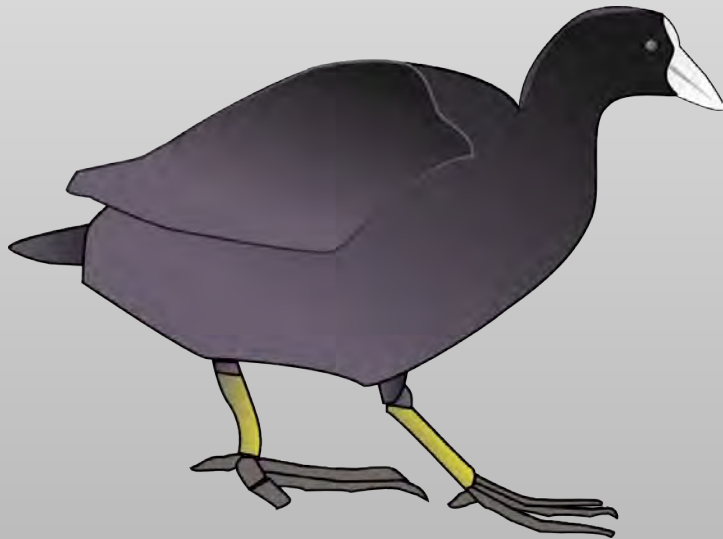


Ligands, Validation, ... and more model-building in *Coot*



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Bernhard.Lohkamp@ki.se

Tsukuba PF November 2014

Ligands in Coot

- Ligand fitting
- Importing and building ligand from scratch
 - PRODRG, LIBCHECK
- Representation
- Analysis
- Validation

Ligand building

Optimize with "ligand expert" options

Ligand Type

Known

Cocktail

Unknown

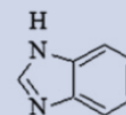
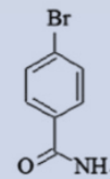
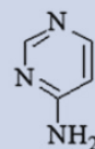
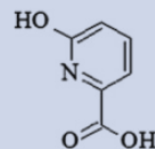
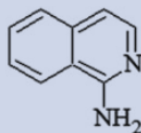
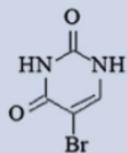
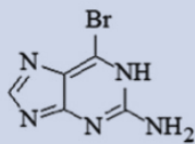
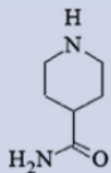
Ligand Site

Known

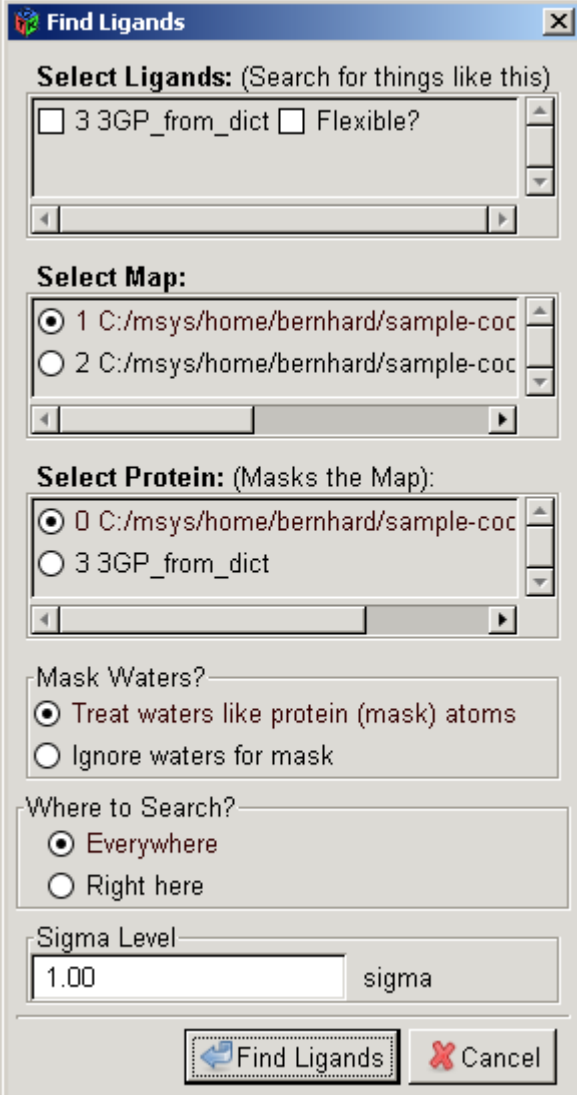
Unknown

✓	✓
✓	✓
✗	✗

Cocktail Examples



Ligand search in Coot



Find Ligands

Select Ligands: (Search for things like this)

☐ 3 3GP_from_dict ☐ Flexible?

Select Map:

☒ 1 C:/msys/home/bernhard/sample-coc
☐ 2 C:/msys/home/bernhard/sample-coc

Select Protein: (Masks the Map):

☒ 0 C:/msys/home/bernhard/sample-coc
☐ 3 3GP_from_dict

Mask Waters?

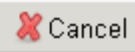
☒ Treat waters like protein (mask) atoms
☐ Ignore waters for mask

Where to Search?

☒ Everywhere
☐ Right here

Sigma Level

1.00 sigma

Ligand Fitting

- *c.f.* Oldfield (2001) *Acta Cryst. D* X-LIGAND
- Somewhat different torsion search algorithm
- Build in crystal-space

REFMAC Monomer Library

chem_comp_bond

loop_

_chem_comp_bond.comp_id

_chem_comp_bond.atom_id_1

_chem_comp_bond.atom_id_2

_chem_comp_bond.type

_chem_comp_bond.value_dist

_chem_comp_bond.value_dist_esd

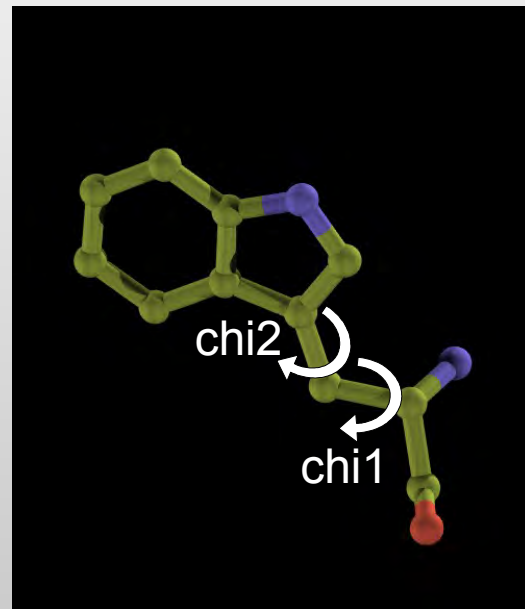
ALA	N	H	single	0.860	0.020
ALA	N	CA	single	1.458	0.019
ALA	CA	HA	single	0.980	0.020
ALA	CA	CB	single	1.521	0.033
ALA	CB	HB1	single	0.960	0.020
ALA	CB	HB2	single	0.960	0.020
ALA	CB	HB3	single	0.960	0.020
ALA	CA	C	single	1.525	0.021
ALA	C	O	double	1.231	0.020

REFMAC Monomer Library

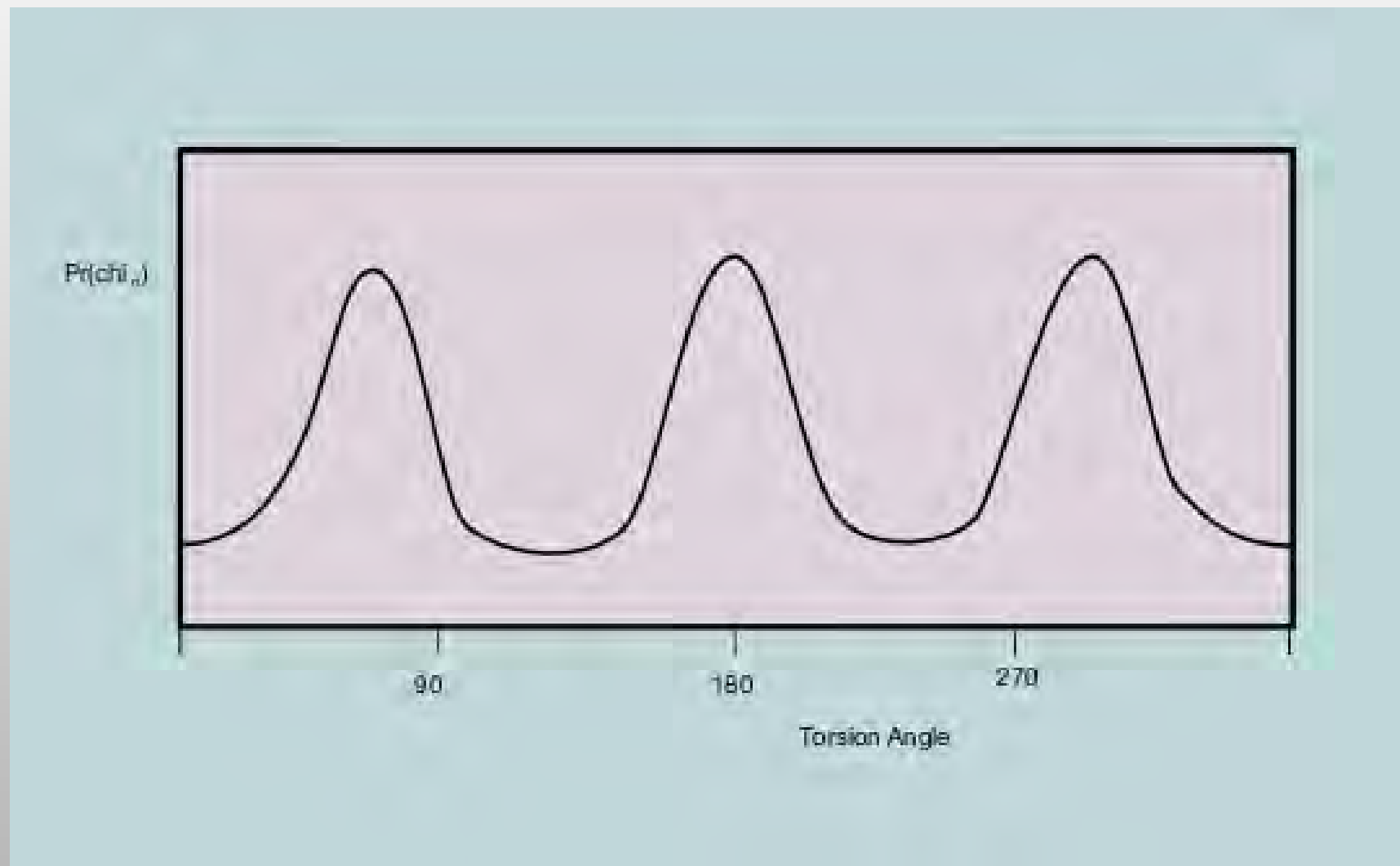
chem_comp_tor

```
loop_  
_chem_comp_tor.comp_id  
_chem_comp_tor.id  
_chem_comp_tor.atom_id_1  
_chem_comp_tor.atom_id_2  
_chem_comp_tor.atom_id_3  
_chem_comp_tor.atom_id_4  
_chem_comp_tor.value_angle  
_chem_comp_tor.value_angle_esd  
_chem_comp_tor.period
```

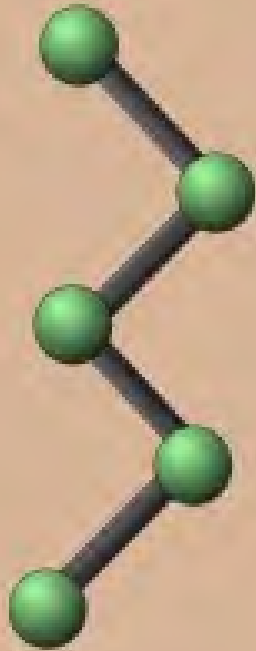
TRP	chi1	N	CA	CB	CG	180.000	15.000	3
TRP	chi2	CA	CB	CG	CD1	90.000	20.000	2



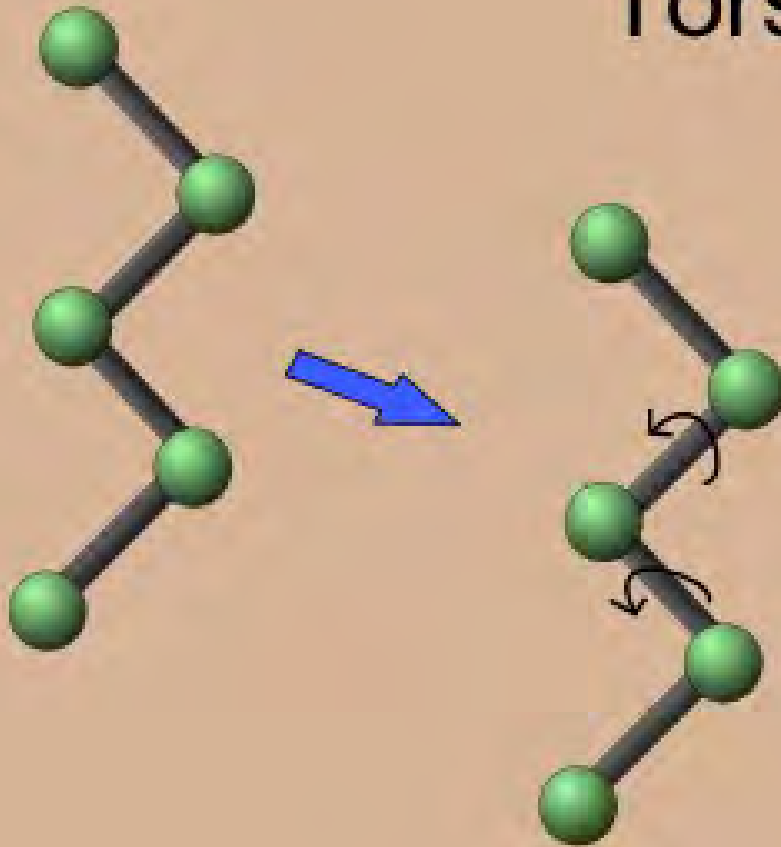
Ligand Torsionable Angle Probability from CIF file



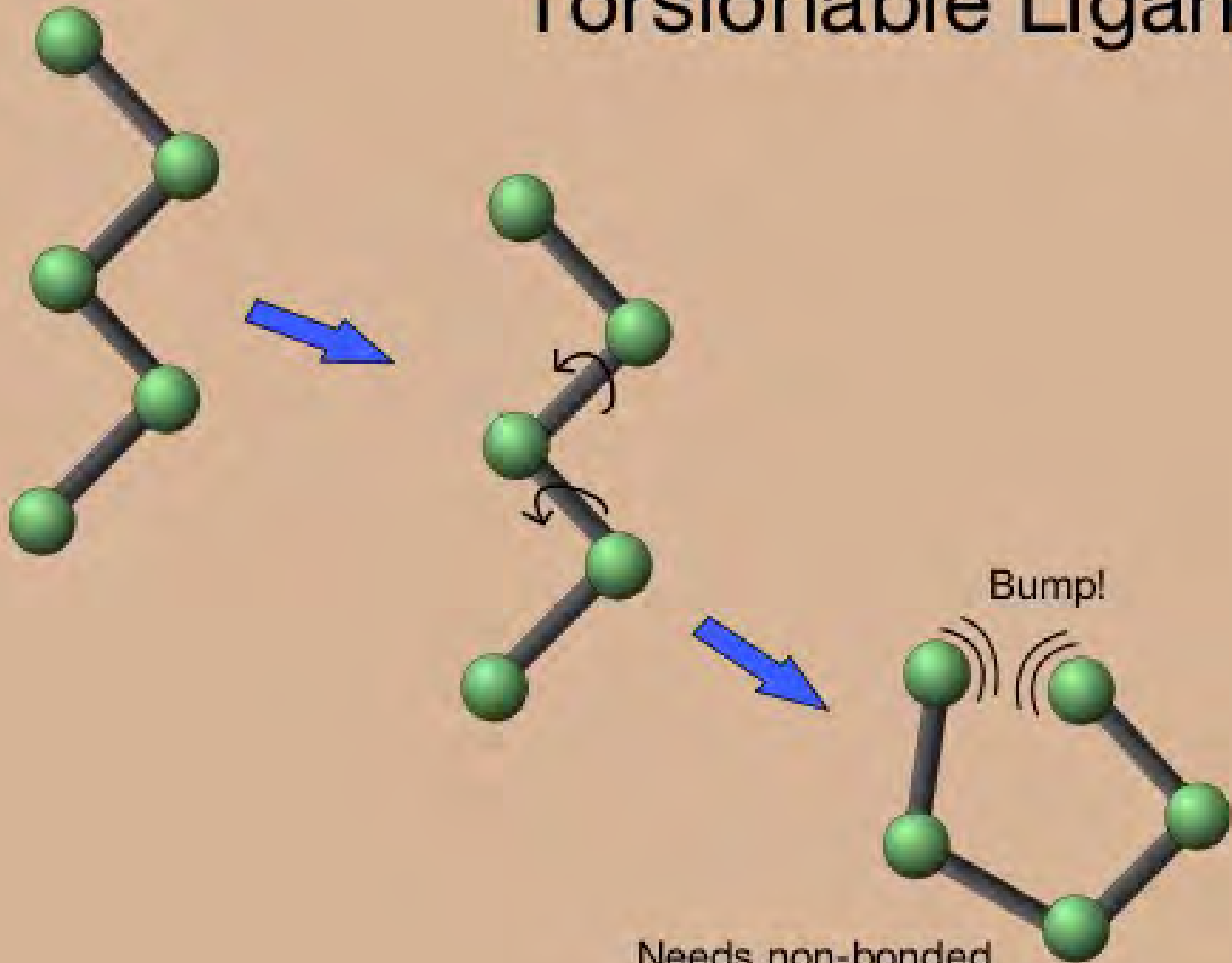
Torsionable Ligands



Torsionable Ligands

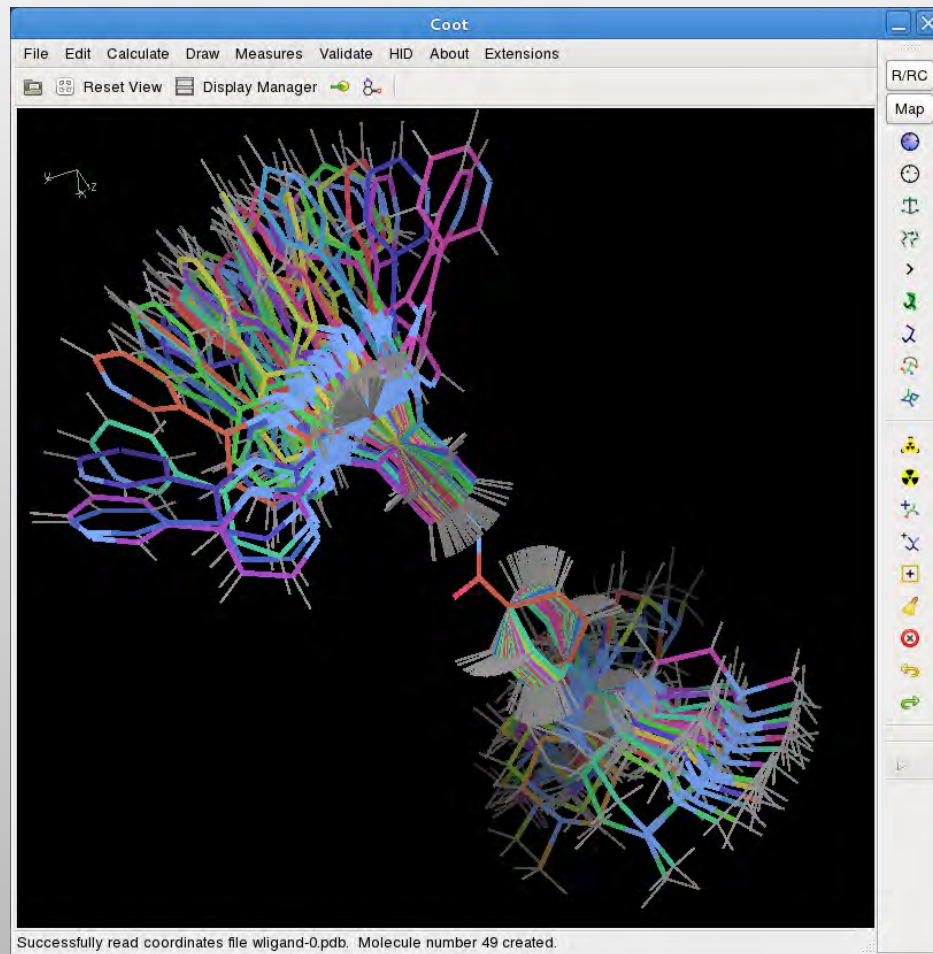


Torsionable Ligands



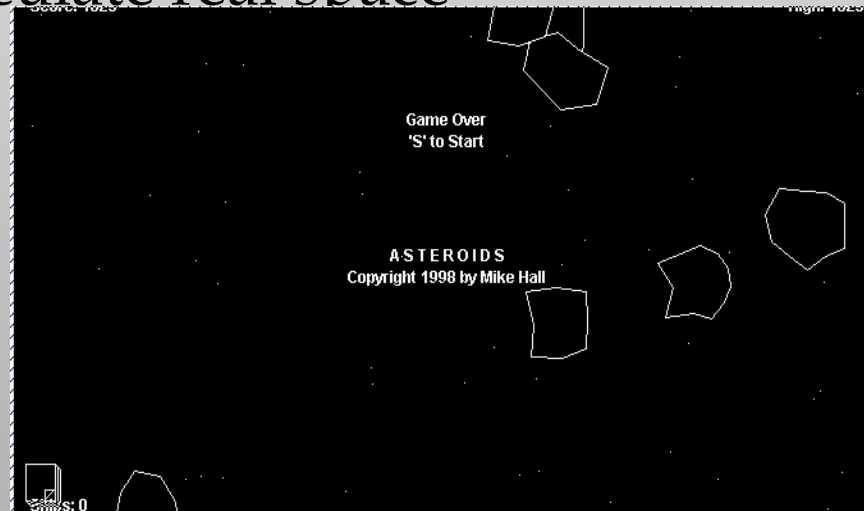
Needs non-bonded
contact idealization

Conformer Generation

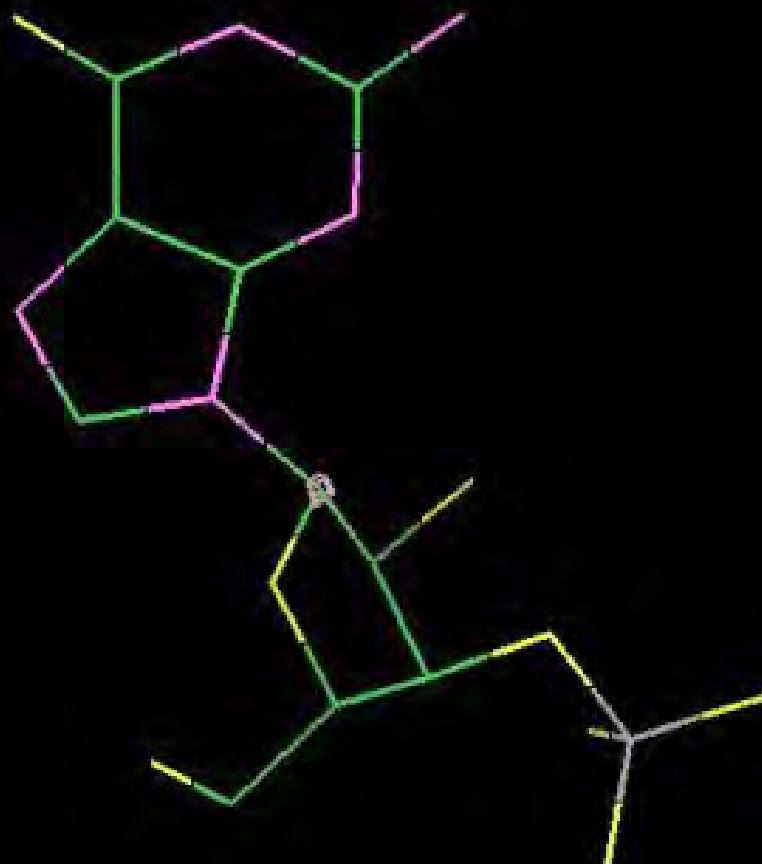


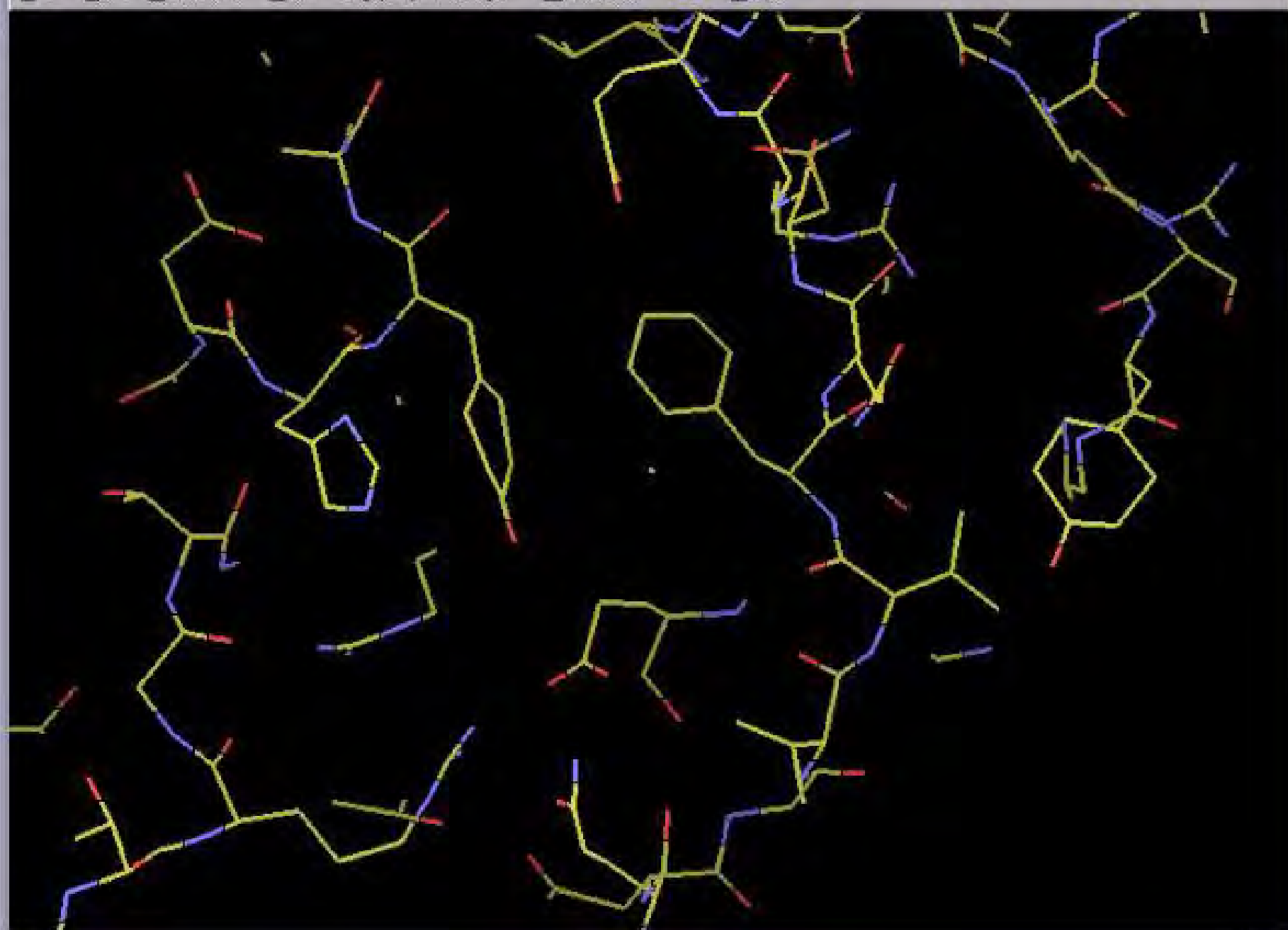
Crystal Space

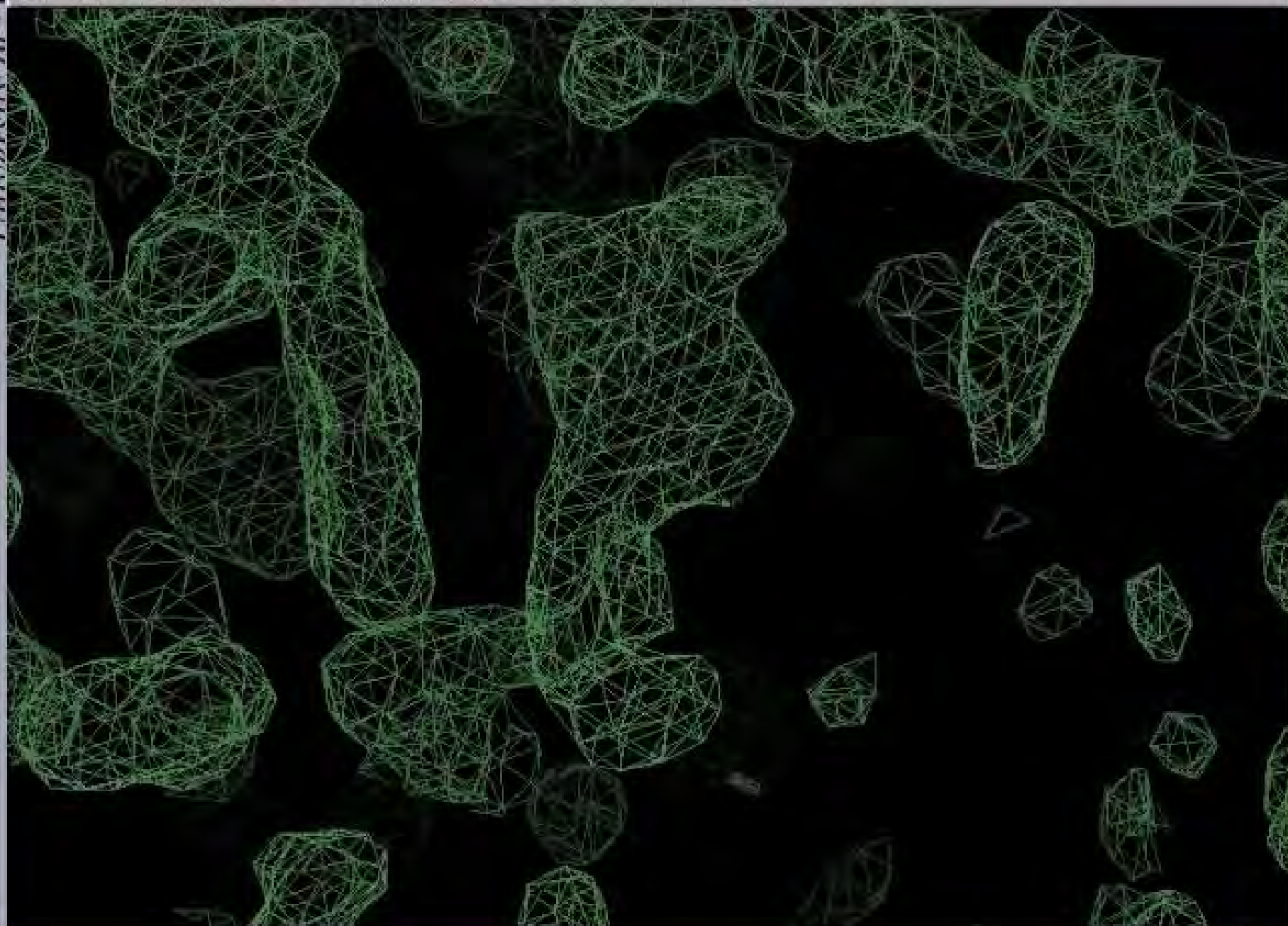
- Build in “crystal space”
 - Like real-space, but wrapped by crystal symmetry
 - Like “Asteroids”
 - Assures only one real-space representation of map features
 - Build everything only once
 - No symmetry clashing
 - However, more difficult to calculate real space geometries
 - ...such as bonds, torsions

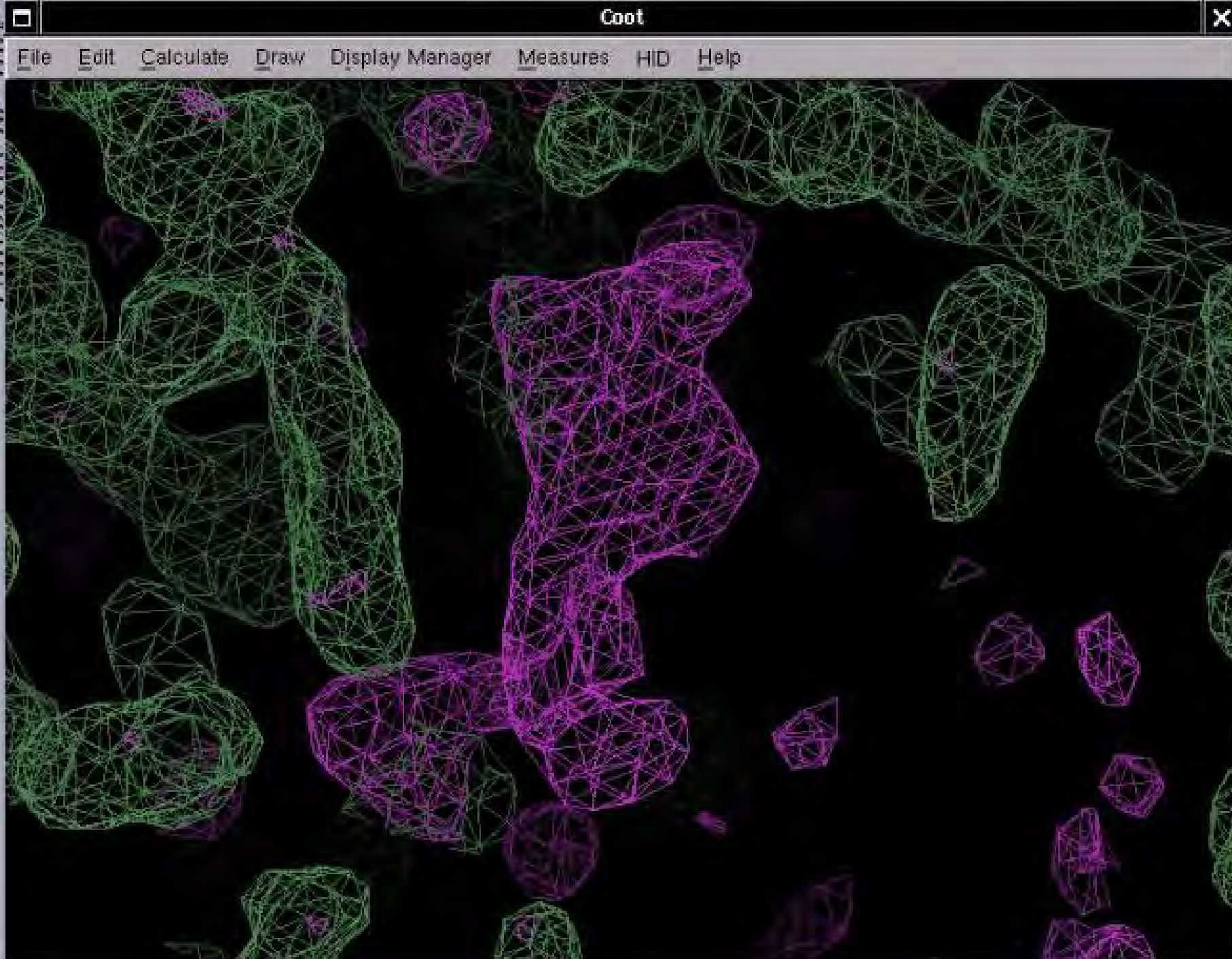


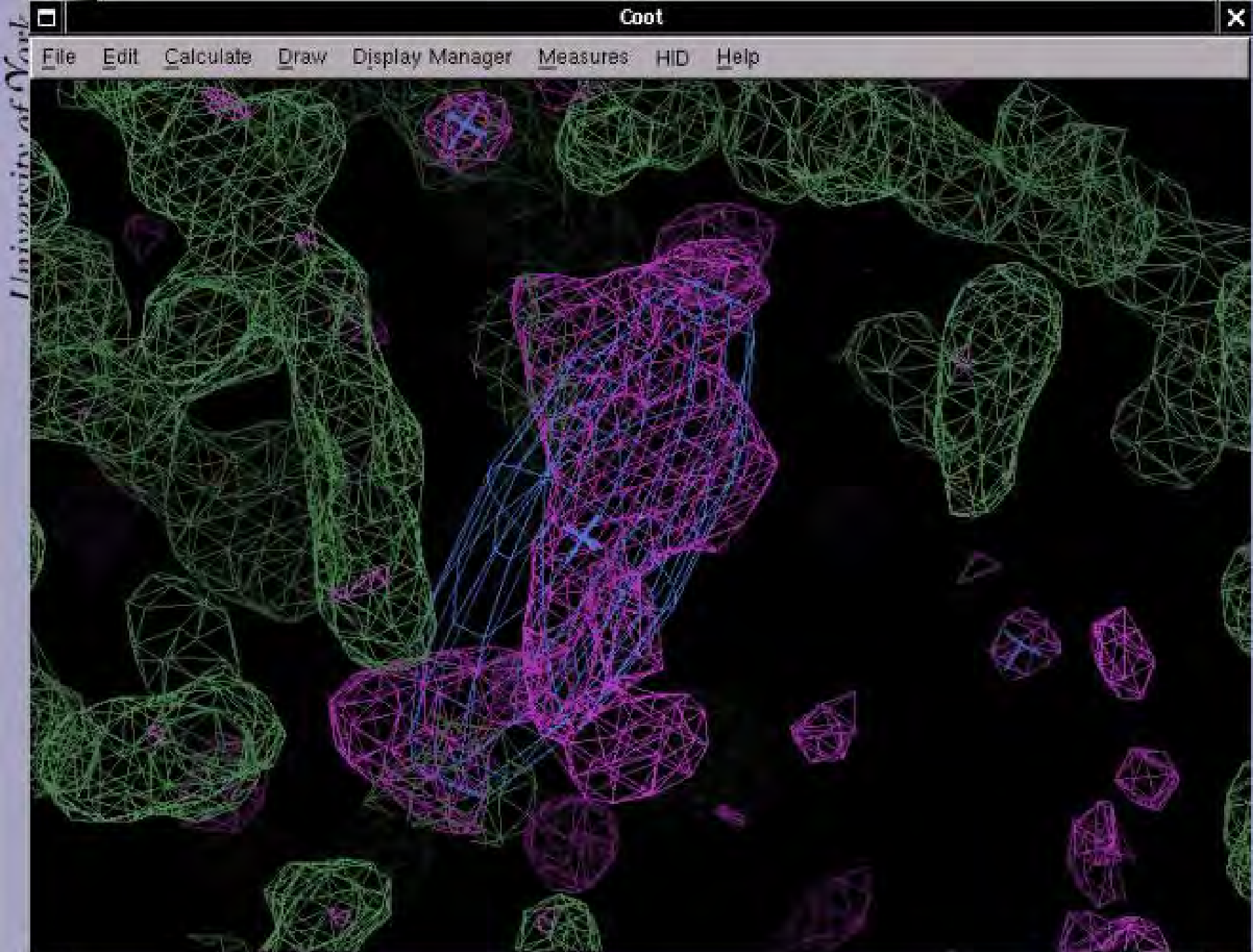
File Edit Calculate Draw Display Manager Measures HID Help

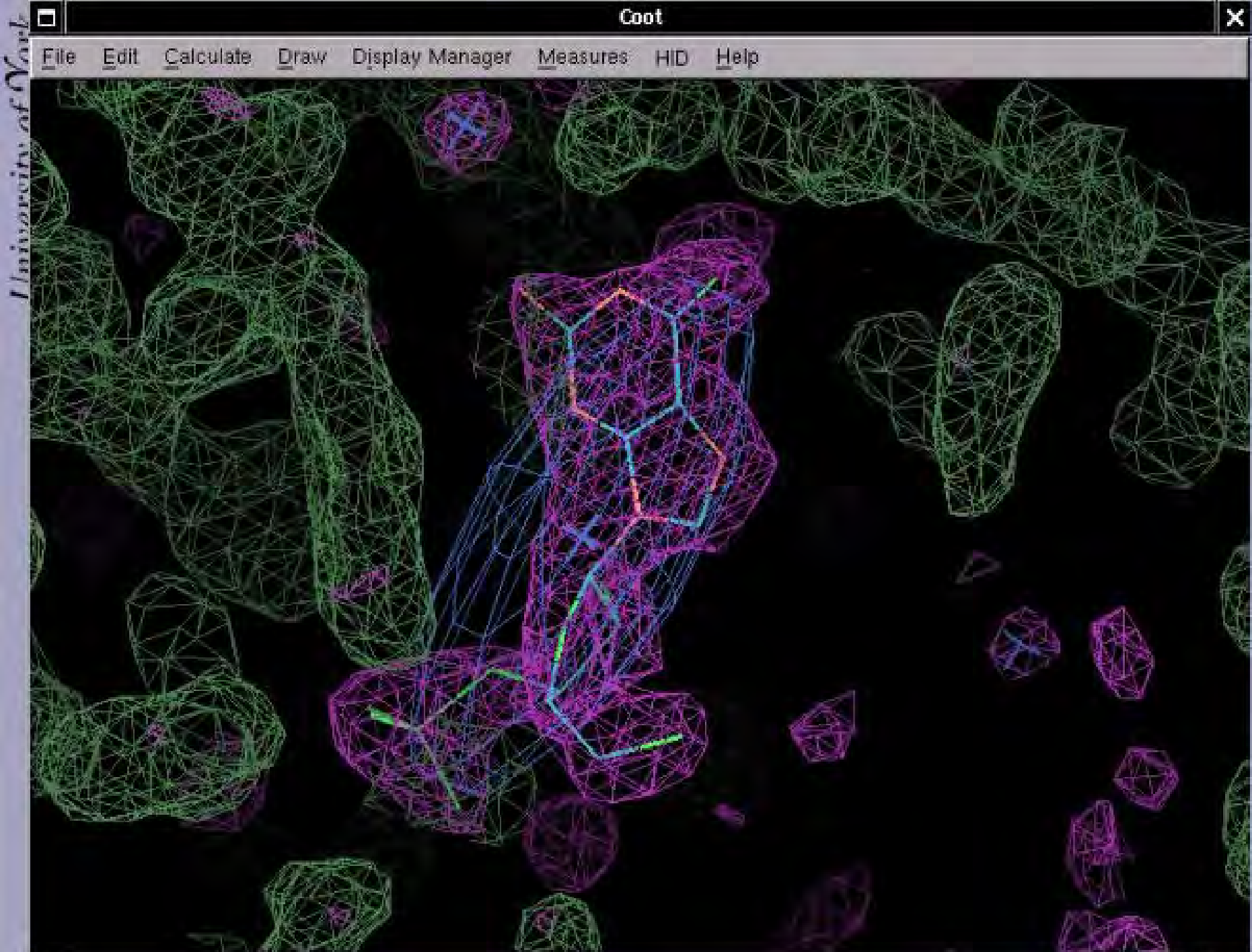


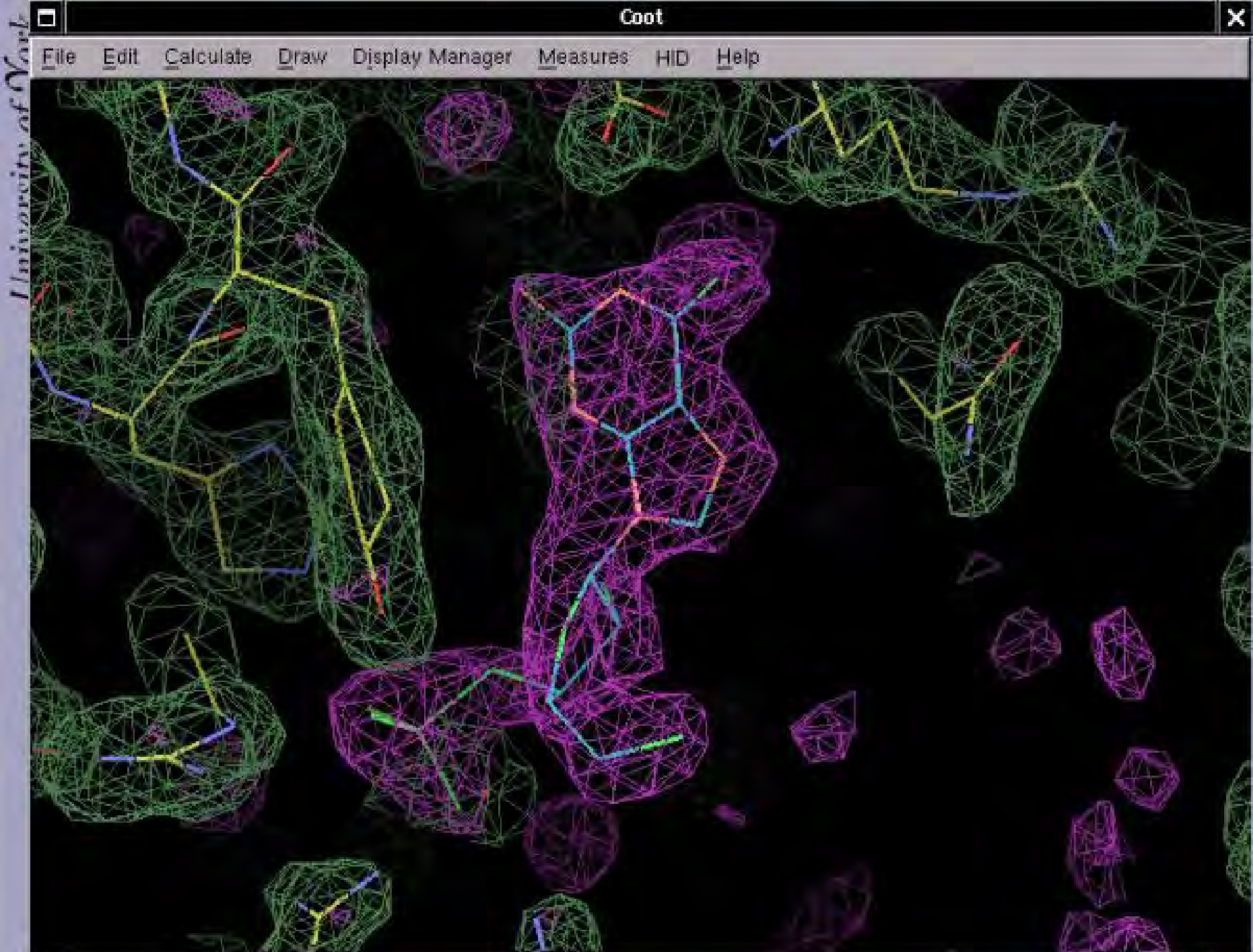










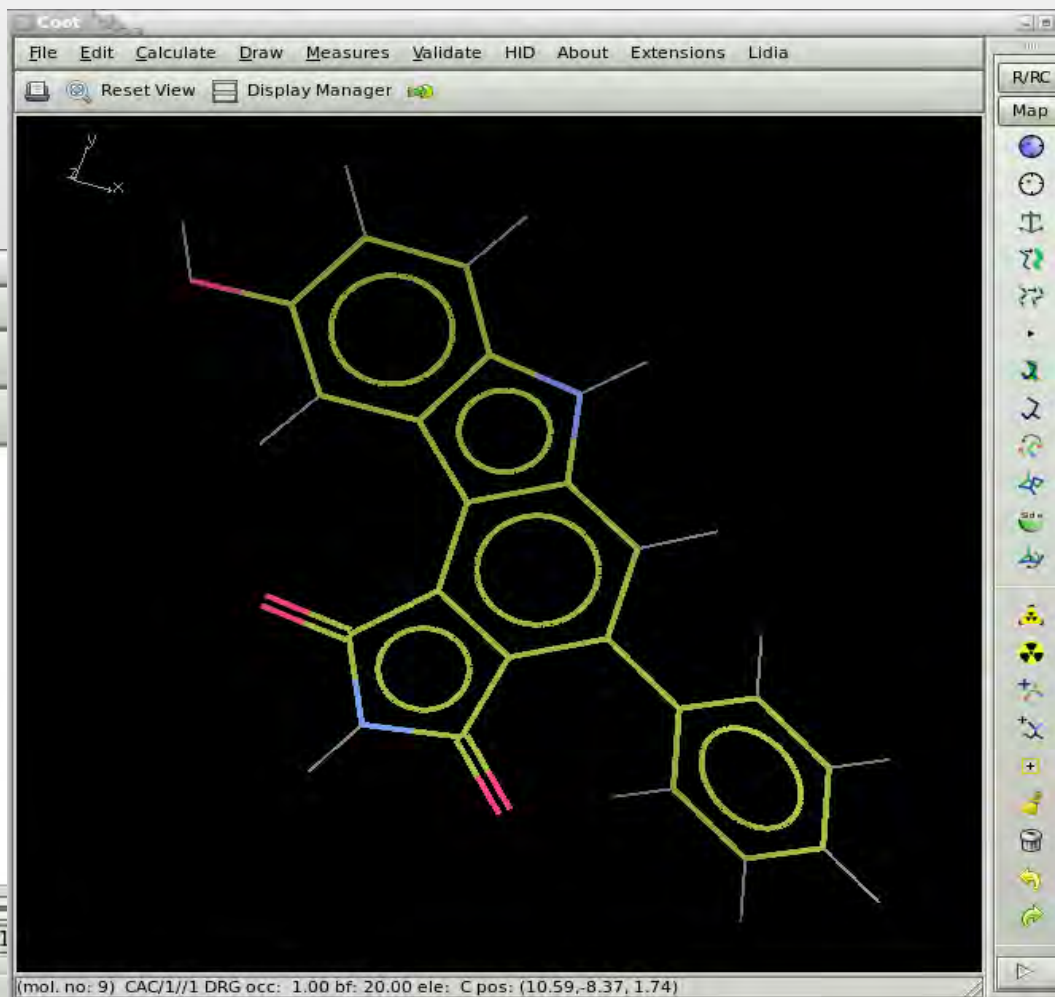
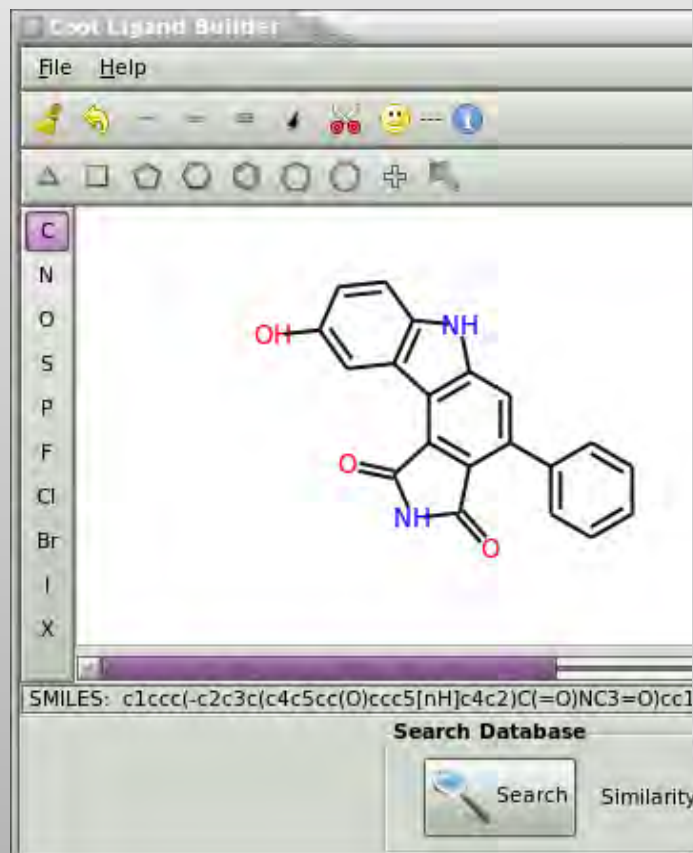


Conformation Idealization

- Each conformer is passed through the “Regularization” function of Coot
 - Non-bonded terms included
- Better to have hydrogen atoms on the model
- Slows things down a good deal...
 - May not be the best method to explore conformational variability for many rotatable bonds

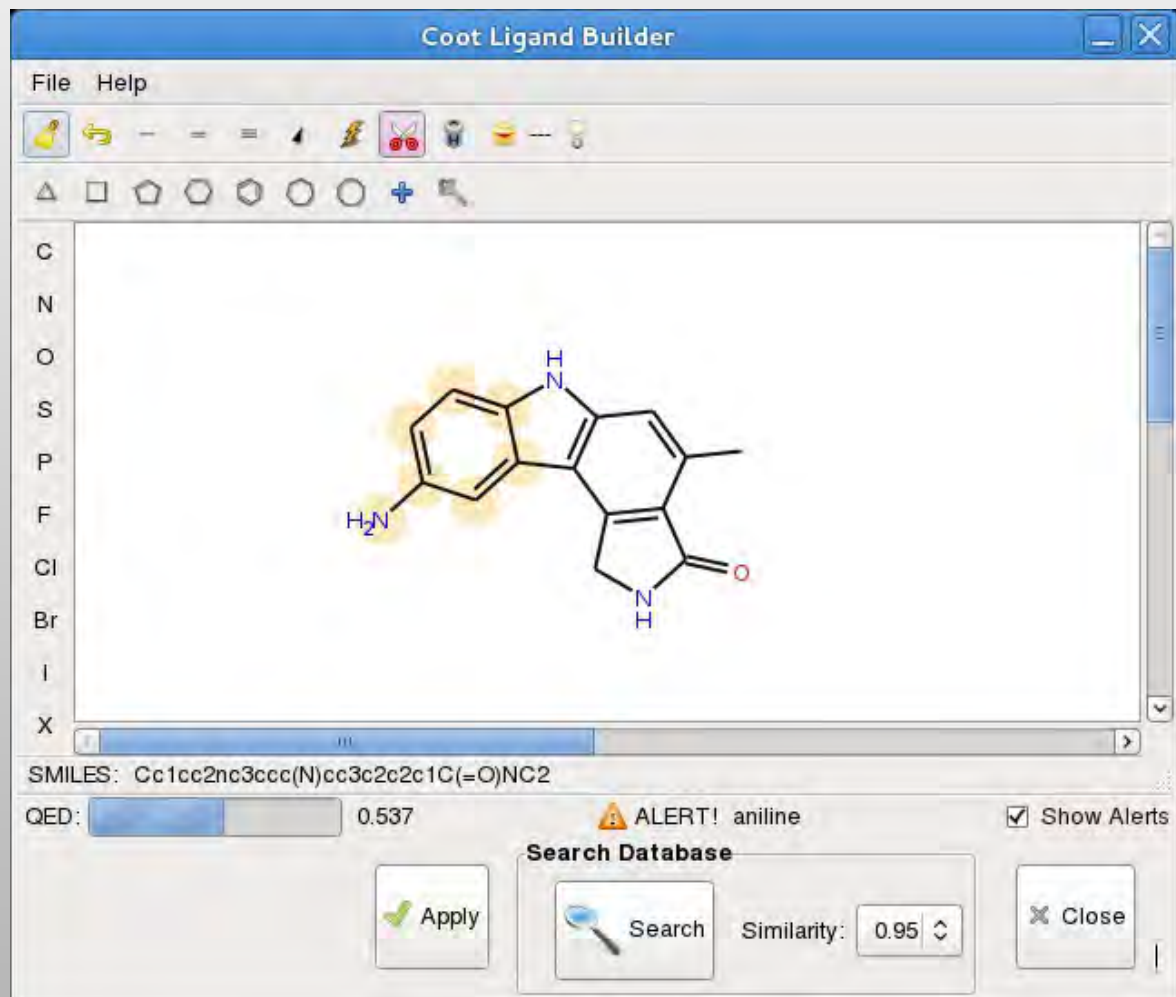
2D Ligand Builder

- Free sketch
- PRODRG



2D Sketcher

- Structural Alerts



200 built-in alerts
with the ability to
any additions

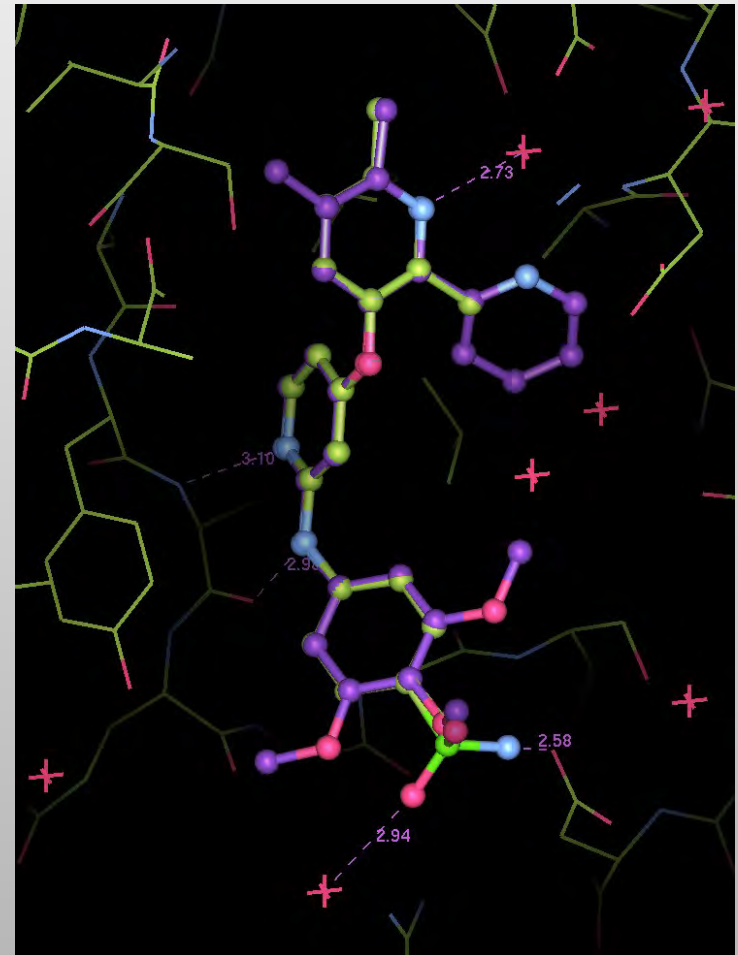
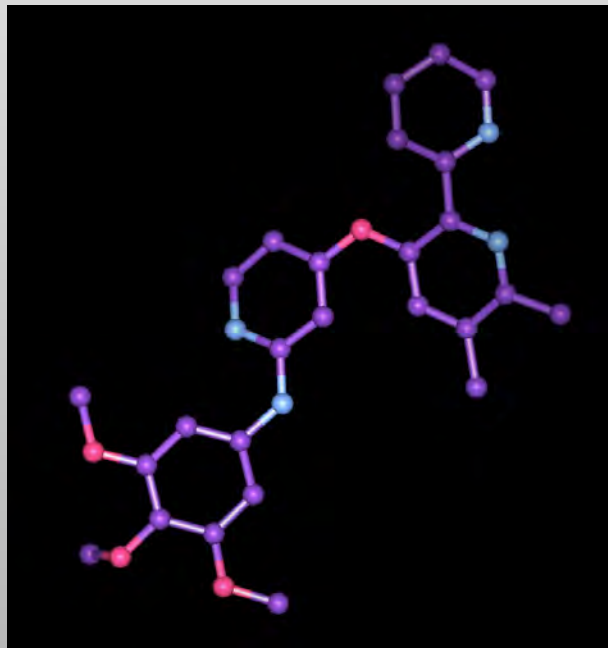
Ligand Overlay

- Algorithm and Code by Eugene Krissinel
- Tries to overlay different ligands/monomers by graph matching
- Useful for “database” ligands where atom names are not selected by hand
- Has been used as the basis of the function which “mutates” residues to alternative monomer types
 - e.g. phosphorylation

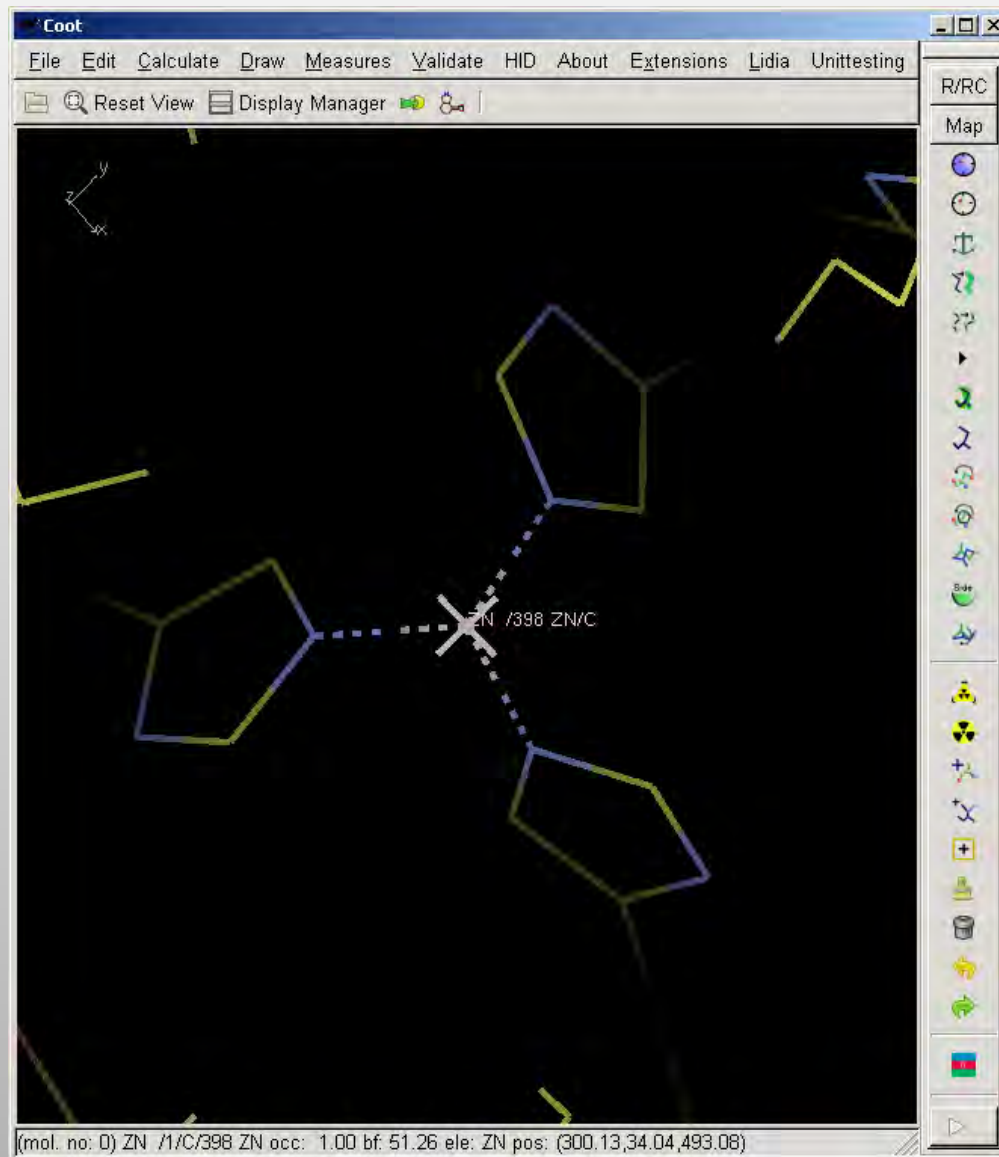
"Yesterday's" Ligand

Common subgraph isomorphism, Krissinel & Henrick (2004)

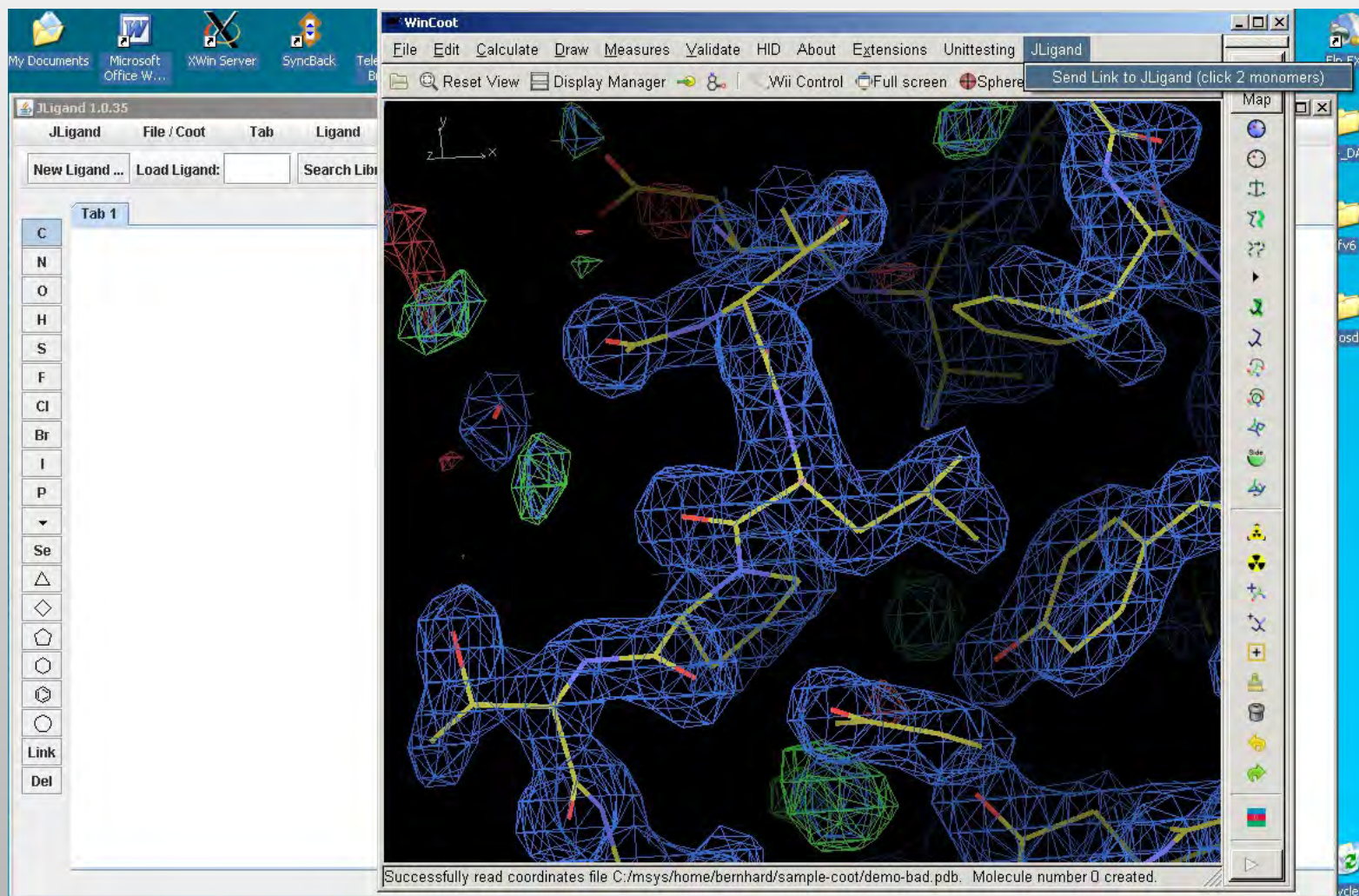
- Atom name matching
- Torsion matching
- Ligand overlay



Displaying LINKs



Interface with JLigand to create LINKs

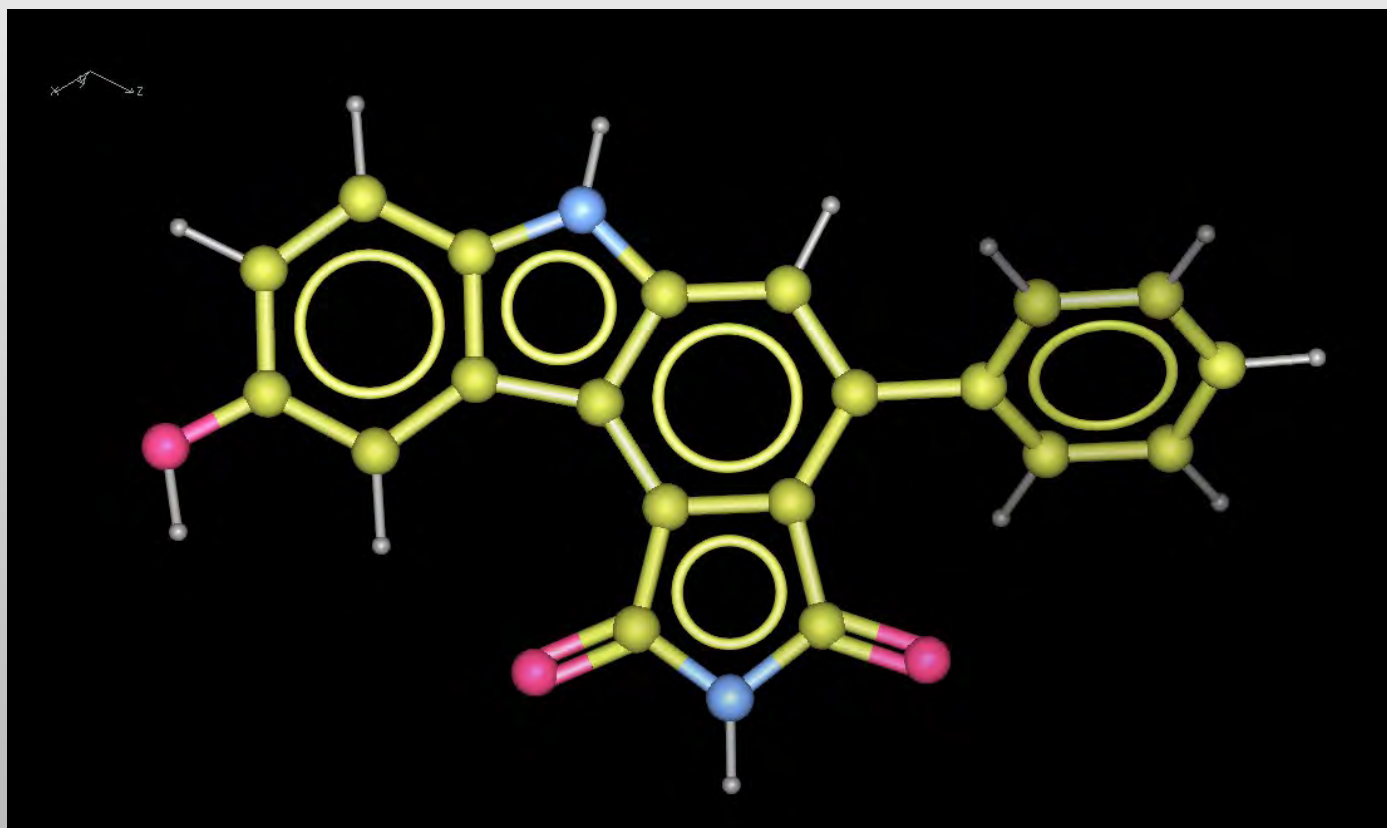


First: Extensions → Modelling → Launch JLigand

Ligand representation

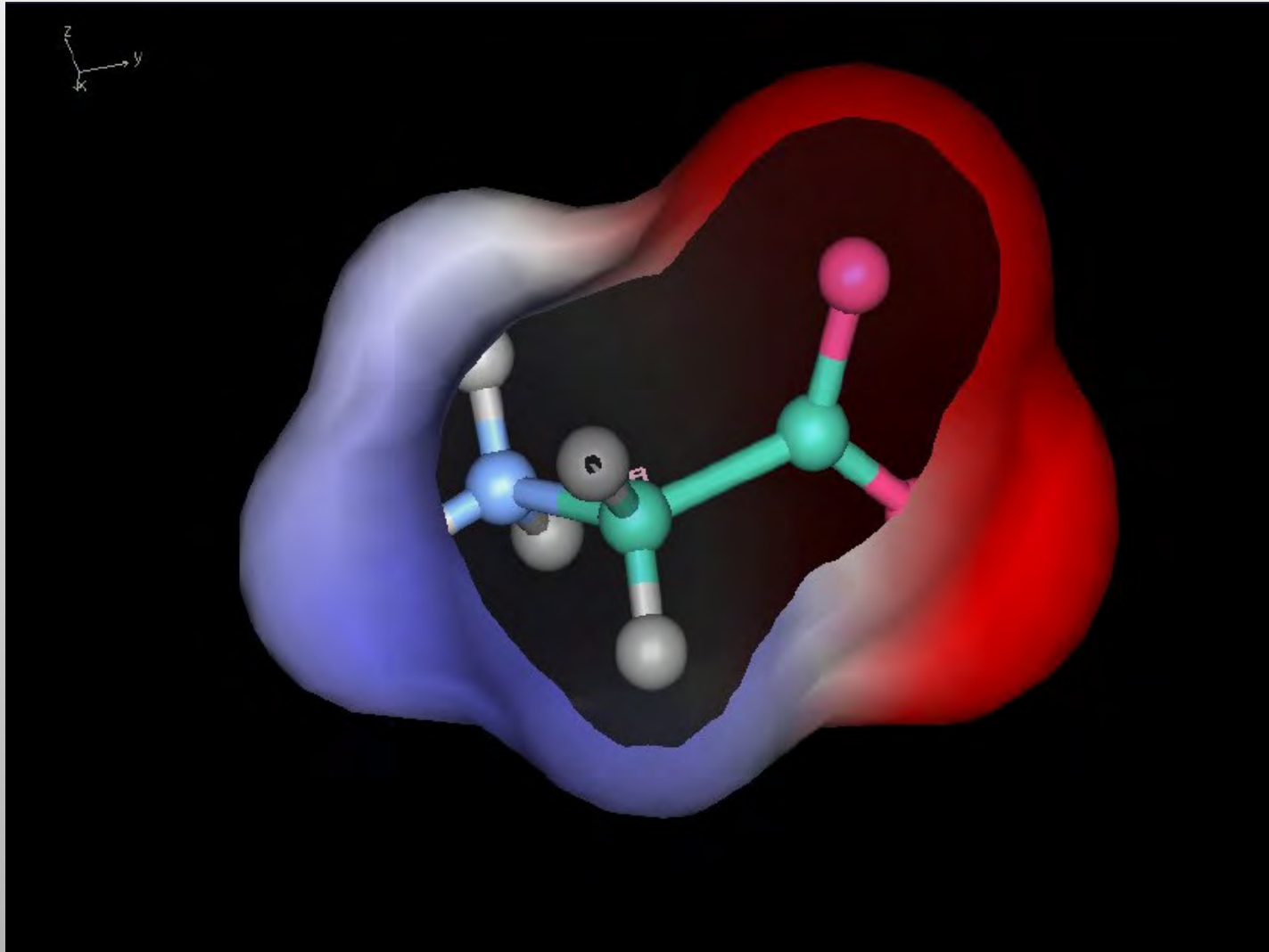
Ligand Representation

- Bond orders (from dictionary)



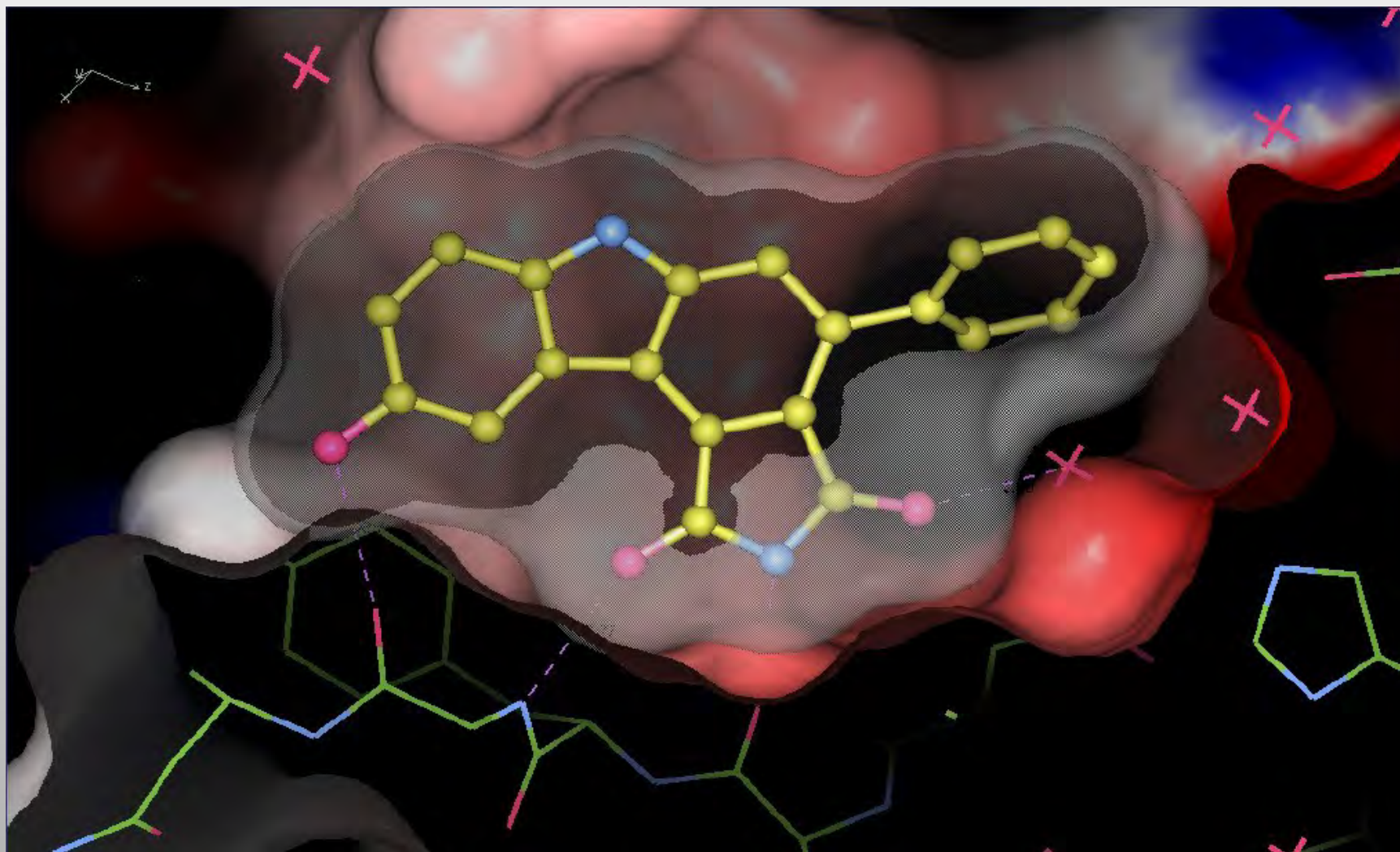
Surfaces

- Partial charges



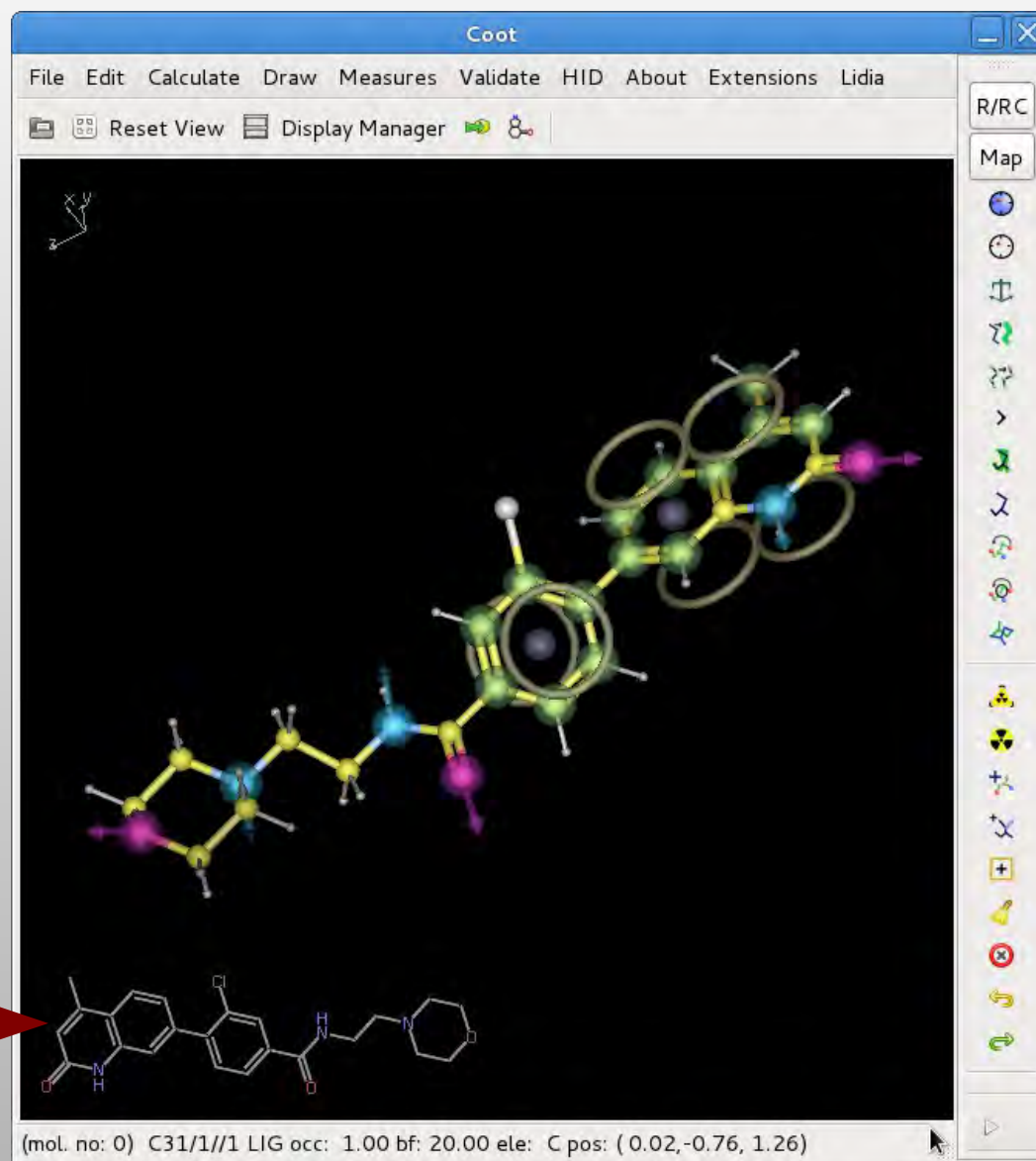
Surfaces

- Transparent surfaces – surface complementarity



Chemical Features

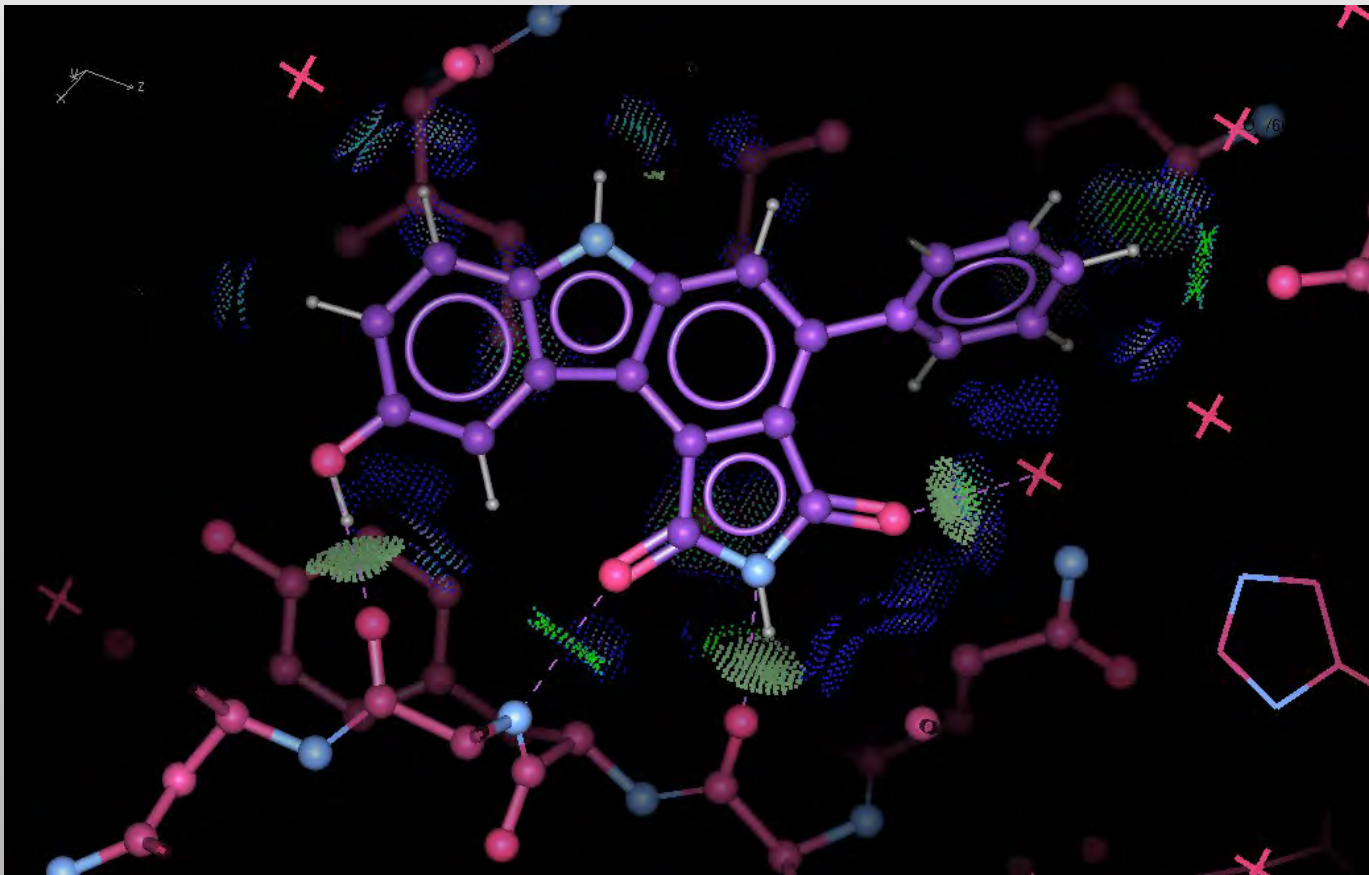
and chemical
diagram thumbnails



Ligand representation

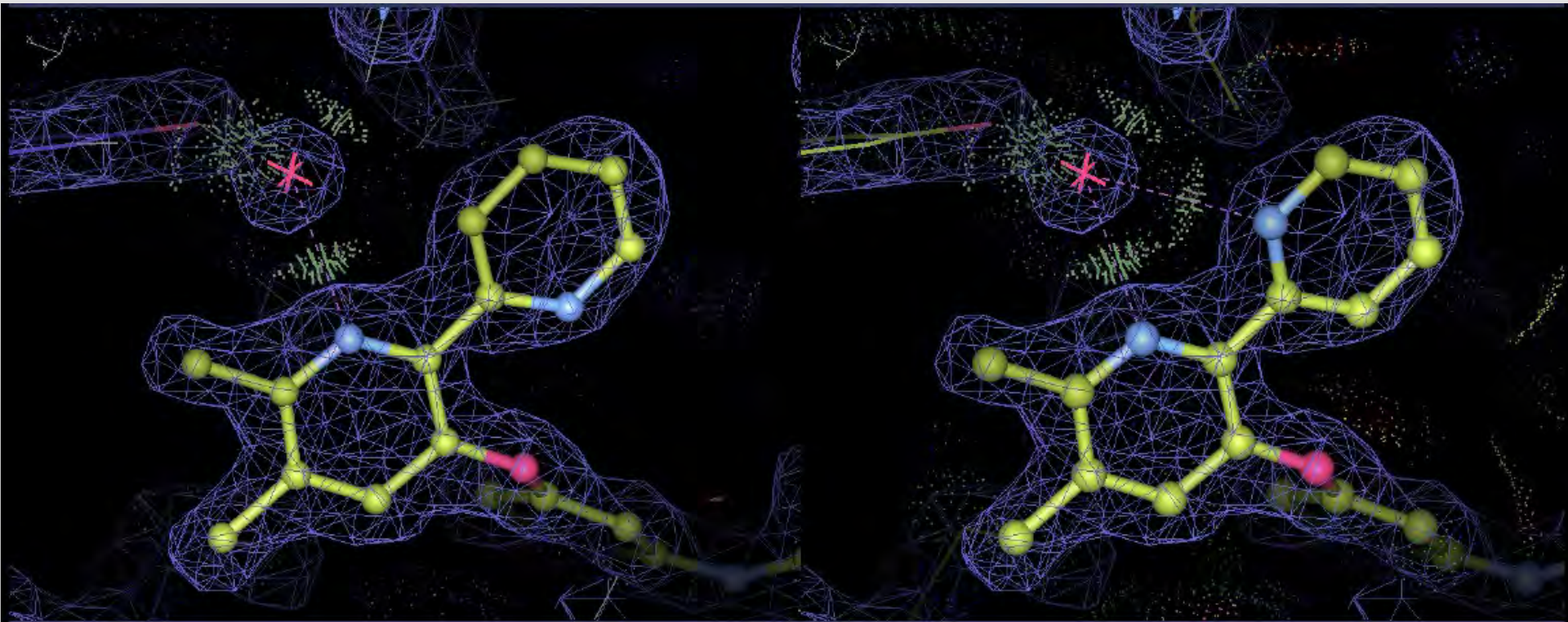
Binding mode analysis

- Binding site highlighting
- Isolated Molprobit probe dots



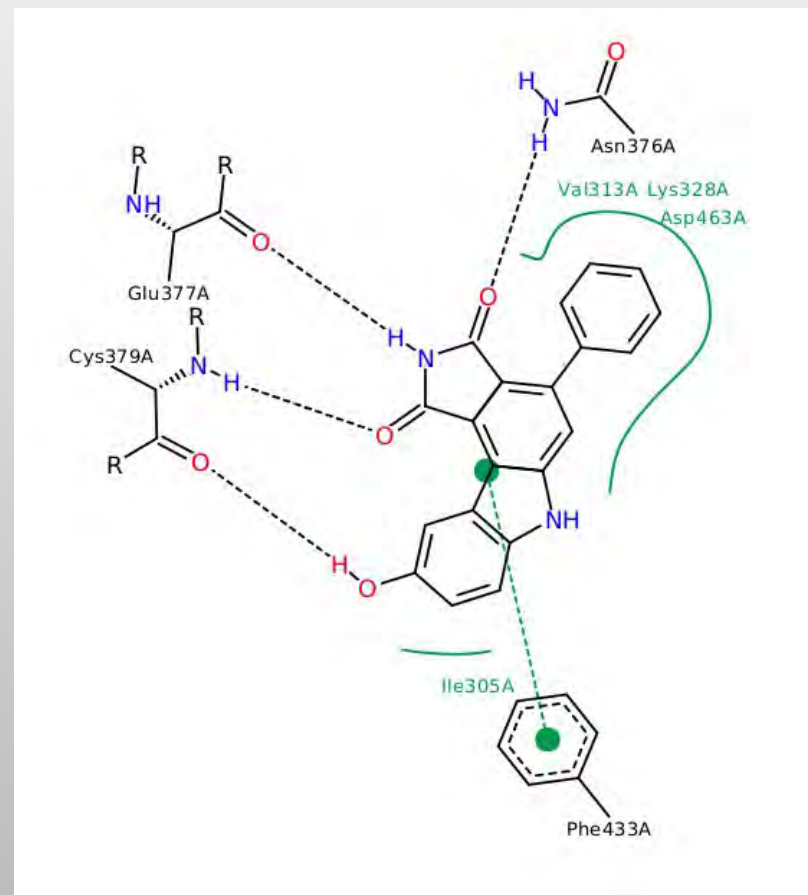
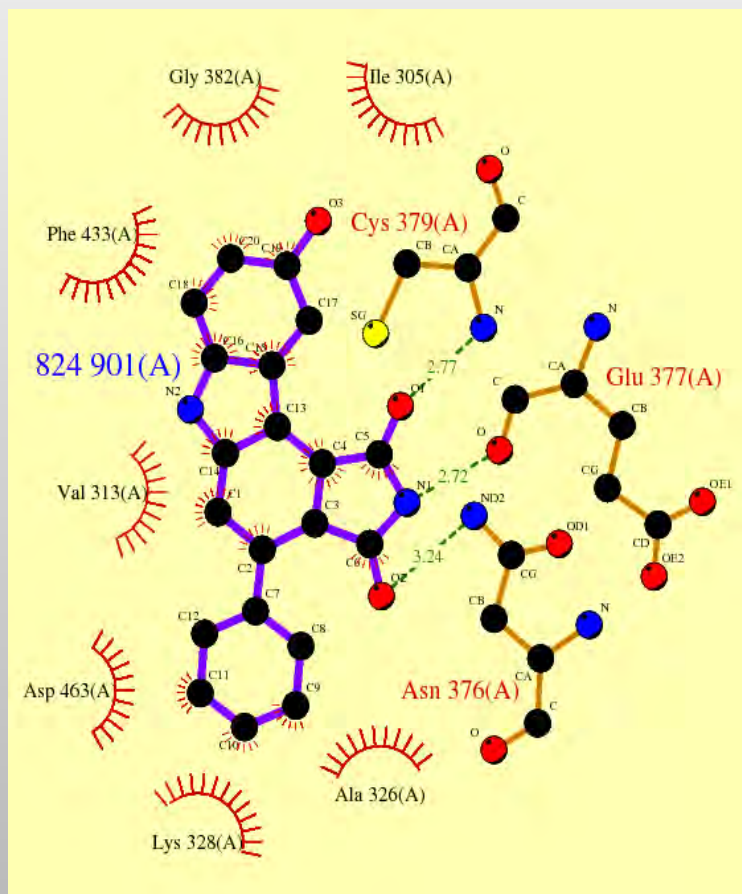
Binding mode analysis

- Binding site highlighting
- Isolated molprobability dots



Ligand Environment Layout

- 2d Ligand pocket layout (ligplot, poseview)



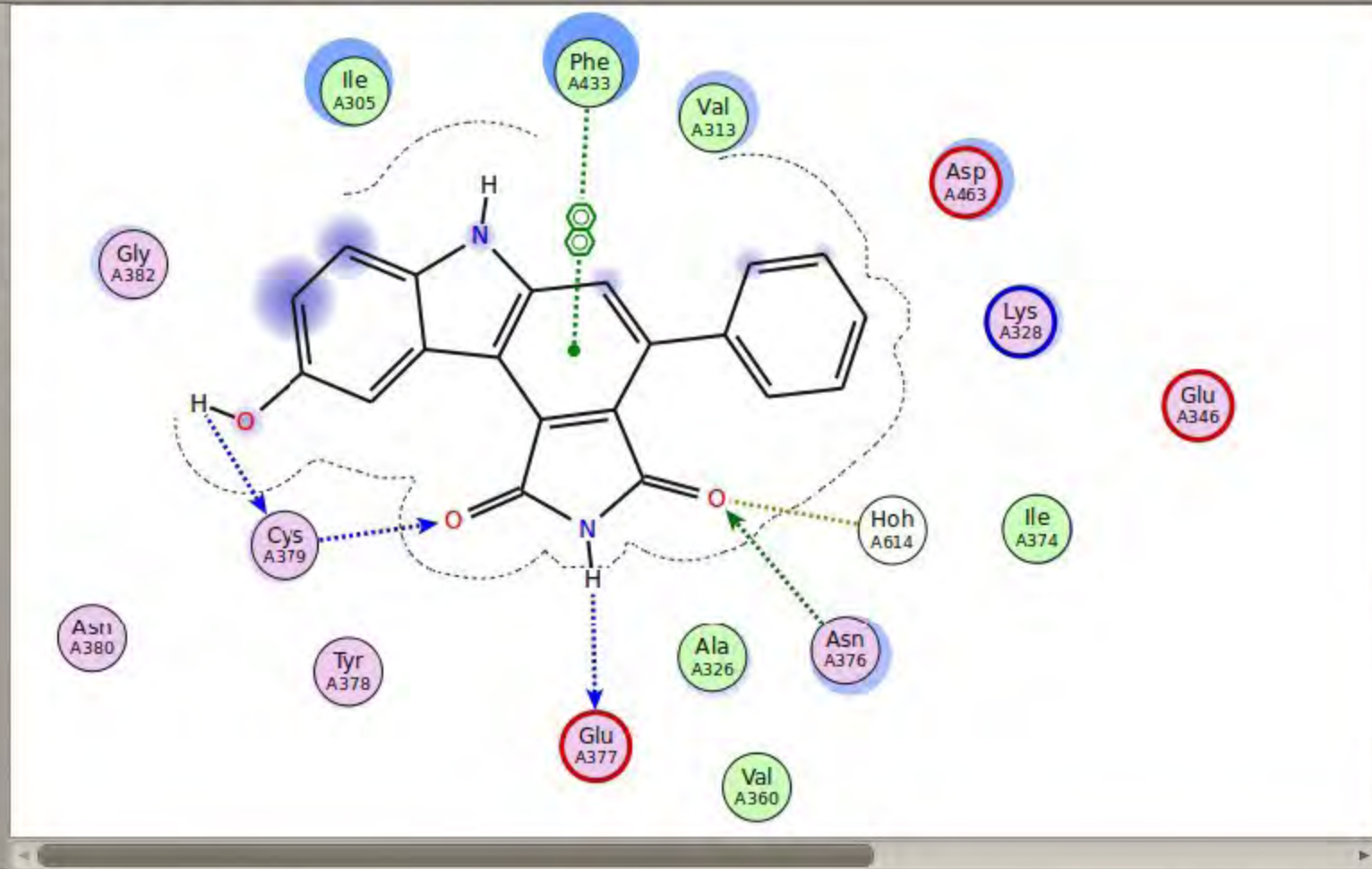
Can we do better? - Interactivity?

Ligand Environment Layout

- Binding pocket residues
- Interactions
- Solvent accessibility
 - halos
- Solvent exclusion by ligand
- Substitution contour

Ligand Environment Layout

- Considerations
 - 2D placement and distances should reflect 3D metrics (as much as possible)
 - H-bonded residues should be close the atoms to which they are bonded
 - Residues should not overlap the ligand
 - Residues should not overlap each other
 - *c.f.* Clark & Labute (2007)
(work in progress)

C
N
O
S
P
F
Cl
Br
I
X

Search Database



Search

Similarity:

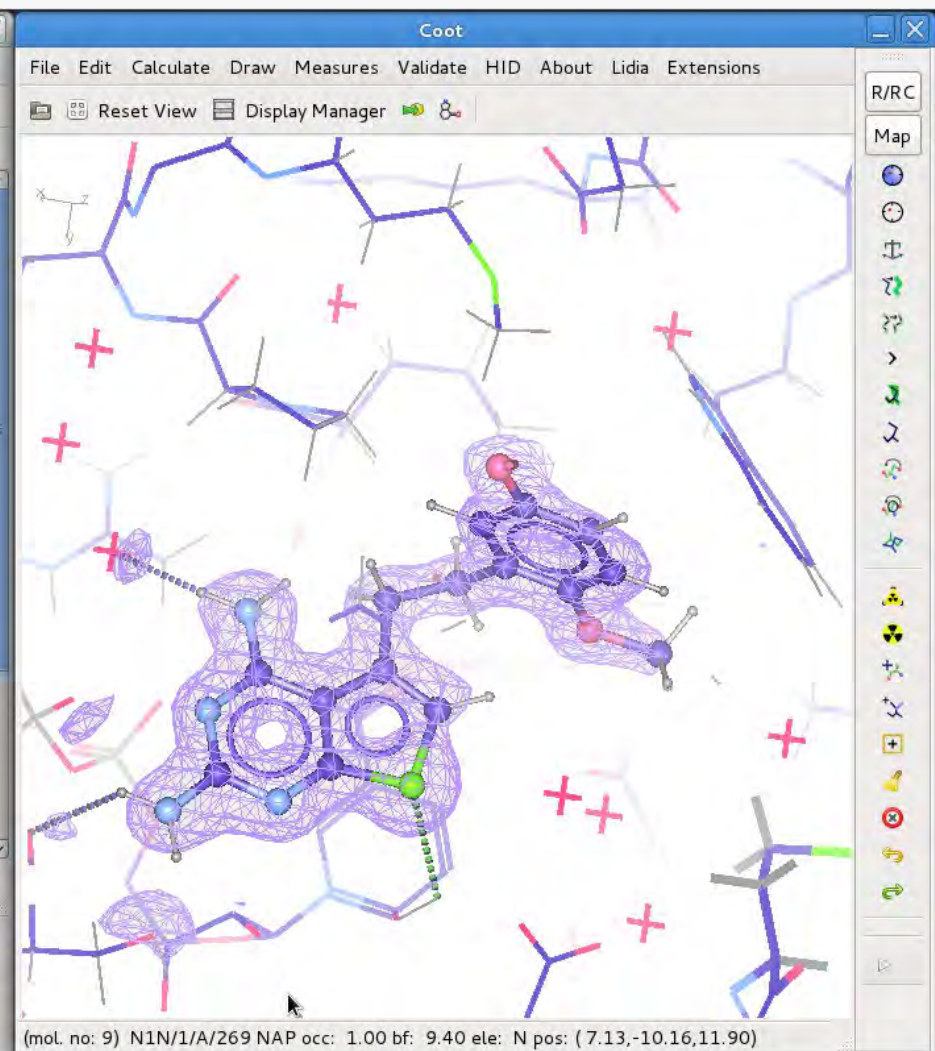
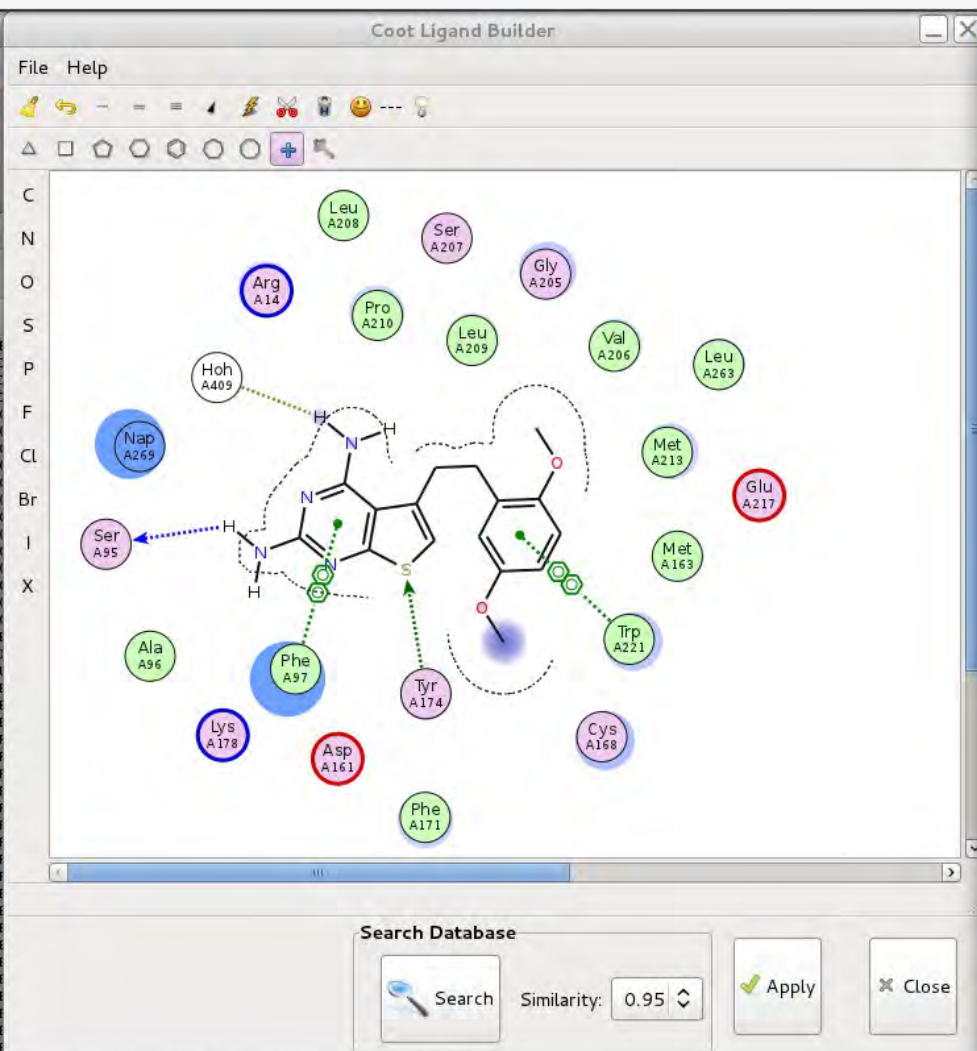
0.75



Apply



Close



Ligand futures

- Ligand validation with CSD (Mogul or similar)
- Dynamic hydrogen bonding and non-bonding contacts
- pyrogen?!
- Ace-drg

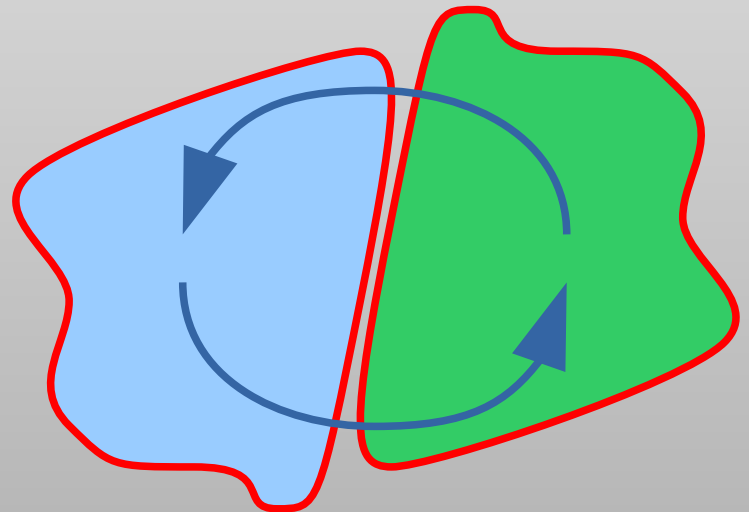
...and more?

- NCS
- Validation

Handling NCS...

What is Non-Crystallographic Symmetry (NCS)?

- 2 or more copies of a molecule in the unit cell not related by crystallographic symmetry
- NCS related molecules provide different representations of the same molecule

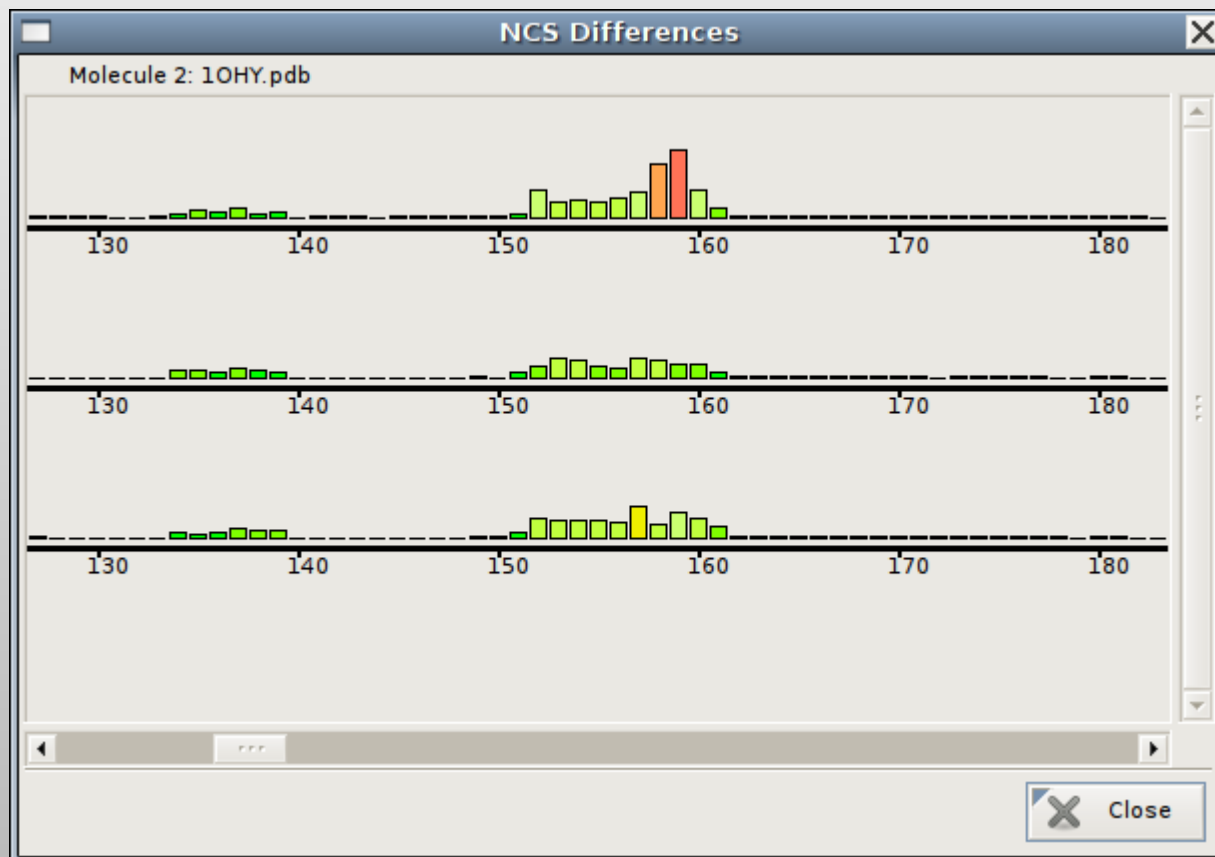


Handling NCS

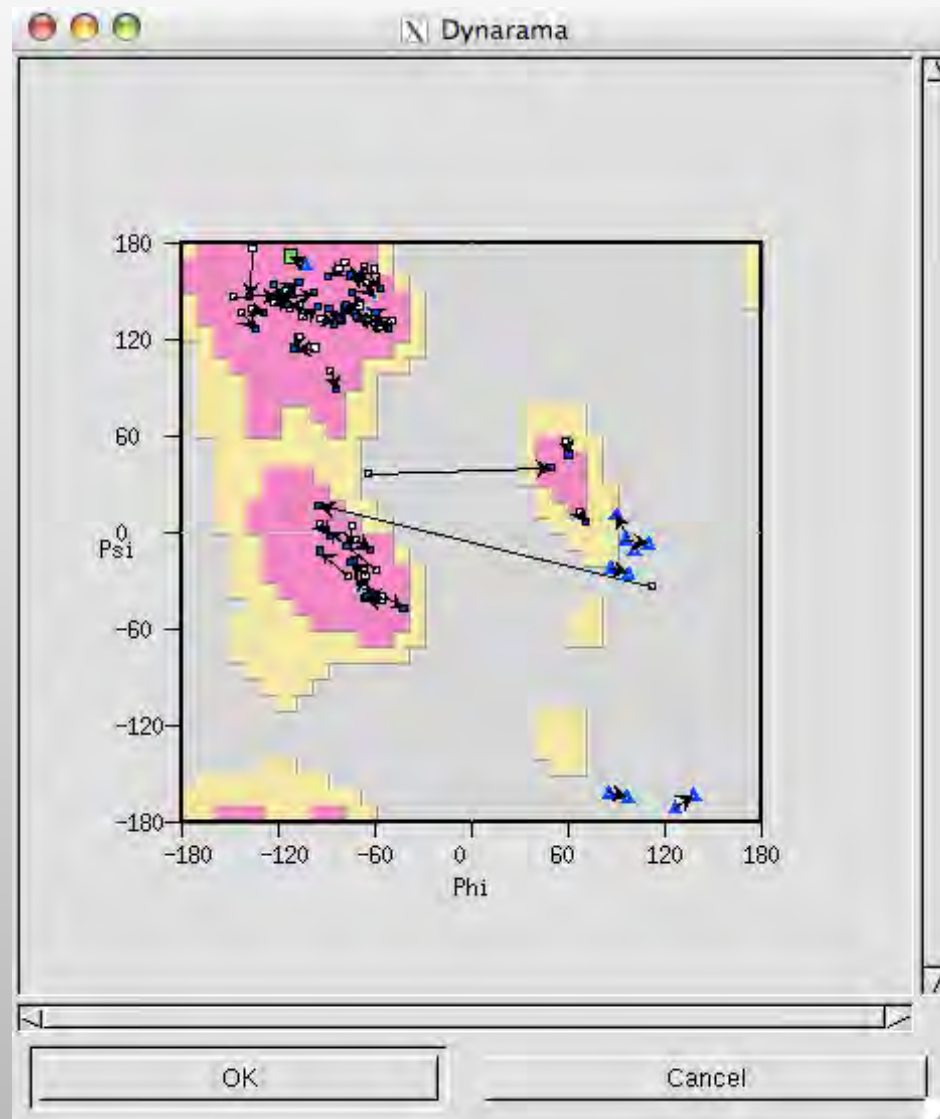
Typical Scenario:

- I have done an LSQ overlap of my NCS-related molecules and from the graph, have seen significant deviations in the positions of some side-chains.
- Why are they different?

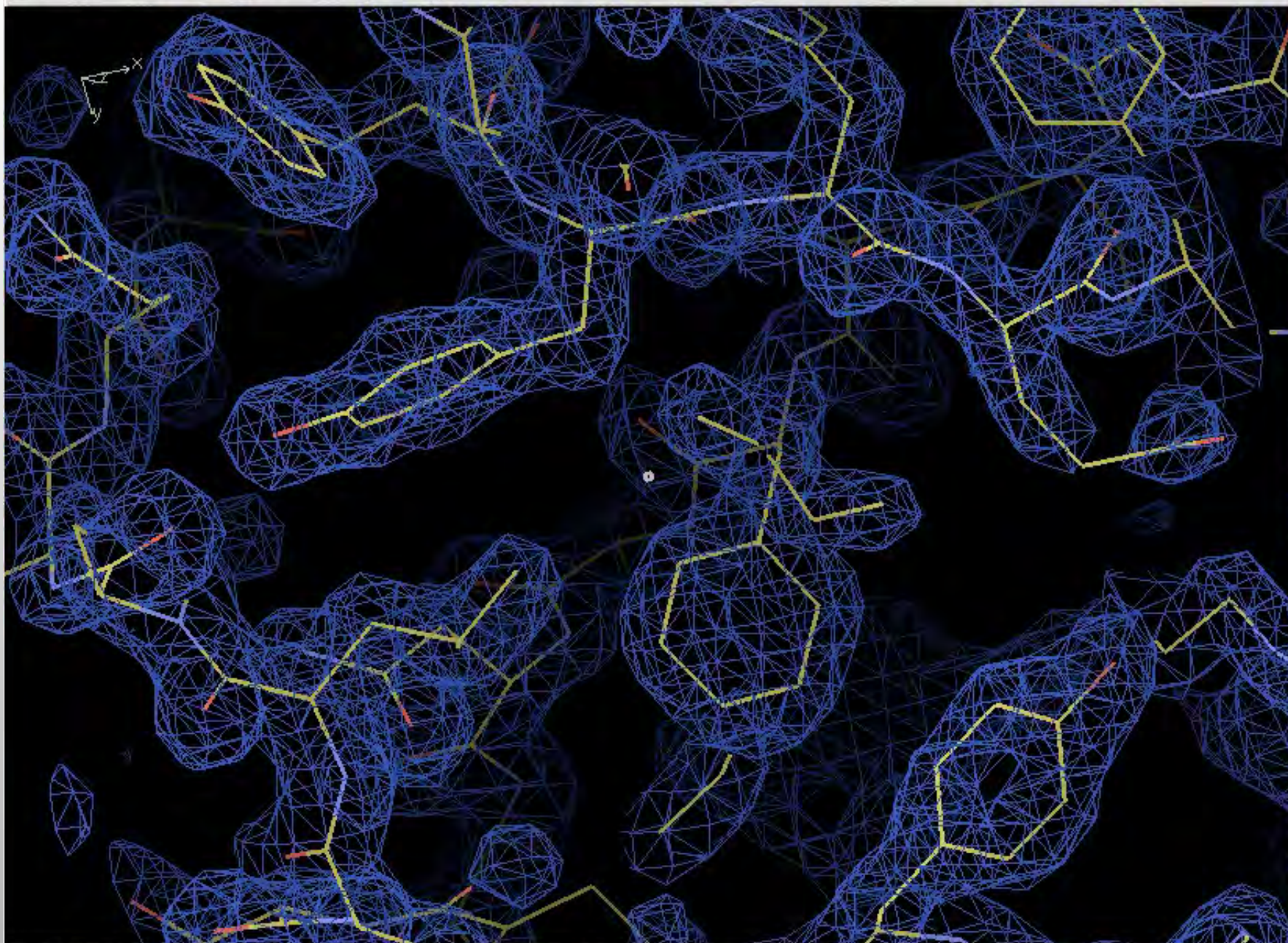
...or NCS Differences graph



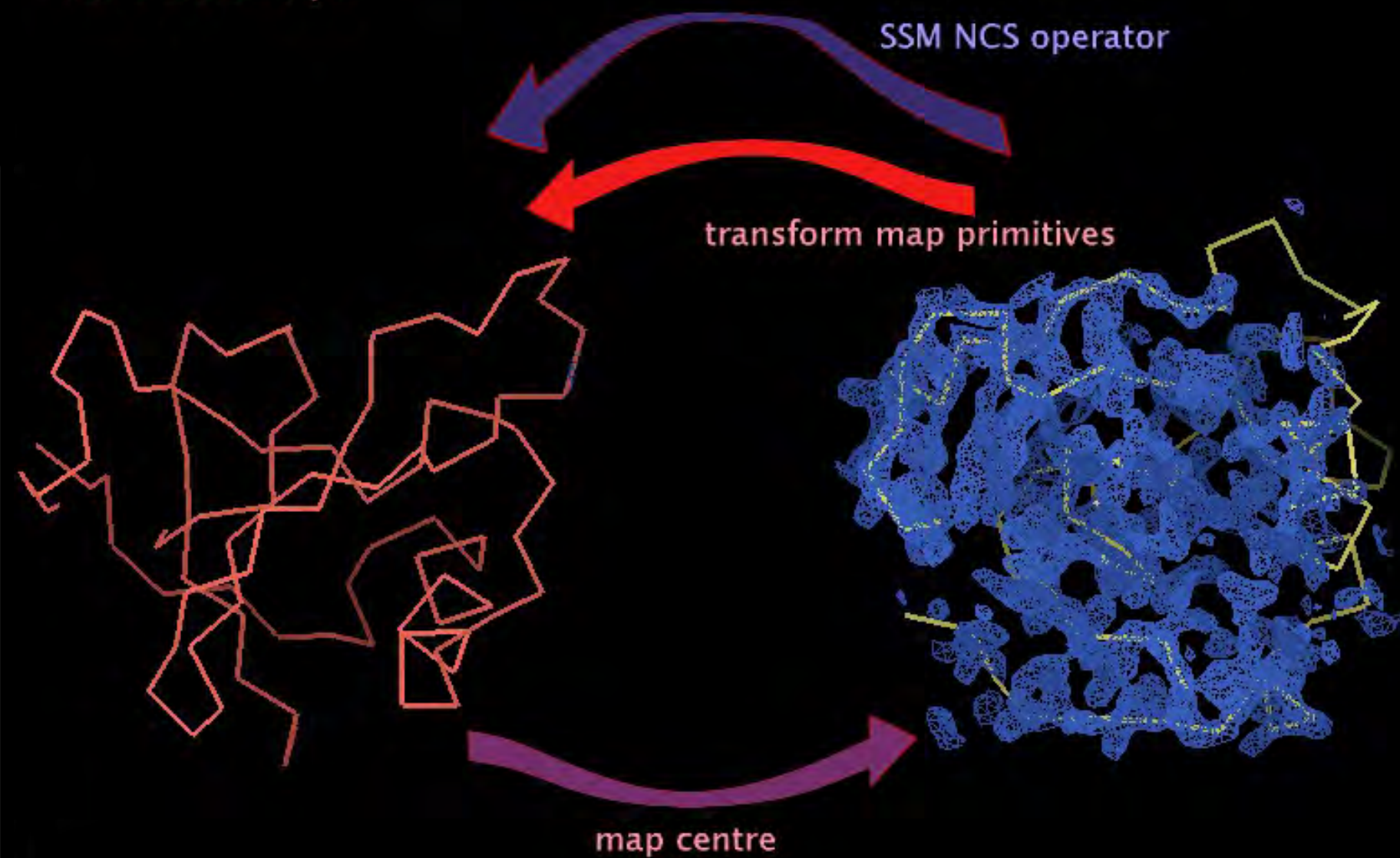
...or Kleywegt Plots[*]

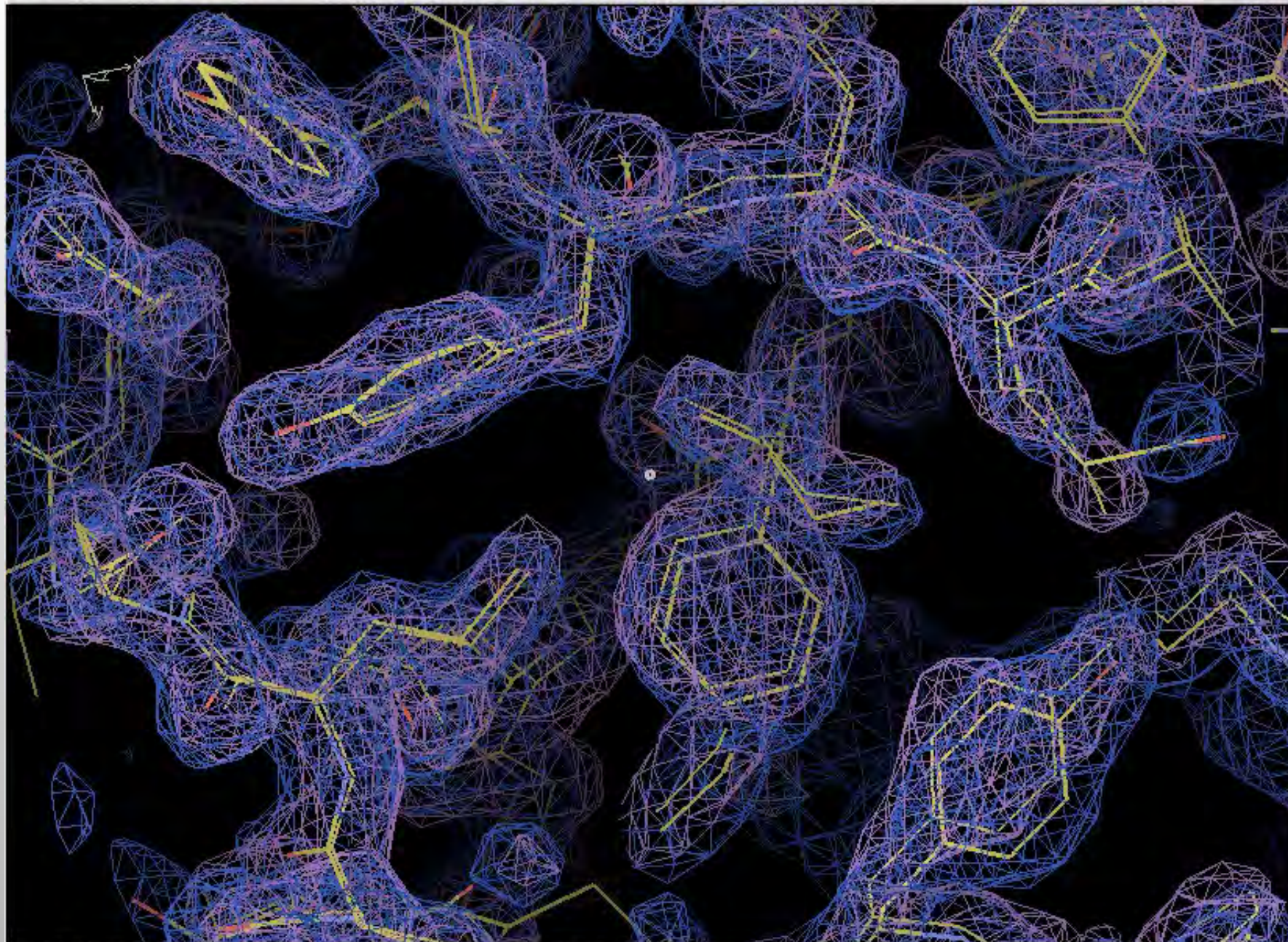


[*] Named by George Sheldrick



NCS Overlays





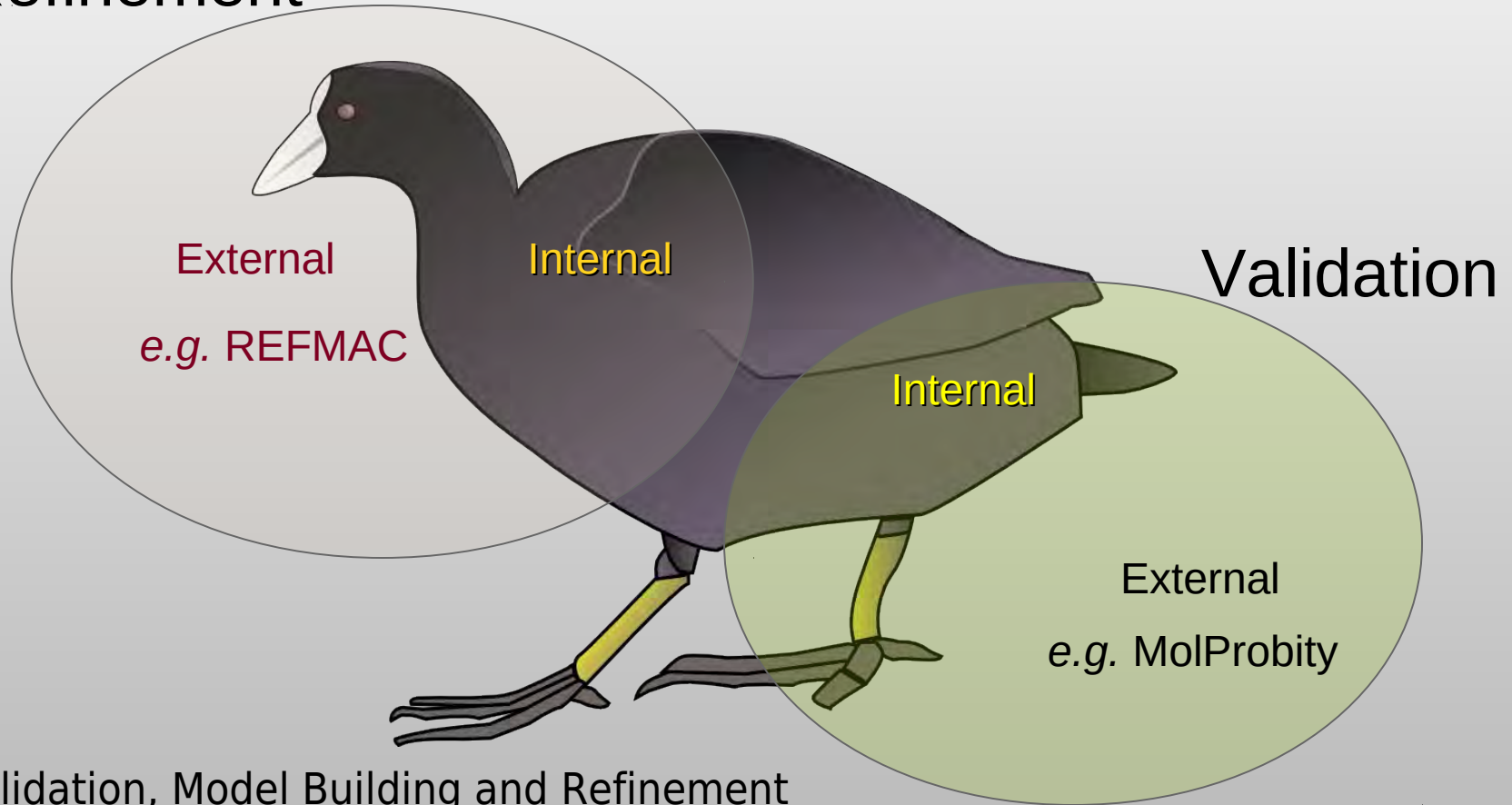
NCS Model-modification Tools

- Automatic detection of NCS
 - And their operators
- Copy Master NCS molecule to others
 - Applies NCS transformation
- Copy NCS Master residue-range
- Change NCS Master chain
- NCS Skipping ('o' key)

Validation...

Feature Integration

Refinement



Validation, Model Building and Refinement
should be used together

What is Validation?

- Comparison of various aspects of the model with pre-conceived notions of “good quality”
 - Includes **unrestrained** and **restrained** criteria
 - Many aspects of validation overlap with refinement and model-building
 - Global and local indicators

Why Validate?

- Model-building is error-prone
 - (although automated methods seem to do better)
- Map interpretation is subjective
 - Someone else did the model-building
- The model was built several years ago
 - and the notion of “good quality” has changed
- Deposition requires validation

A “good” model

- Makes statistical sense
 - The reciprocal space representation agrees tolerably well with the observations (R-factor)
 - No meaningful difference map peaks (no under- or over-fitting)
- Makes chemical sense
 - Model geometry is consistent with the restraints
 - Ramachandran Plot has less than 1% outliers
 - A good **clashscore**
- Make physical sense
 - Crystal packing and contacts
- Makes biological sense
 - Residues in chemically sensible environment
 - Is consistent (on the whole) with external biochemistry observations (active site residues)

Quick Bayes

- Bayes Eq:
- $\Pr(\text{model}|\text{data}) \propto \Pr(\text{data}|\text{model}) * \Pr(\text{model})$
- $\Pr(\text{data}|\text{model})$ is also called the model likelihood,
 $L_M(\text{model}|\text{data})$

Validation Tools - Pr(model)

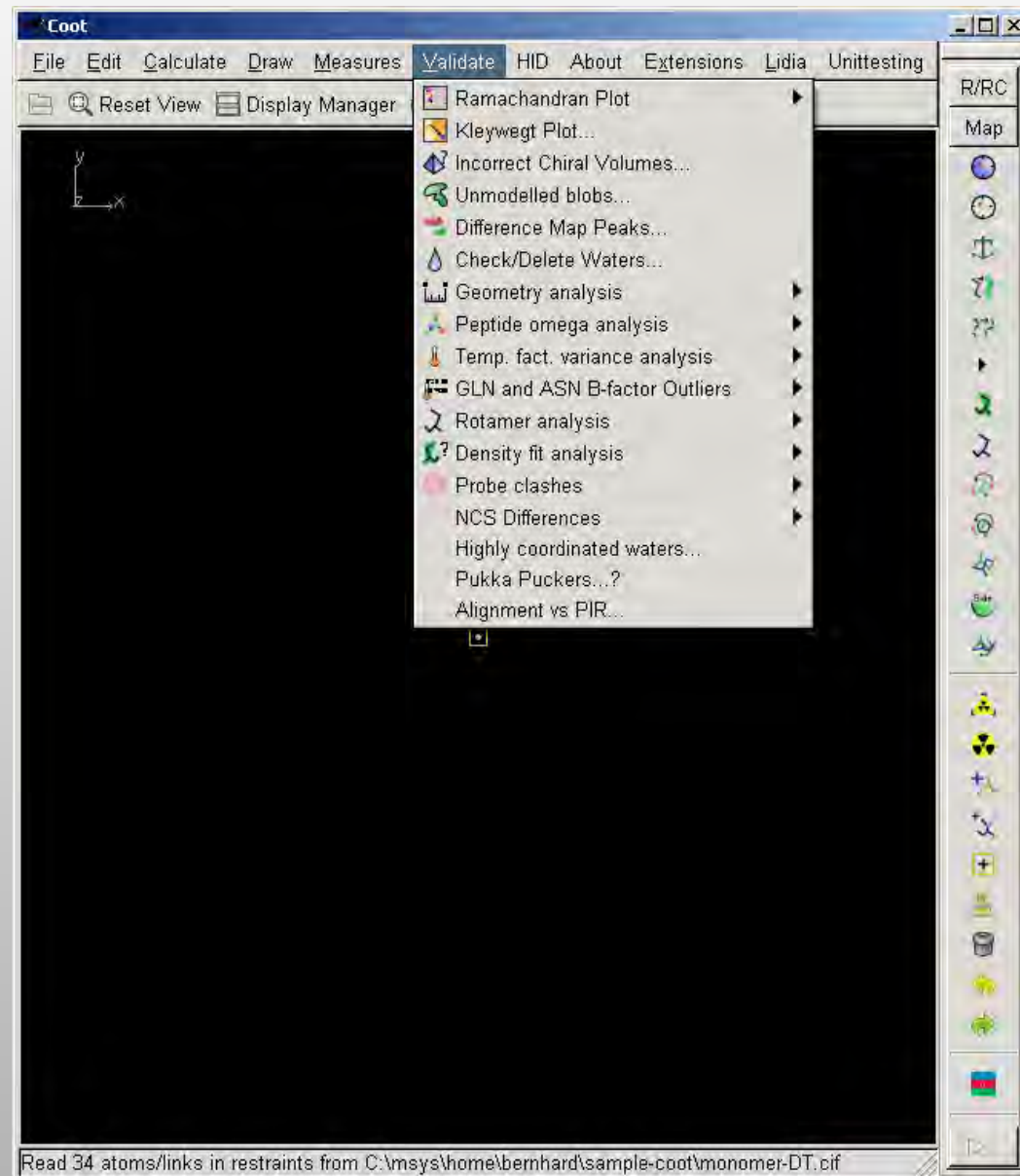
- Ramachandran Plot
 - Kleywegt Plot (NCS differences)
- Geometry Analysis
- Peptide ω Analysis
- Temperature Factor Analysis
- Rotamer Analysis
- Sequence Analysis
- Clashes

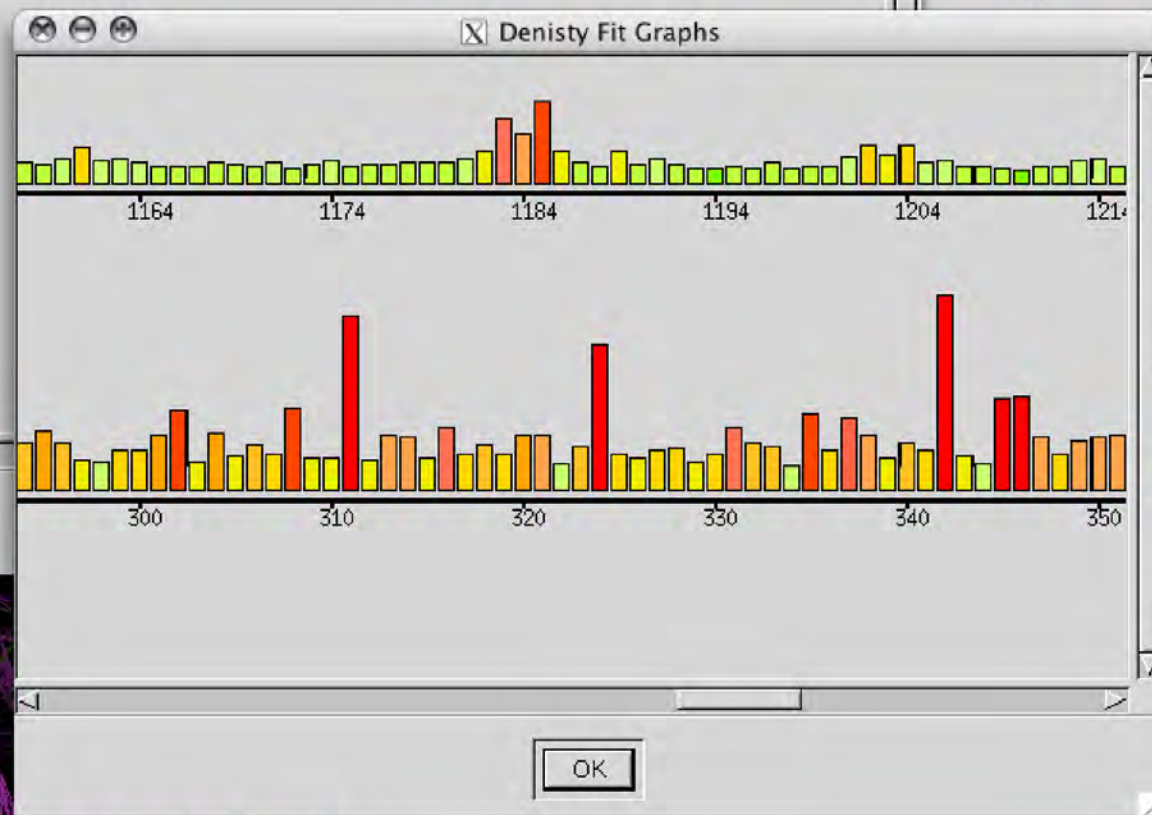
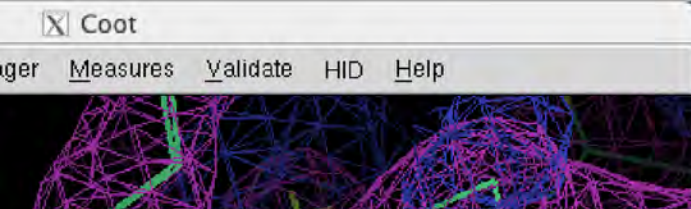
Validation Tools - $\text{Pr}(\text{data}|\text{model})$

- Density Fit Analysis
- Difference Map Peaks
 - Variance analysis at water positions
- Unmodelled blobs

Validation tools

- Geometry Distortion
 - Bonds, Angles, Planes
- B-factor variance
- Density Fit
- Peptide Omega angle
- Rotamers
- Water check
 - By distance/map density/B-factor
 - By difference map variance
- Un-modelled density blobs
- Ramachandran Plot





Pathological

Applications Places System 996 MHz Thu 21 Feb, 4:34 AM

Coot

File Edit Calculate Draw Measures Validate HID About Extensions

Reset View Display Manager

Successfully read coordinates file coot-molprobity/tutorial-modern-with-H.pdb. Molecule number 3 created.

MolProbity Multi-Chart

problems near A 68 ARG

Cluster Features

- Clash at A 68 ARG (0.594 Å)
- Clash at 220 HOH (0.594 Å)
- Clash at A 11 LEU (0.518 Å)
- Clash at A 6 VAL (0.518 Å)
- Clash at A 93 ASP (0.513 Å)
- Clash at A 5 THR (0.513 Å)
- Bad rotamer A 13 PRO (0.9%)

problems near A 41 GLU

Cluster Features

- Clash at A 41 GLU (0.566 Å)
- Clash at 194 HOH (0.566 Å)
- Clash at A 84 ASP (0.45 Å)
- Clash at A 87 ALA (0.45 Å)
- C-beta deviation A 88 THR (0.266 Å)

problems near B 69 ARG

Cluster Features

- Clash at B 69 ARG (0.424 Å)
- Clash at B 82 THR (0.424 Å)
- C-beta deviation B 3 SER (0.352 Å)

problems near A 49 TYR

Cluster Features

- Clash at A 49 TYR (0.597 Å)

Close

Generic Objects

- ☒ 0 wide contact
- ☒ 1 close contact
- ☒ 2 small overlap
- ☒ 3 bad overlap
- ☒ 4 H-bonds

Molprobity Prob

Clash gap: -1.66 : A 2 CA

Clash gap: -1.53 : A 89 CD

Clash gap: -1.32 : A 41 CA

Clash gap: -0.97 : A 39 C

Clash gap: -0.94 : A 71 C

Clash gap: -0.93 : A 72 CA

Clash gap: -0.85 : A 40 N

Clash gap: -0.76 : A 90 CA

OK

Other Programs

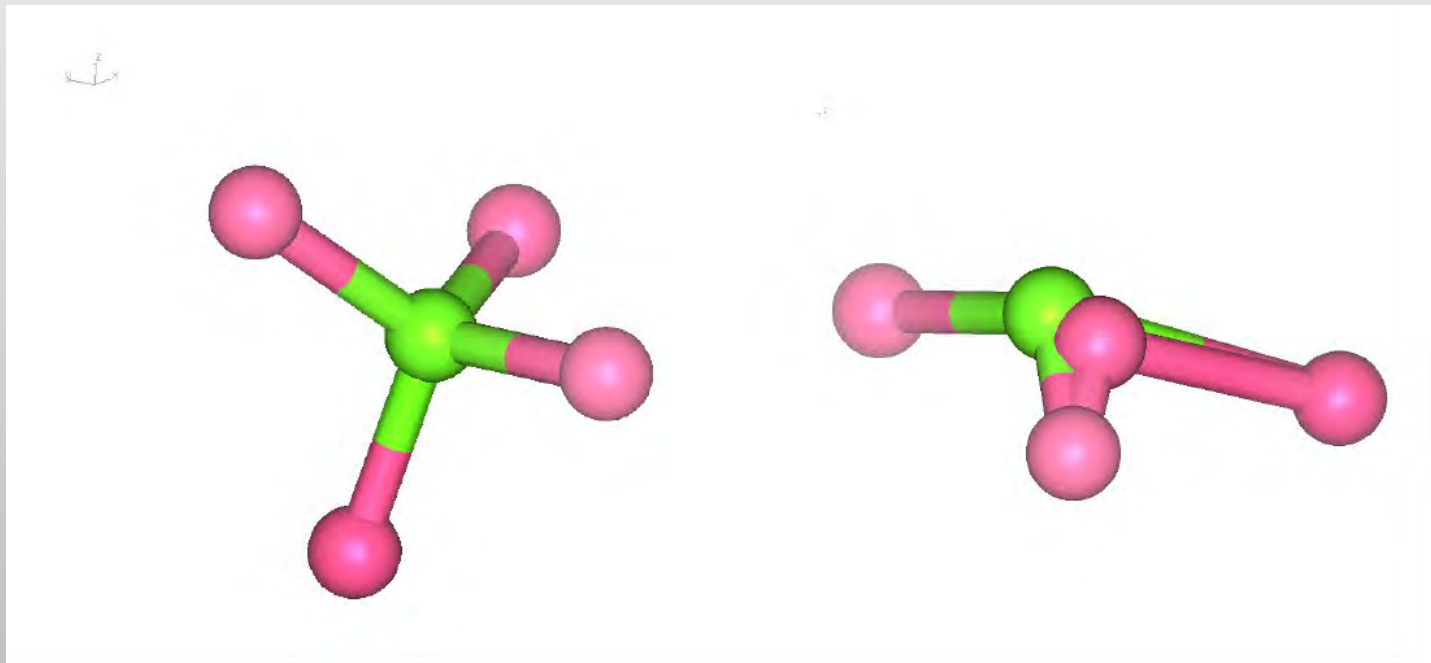
- Molprobit Suite
 - molprobit.biochem.duke.edu
- WHATCHECK
- VERIFY-3D

Ligand validation

Ligand validation

Why validate?

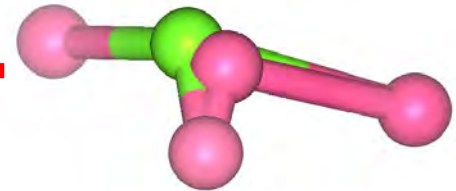
- 70.000 structures in PDB
- 11.000 'ligands' in 50.000 structures



- R/Rfree 0.15/0.19

Ligand validation

- How to validate ligand geometry?
 - Compare observed structure to restraints
 - Coot: Validate → Geometry analysis



Parametrisation issues... (what if they are wrong?)

- Perfect refinement with incorrect parameters → distorted structure
- Need independent parameters to test against

How to validate ligand geometry?

- QM
 - CPU hungry
 - In vacuo
 - Low energy $\neq$ (?) bound ligand
- PDB (e.g. ValLigURL)
 - Good cofactor structures
 - Less useful for novel ligands
 - (occ.) questionable quality
- Databases [Cambridge Structural Database (CSD) or COD]
 - Using small molecule geometries
 - e.g. Mogul

Further information

- Coot WIKI
 - <http://strucbio.biologie.uni-konstanz.de/ccp4wiki/index.php/Coot>
- Coot BB (mailing list)
 - <http://www.jiscmail.ac.uk/lists/coot.html>
- Coot documentation
 - <http://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/web/docs/>

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- Kevin Cowtan
- Eleanor Dodson
- Keith Wilson

<http://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/>

or

Google: Coot

or for WinCoot

<http://www.ysbl.ac.uk/~lohkamp/coot>

- Libraries, dictionaries
 - Alexei Vagin, Eugene Krissinel, Stuart McNicholas
 - Dunbrack, Richardsons
- Coot Builders and Testers
 - William Scott, Ezra Peisach
 - York YSBL, Dundee, Glasgow (early adopters)
 - Coot Mailing List subscribers