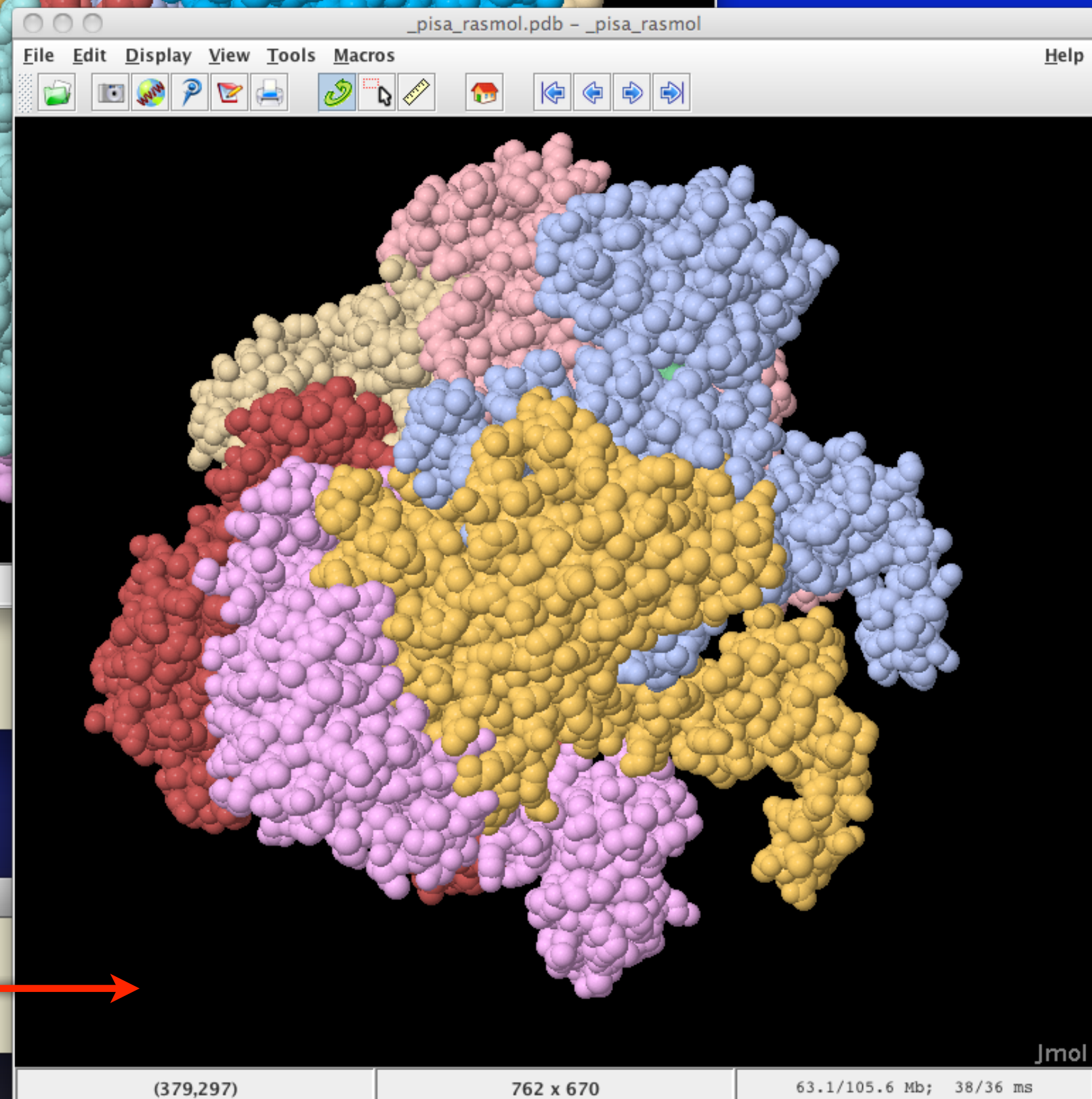
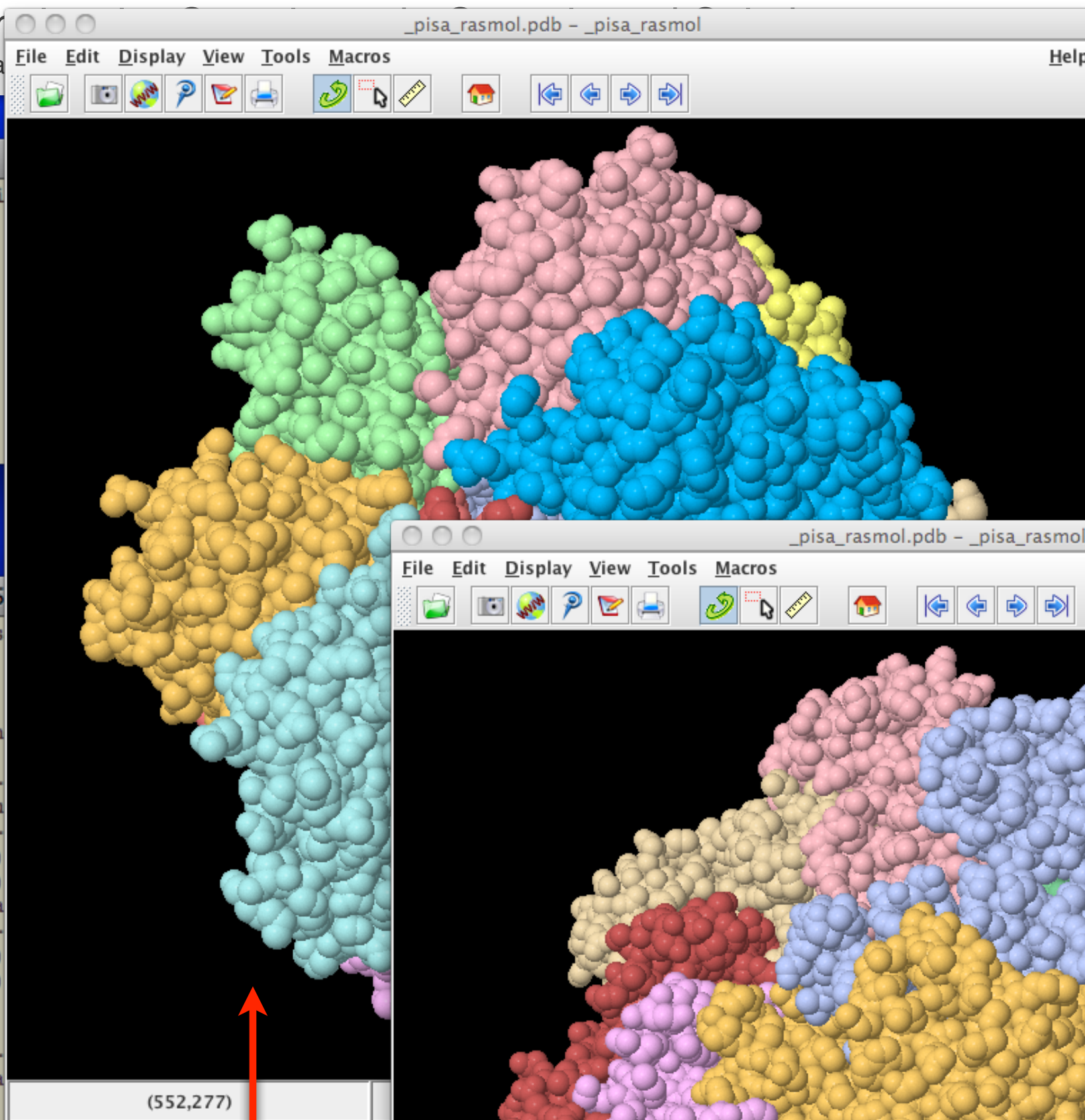
```
Terminal — bash — 90x25
rcccp4mw001:setup Eugene$ ./pisa 1e94 -list assemblies pis

PISA v1.06 << 16/01/2010 >> Session 1e94

Analysis of protein interfaces suggests that the following
quaternary structures are stable in solution
```

Set	No	Size	Id	ASA	BSA	DGdiss	Formu
1	1	12	1	77218.0	25619.7	31.6	A(12)
	2	12	1	77204.0	25139.2	22.4	A(12)
	3	6	2	104504.0	32216.1	69.2	A(6)a
2	4	12	1	77218.0	25619.7	31.6	A(12)
	5	12	1	77204.0	25139.2	22.4	A(12)
	6	1	3	21852.2	852.8	-0.0	Aa
	7	1	3	22032.3	836.1	-0.0	Aa
3	8	6	2	104504.0	32216.1	69.2	A(6)a
	9	1	4	8561.7	0.0	-0.0	A
	10	1	4	8459.0	0.0	-0.0	A
	11	1	4	8598.2	0.0	-0.0	A
	12	1	4	8578.0	0.0	-0.0	A

```
Terminal — bash — 91x5
rcccp4mw001:setup Eugene$ ./pisa 1e94 -view assembly 1 pisa.cfg > /dev/null &
[1] 1189
rcccp4mw001:setup Eugene$ ./pisa 1e94 -view assembly 3 pisa.cfg > /dev/null &
[2] 1202
rcccp4mw001:setup Eugene$
```

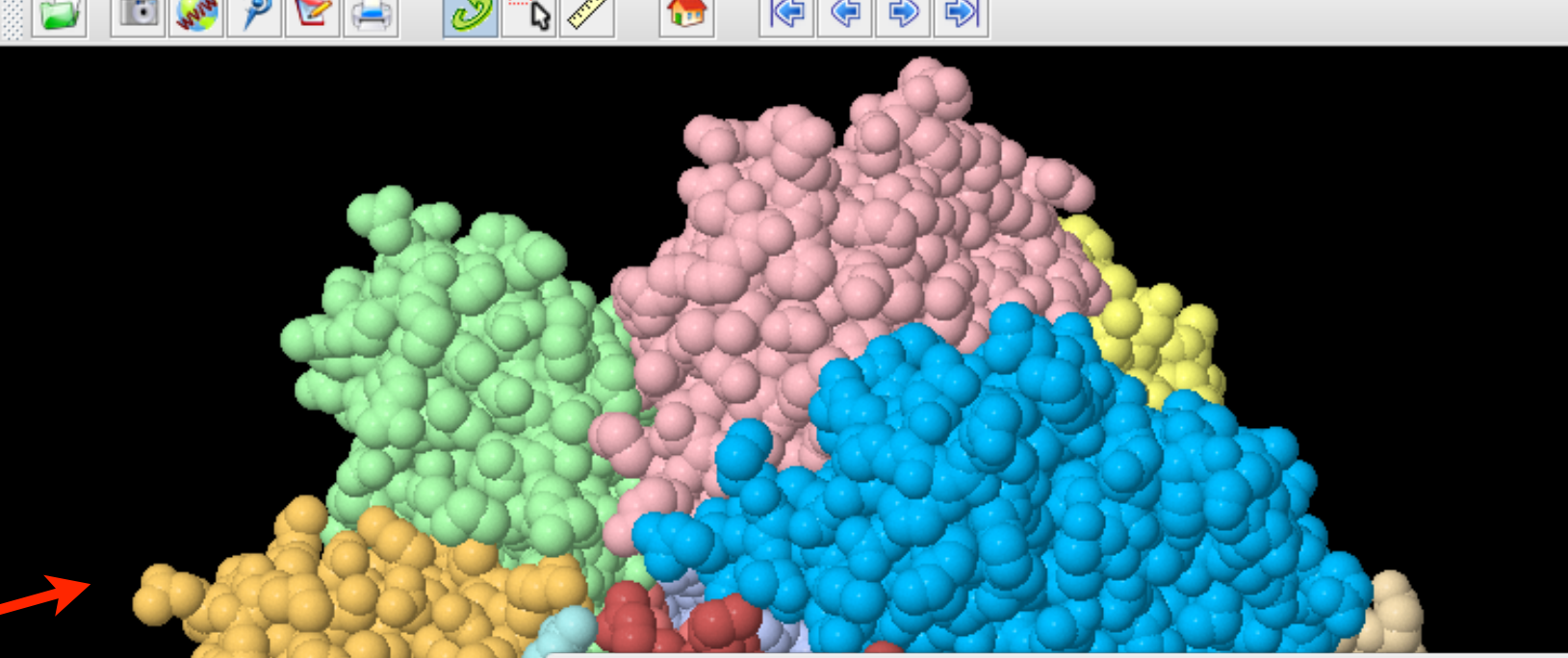


QtPISA 1.12 25/06/2012

File: PDB =>
Title: HSLV-H
Space group: P 63 2 2
Cell: 172.022

ASU (File) contents:
Protein chains: 6 (6)
DNA/RNA chains: 0 (0)
Ligands: 2 (2)
NCS-mates: 0

Contents
Data
Monomers
Interfaces
Assemblies
Stable
Grey area
Unstable
As given

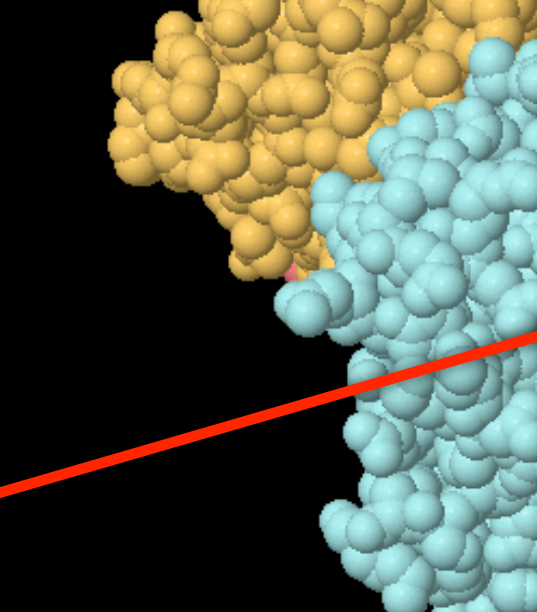


QtPISA 1.12

The following quaternary structures appear to be stable in solution

Set	Size	Type
1	12	1
2	12	1
3	6	2
4	12	1
5	12	1
6	1	3
7	1	3

Contents
Data
Monomers
Interfaces
Assemblies
Stable
1. A(12)
2. A(12)
3. A(6)a(6)

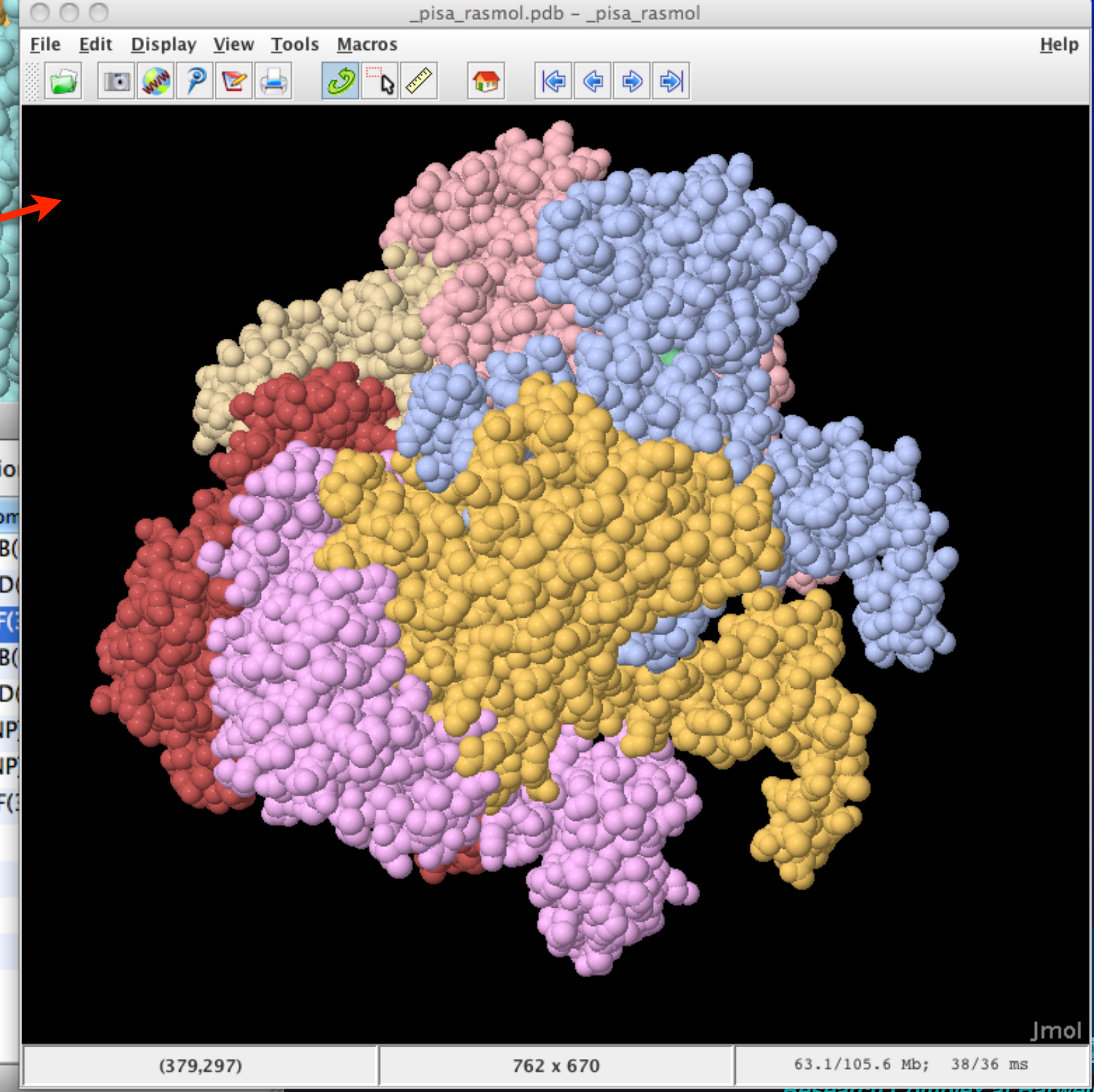


QtPISA 1.12 25/06/2012 [PDB => 1e94]

The following quaternary structures appear to be stable in solution

Set	Size	Type	ASA	BSA	Delta G	Formula	Comp
1	12	1	77218.4	25619.7	31.6	A(12)	A(6)B(6)
2	12	1	77204.3	25139.2	22.4	A(12)	C(6)D(6)
3	6	2	104504.1	32216.1	69.2	A(6)a(6)	E(3)F(3)
4	12	1	77218.4	25619.7	31.6	A(12)	A(6)B(6)
5	12	1	77204.3	25139.2	22.4	A(12)	C(6)D(6)
6	1	3	21852.2	852.8	-0.0	Aa	E[ANP]
7	1	3	22032.3	836.1	-0.0	Aa	F[ANP]
8	3	6	104504.1	32216.1	69.2	A(6)a(6)	E(3)F(3)
9	1	4	8561.7	0.0	-0.0	A	B
10	1	4	8459.0	0.0	-0.0	A	C
11	1	4	8598.2	0.0	-0.0	A	D
12	1	4	8578.0	0.0	-0.0	A	A

Contents
Data
Monomers
1. A
2. B
3. C
4. D
5. E
6. F
7. [ANP]E:500
8. [ANP]F:501
Interfaces
Assemblies
Stable
Grey area
Unstable
As given



Detection of Biological Units in Crystals: PISA summary

1. Enumerate all possible assemblies in crystal packing, subject to crystal properties: space symmetry group, geometry and composition of the Asymmetric Unit

- Achieved with graph-theoretical techniques, by representing crystal as an infinite periodic graph of connected macromolecules

2. Evaluate assemblies for chemical stability:

$$\Delta G_0 = -\Delta G_{\text{int}} - T\Delta S > 0$$

3. Leave only sets of stable assemblies in the list, and range by chances to be a biological unit:

- Larger assemblies take preference
- Single-assembly sets take preference
- Otherwise, assemblies with higher ΔG_0 take preference



Classification of Protein Assemblies

Assembly classification on the benchmark set of 218 protein structures published in

Ponstingl, H., Kabir, T. and Thornton, J. (2003) Automatic inference of protein quaternary structures from crystals. J. Appl. Cryst. 36, 1116-1122.

	1mer	2mer	3mer	4mer	6mer	Other	Sum	Correct
1mer	49	3	0	1	1	1	55	89%
2mer	3	71+11	0	2+1	0	0	76+12	93%
3mer	1	0	22	0	1	0	24	92%
4mer	2	2+1	0	26+6	0	1	31+7	84%
6mer	0	0	0	1	10+2	0	10+3	92%
196+22 $\Leftrightarrow$ 196 homomers and 22 heteromers							Total: 196+22	90%

Classification error in ΔG_0 : ± 5 kcal/mol



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Classification of Protein-DNA Complexes

Assembly classification on the benchmark set of 212 protein-DNA complexes published in

Luscombe, N.M., Austin, S.E., Berman, H.M. and Thornton, J. (2000) An overview of the structures of protein-DNA complexes. Genome Biol. 1, 1-37.

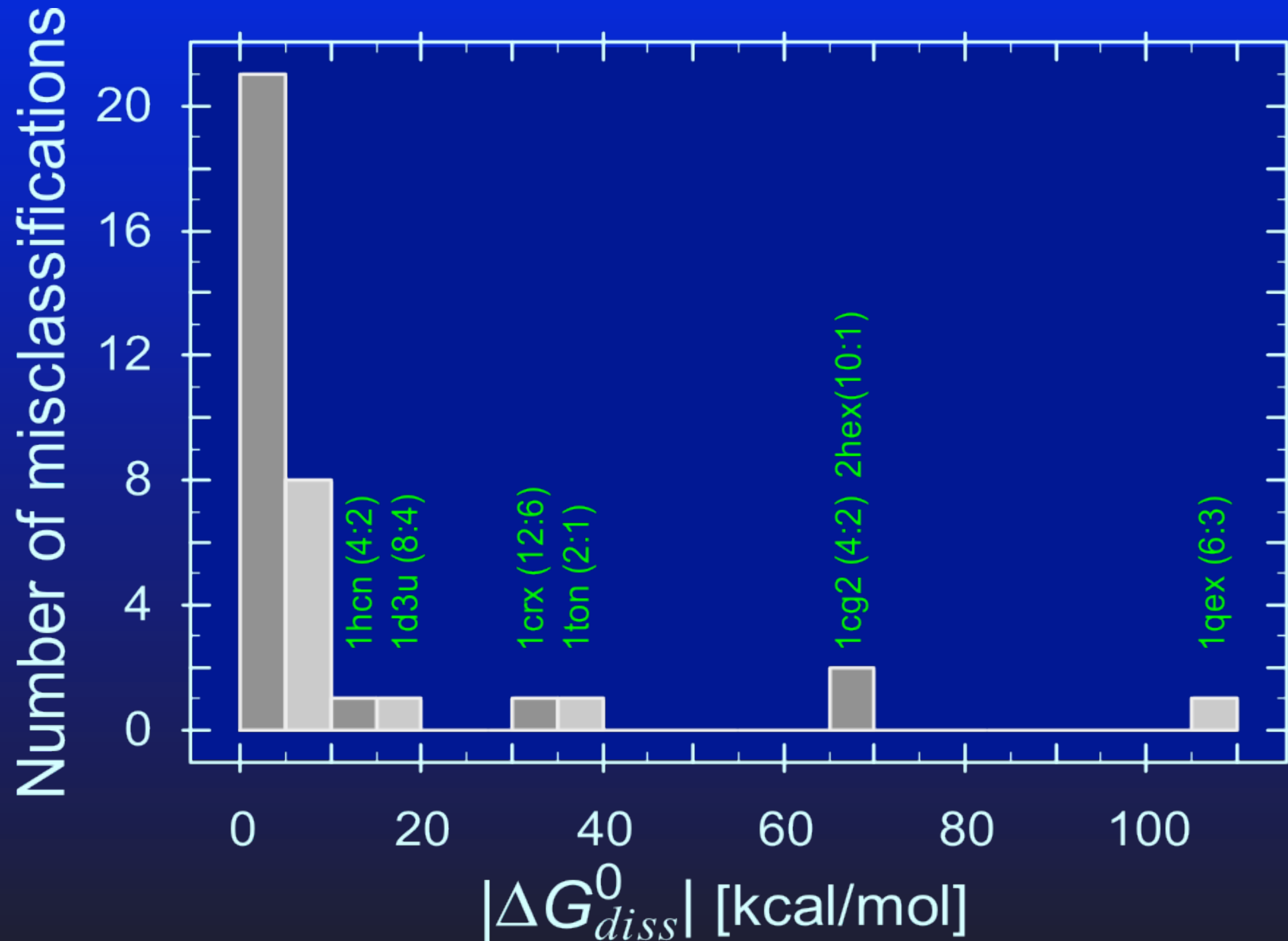
	2mer	3mer	4mer	5mer	6mer	10mer	Other	Sum	Correct
2mer	1	0	0	0	0	0	0	1	100%
3mer	6	96	0	0	1	0	2	105	91%
4mer	0	2	83	0	0	0	0	85	98%
5mer	0	0	2	3	0	0	0	5	60%
6mer	1	0	0	0	13	0	1	15	87%
10mer	0	0	0	0	0	1	0	1	100%
Total:								212	93%

Classification error in ΔG_0 : ± 5 kcal/mol



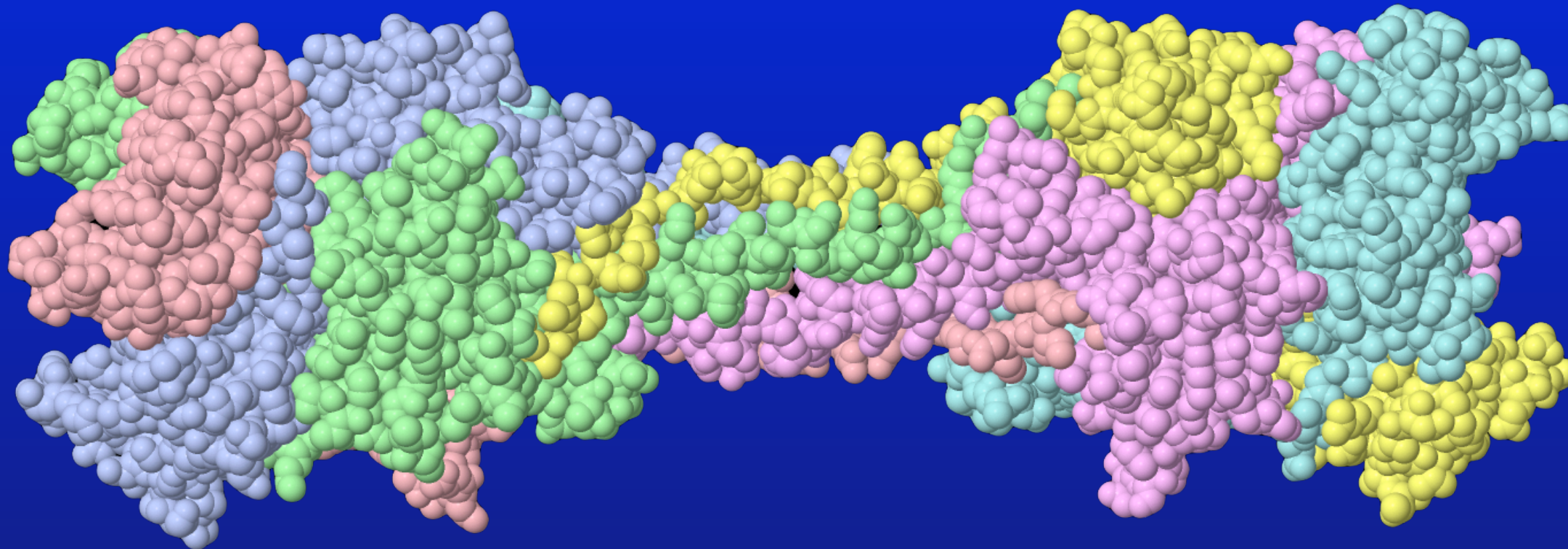
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Free Energy Distribution of Misclassifications

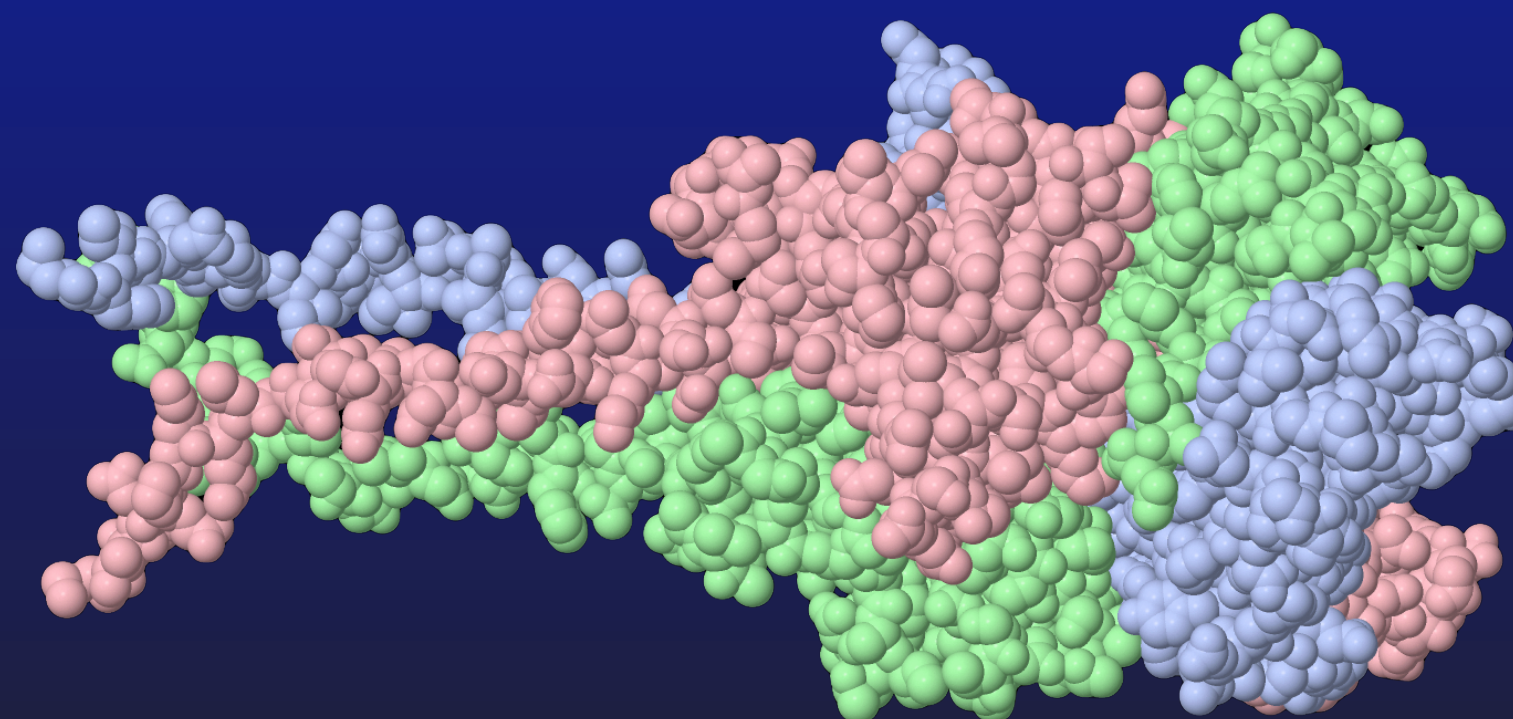


Example of misclassification: 1QEX

BACTERIOPHAGE T4 GENE PRODUCT 9 (GP9), THE TRIGGER OF TAIL CONTRACTION AND THE LONG TAIL FIBERS CONNECTOR



Predicted: homohexamer
Dissociates into 2 trimers
 $\Delta G_0 \approx 106$ kcal/mol

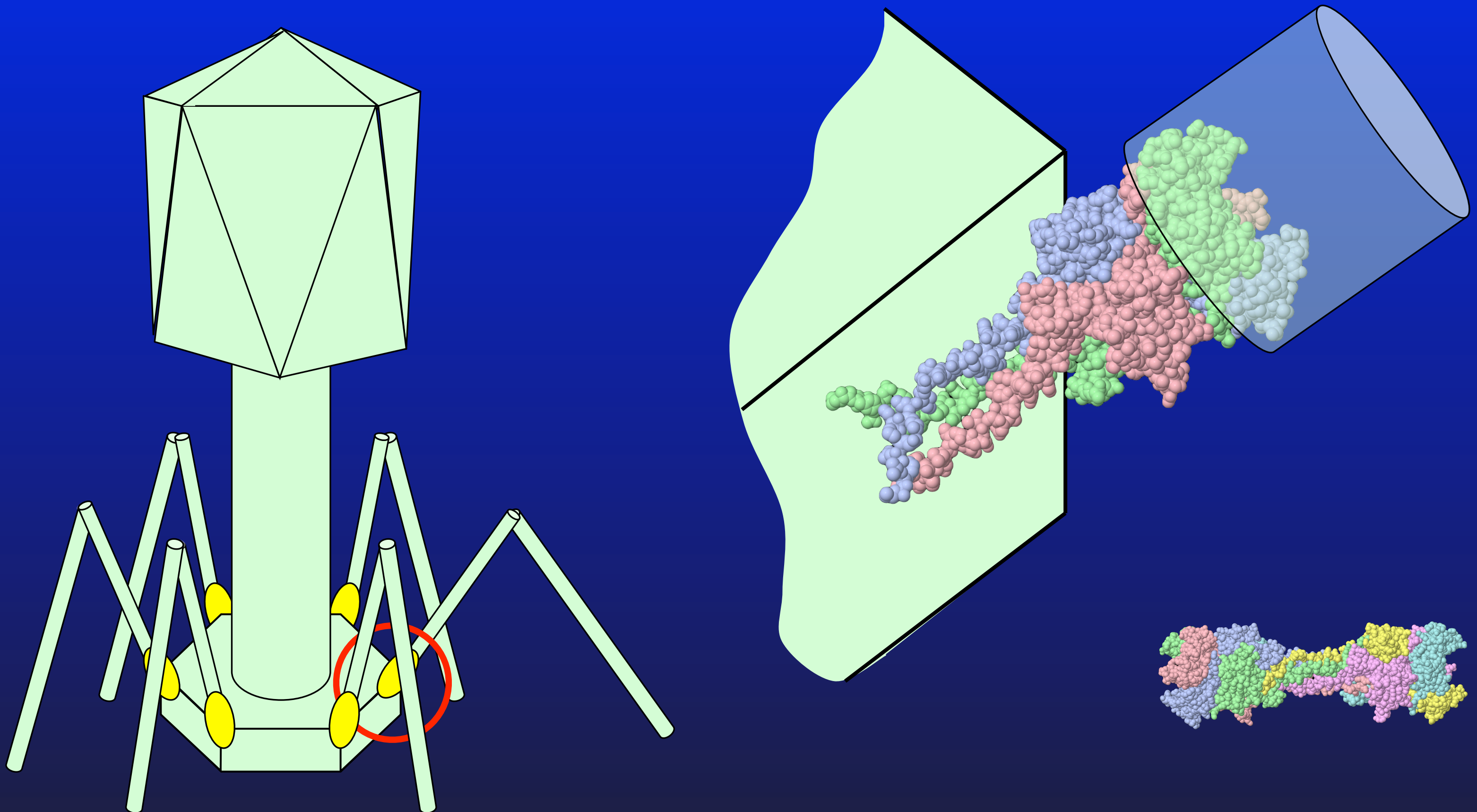


Biological unit: homotrimer
Dissociates into 3 monomers
 $\Delta G_0 \approx 90$ kcal/mol



Example of misclassification: 1QEX

BACTERIOPHAGE T4 GENE PRODUCT 9 (GP9), THE TRIGGER OF TAIL CONTRACTION AND THE LONG TAIL FIBERS CONNECTOR



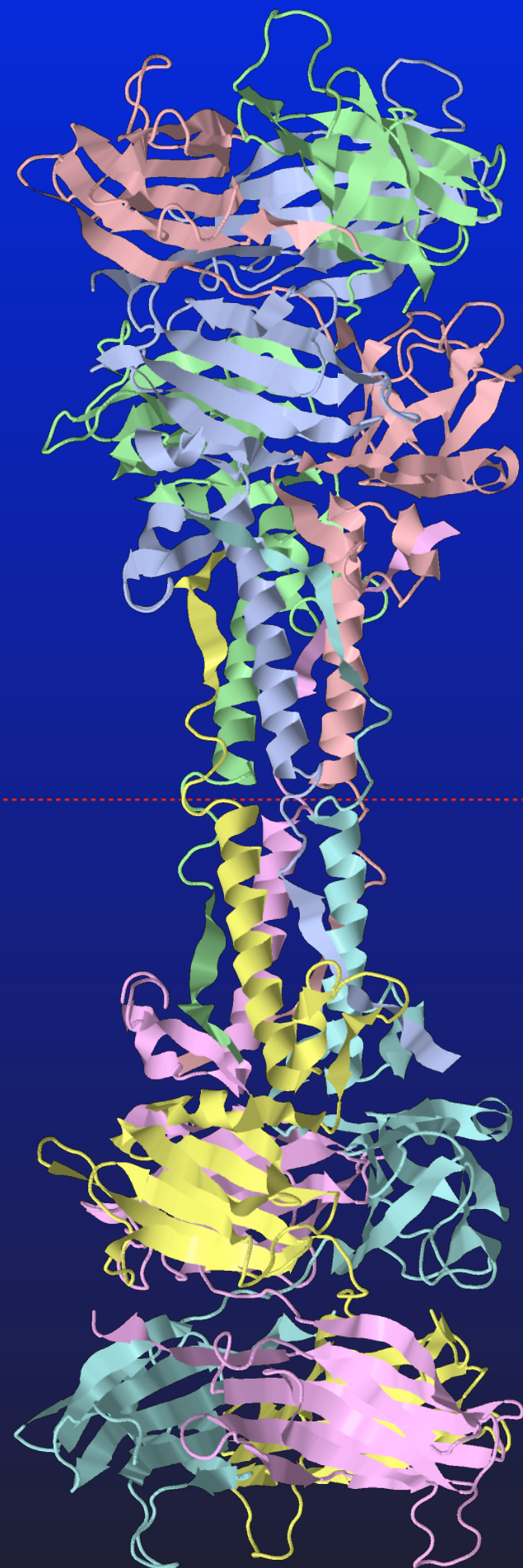
Rossmann M.G., Mesyanzhinov V.V., Arisaka F and Leiman P.G. (2004) *The bacteriophage T4 DNA injection machine*.
Curr. Opin Struct. Biol. 14:171-180.



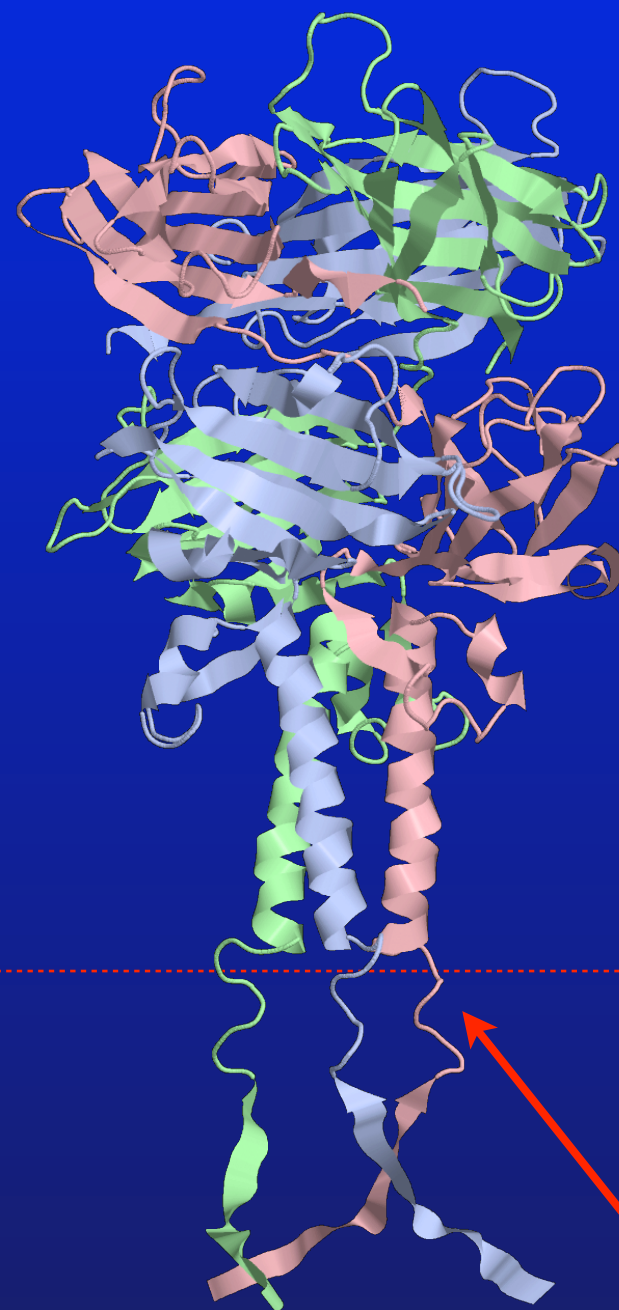
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Example of misclassification: 1QEX

BACTERIOPHAGE T4 GENE PRODUCT 9 (GP9), THE TRIGGER OF TAIL CONTRACTION AND THE LONG TAIL FIBERS CONNECTOR

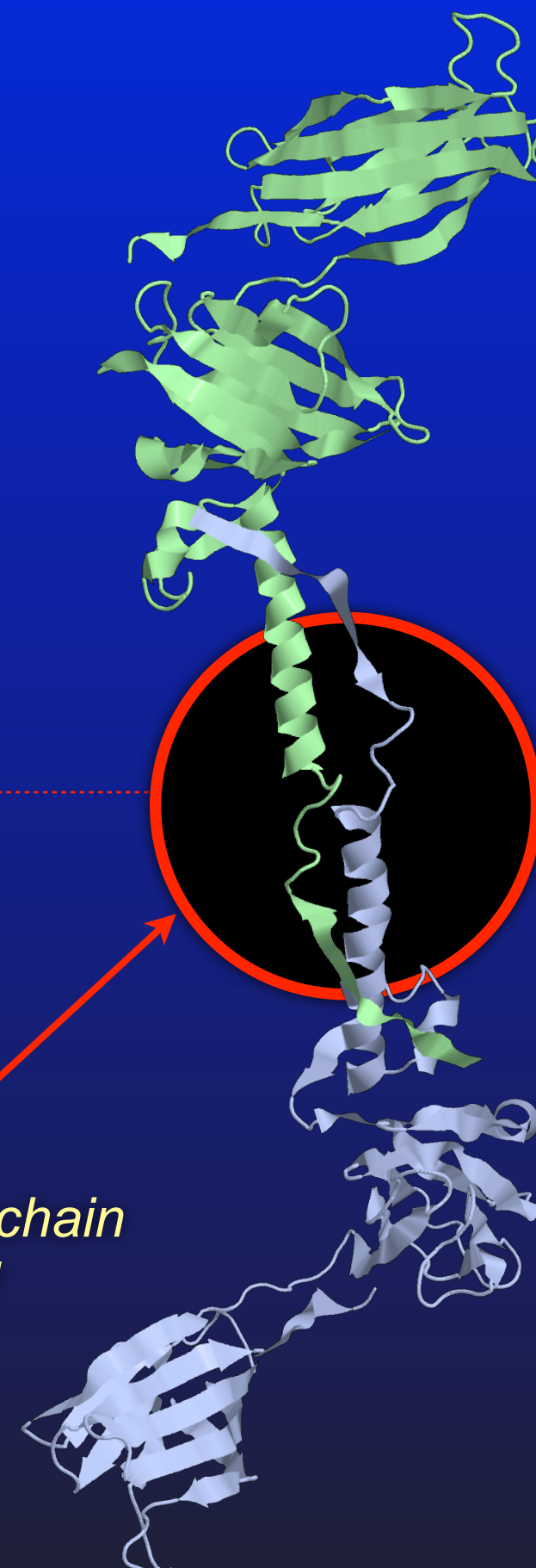


1QEX hexamer

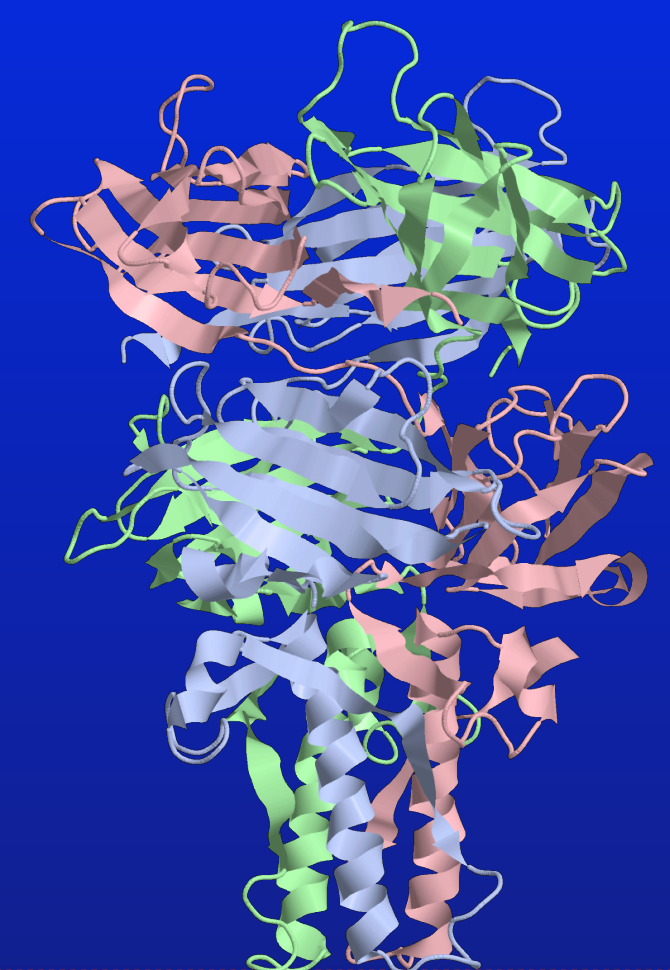


1QEX trimer

Wrong mainchain tracing!



1QEX asu

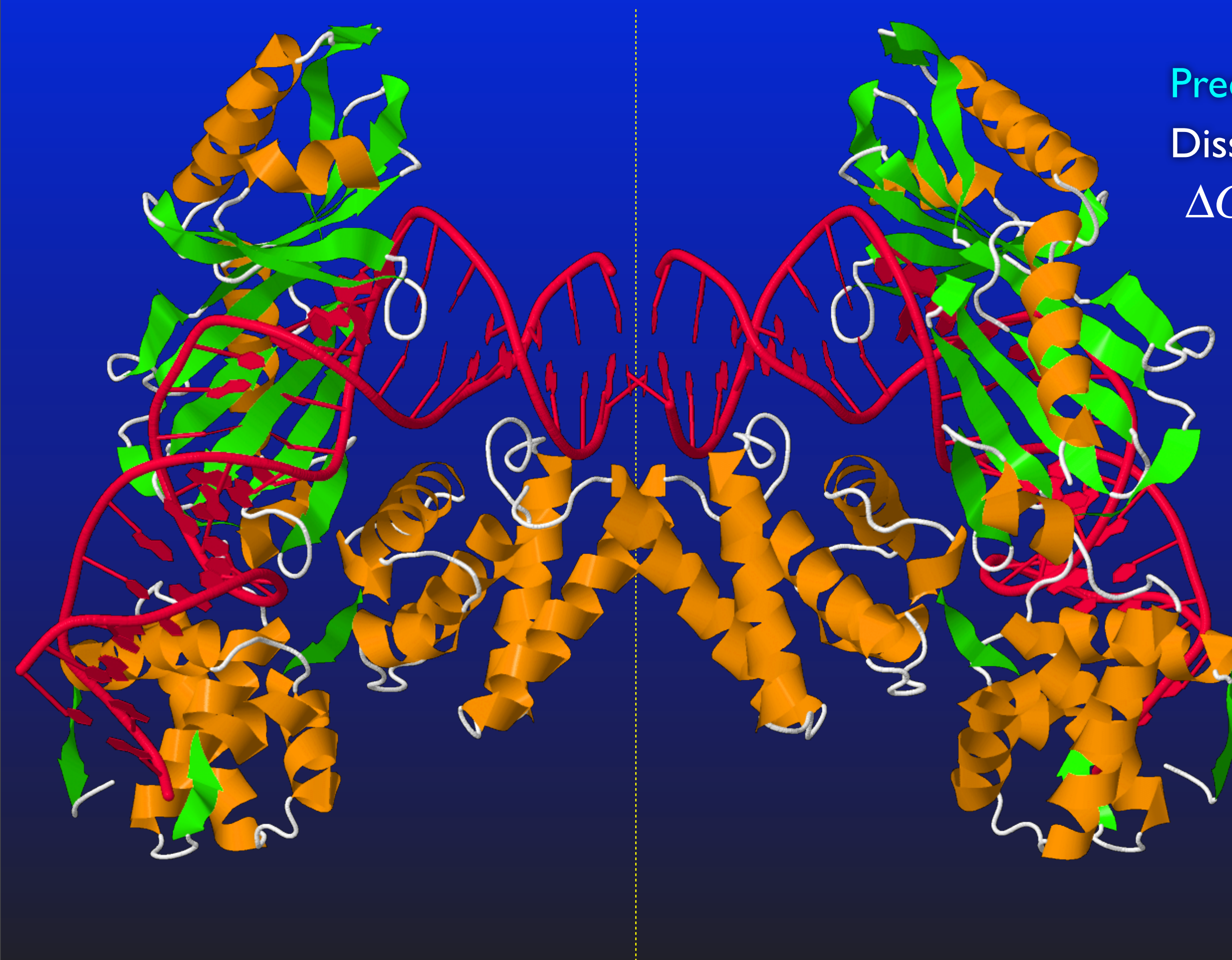


1S2E trimer
Correct mainchain tracing
Identified correctly



Example of misclassification: 1D3U

TATA-BINDING PROTEIN / TRANSCRIPTION FACTOR

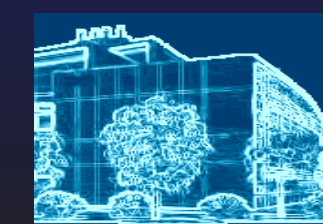


Predicted: octamer

Dissociates into 2 tetramers

$$\Delta G_0 \approx 20 \text{ kcal/mol}$$

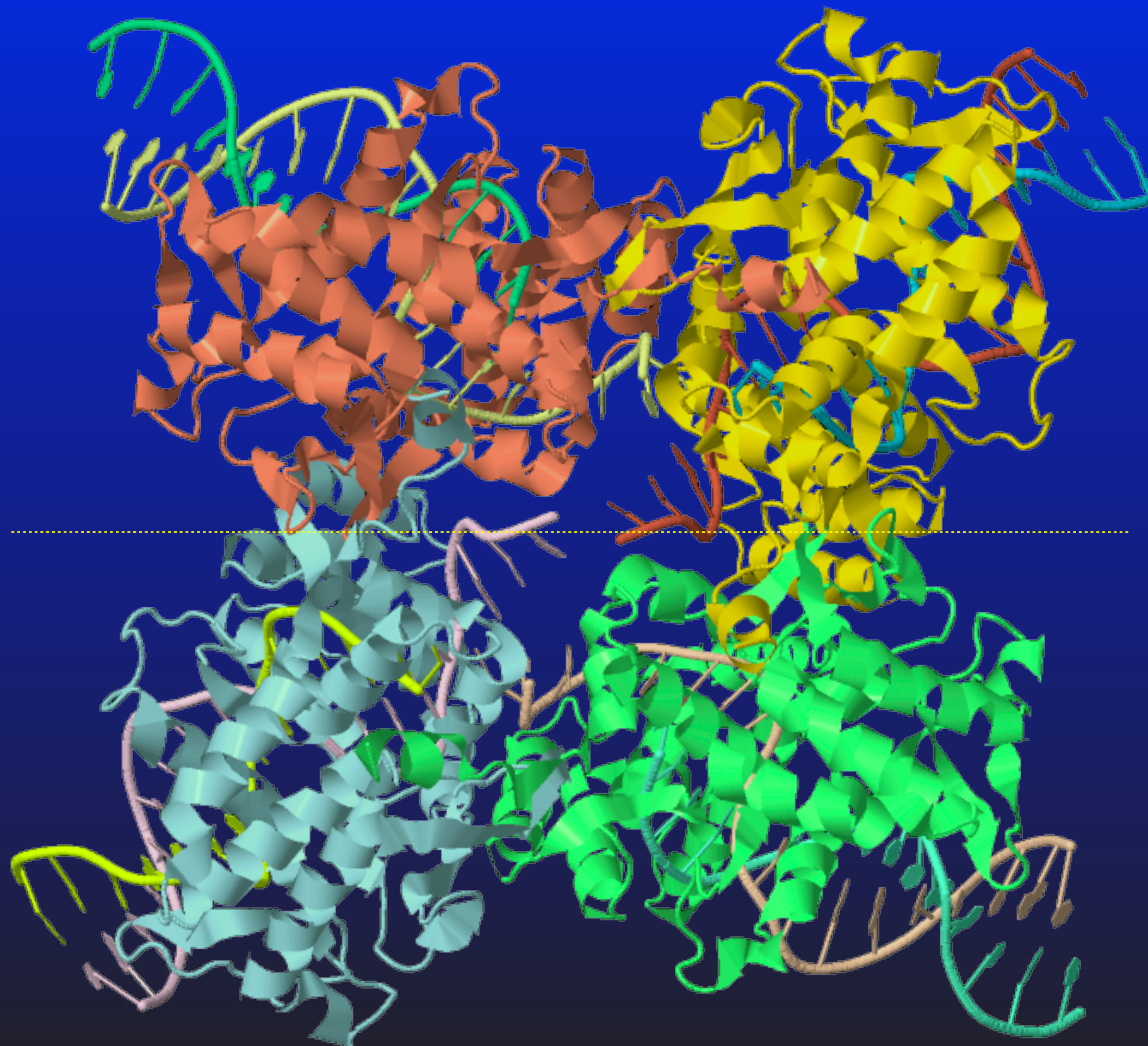
Functional unit:
tetramer



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Example of misclassification: 1CRX

CRE RECOMBINASE / DNA COMPLEX REACTION INTERMEDIATE



Predicted: dodecamer

Dissociates into 2 hexamers

$$\Delta G_0 \approx 28 \text{ kcal/mol}$$

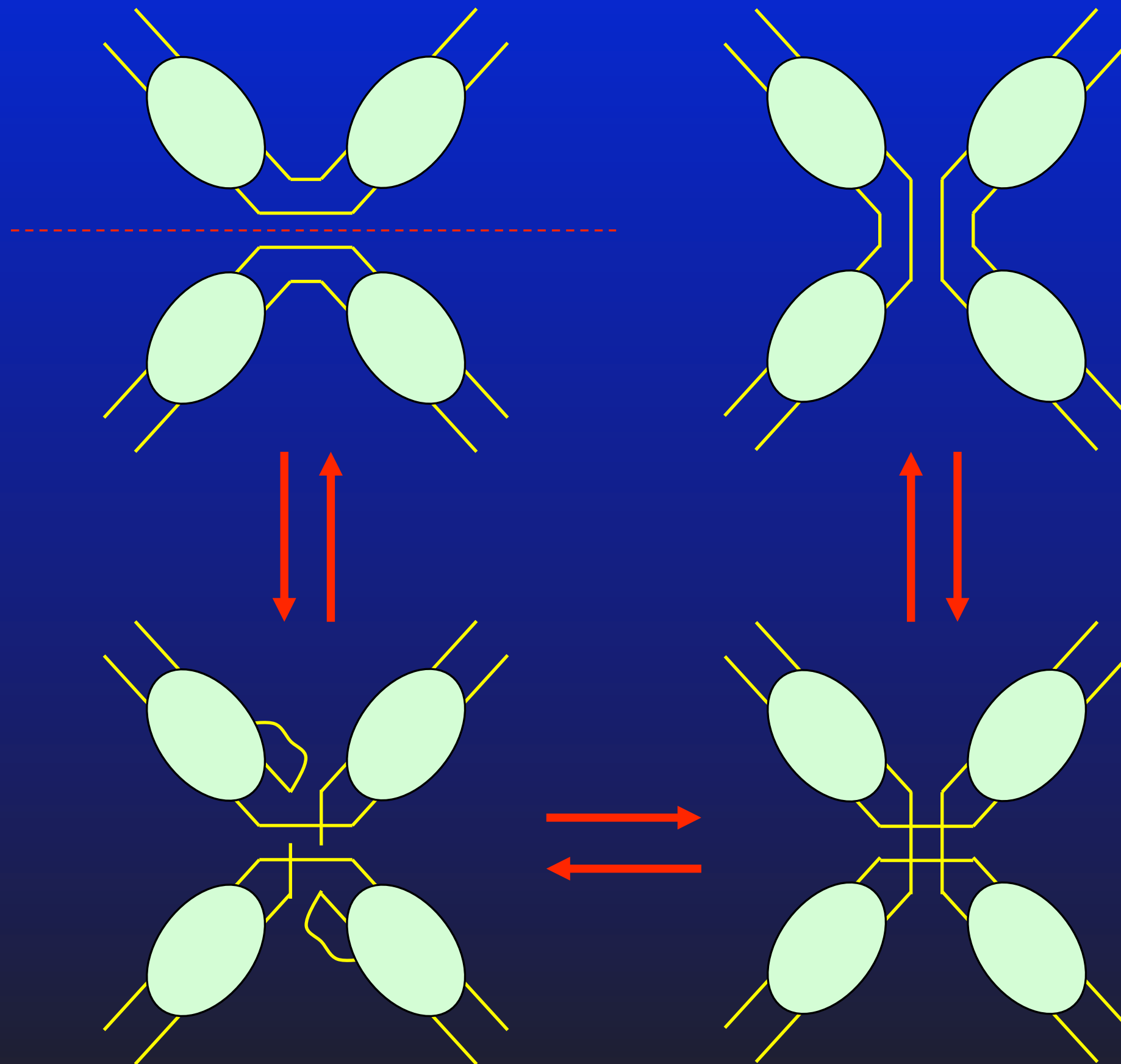
Functional unit: trimer



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Example of misclassification: 1CRX

CRE RECOMBINASE / DNA COMPLEX REACTION INTERMEDIATE



Guo F., Gopaul D.N. and van
Duyne G.D. (1997)

*Structure of Cre recombinase
complexed with DNA in a site-
specific recombination
synapse.*

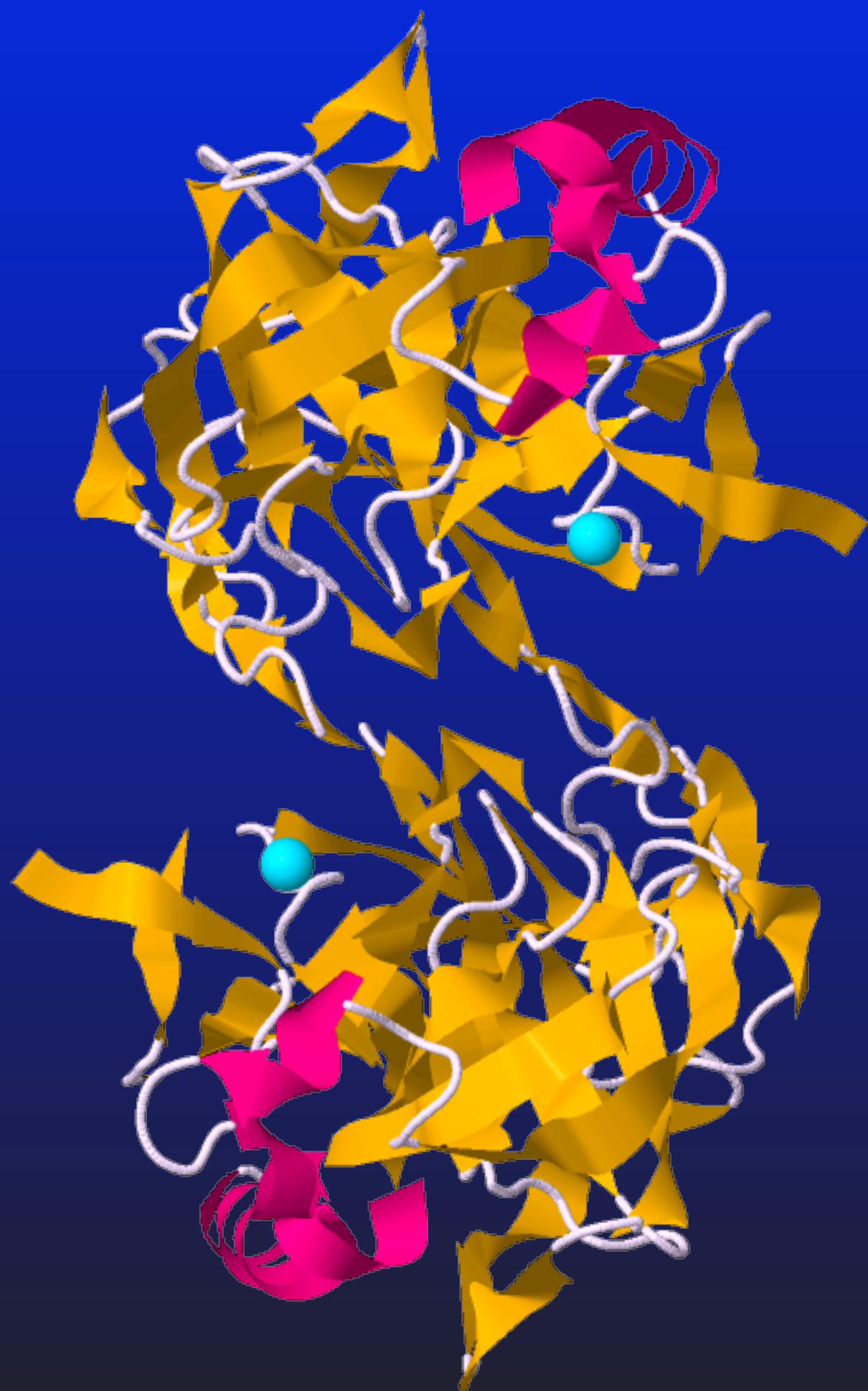
Nature 389:40-46.



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Example of misclassification: 1TON

TONIN



Predicted: dimer

Dissociates at

$$\Delta G_0 \approx 37 \text{ kcal/mol}$$

Biological unit: monomer

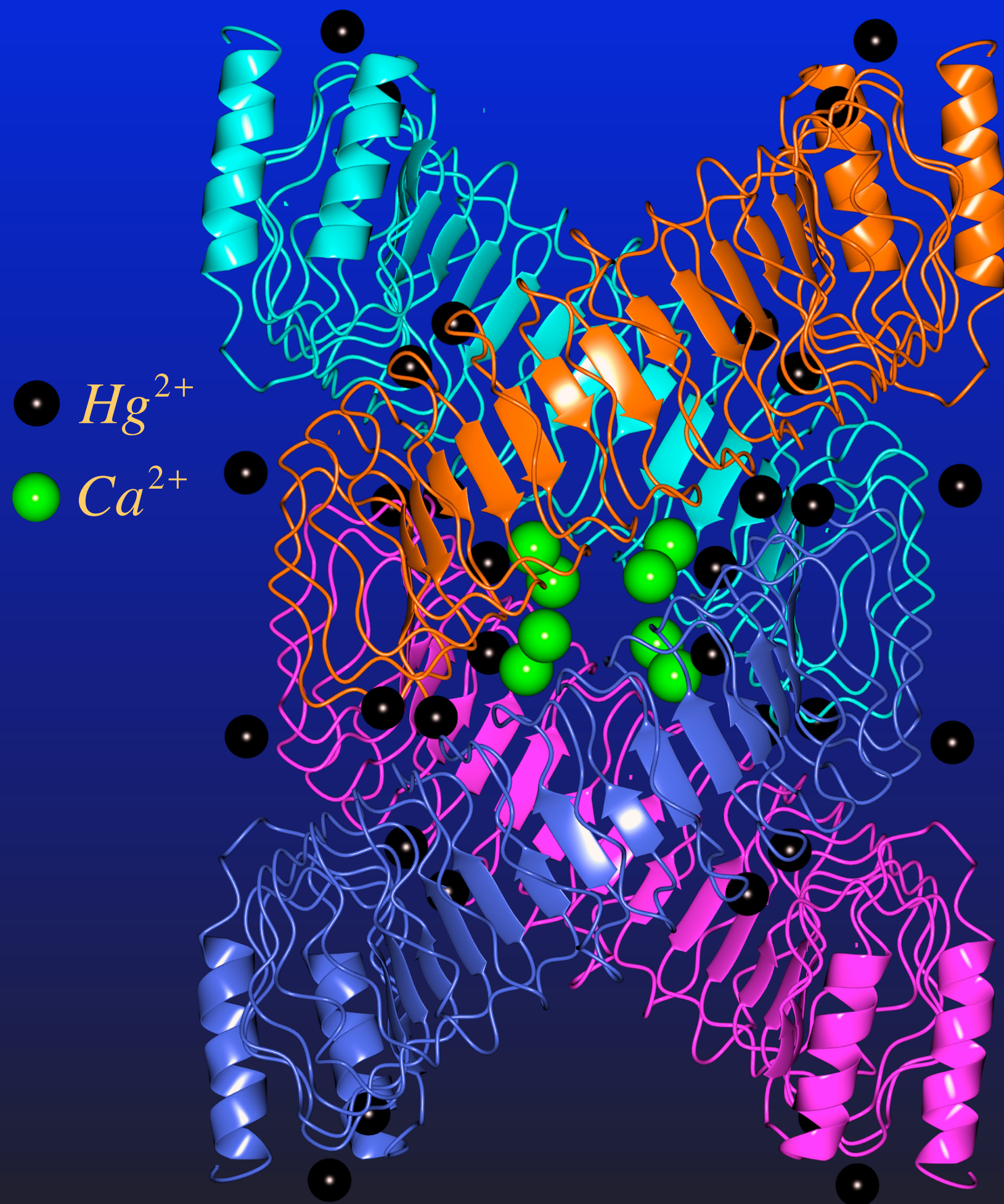
Apparent dimerization is an artefact due to the presence of Zn^{+2} ions added to the buffer to aid crystallization. Removal of Zn from the file results in $\Delta G_0 \approx 3 \text{ kcal/mol}$

Fujinaga M., James M.N.G. (1997) *Rat submaxillary gland serine protease, tonin structure solution and refinement at 1.8 Å resolution.* J.Mol.Biol. 195:373-396.



Example of ion effect: 1G9U vs 1JL5

Y. PESTIS CYTOXIN YopM



Predicted: homotetramer in form of a superhelix featuring a hollow cylinder with an inner diameter of ~ 35 Å.

	1G9U	1JL5
Space Group	$P4_222$	$I4_122$
ΔG_0 , kcal/mol	37	3
Number of ions	40	16

Biological unit: monomer

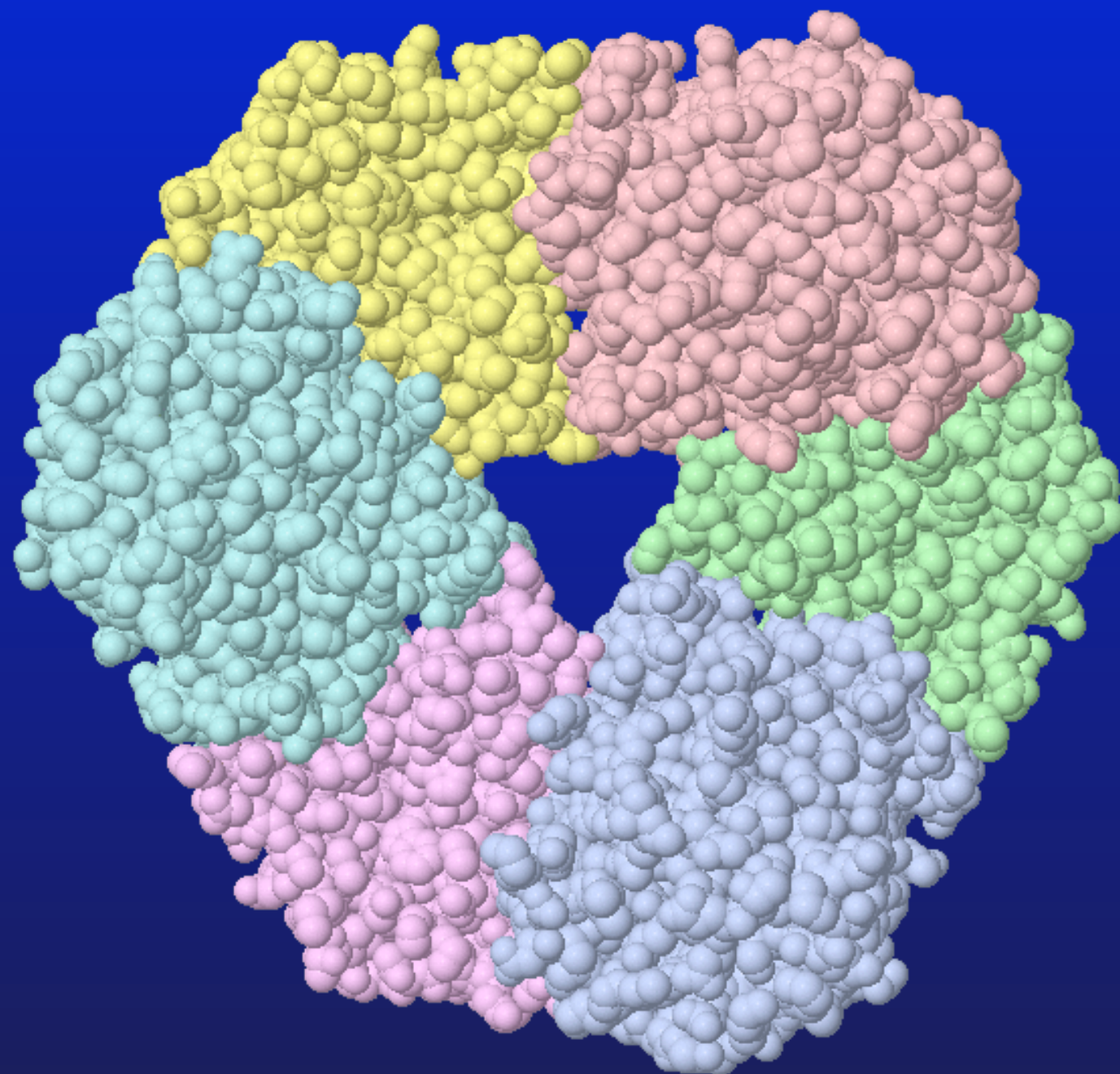
Evdokimov, A. G., Anderson, D. E., Routzahn, K. M. & Waugh, D. S. (2001). J. Mol. Biol. 312, 807–821

Removal of ions makes the structure monomeric in PISA estimates

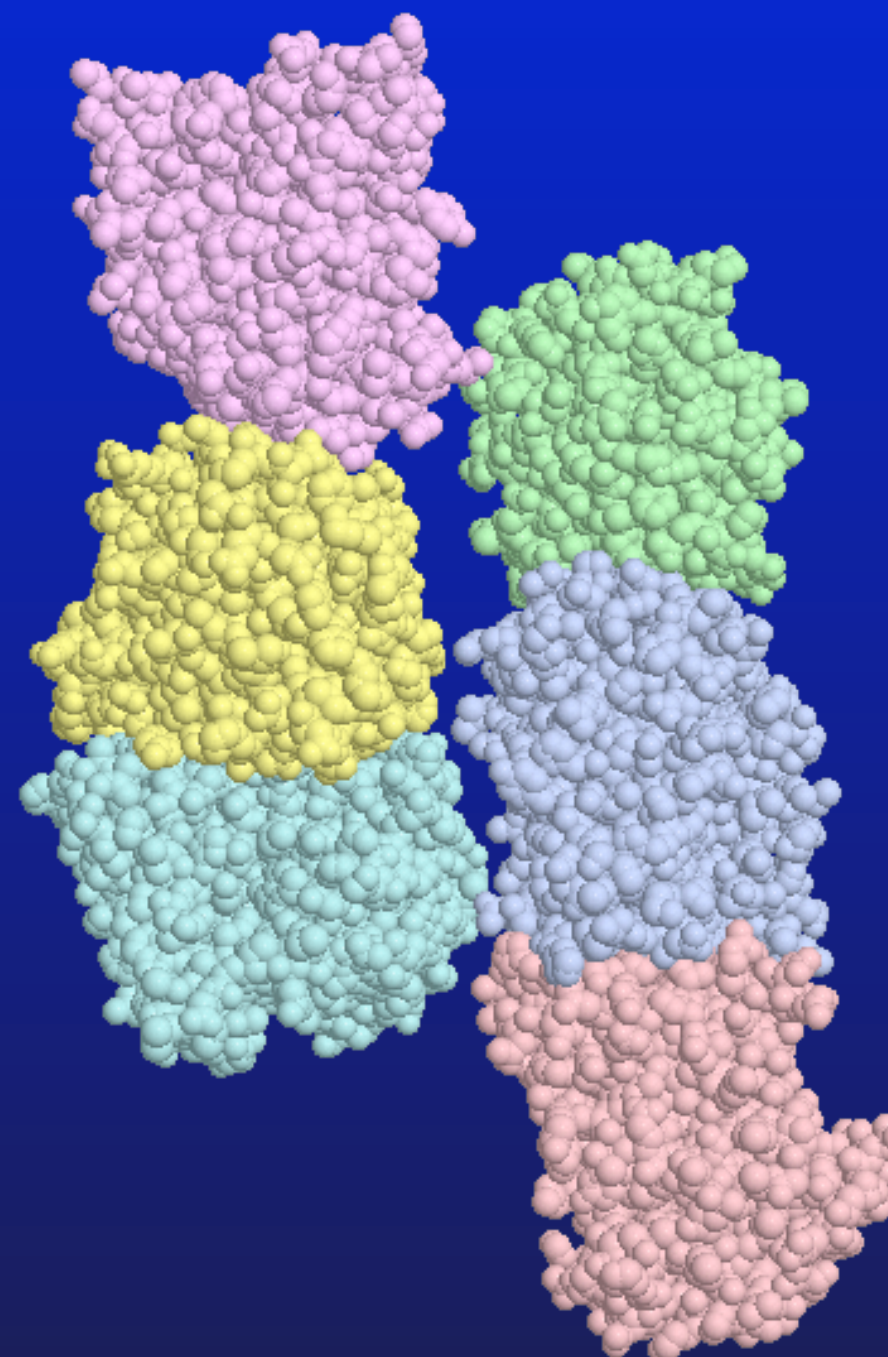


Research Complex at Harwell

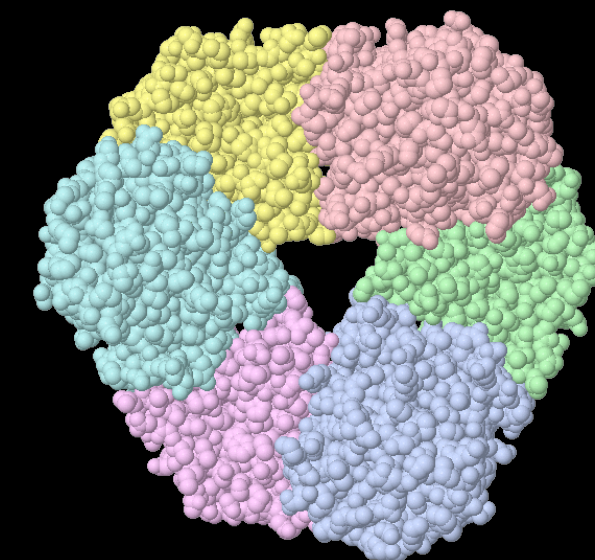
Example of misclassification: 1YWK



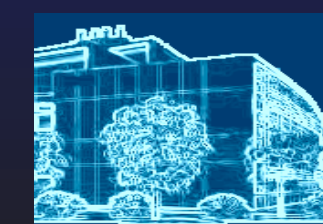
Predicted: homohexameric
Dissociates into 3 dimers at
 $\Delta G_0 \approx 4.4$ kcal/mol



Believed to be:
monomeric,
6 units in ASU

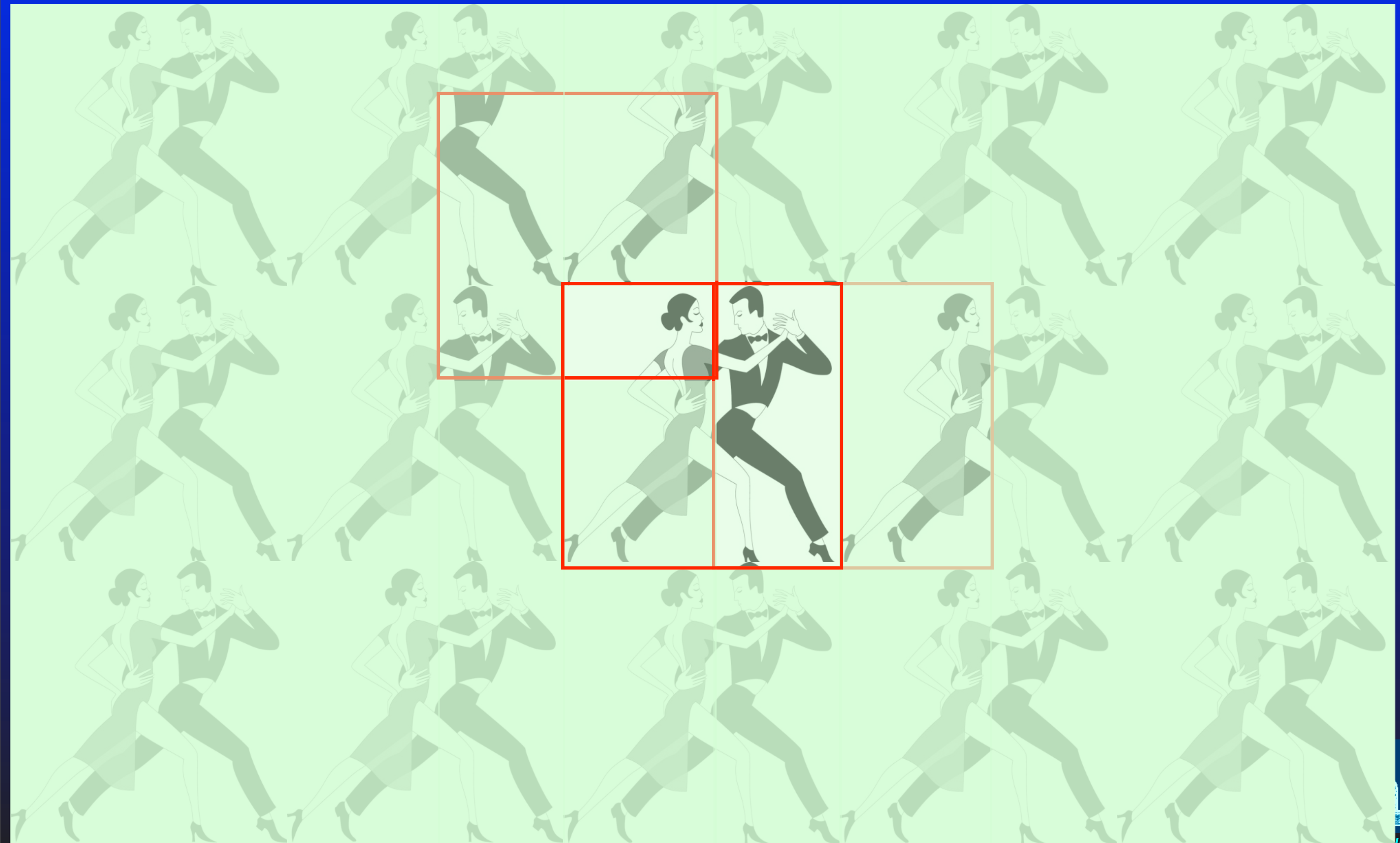


Structural homologue
IXRU:
RMSD = 0.9 Å
Seq.Id = 50%
Homohexameric with
 $\Delta G_0 \approx 20$ kcal/mol

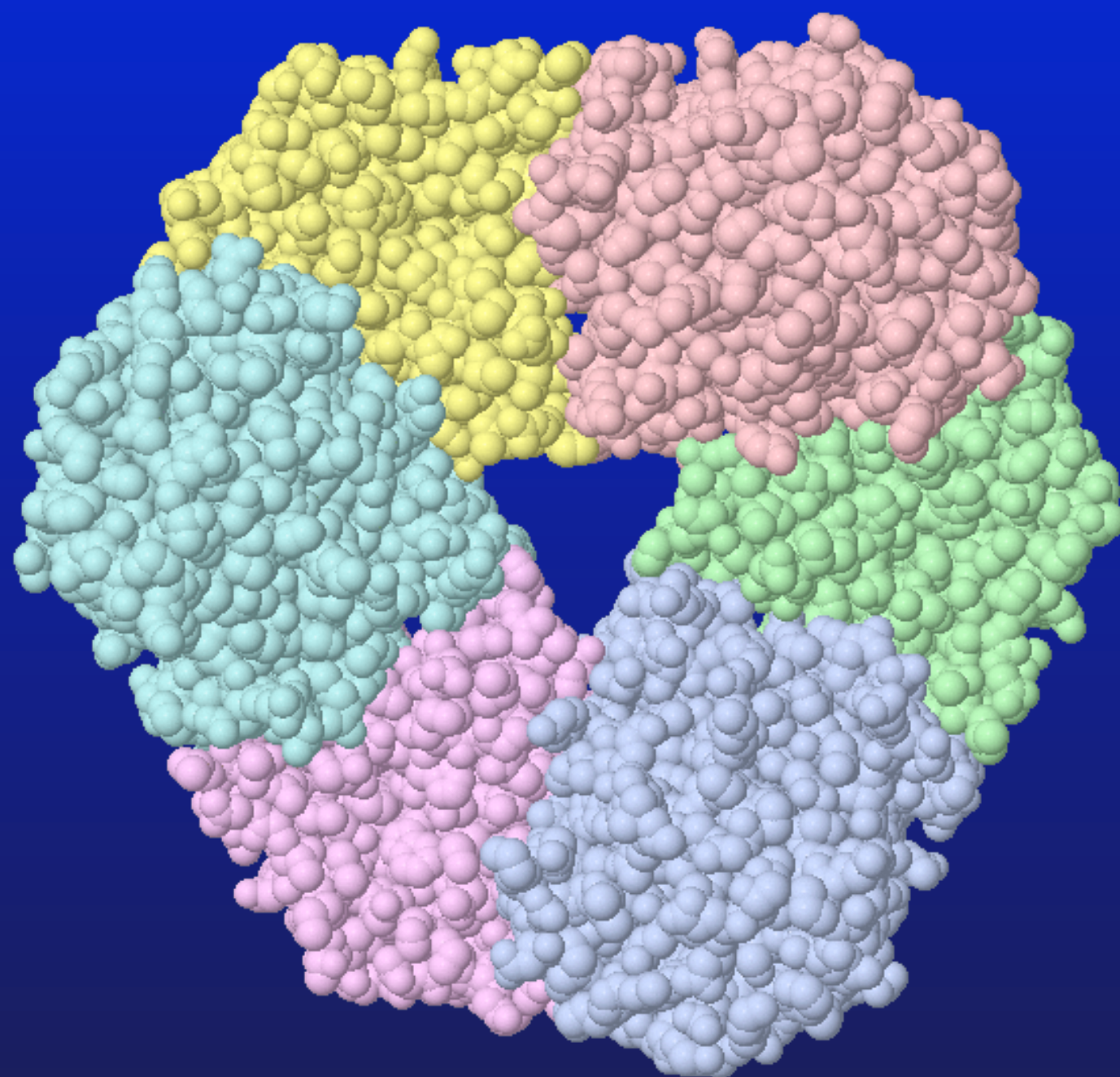


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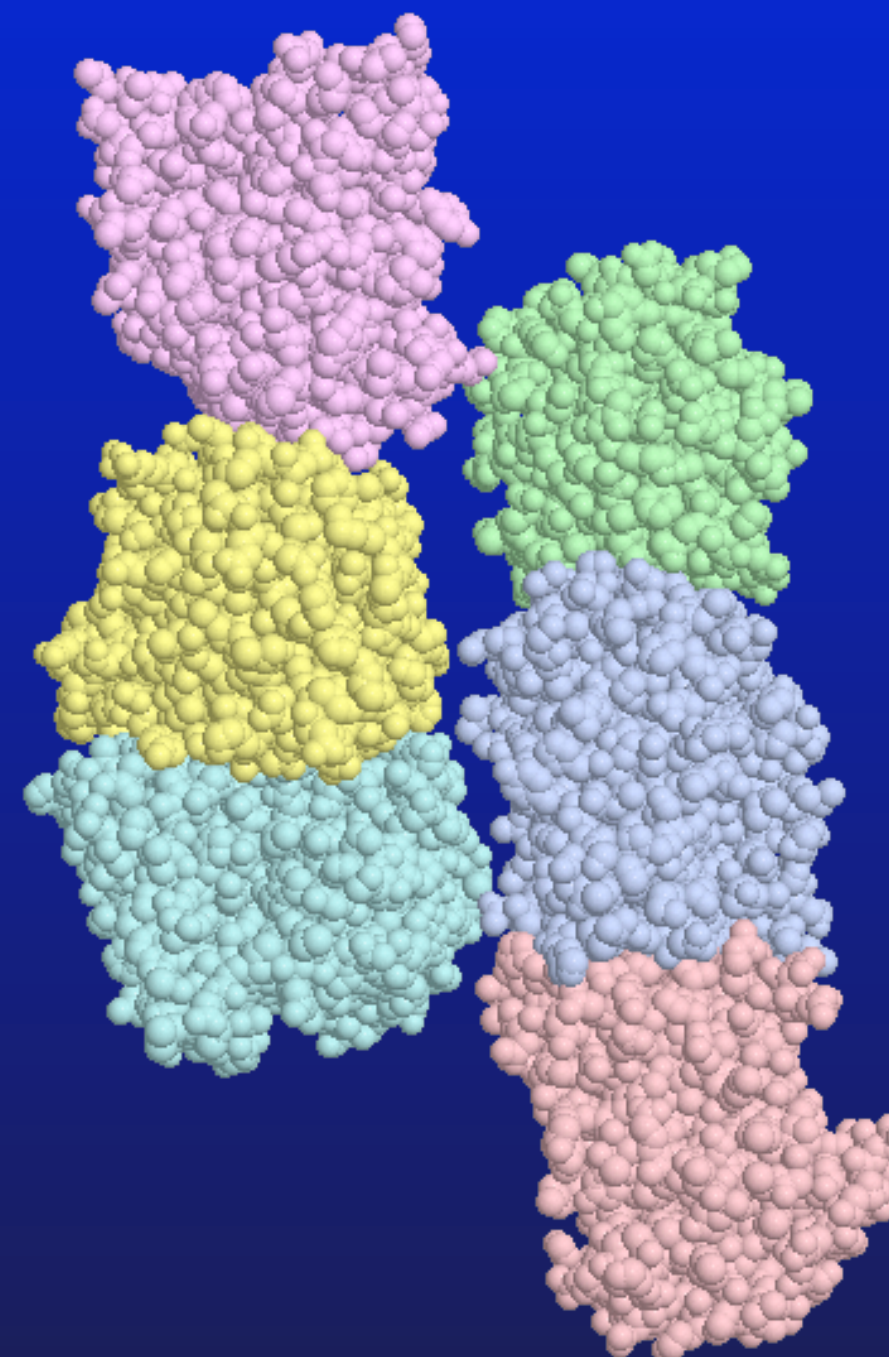
Choice of ASU



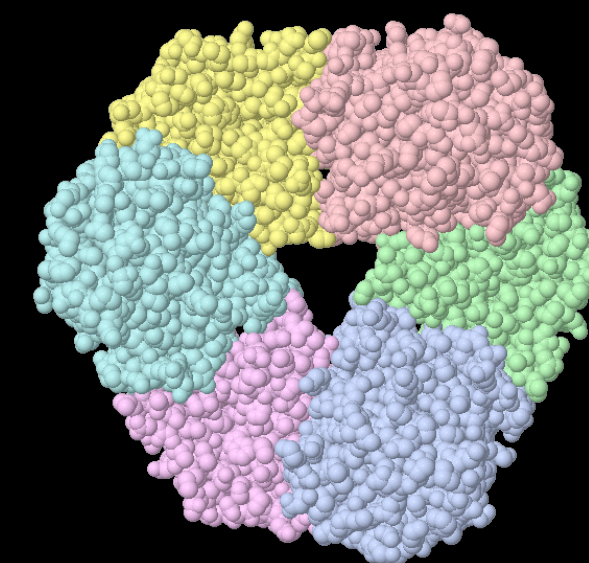
Example of misclassification: 1YWK



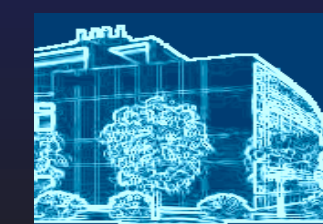
Predicted: homohexameric
Dissociates into 3 dimers at
 $\Delta G_0 \approx 4.4$ kcal/mol



Believed to be:
monomeric,
6 units in ASU



Structural homologue
IXRU:
RMSD = 0.9 Å
Seq.Id = 50%
Homohexameric with
 $\Delta G_0 \approx 20$ kcal/mol



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Does it really work?

- ★ PISA appears to work quite well, which seems to be a “problem”
 - ➔ 90% success rate achieved on the benchmark set
 - ➔ in 2007, wwPDB adopted PISA as a mandatory processing tool for all depositions
 - ➔ since that, feedback from wwPDB curators suggests that up to 95% of classifications made by PISA agree with experimental data on oligomeric state, where available, and with intuitive and common-sense considerations where experimental evidence is not given

- ★ Why it should work well? Two reasons:

Energy models and calculations are quite accurate

PISA relies on geometry of interactions given by crystal packing. PISA does not dock monomeric units; rather, it uses crystal contacts as “nature’s dockings” assuming that they are correct.

Obviously wrong

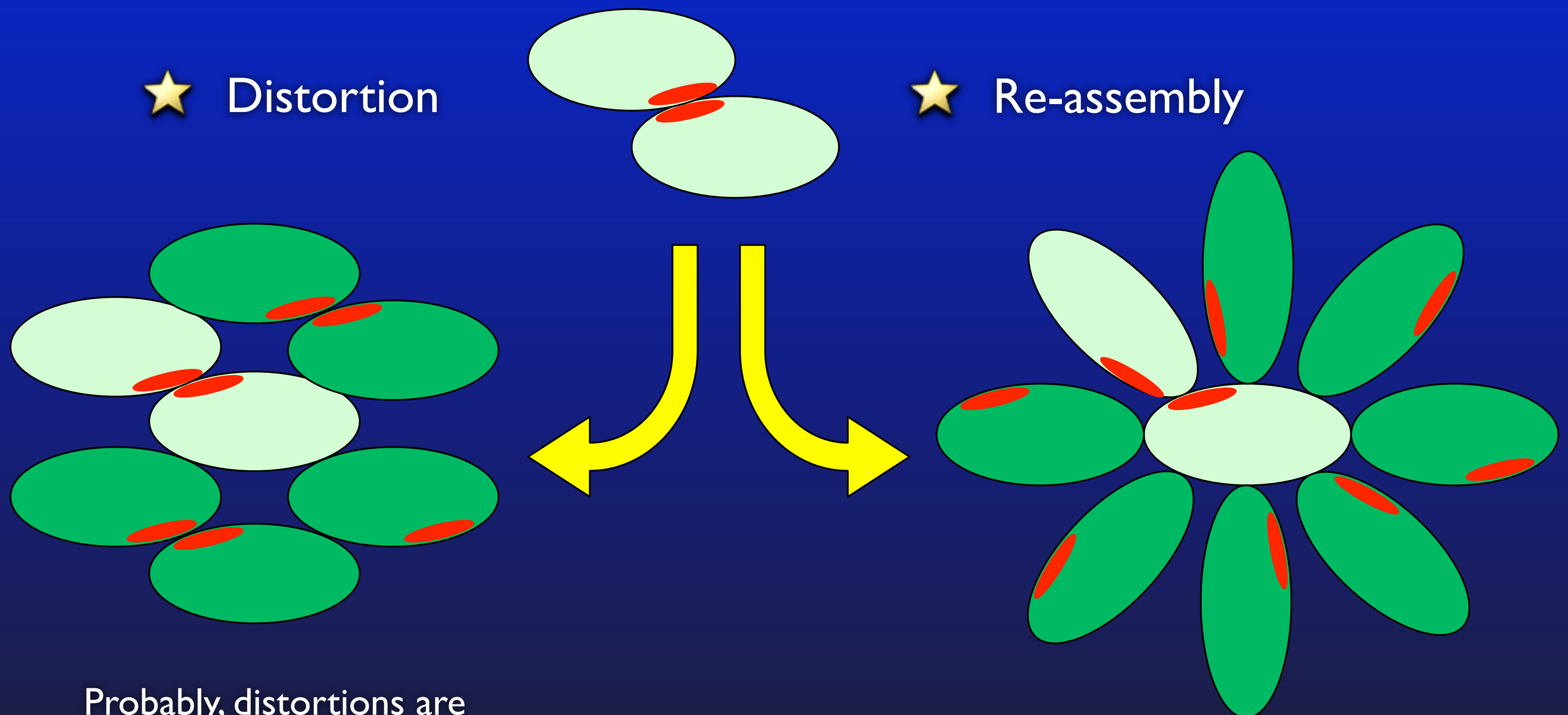
Probably correct



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Distortions and Re-assembly

- ★ Crystal optimizes energy globally, therefore it may sacrifice biologically relevant interaction in favour of unspecific crystal contacts



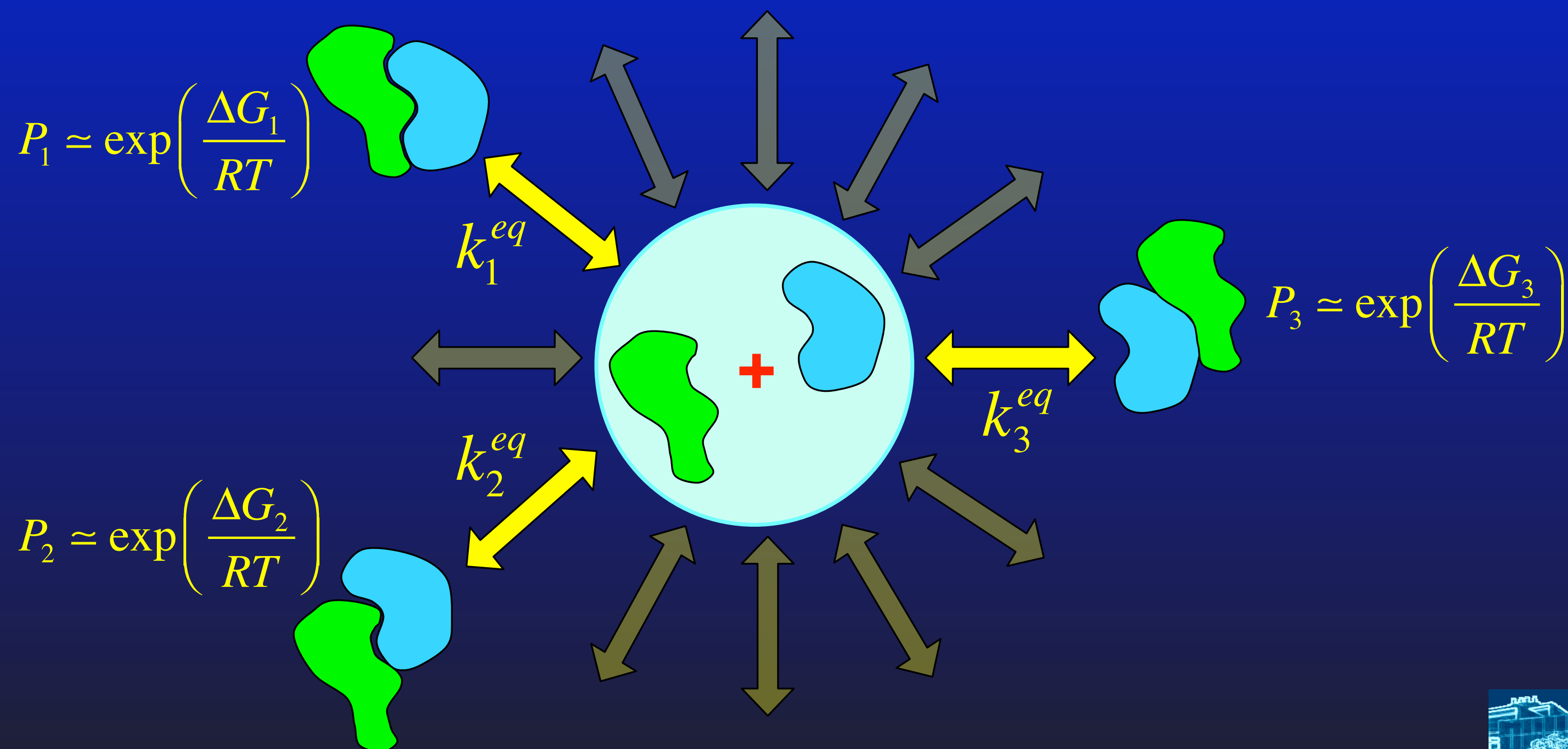
Probably, distortions are always there

There is a chance for re-assembly if interaction is weak



Alternative assemblies

- ★ All complexes (assemblies) have right to exist in solvent, however with different occurrence probabilities. These probabilities may differ of those in crystal environment, e.g., in case of substantially assisted crystallisation.



Real and superficial crystal contacts

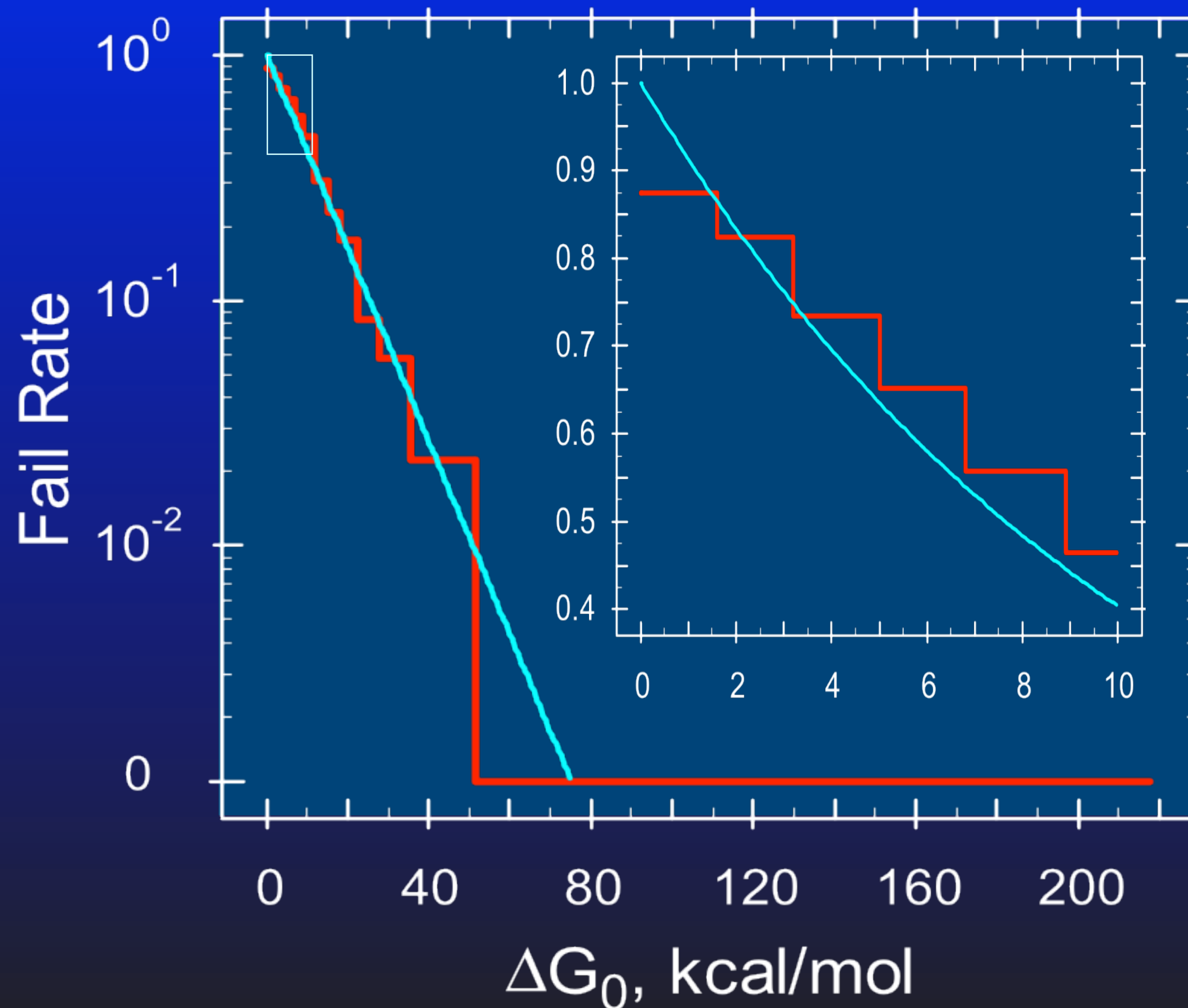
- ★ If a crystal contact remains thermodynamically preferential in solution, the chances are that it represents a biochemically relevant interaction
- ★ Experimental data on structure of complexes in solution is rather sparse
- ★ One can hope to get some clues using computational docking, assuming that docking approximates in-solvent situation
- ★ Being applied to 4065 non-redundant dimers from the PDB, docking **fails** to arrive at crystal interface in **38%** of instances

E. Krissinel (2010) J. Comp. Chem. 31, 133-143



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Fail rate of docking



The plot shows the probability of docking not to arrive at crystal interface, as a function of interface free energy.

The probabilities are calculated using equipopulated bins.

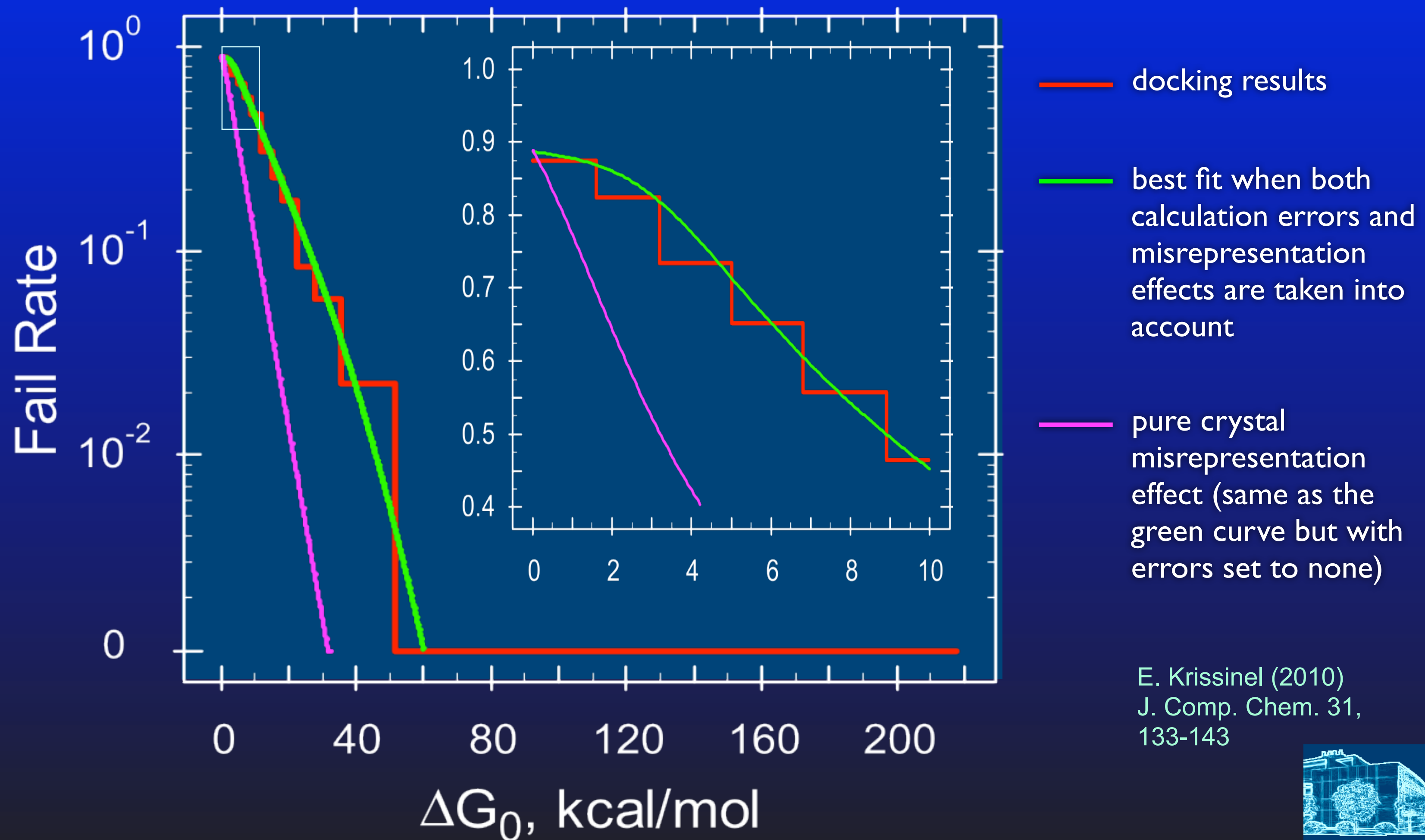
Overall, 38% of failures.

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Calculation errors and crystal misrepresentation effects



E. Krissinel (2010)
J. Comp. Chem. 31,
133-143



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So what is the practicality of all this?

- ★ PISA *is not* a substitution for experiments on the identification of protein's oligomeric state
 - both the software and (much less likely) experiment may give wrong results
 - but, in difference of experimental results, calculations do not make a scientific evidence!
- ★ PISA may be used for choosing complex models for molecular replacement
 - already done in CCP4's BALBES and MrBUMP automatic molecular replacement pipelines
- ★ PISA may be used for interpretation of experimental results when evidence is not sufficient for a definite answer
 - which dimer?
 - inconclusive evidence (e.g. oligomeric state highly dependent on concentration/temperature/ion presence etc.)
- ★ PISA may be used for sanity checks, comparative analysis and flag raising
 - is proposed complex structure compatible with crystal packing?
 - is proposed complex different from close homologs?
 - is there a strong disagreement with biological/biochemical expectations?



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Kim Henrick <i>European Bioinformatics Institute</i>	General introduction and PQS expertise
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CCP4 <i>Daresbury-York-Oxford</i>	Encouragement and publicity
~10,000 PISA users <i>Worldwide</i>	Using PISA and feedback
Biotechnology and Biological Sciences Research Council <i>(BBSRC) UK</i>	Research grant No. 721/B19544



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