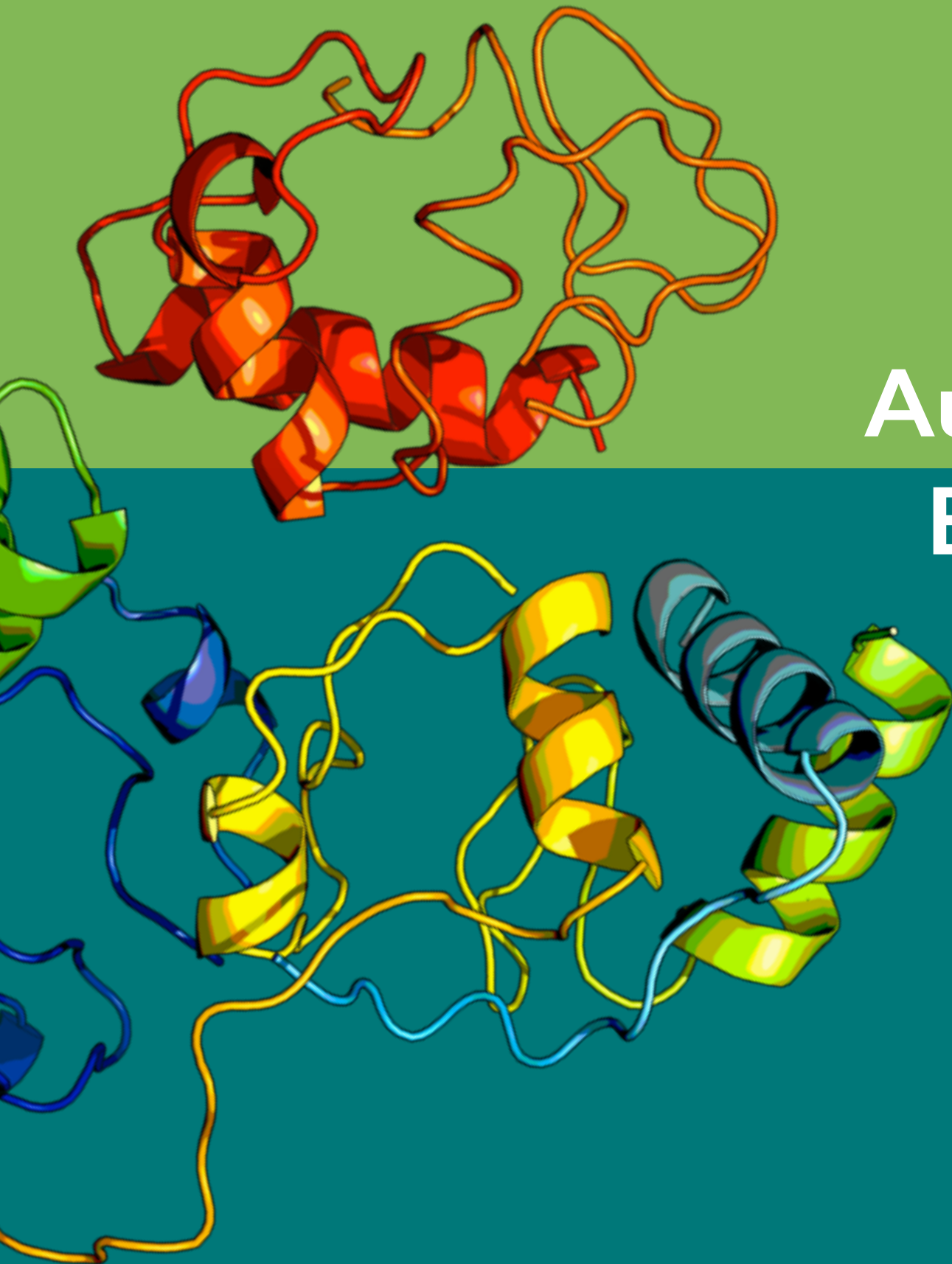


Joana Pereira
Lamzin Group
EMBL Hamburg, Germany

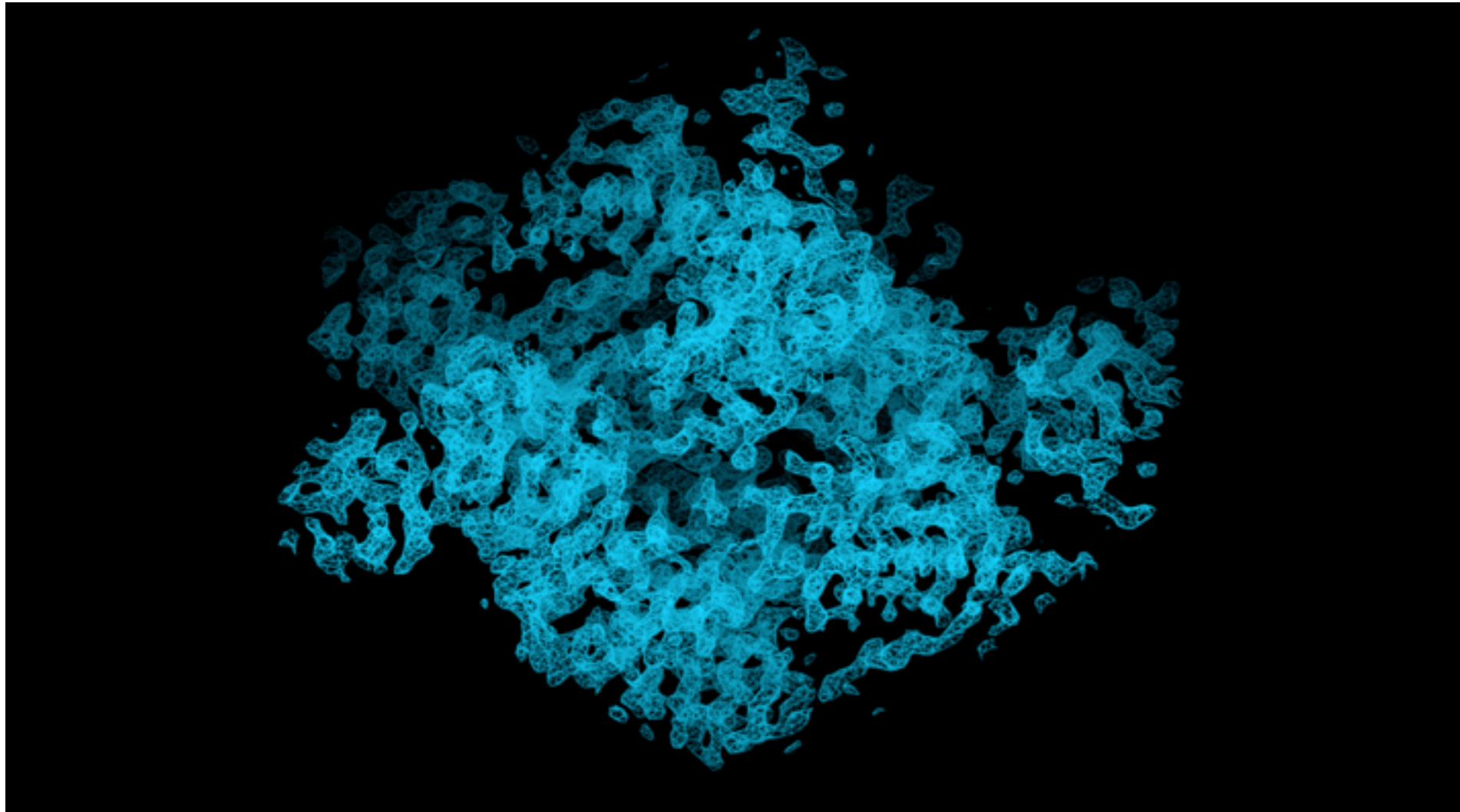
Automated Protein Model Building with ARP/wARP

(and nucleic acids too)



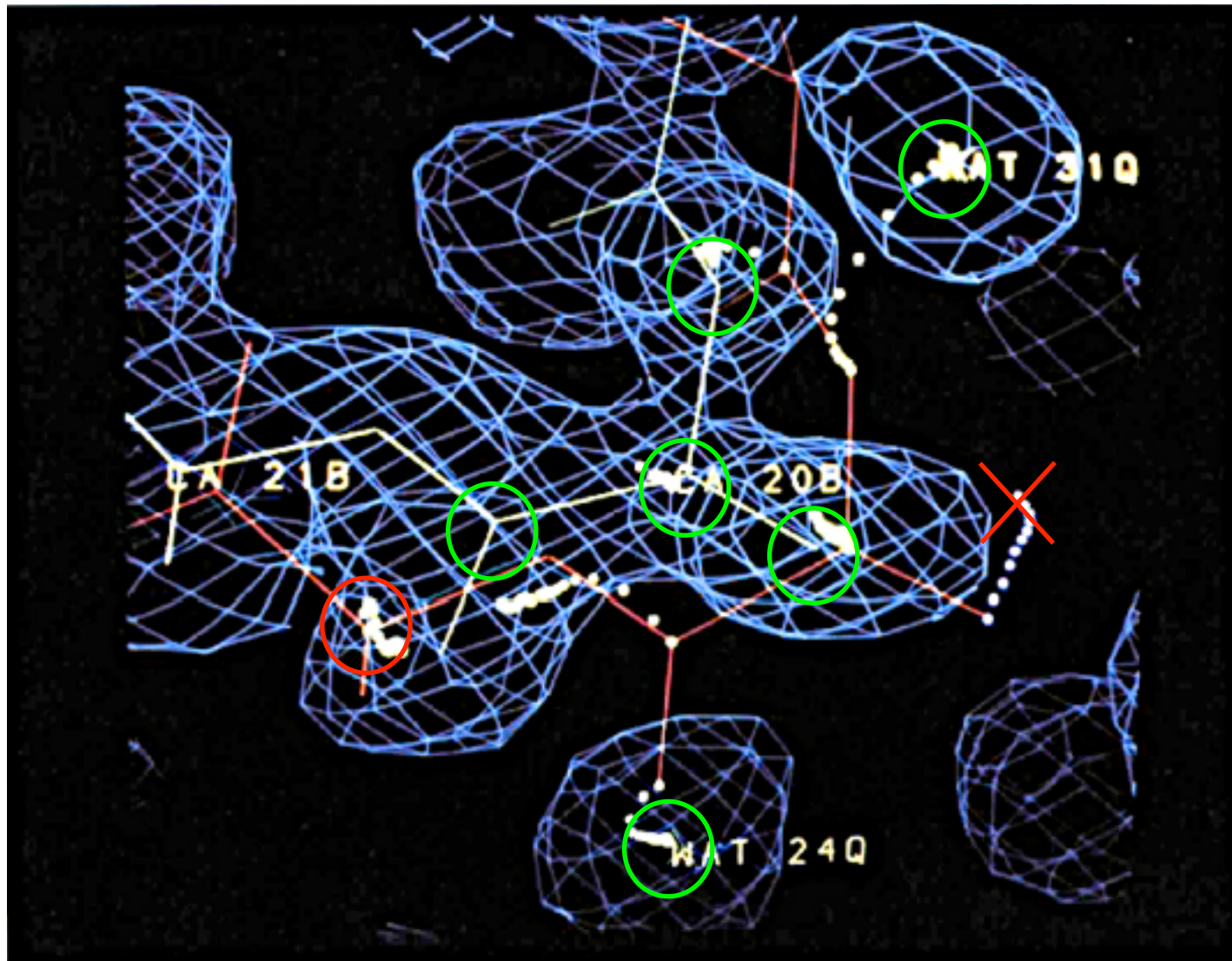
Electron density map: the ultimate result of a crystallography experiment

How would you do to interpret this map?



Building a molecular model from the electron density map calls for automation, it can take a long time when done by hand!

Developed in Hamburg by the Lamzin group, it is:

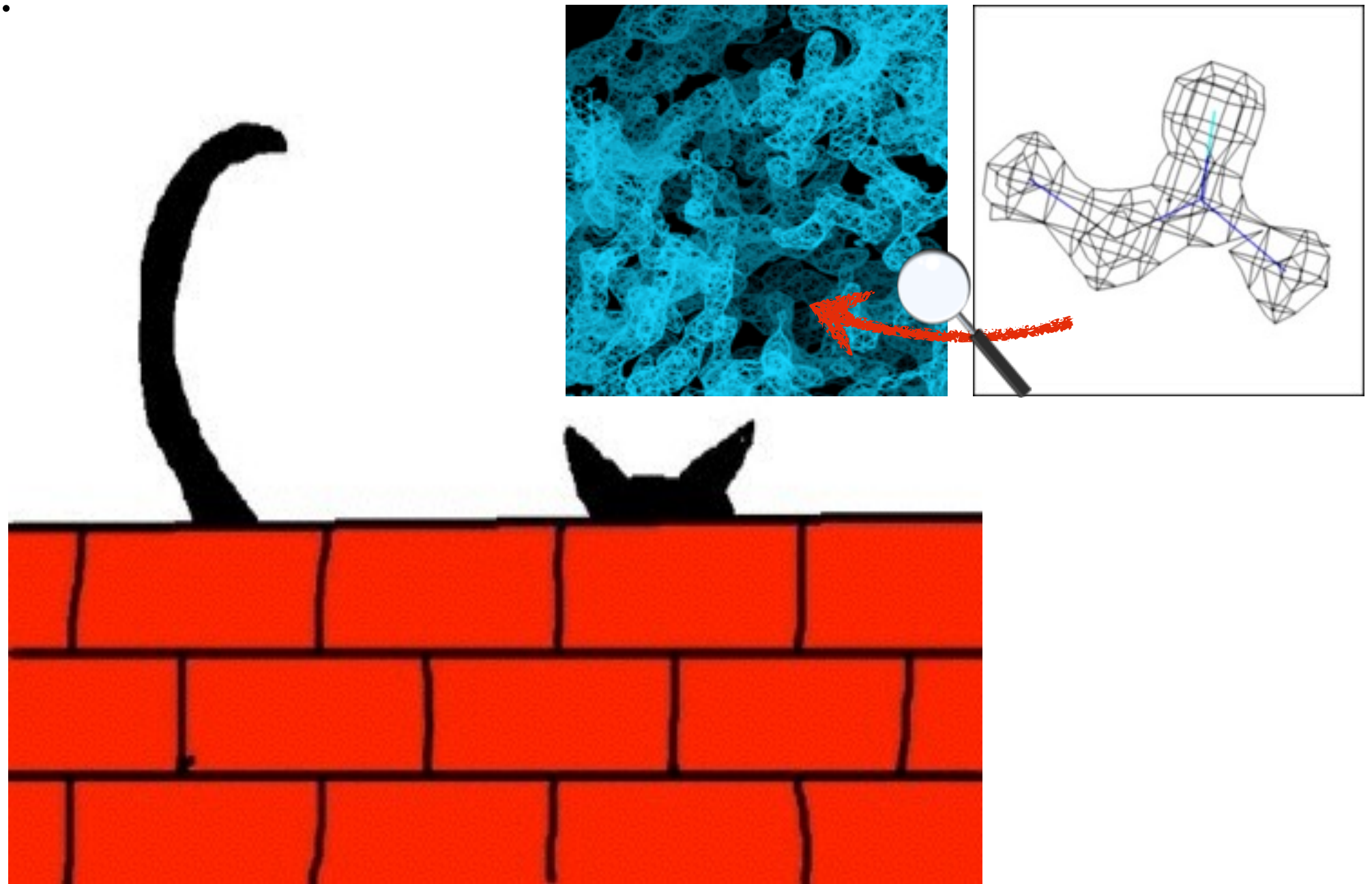


- one of the leading **MX** software projects
- used to build structures of **proteins**, **nucleotides**, **ligands** and their **complexes**
- making use of pattern recognition to build **models** from MX **electron density maps**

Lamzin, V.S. & Wilson, K.S. (1993)
Automated refinement of protein
models. *Acta Cryst.* D49, 129-147.

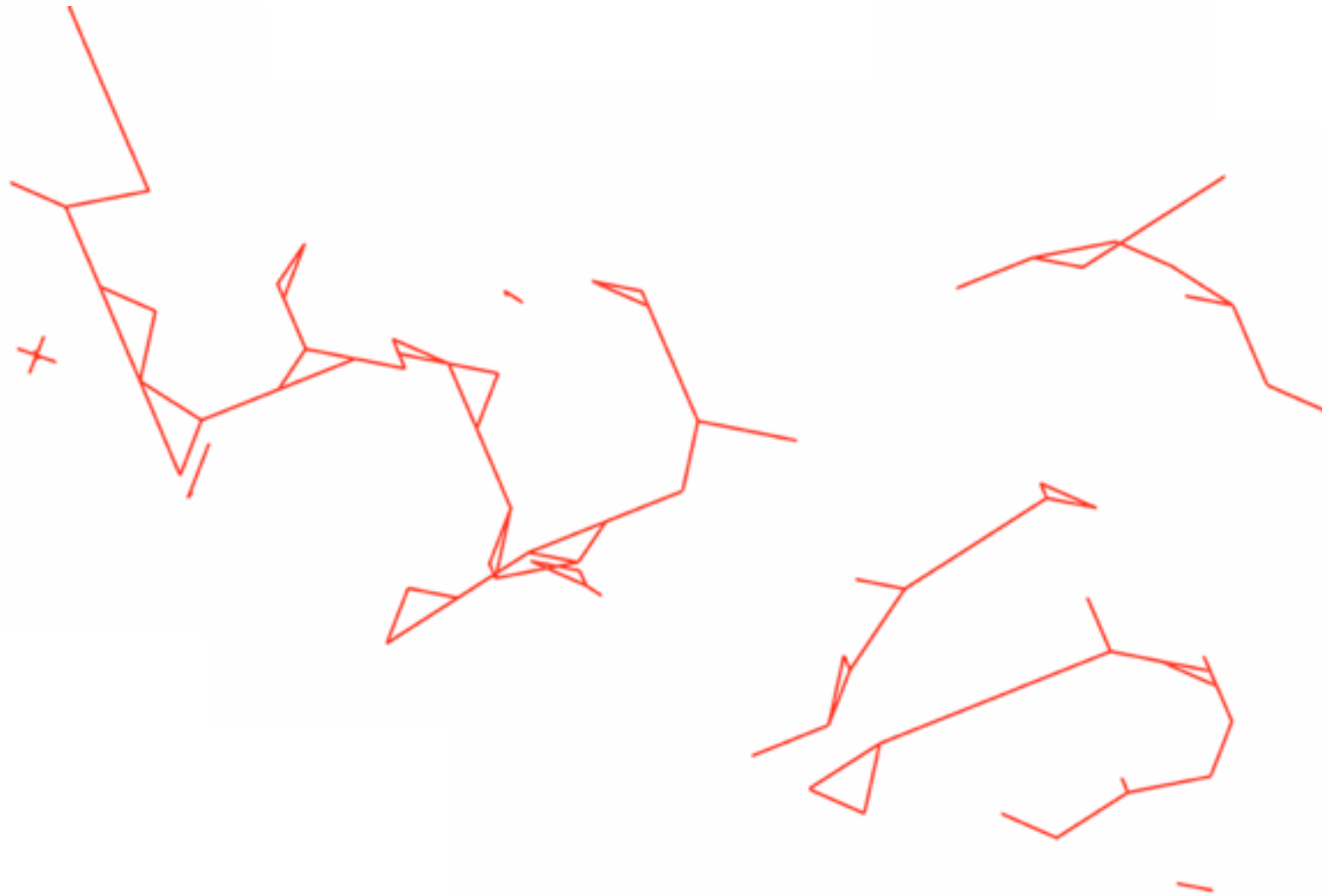
Pattern Recognition

It allows the transformation (reduction) of information from **signals** to **meaning** given a *priori* available knowledge:



Methods for electron density map interpretation

Skeletonisation: tracing the path of the macromolecular chain using the connectivity of regions of high electron density



It gives a picture of how one can connect the electron density and where the main chain may run.

Methods for electron density map interpretation

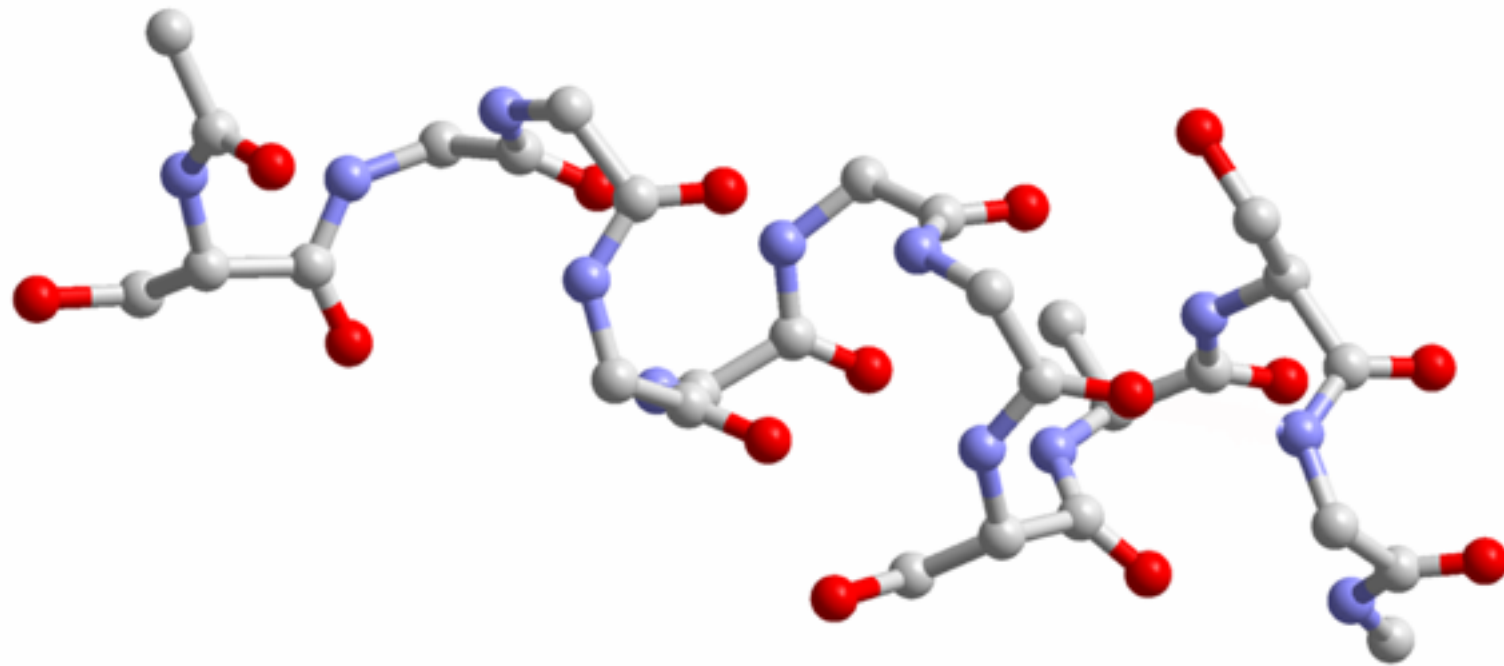
Secondary structure templates: using penta-alanine templates in ideal α -helix and β -sheet conformations to find secondary structure elements in the density



It shows where we can find the secondary structure elements and then one has to try to connect them.

Methods for electron density map interpretation

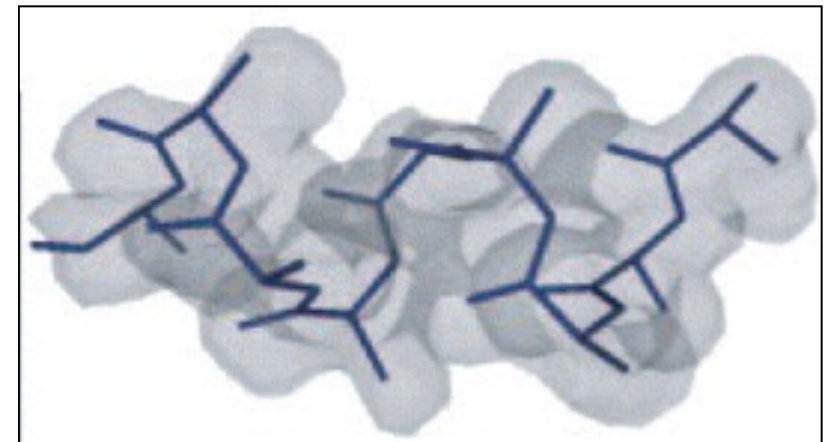
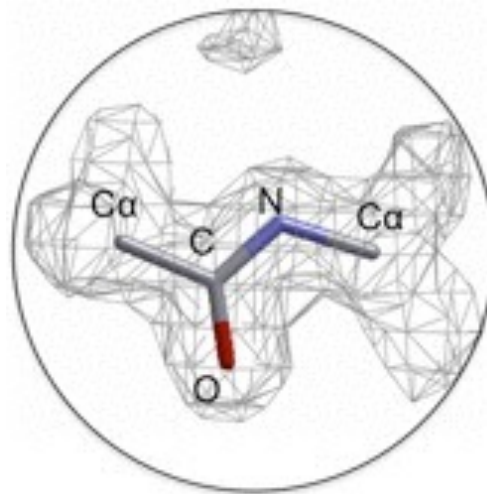
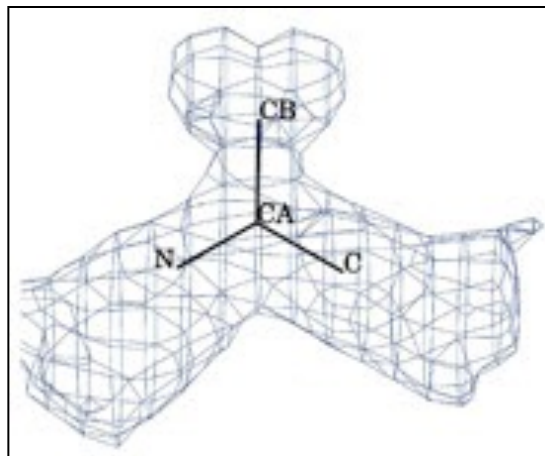
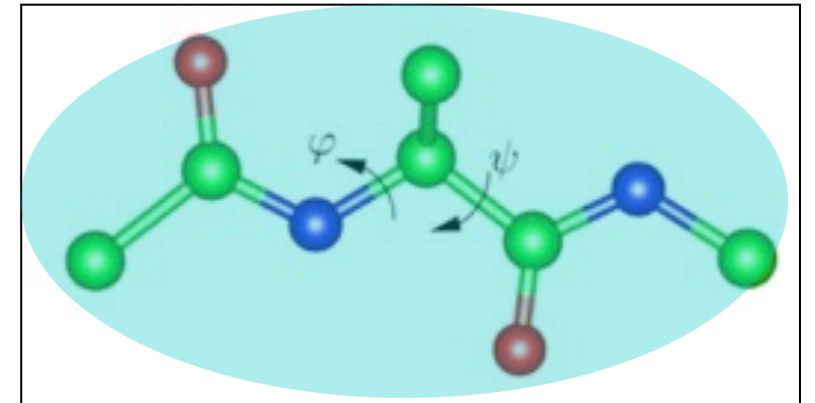
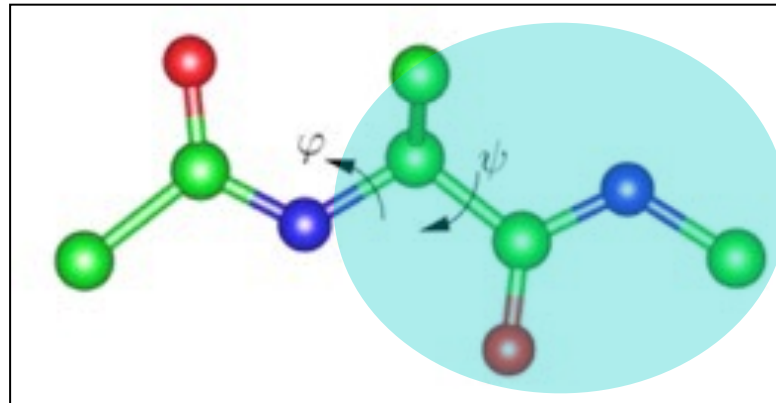
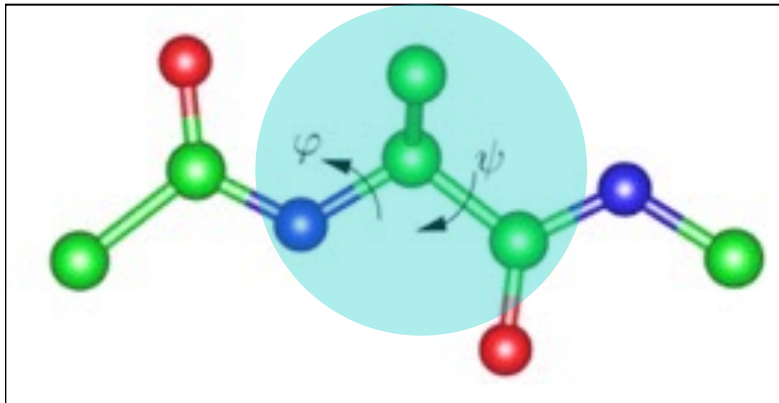
Free atoms: describing the electron density by a set of free atoms without chemical identity that can then be connected to form a main chain

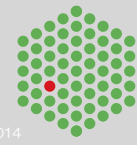


Searching for free atoms in possible C α positions, the main chain can be found.

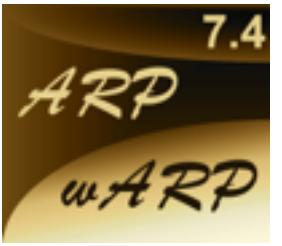
Methods for automated model building

Most current methods search for different patterns that are then used to build and extend a main chain:

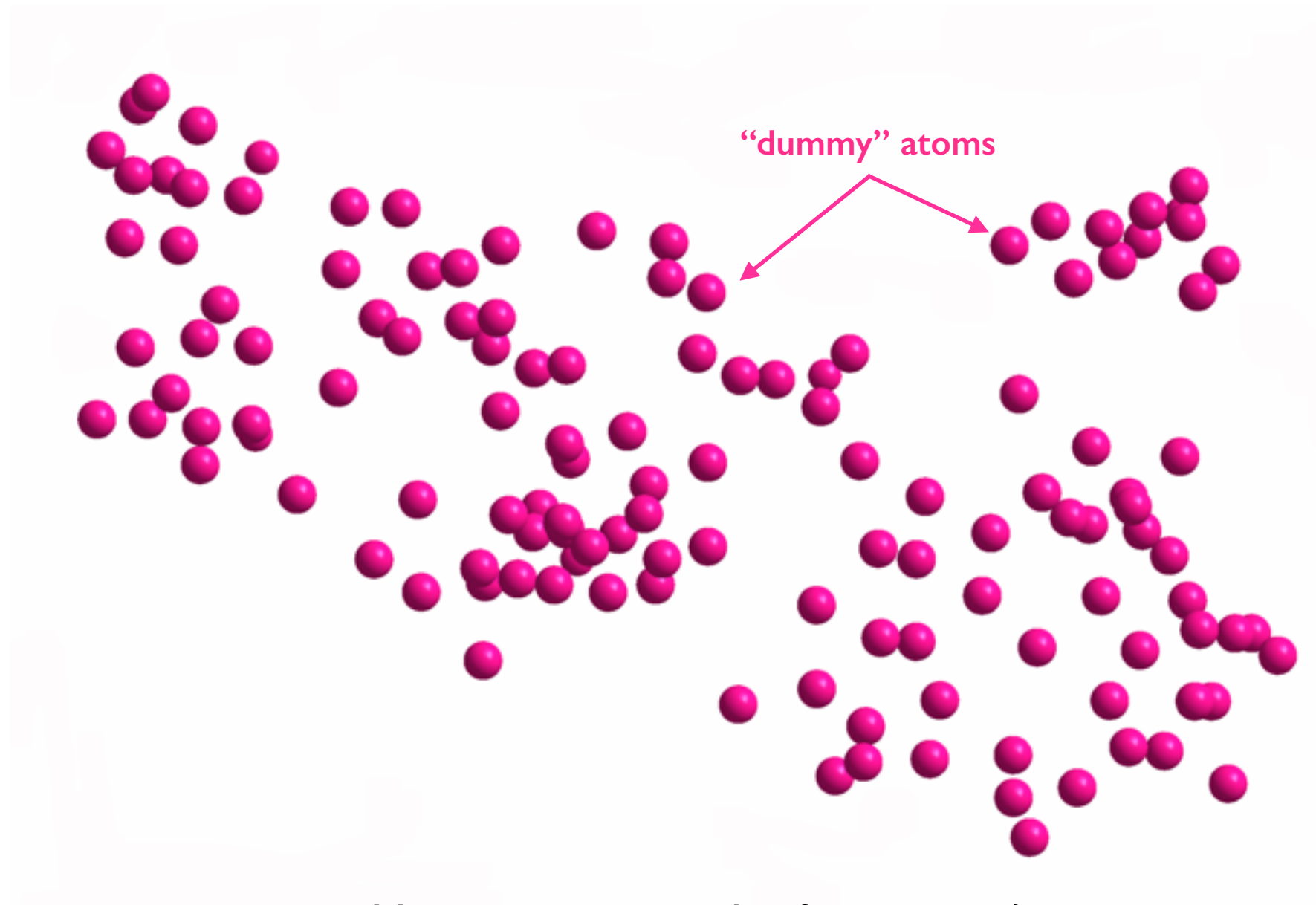


TEXTAL / Buccaneer	ARP/wARP 	RESOLVE / ACMI
Cα	(Di-) Peptides	Fragments

The initial model

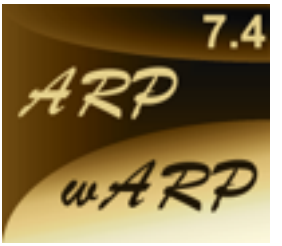


Given an electron density map and the protein sequence, ARP/wARP describes the electron density map as a set of free atoms without chemical identity:

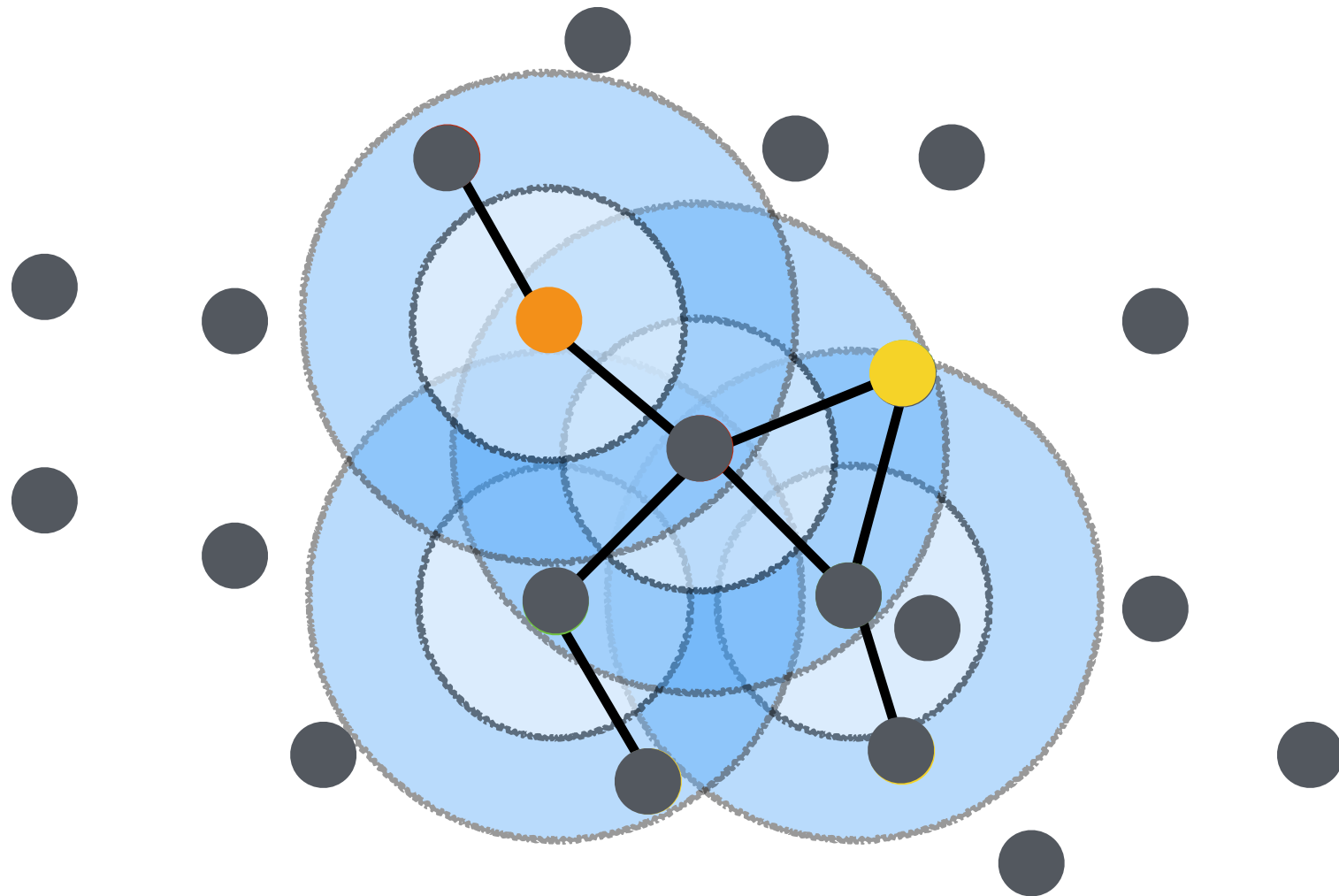


How to connect the free atoms?

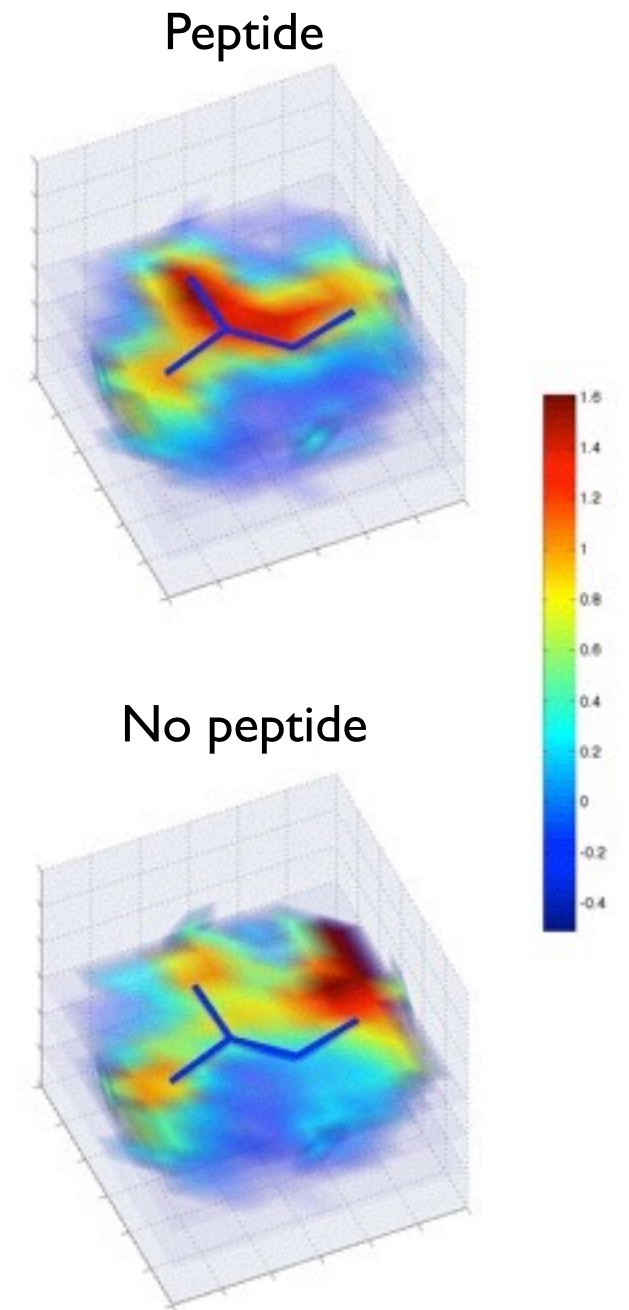
Finding peptides



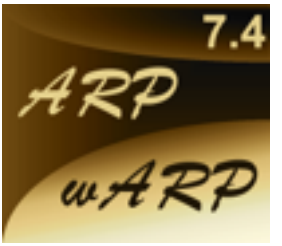
Free atoms at distances close to the ones observed in peptides are searched:



Peptides that share one C α atom can be connected and dipeptides with correct conformation connected to build the longest fragment

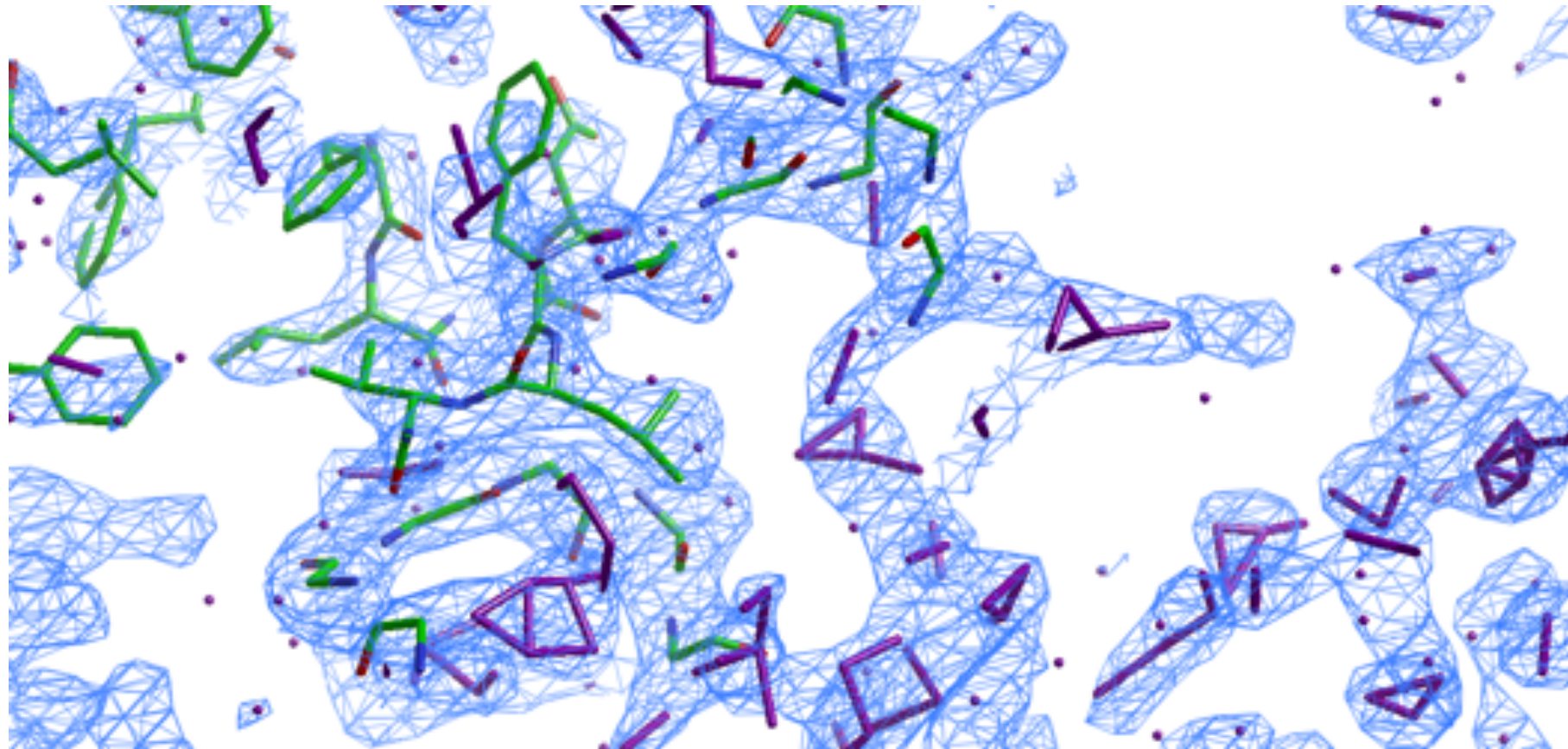


The hybrid model



As only a small part of the initial free atoms model is now recognised as a putative protein, we have an hybrid model that combines 2 sources of information:

- Chemical knowledge from the partially built model
- Free atoms continue to describe the electron density



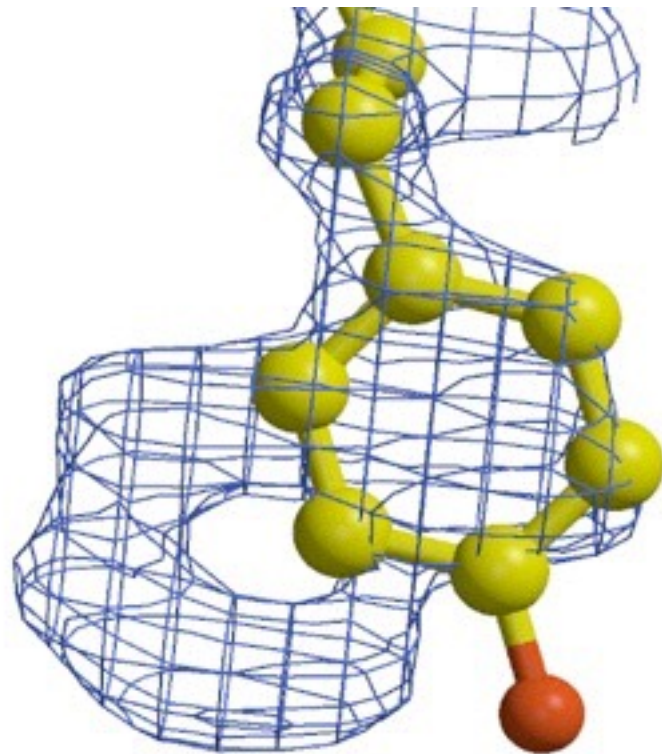
It can provide restraints for further **refinement**



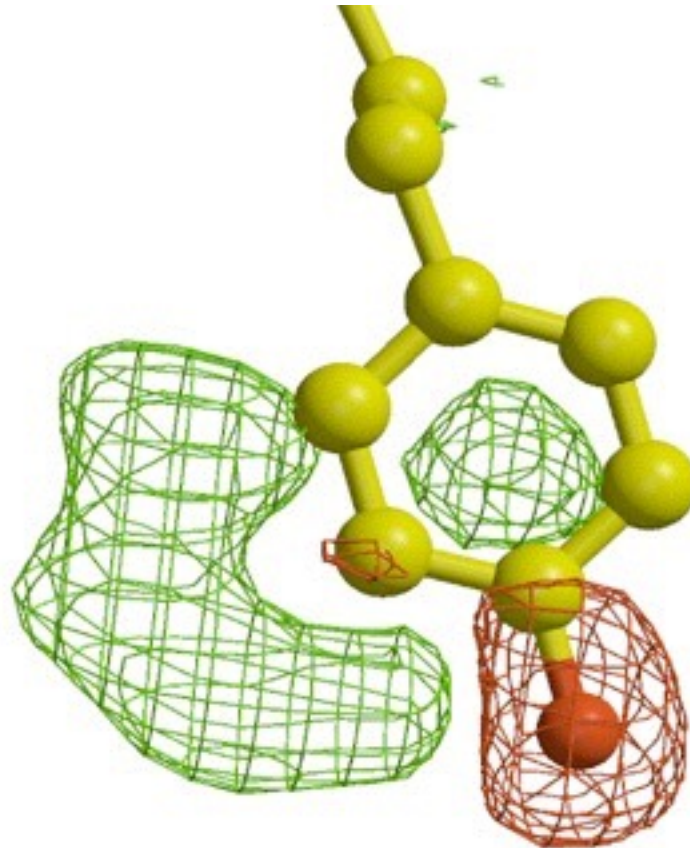
Allows the improvement of the map and consequent better model building

Model update

As the density is improved by refinement, the model can be updated by removing and addition of atoms:



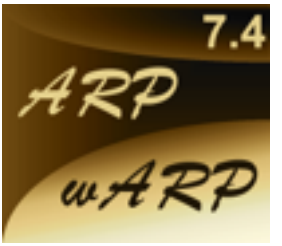
2Fo-Fc map



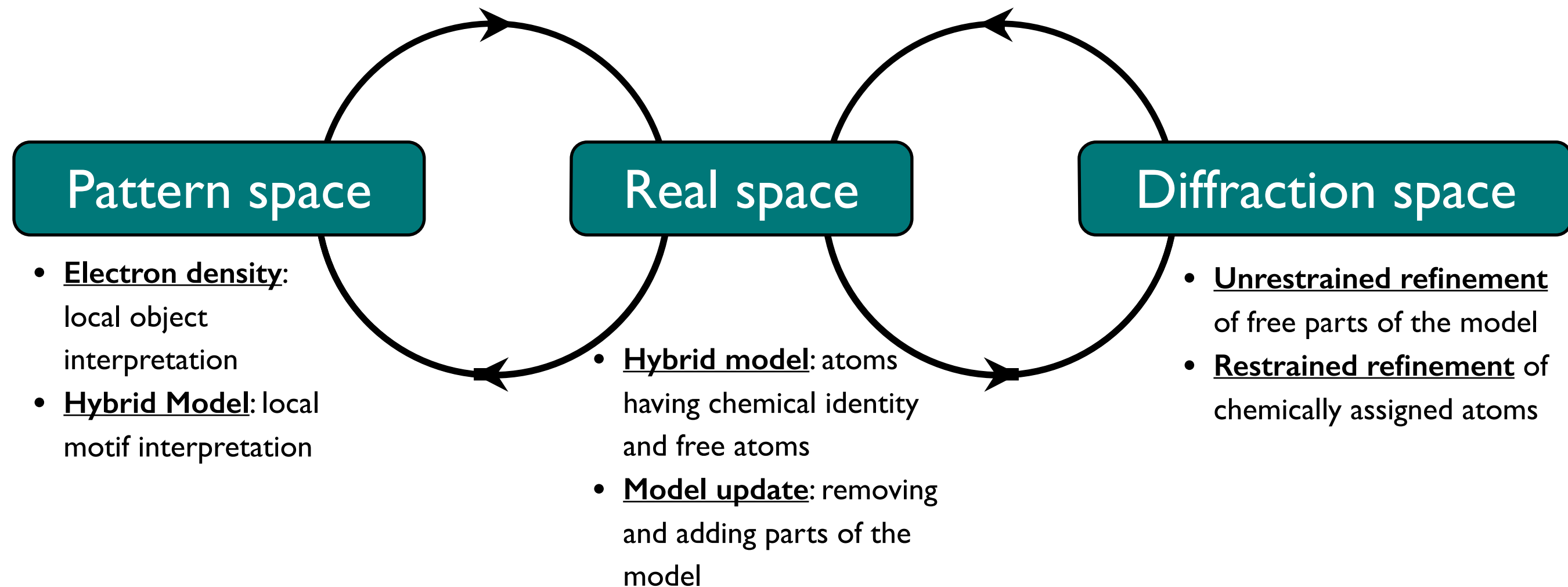
Fo-Fc map

```
ARP/wARP will be iterated with REFMAC5  
50 refinement / model update cycles will be run in total.  
Atoms will be removed below 1.0 sigma in 2mFoDFc map and added above 3.2 sigma in mFoDFc  
map.
```

The fundamental ARP/wARP concept



ARP/wARP circulates between three spaces, forming a unified process of model building:



Scheme of restraints and free atoms are iteratively updated
Hybrid model is converging to the final model

ARP/wARP workflow

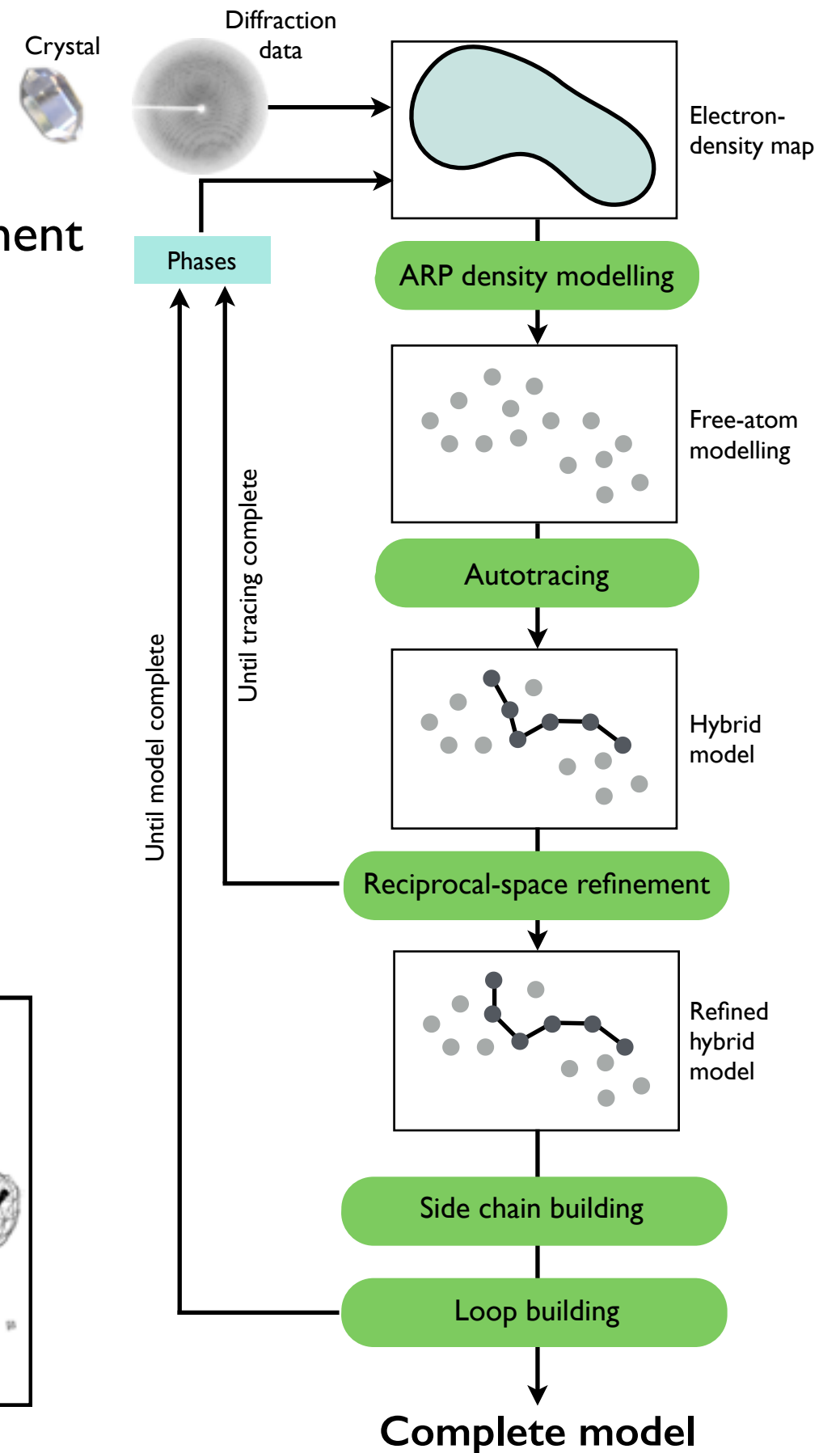
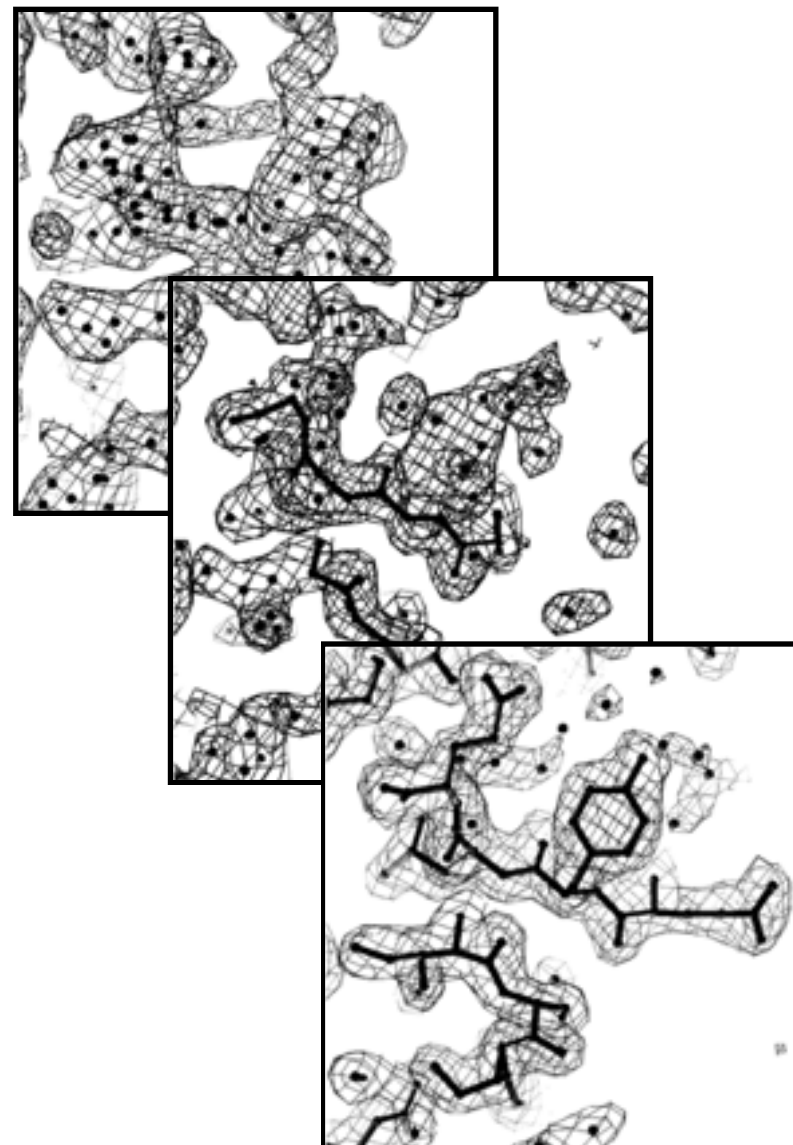
ARP/wARP iterates between main chain tracing, refinement and model update to build models that converge to the final model:

Free Atoms

Pattern Recognition

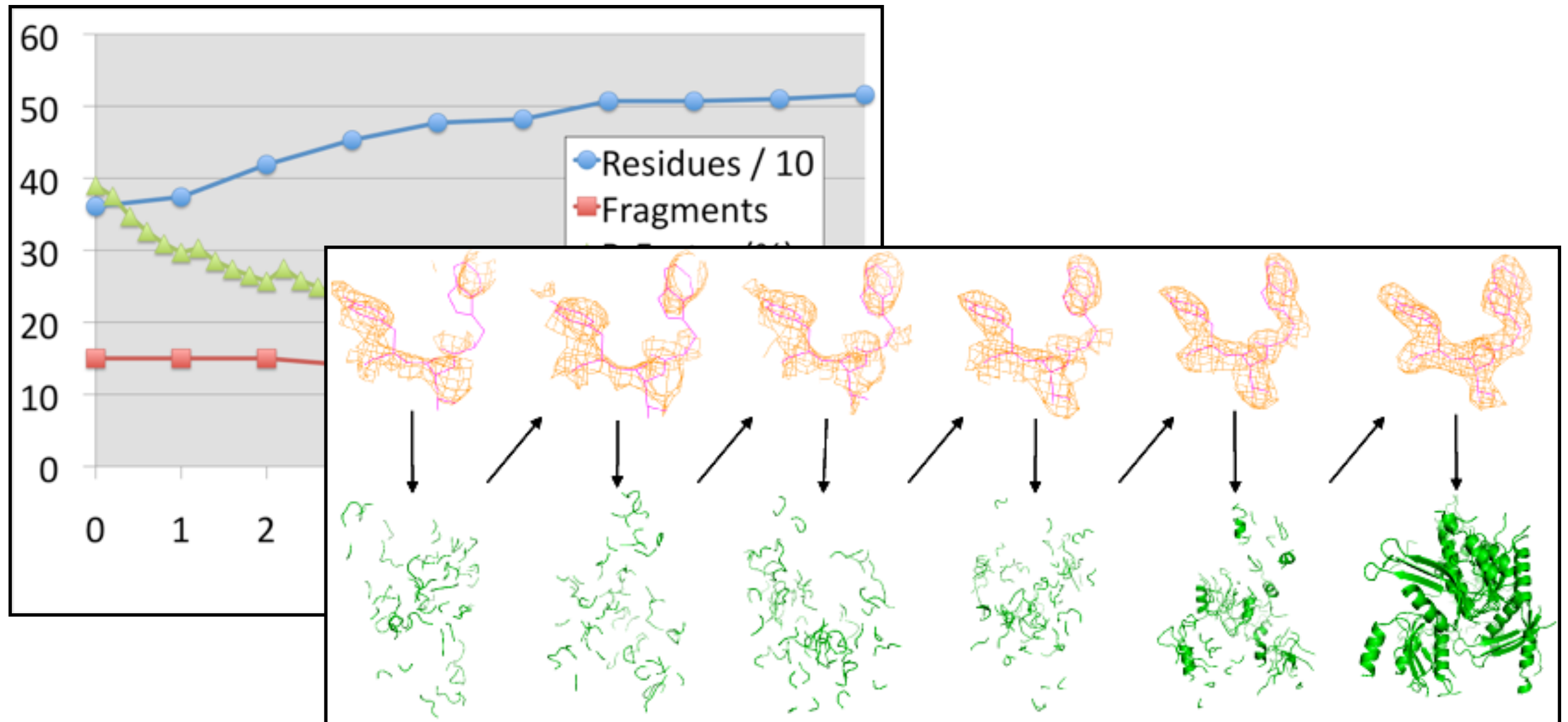
Hybrid Models

Iterations



ARP/wARP workflow

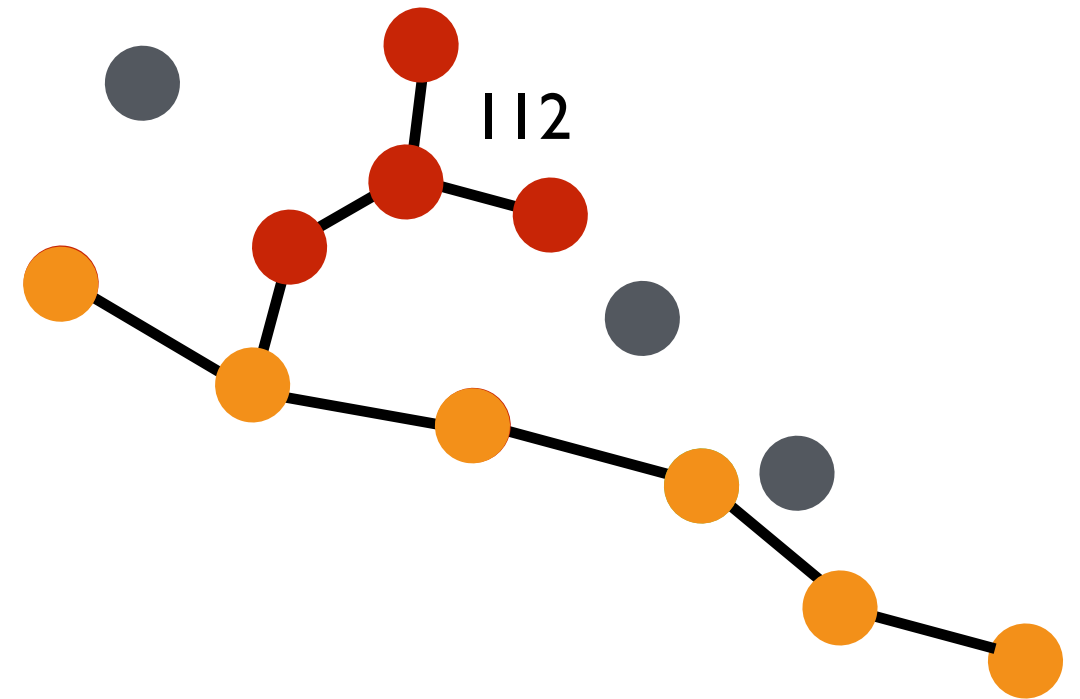
ARP/wARP iterates between main chain tracing, refinement and model update to build models that converge to the final model:



Sequence docking and side chain building

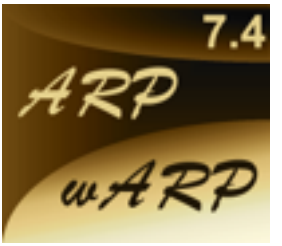
After main-chain tracing, one gets a set of main-chain fragments and the **side chain-building module** has to:

1. **Sequence docking** using connectivity vectors
(e.g., serine “11”, aspartate “112”)
2. **Side chain building** using the Richardson’s rotamer database



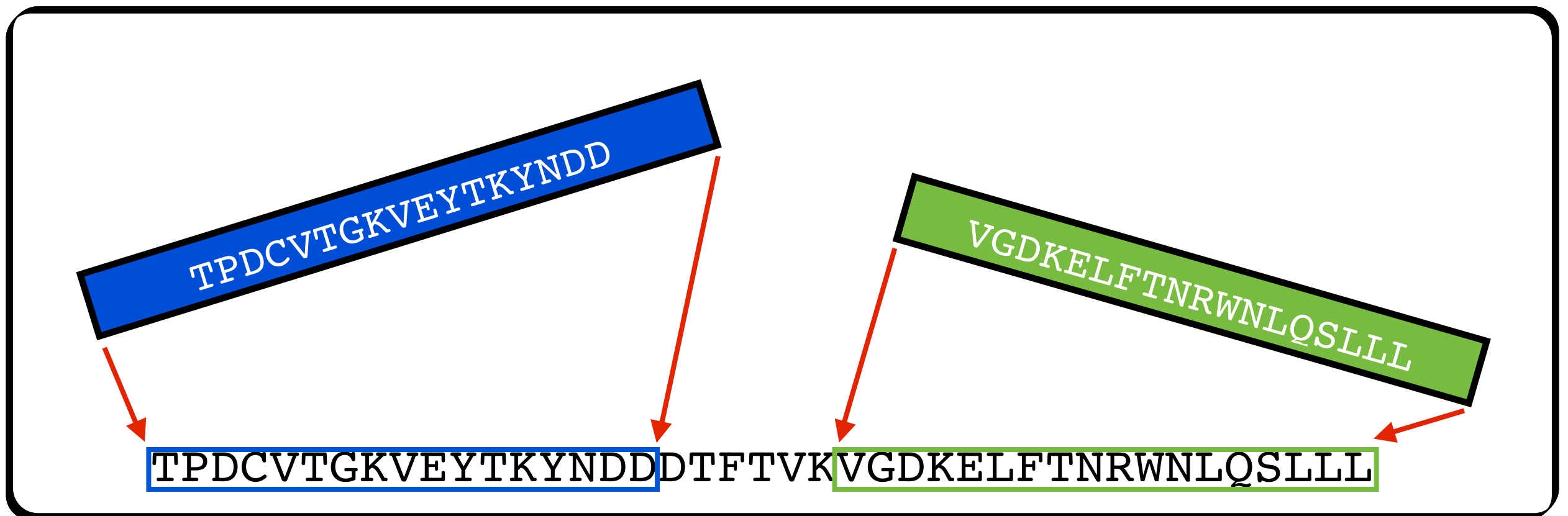
This allows the construction of loops!

Loop building



Even after a successful automated model-building run, some regions of known start, end, length and amino-acid sequence remain to be built, and ARP/wARP builds these **loops** by:

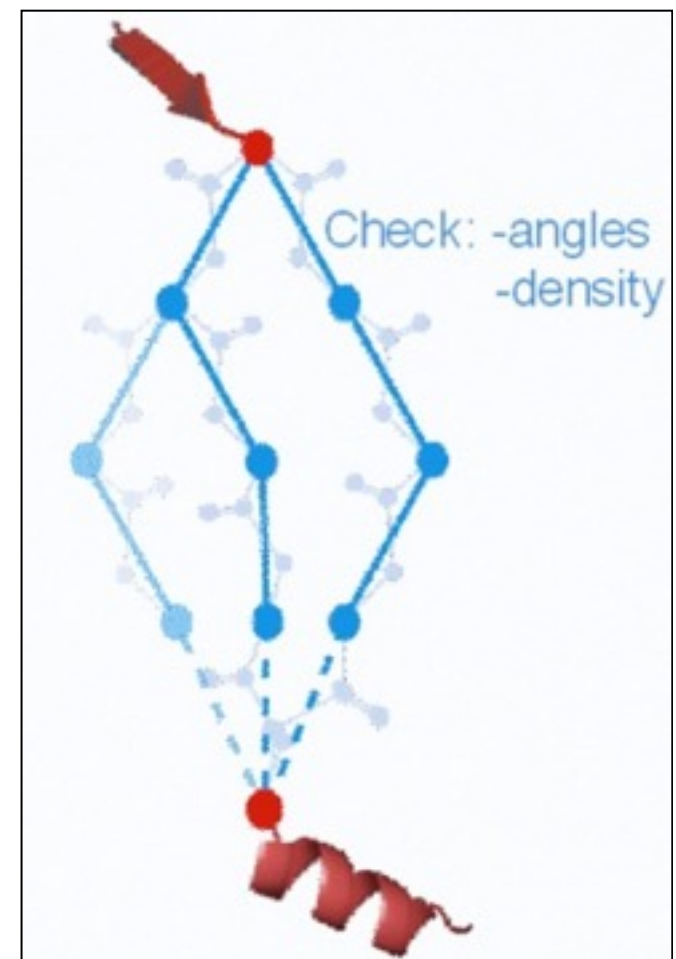
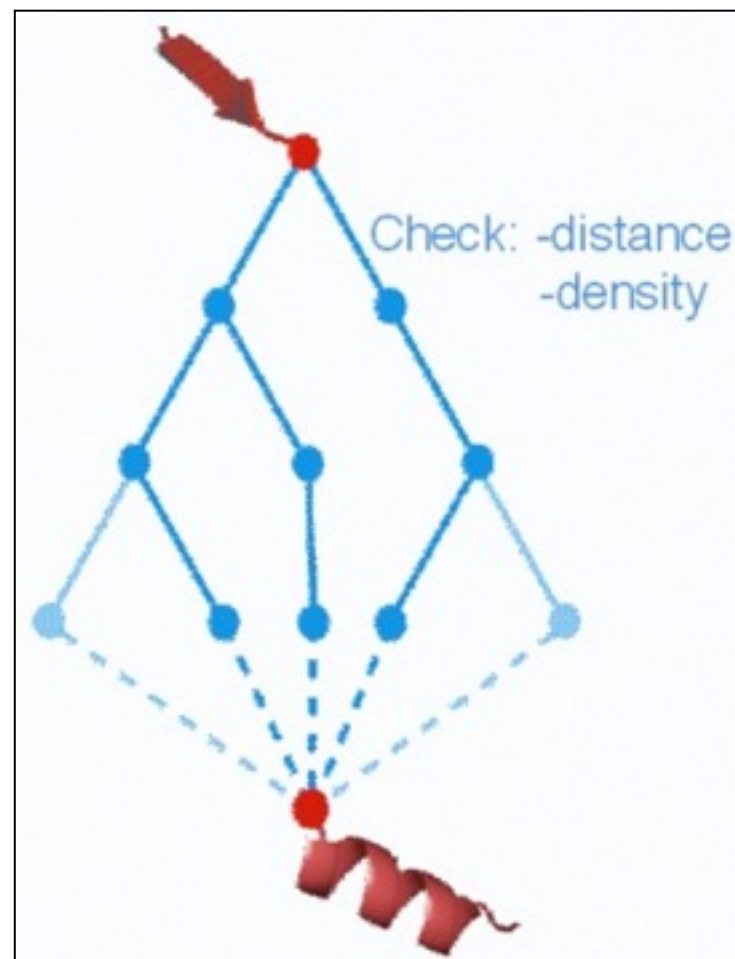
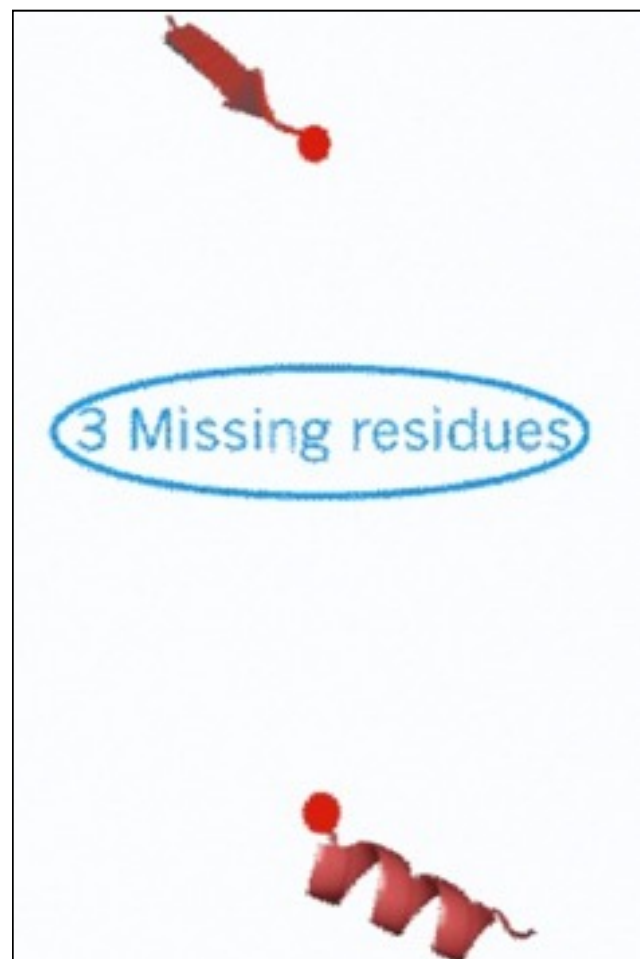
- Using knowledge of angles between anchors
- Check with density to see if modelled loops are valid



Loop building

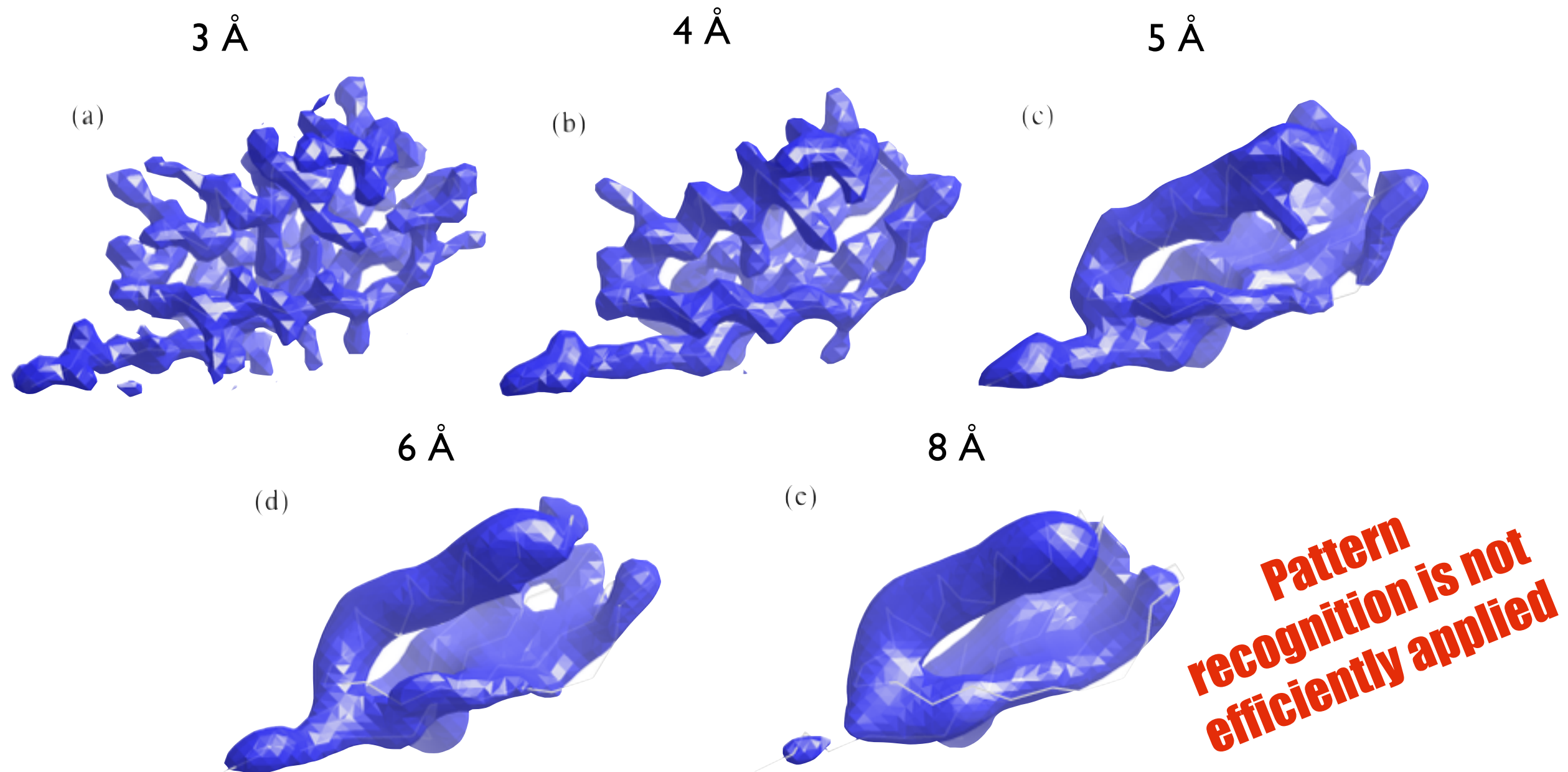
Even after a successful automated model-building run, some regions of known start, end, length and amino-acid sequence remain to be built, and ARP/wARP builds these **loops** by:

- Using knowledge of angles between anchors
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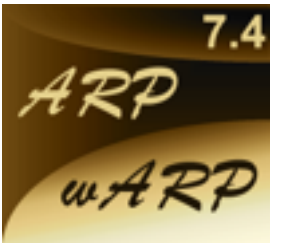


Low resolution as a limit

The primary factor determining the accuracy of an experimental protein structure is the **resolution of the X-ray data**:



ARP/wARP and resolution



Model building methods were historically developed for and work considerably better with high-resolution diffraction data:

Results from the ARP/wARP webservice (tracing performance)

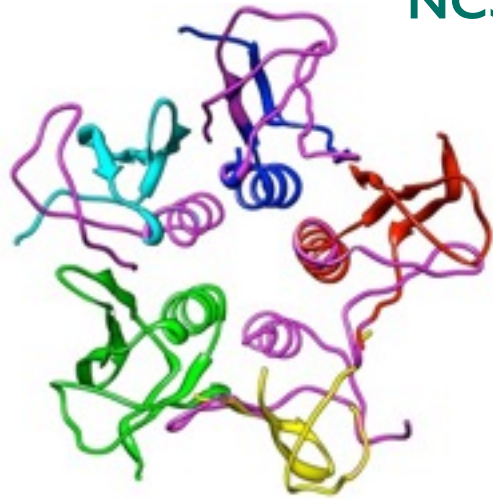
Resolution	Estimated fraction of automatically built protein structure (October 2011 – March 2012)
1.5 Å and higher	98% and higher
1.5 - 2.0 Å	91 – 98%
2.0 - 2.5 Å	81 - 91%
2.5 - 3.0 Å	67 - 81%
3.0 – 3.5 Å	51 - 67%

Protein model building at medium-to-low resolution in the best case generates incomplete and fragmented macromolecular models!

Methods to improve modelling at lower resolution

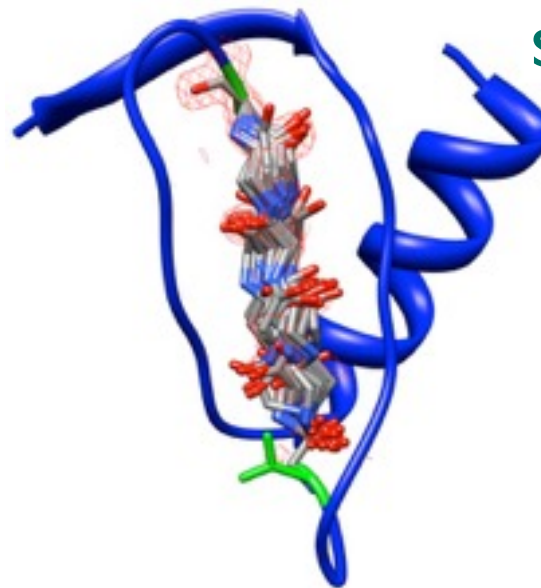
There's a big effort now to improve automated model building at medium-to-low resolution:

NCS detection and extension



- Detection of NCS related fragments and extension by “copy-paste”
- Improvements at 2.0-3.5 Å
- PNSextender

Sequence-independent loop fitting



- Secondary structure prediction docking to find fragments that can be connected
- Search for links based on the gap size probability and a database of fragments
- Improvements at 3.0-3.8 Å

NCS Detection and Extension

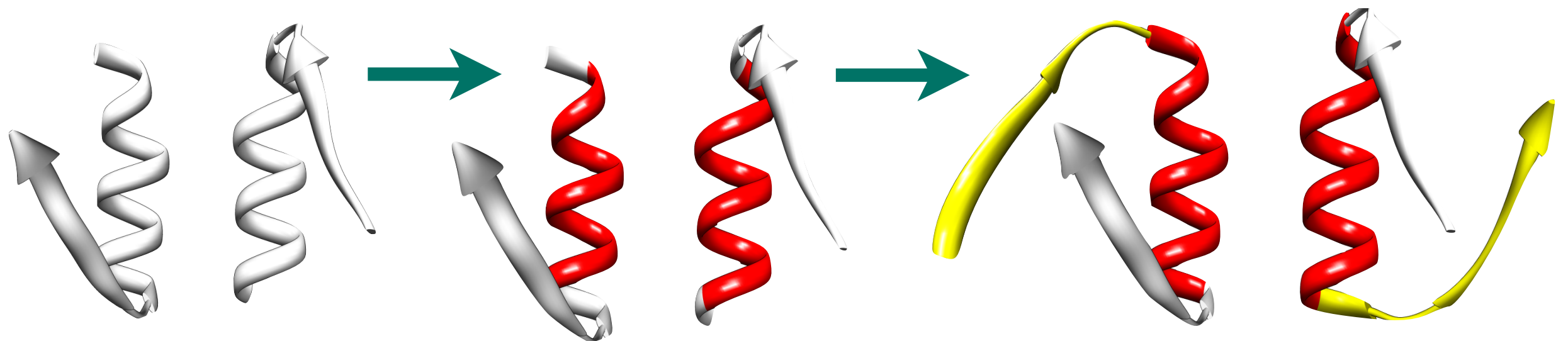
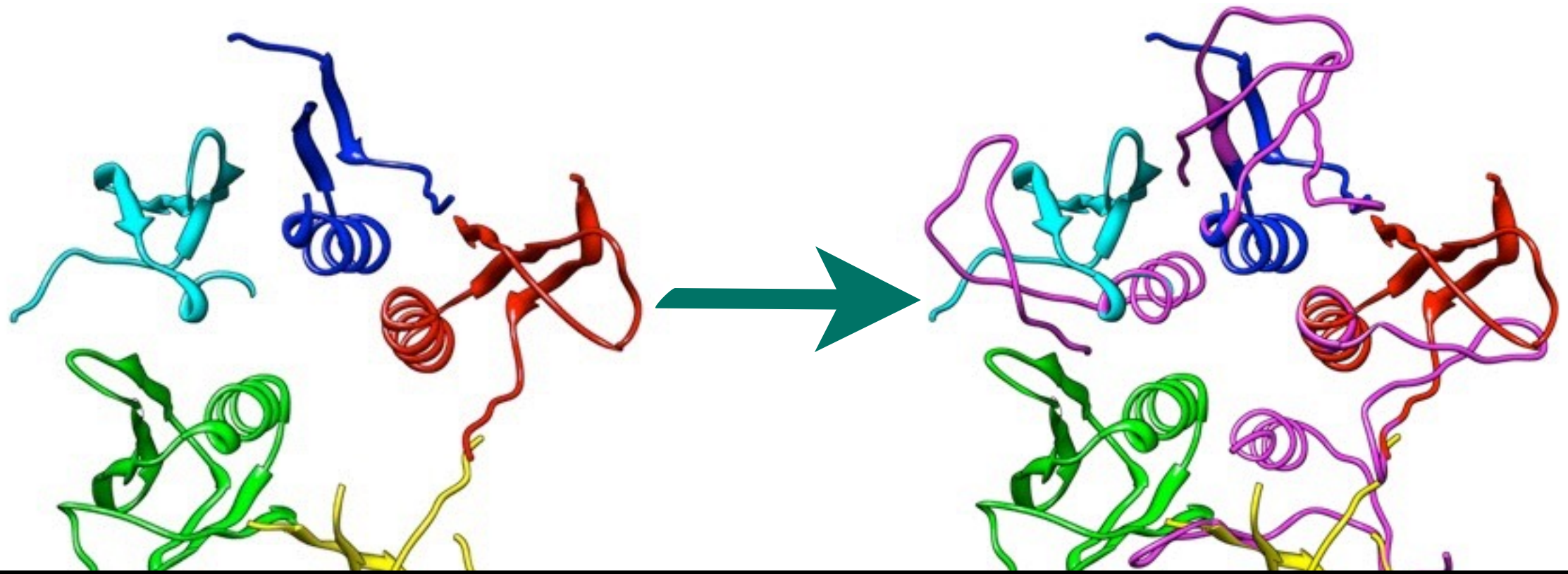
Non-Crystallographic Symmetry occurs if there are multiple subunits in the asymmetric unit of a crystal and some of them adopt almost the same tertiary structure:



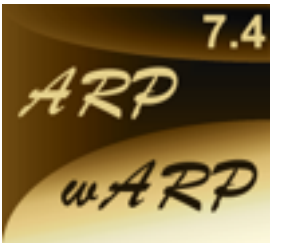
Basically like dogs!

NCS Detection and Extension

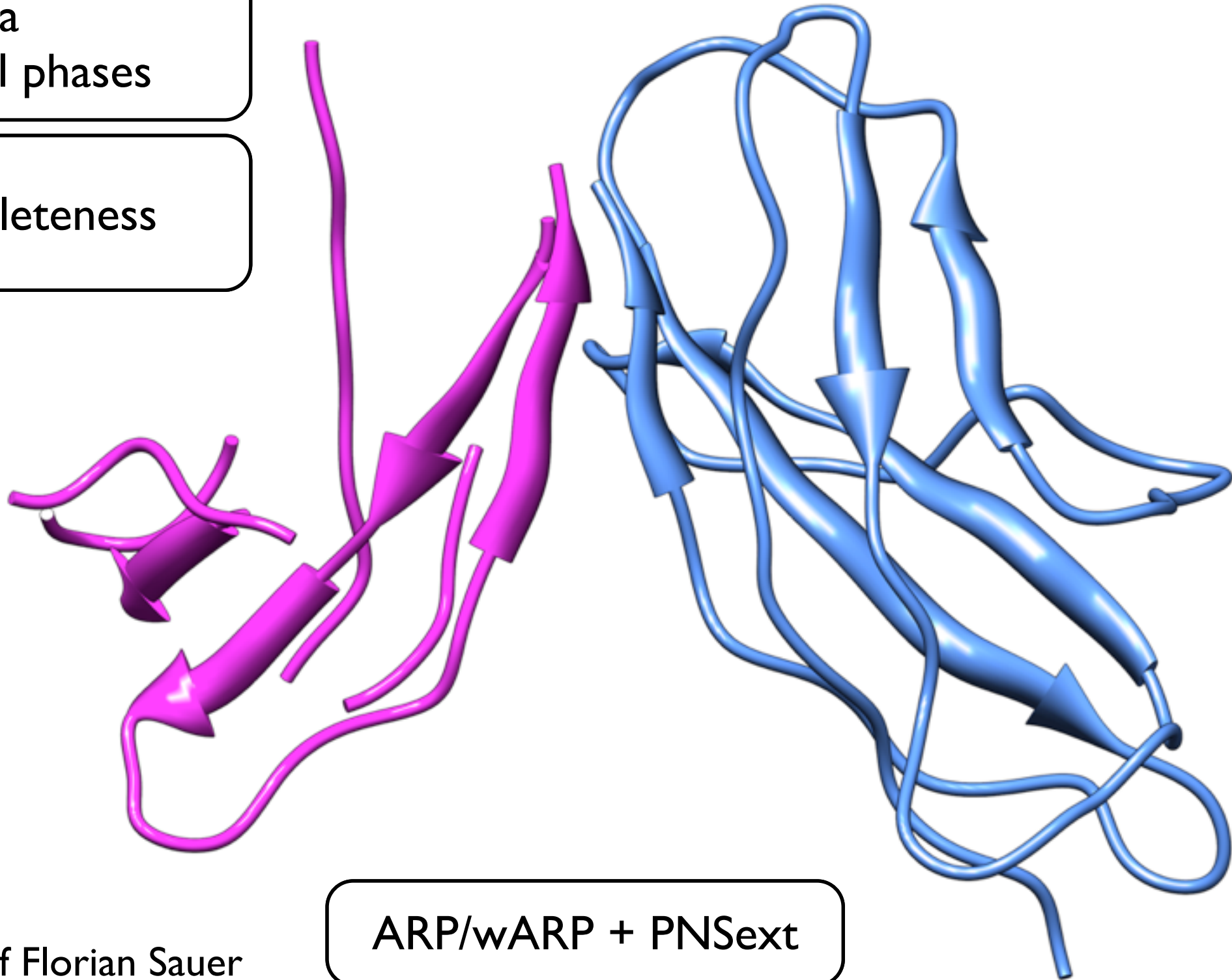
We can use this information to extend fragmented models!



NCS Detection and Extension



- 1.95 Å data
 - poor initial phases
-
- 75% completeness



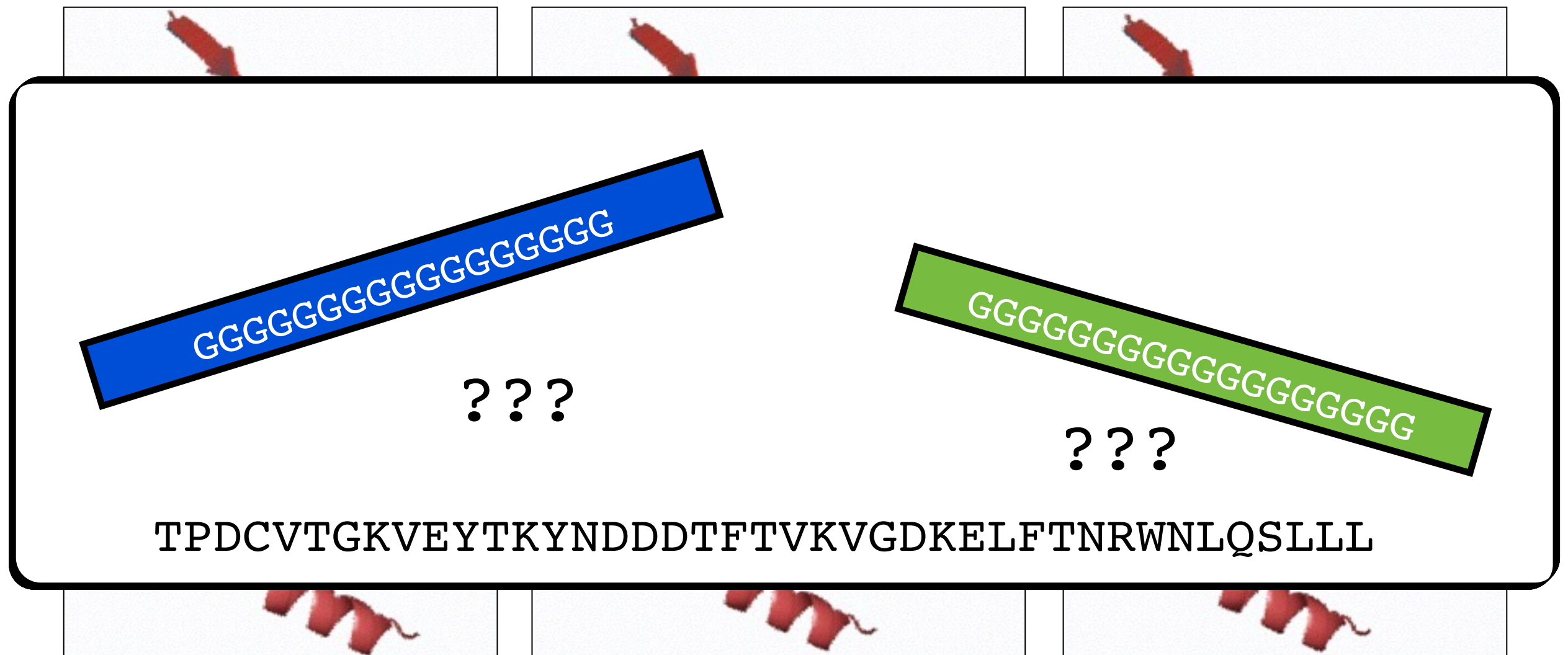
ARP/wARP + PNSext

Data courtesy of Florian Sauer

Modelling Loops With *a priori* Knowledge

Chain fragments are built and docked to sequence

- using knowledge of angles between anchors
- check with density to see if modeled loops are valid

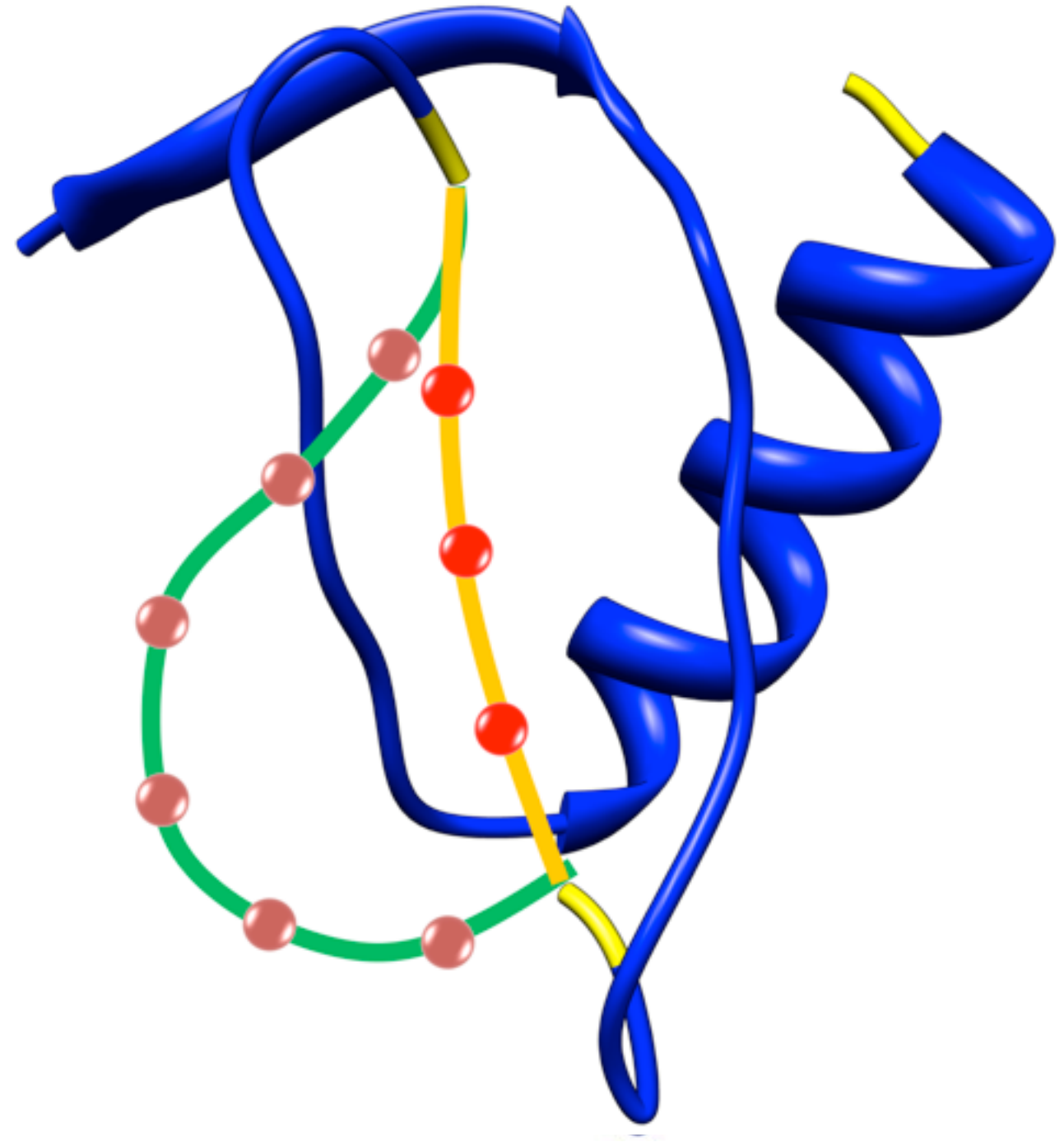


Modelling Loops With *a priori* Knowledge

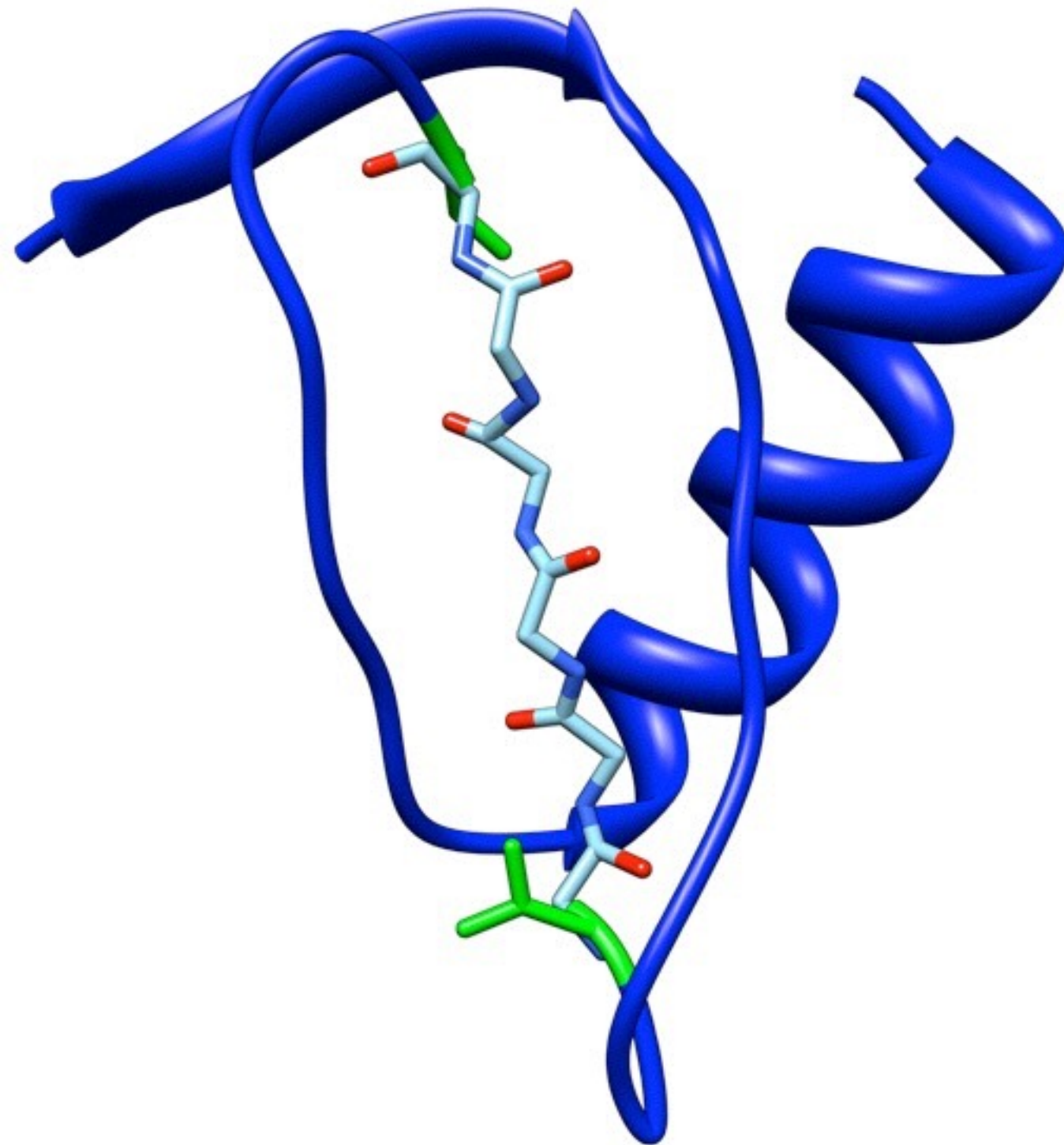


When no sequence has been docked - no loop building!

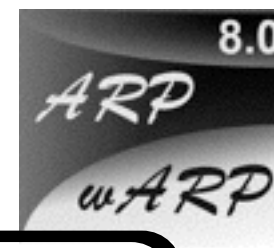
- Search for missing fragments in a structural database
- In development together with Marco Biasini (Schwede group, SWISSmodel)
 - Using the given density information



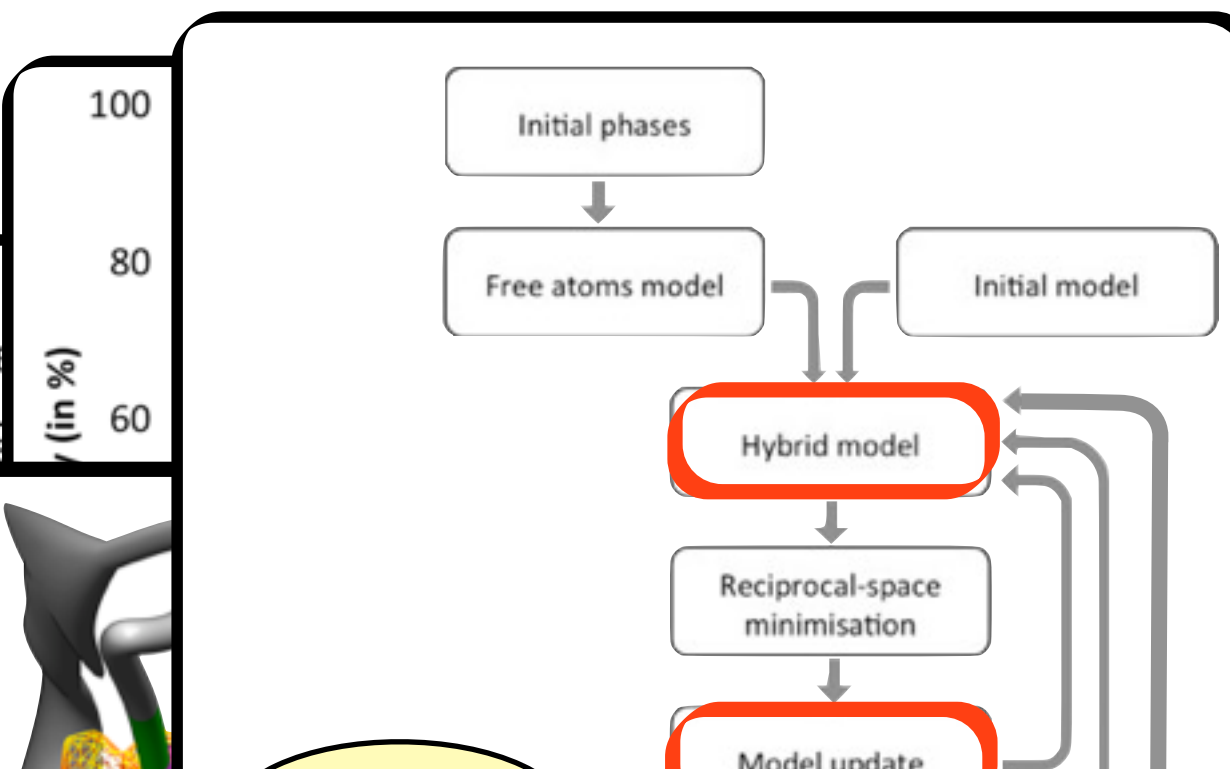
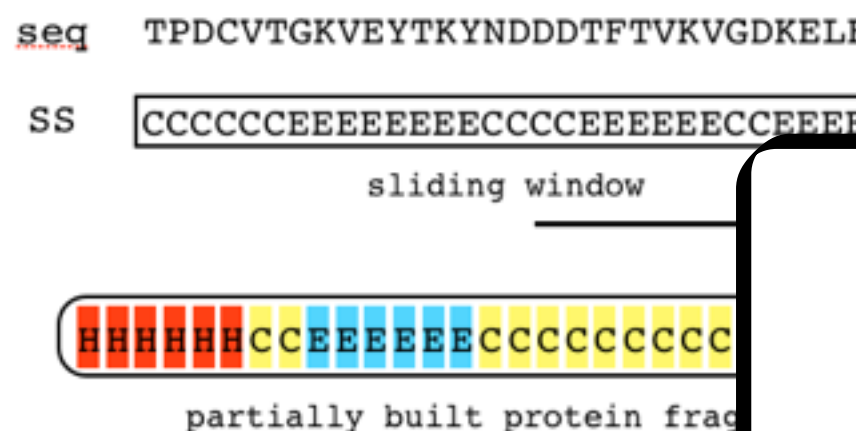
Modelling Loops With *a priori* Knowledge



Modelling Loops With *a priori* Knowledge



Docking results



10 protein test structures with resolution 3.0 to 3.8 Å, up to 300 residues

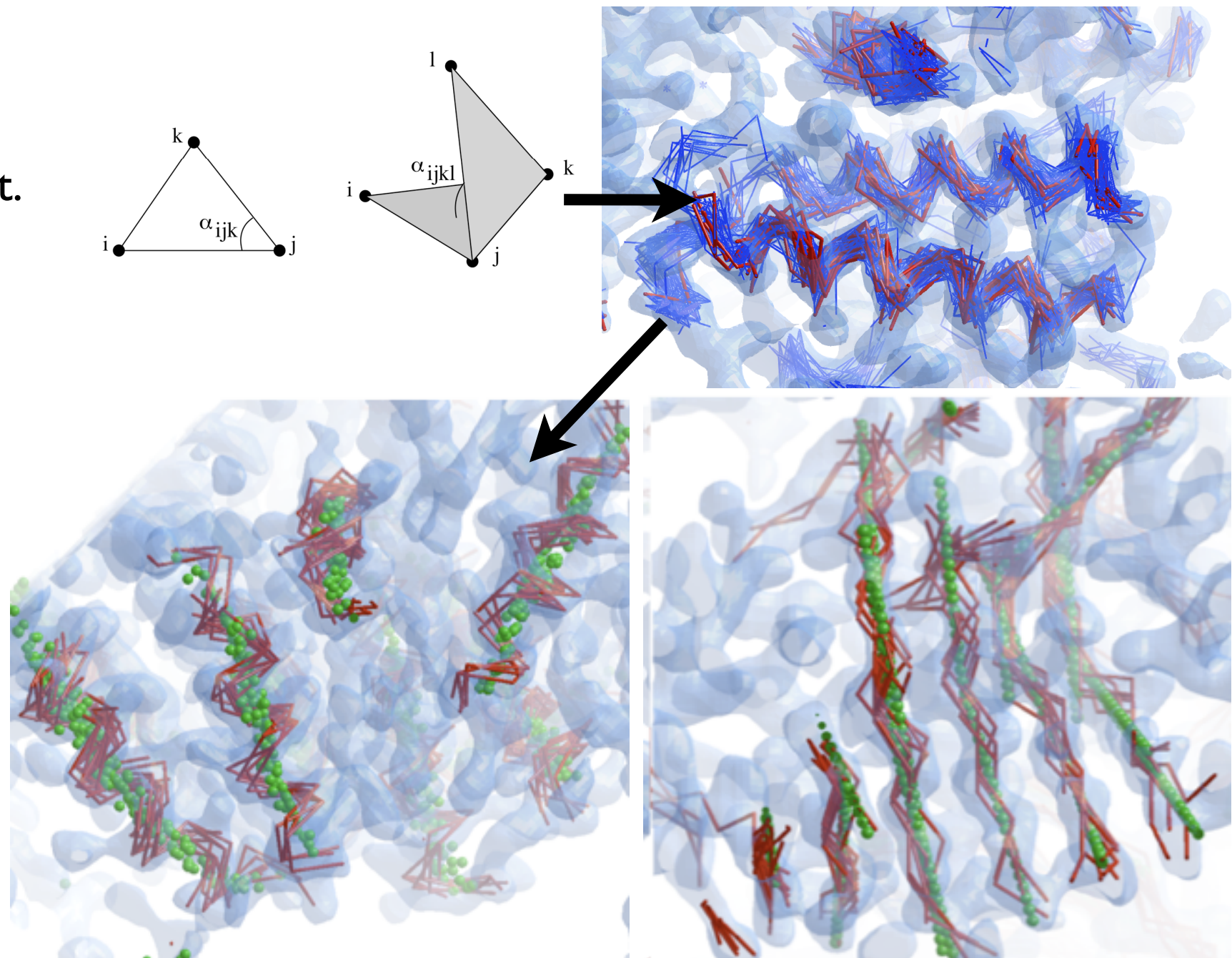
	Model Completeness	Residues / Fragment	Sequence Coverage
Top Results	+ 12%	+ 98%	+ 59%
Average	+ 4%	+ 25%	+ 7%

Secondary structure modelling

At low resolution, one can try to find **secondary structure elements** on the map:

- Short helix/strand fragments (3 to 5 C α candidates) are built.
- Longer traces are formed and the best are kept (in red)
- Traces are clustered
- Assemblies are averaged

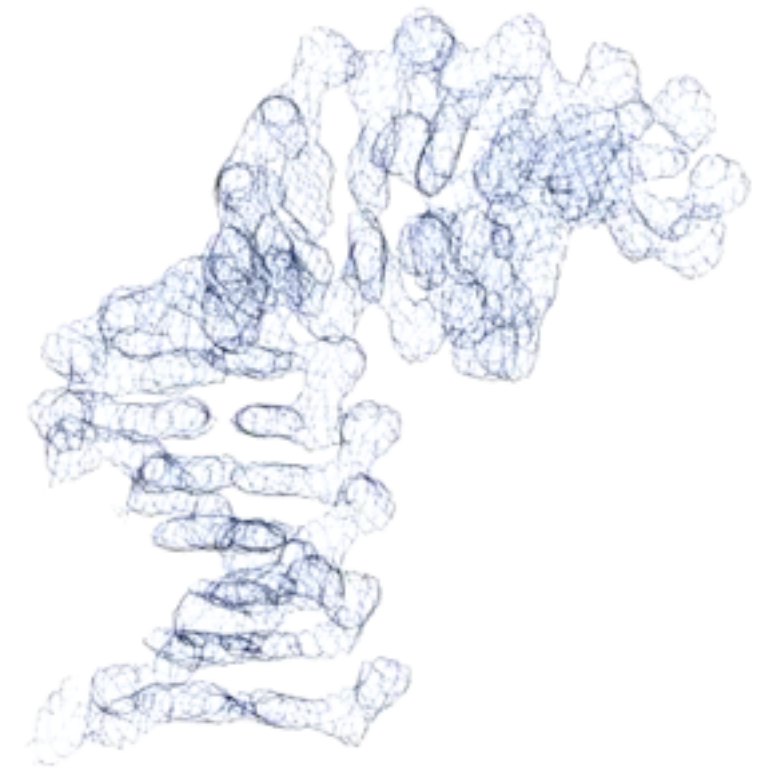
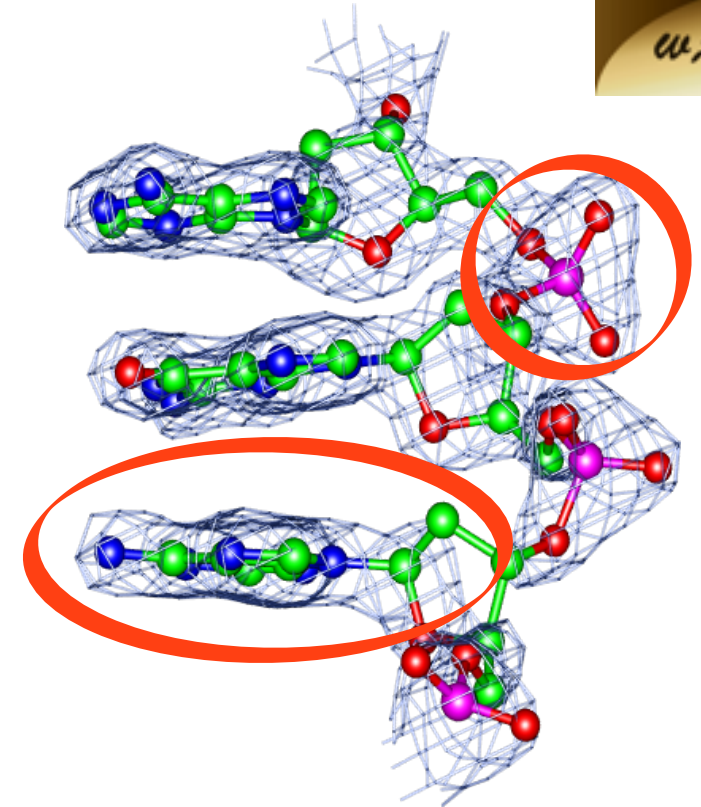
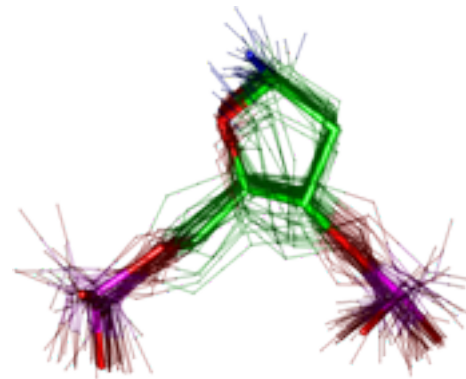
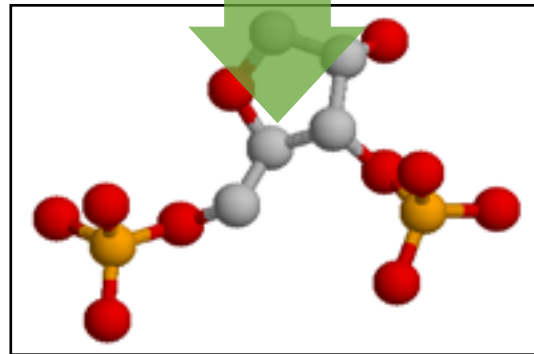
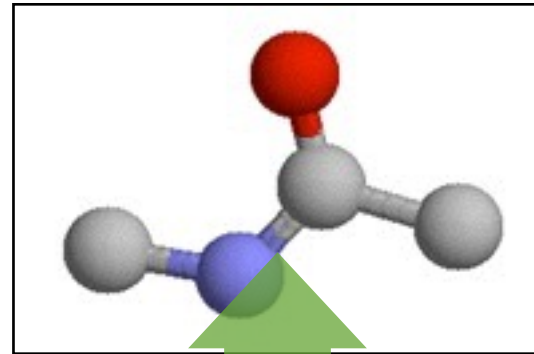
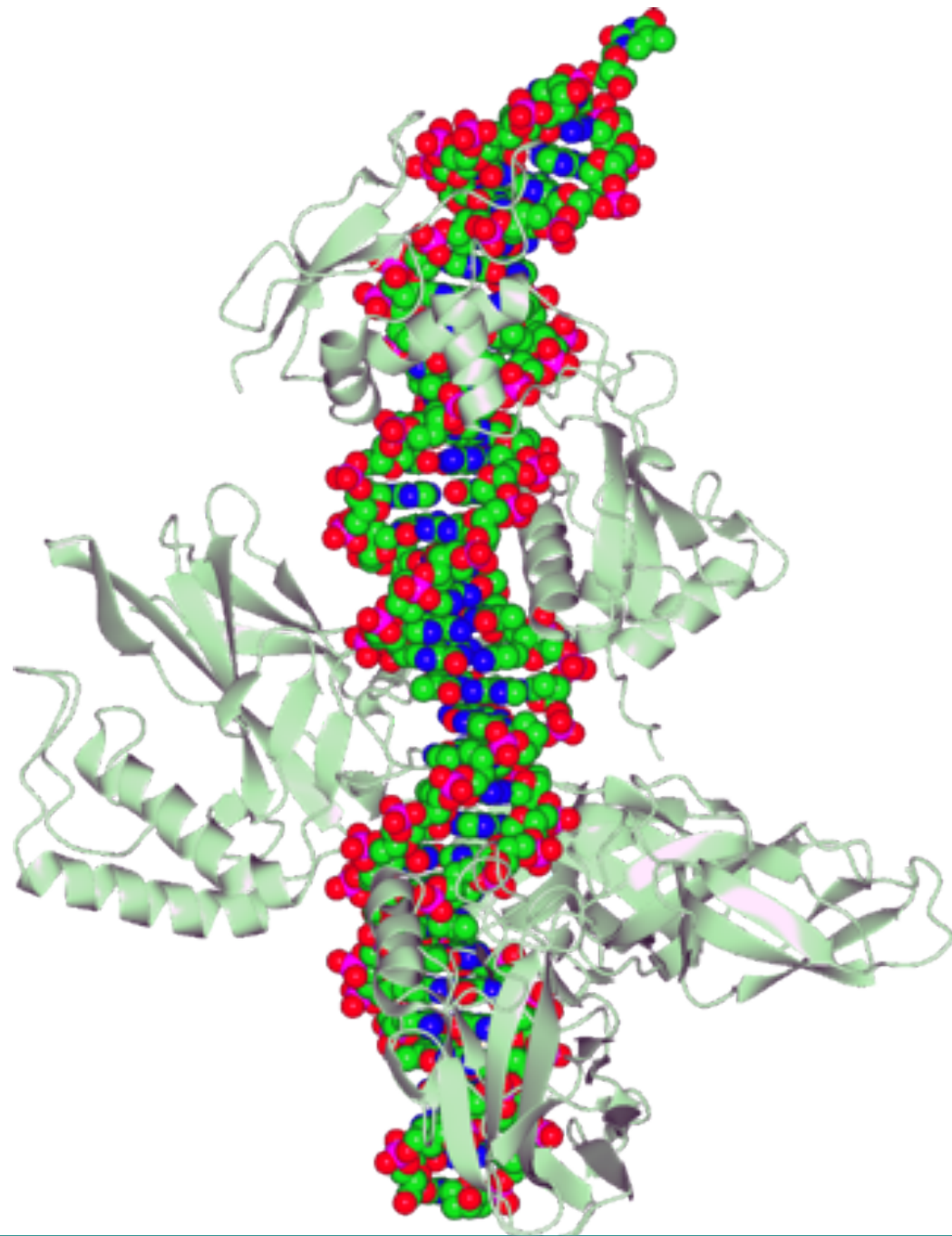
Good to evaluate map quality!



Tracing RNA/DNA Chains

ARP/wARP is just not about building protein models, it can also build **nucleic acids**:

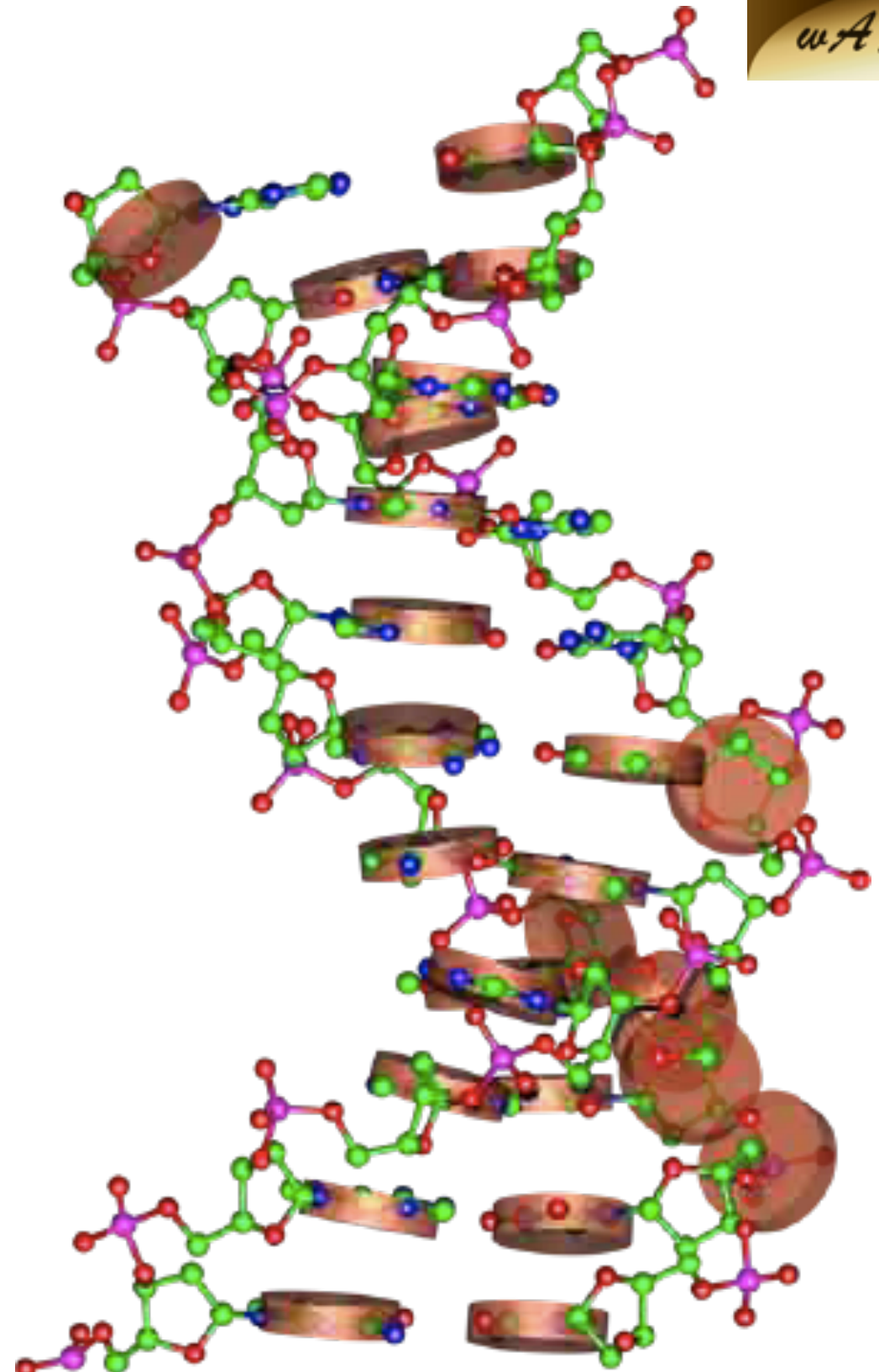
7.4
ARP
wARP



Tracing RNA/DNA Chains

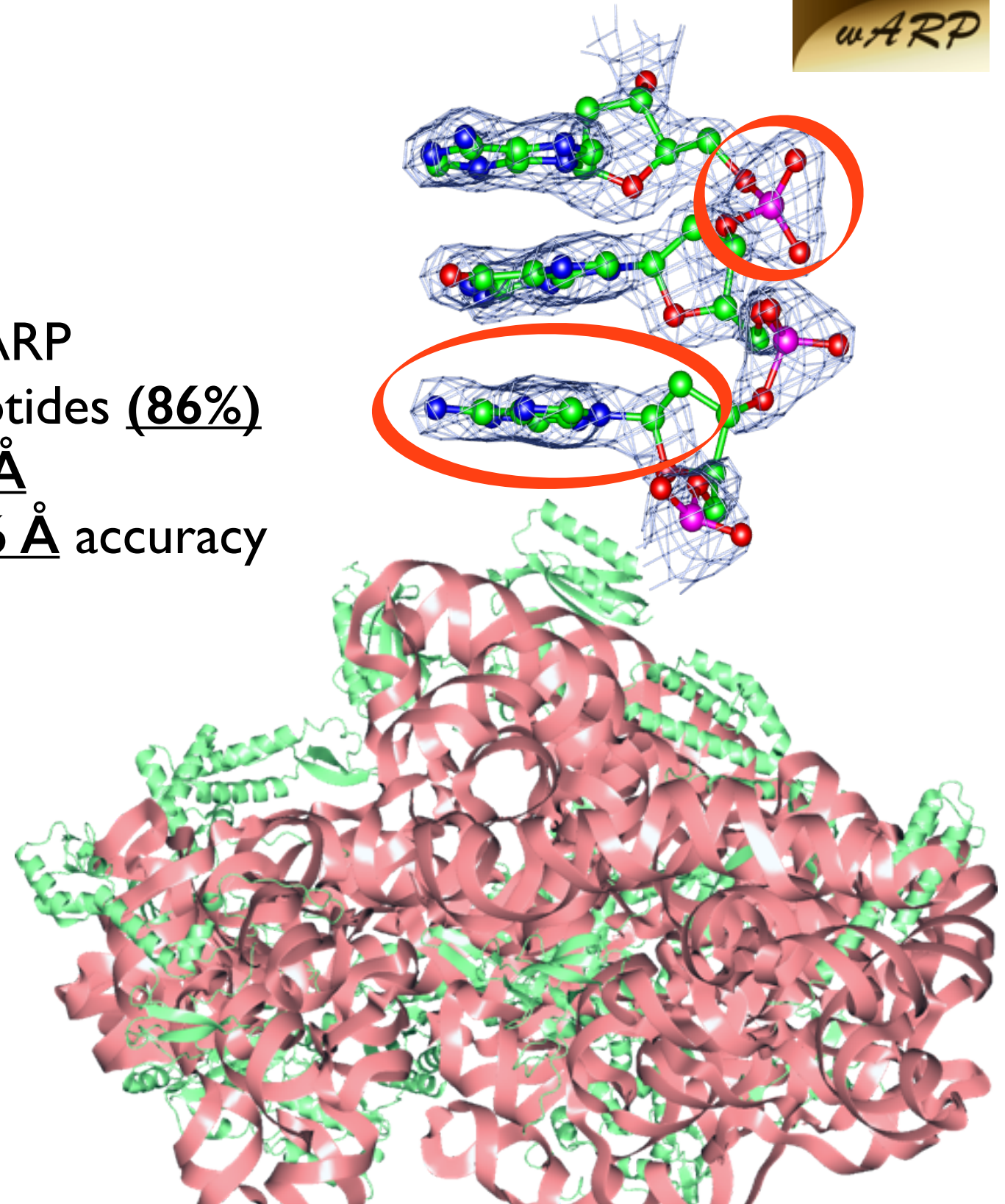
- Detection of
 - phosphate backbone and
 - nucleobase planes
- using pattern recognition algorithms

1p7h (2.5 Å)	
Completeness	0,88
Noise level	0,69
Positional r.m.s.d.:	0.5 Å
Angular Difference	5°

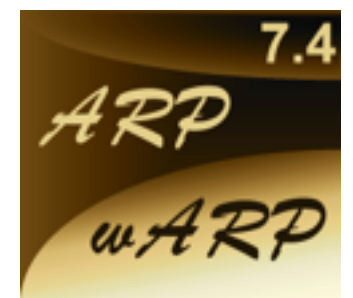
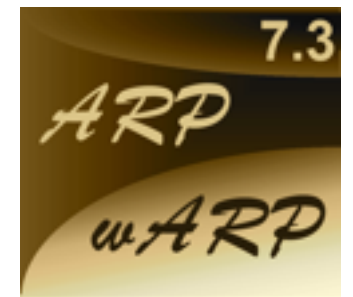
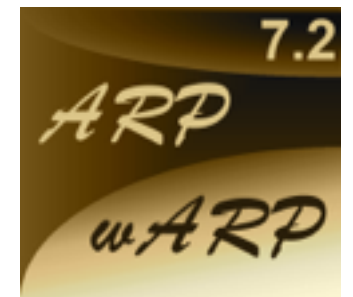
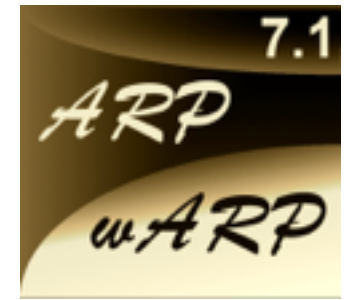
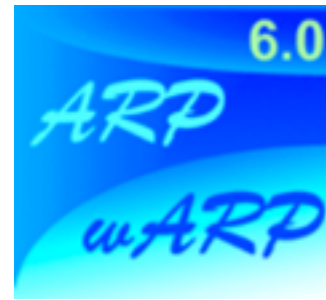


Tracing RNA/DNA Chains

- The 30S ribosomal subunit
 - Resolution: 3.05 Å
 - Experimental phases (MCC 0.73)
- Automatic model building with ARP/wARP
 - modelled 1,302 out of 1,513 nucleotides (86%)
 - backbone r.m.s.d. to reference: 0.7 Å
 - Located 1,121 nucleobases with 0.6 Å accuracy in location and 12° in orientation
- Backbone fragmented in 75 chains
- Built in around 6h
- (cf. several months of manual work)



What can I find in ARP/wARP right now?



Changes in Version 7.4

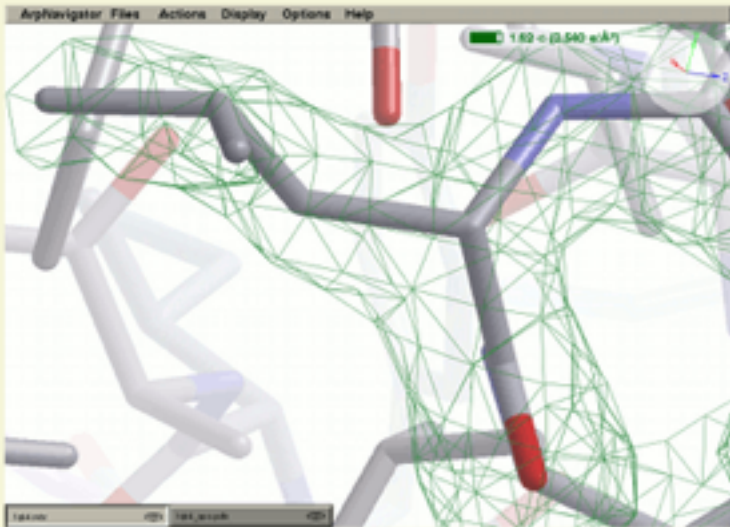
- Accuracy of estimated model correctness has been improved
- Updated method for fitting of partial ligands
- “Fit Ligand” incorporates 84 most common ligands now
 - and uses cif files defining bond, torsion and plane restraints

Where can I find it?

<http://www.embl-hamburg.de/ARP/>



ARP/wARP Copyright 2013
by European Molecular Biology Laboratory



Click on the images above to zoom.

ARP/wARP: Crystallographic Macromolecular Model Building Version 7.4

Builds proteins, RNA/DNA, secondary structure, side chains, loops, solvent and ligands

Download available for Mac and Linux.

Model building also available over the web.

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[View User Guide \(html\)](#)
[Download Tutorial \(pdf\)](#)

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HELP

[Send email to ARP/wARP developers](#)
[Frequently Asked Questions](#)

Useful Links

Auto-Rickshaw	Automated Crystal Structure Determination Platform
ViCi	In-Silico Ligand-Based Drug Design
CCP4	Software for Macromolecular X-Ray Crystallography
Refmac	Macromolecular Refinement Program

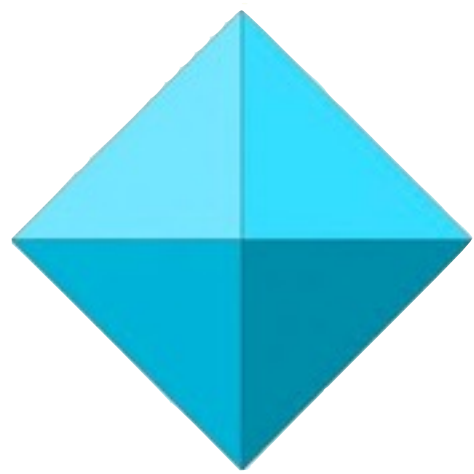
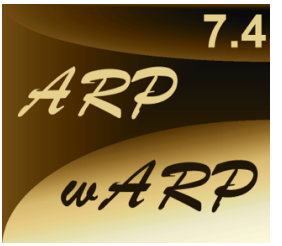
Only for Mac
or Linux!!



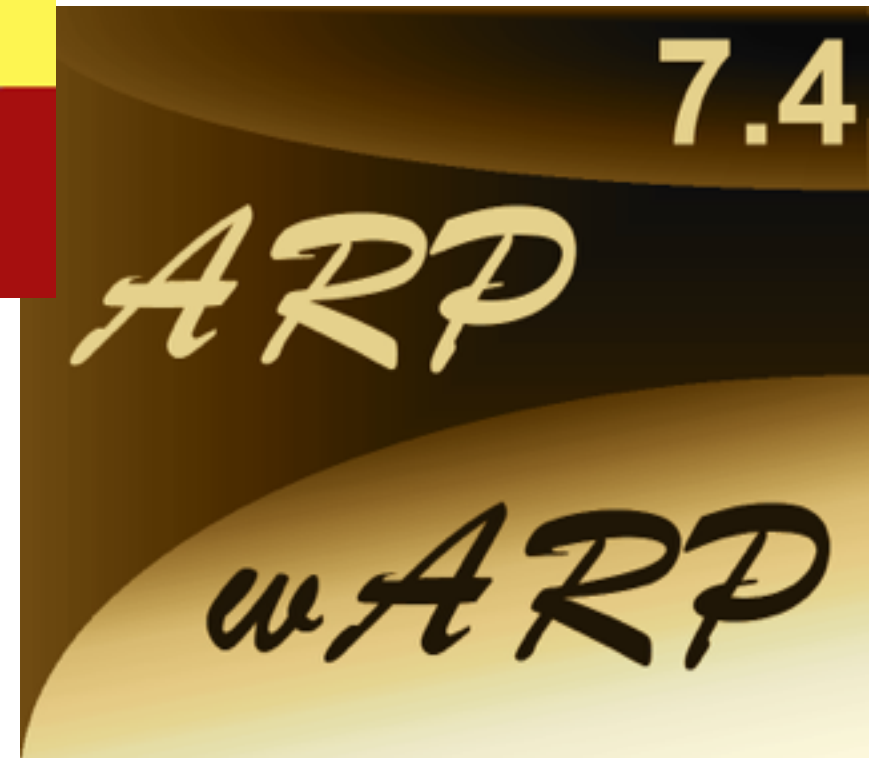
Good for
Windows
users!!



Collaboration with CCP4



CCP4



- Joint release
 - CCP4 6.4 and ARP/wARP 7.4
 - one installer, no worries ;)

ArpNavigator

Last year, we made the cover of Acta Crystallographica D because we launched **ArpNavigator**:

- The primary intuitive interface for interacting with the automated model building processes
- Real time visualisation of model building
- Can be used by both experienced and novice users
- It controls the setup of computational tasks and displays the intermediate steps
- Allows model visualisation, map quality check and figure production

Wait for the tutorial!

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Part 4

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Ostertagia ostertagi ASP-1

dsDNA (6–4) photoproduct

Cytoplasmic domain of
Vibrio cholerae TcpE

Teicoplanin–cell-wall peptide

Caldicellulosiruptor bescii
family 3 pectate lyase

Amyloid zipper groups

SyCH–YopH complex

Wheat cyclophilin TaCypA-1

ω -Aminotransferases

Human cystatin C mutants

PSD-93 interaction partners

SET7/9 AdoMet complexes

Human GLTP dimerization

Vaccinia virus poly(A)
polymerase VP55

Bulk-solvent and overall
scaling

Brazzein

Fatty acid double-bond
hydratase

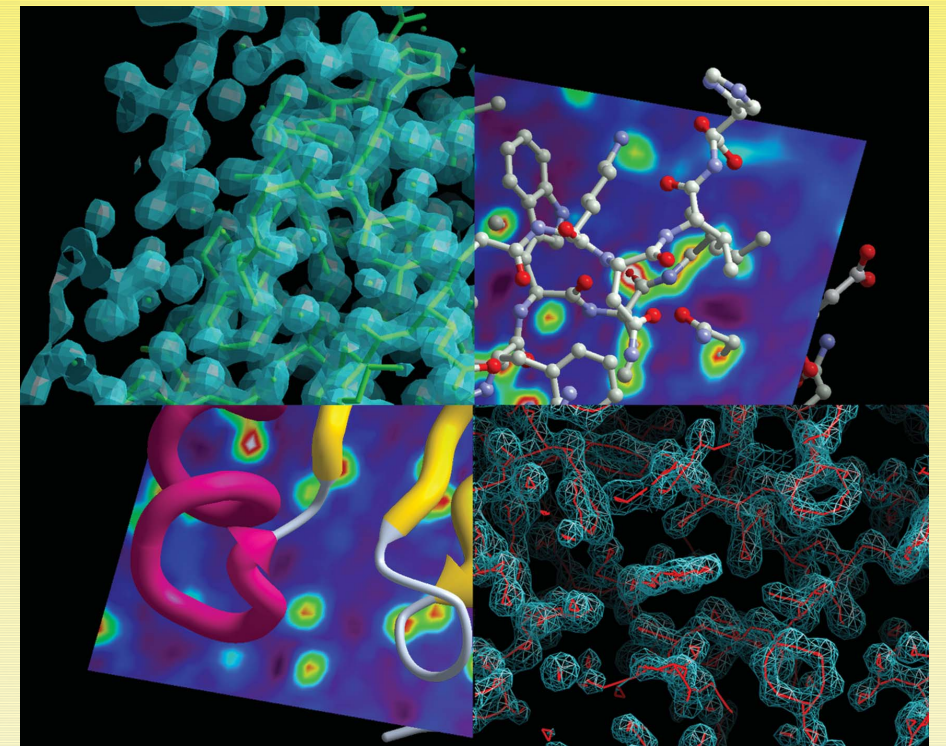
ADP-L-glycero-D-manno-
heptose 6-epimerase

Nm23-H1



Acta Crystallographica Section D **Biological Crystallography**

Editors: E. N. Baker and Z. Dauter

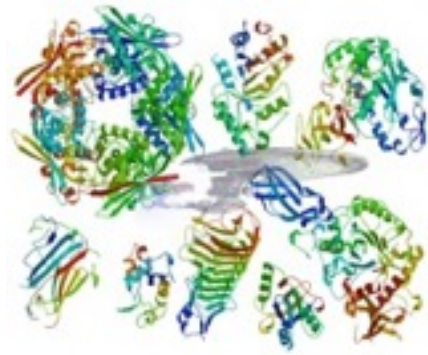
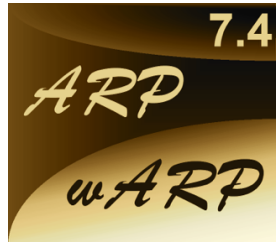


Model-viewing options in *ArpNavigator*, p. 635

journals.iucr.org

International Union of Crystallography
Wiley-Blackwell

Tutorial content



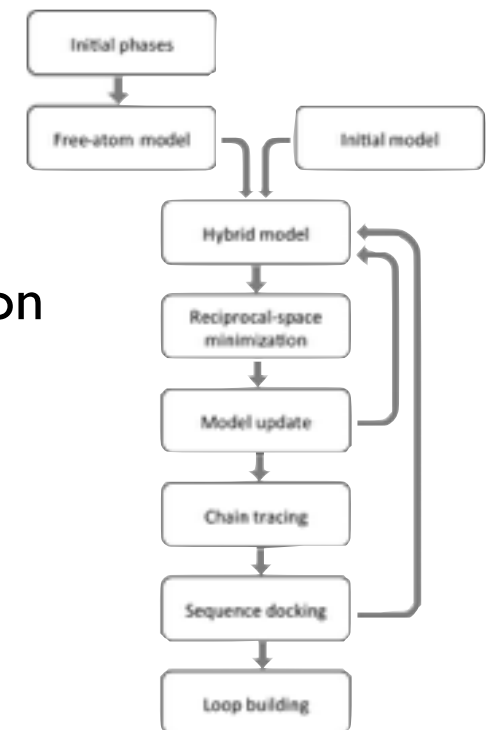
Iterative protein-model building

ARP/wARP – Perrakis *et al.*, (1999)

'Free atoms' – Agarwal and Isaacs (1985)

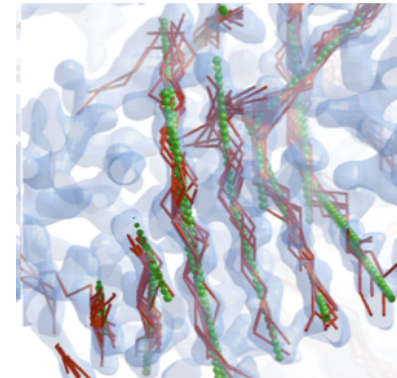
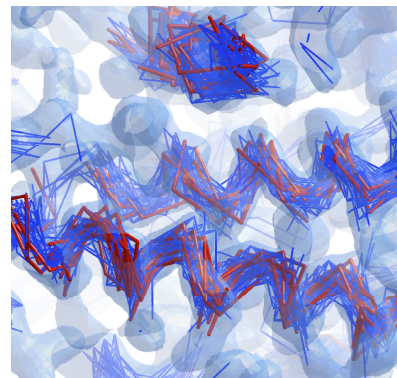
REFMAC5 – Murshudov *et al.*, (1997)

Model extension by automatic NCS detection

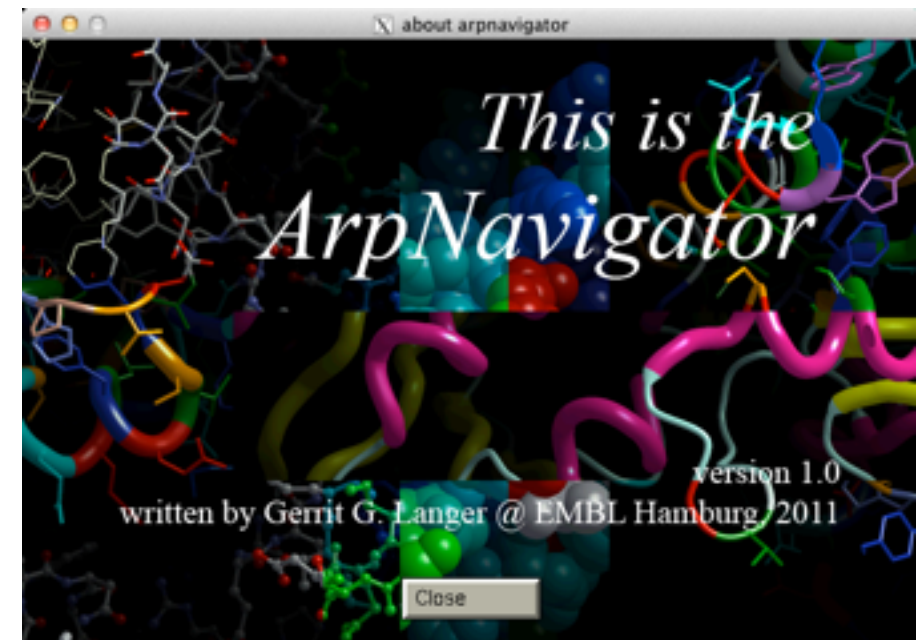


Recognition of secondary structure:

fast approach for helices and sheets at low resolution ($< 4.0 \text{ \AA}$)



... and all this in the **ArpNavigator**



The people - The power!



Collaborators

EMBL Hamburg: Gleb Bourenkov,
Santosh Panjikar

MRC Laboratory, Cambridge :
Garib Murshudov's group

**Rutherford Appleton
Laboratory:** CCP4 team

Previous Members

Ciaran Carolan, Tim Wiegels

Funding



If you are ever in town



The EMBL

EBML Heidelberg (“Main Laboratory”)

- Gene Expression
- Cell Biology
- Structural and Computational Biology
- Developmental Biology

EMBL Monterotondo (“Rome”)

- Mouse Genetics

EMBL Grenoble

- Structural Biology

EMBL / EBI Hinxton (“Cambridge”)

- Bioinformatics

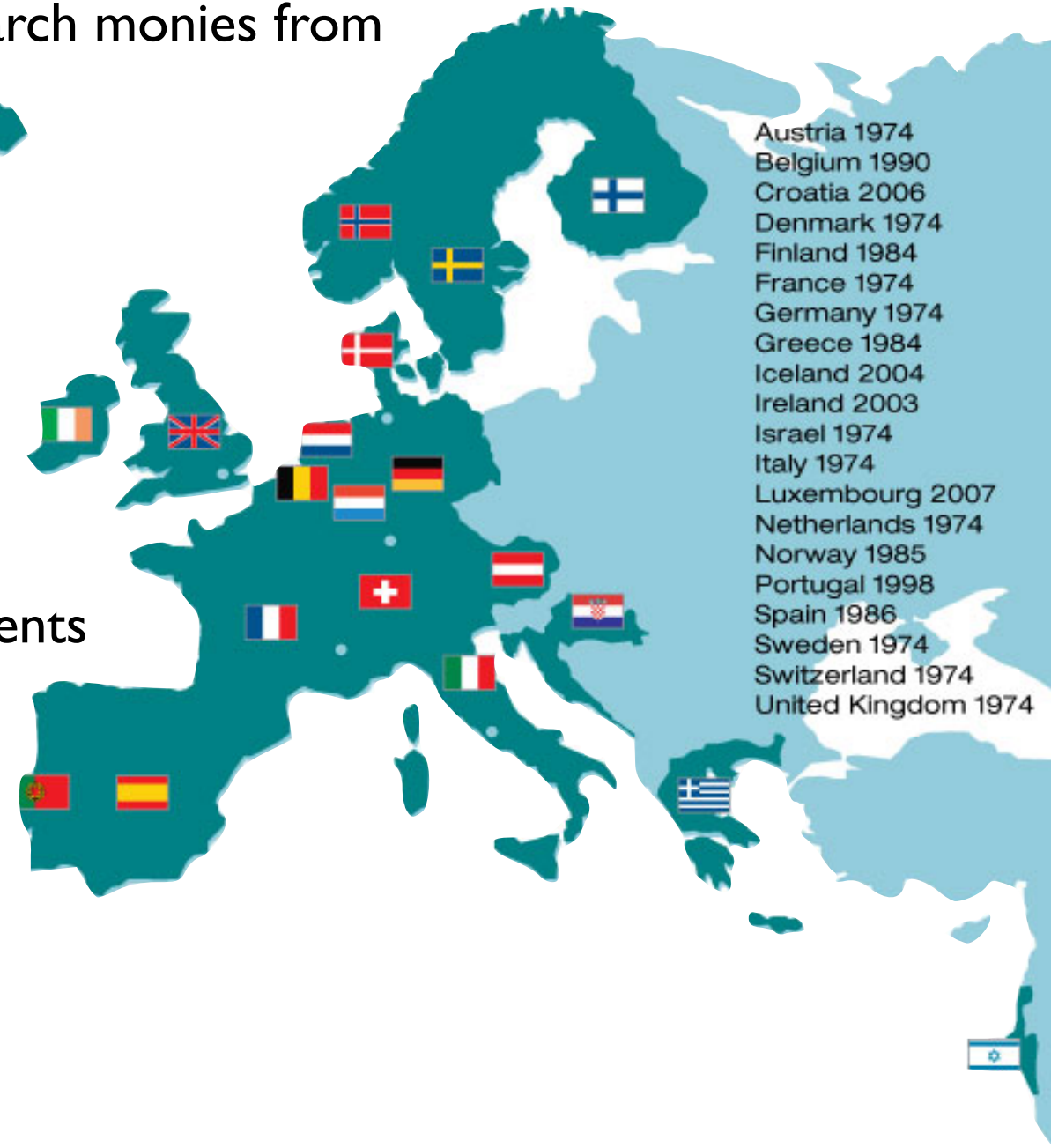
EMBL Hamburg

- Structural Biology



The EMBL

- Established in 1974 to reflect the need for a European centre for research and training in molecular biology
- A basic research institute funded by public research monies from 20 member states
- Over 1300 employees from 60 nations
- Approximately 80 independent research groups



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- Advanced Training at all Levels
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