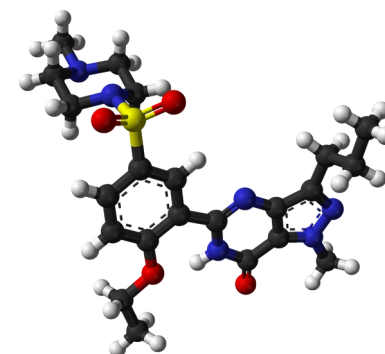
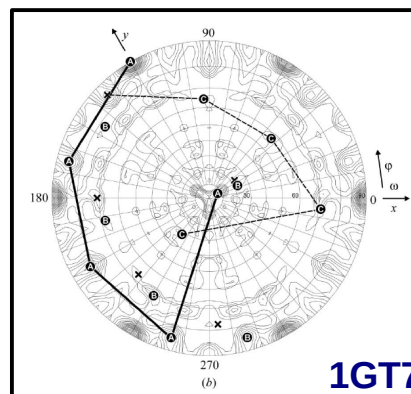
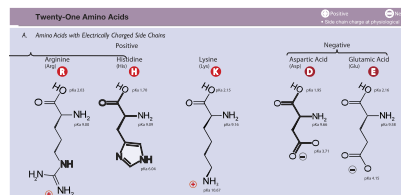


Refinement: The BUSTER perspective

Clemens Vonnrhein
Global Phasing Ltd.

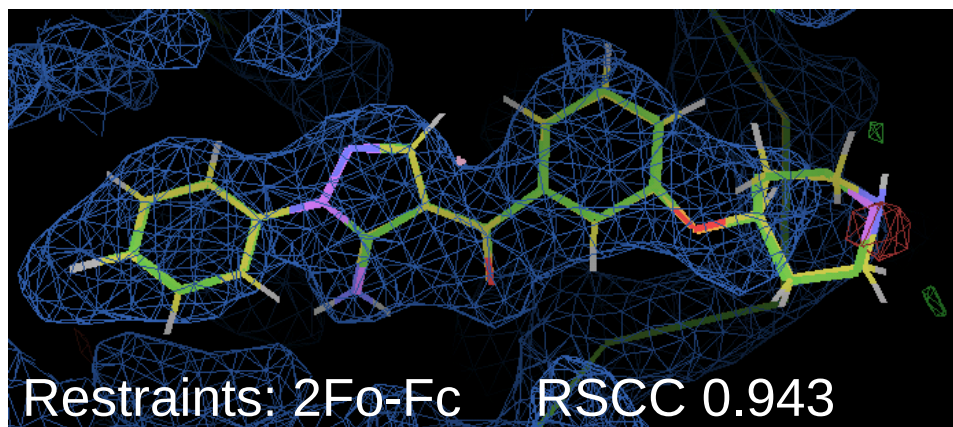
DLS/CCP4 2024

- ❑ Combines prior knowledge and observed (X-Ray) data
- ❑ Chemistry (bonds, angles, ...)
 - Protein (Engh & Huber)
 - Compound/Ligand
 - ...
- ❑ Similarity
 - Within the crystal (NCS)
 - To other structures (“targeting”)
- ❑ Occupancy, disorder (alternate conformations), ...



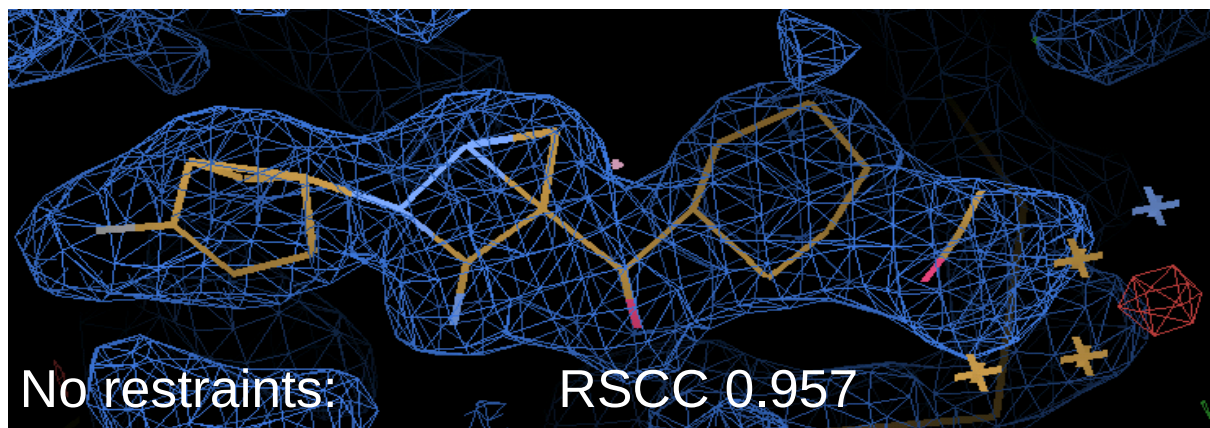
Prior knowledge is essential

2bal kinase 2.1Å resolution.



RSCC is a poor validation criteria here: it tells us that ligand model and map agree, but not if we have a good ligand model that also follows prior chemical knowledge

→ pick your quality criteria carefully!

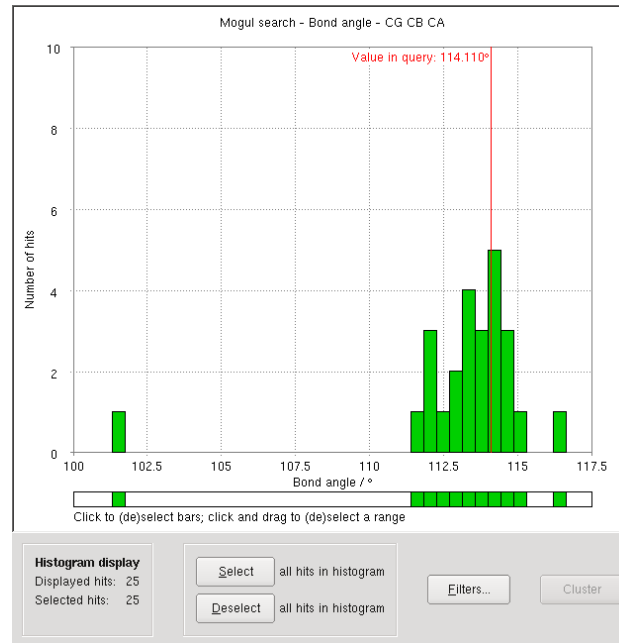
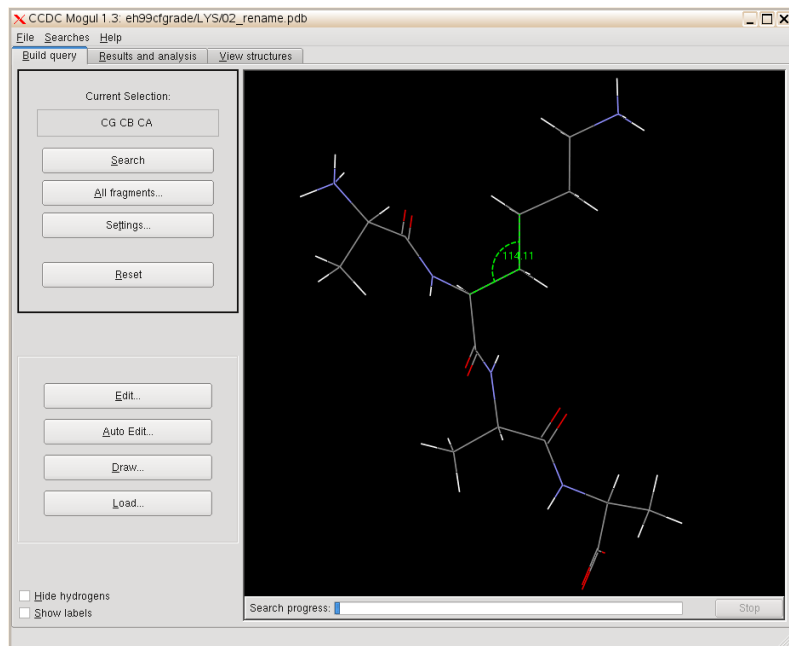


RSCC = real-space correlation coefficient



Ligand dictionaries based on CSD grade.globalphasing.org

- Grade (2011): use CCDC **mogul** program to survey CSD



- Use CSD as **source of information for restraints** (not only in validation)
- Also: AceDRG (Long et al, 2017: COD), eLBOW (Moriarty et al, 2011: CSD)

Grade2 (rewrite, July 2021):

<https://www.globalphasing.com/buster/manual/grade2/manual/>

Restraints - not just for ligands

❑ Standard geometry restraints for bonded atoms

- BOND (1-2)
- ANGLE (1-3)
- TORSION (1-4)
- CHIRAL
- PLANE

❑ Non-bonded atoms

- anti-bumping restraints

❑ B-factor restraints

- bonded atoms
- isotropic/anisotropic B-factor

❑ Similarity restraints

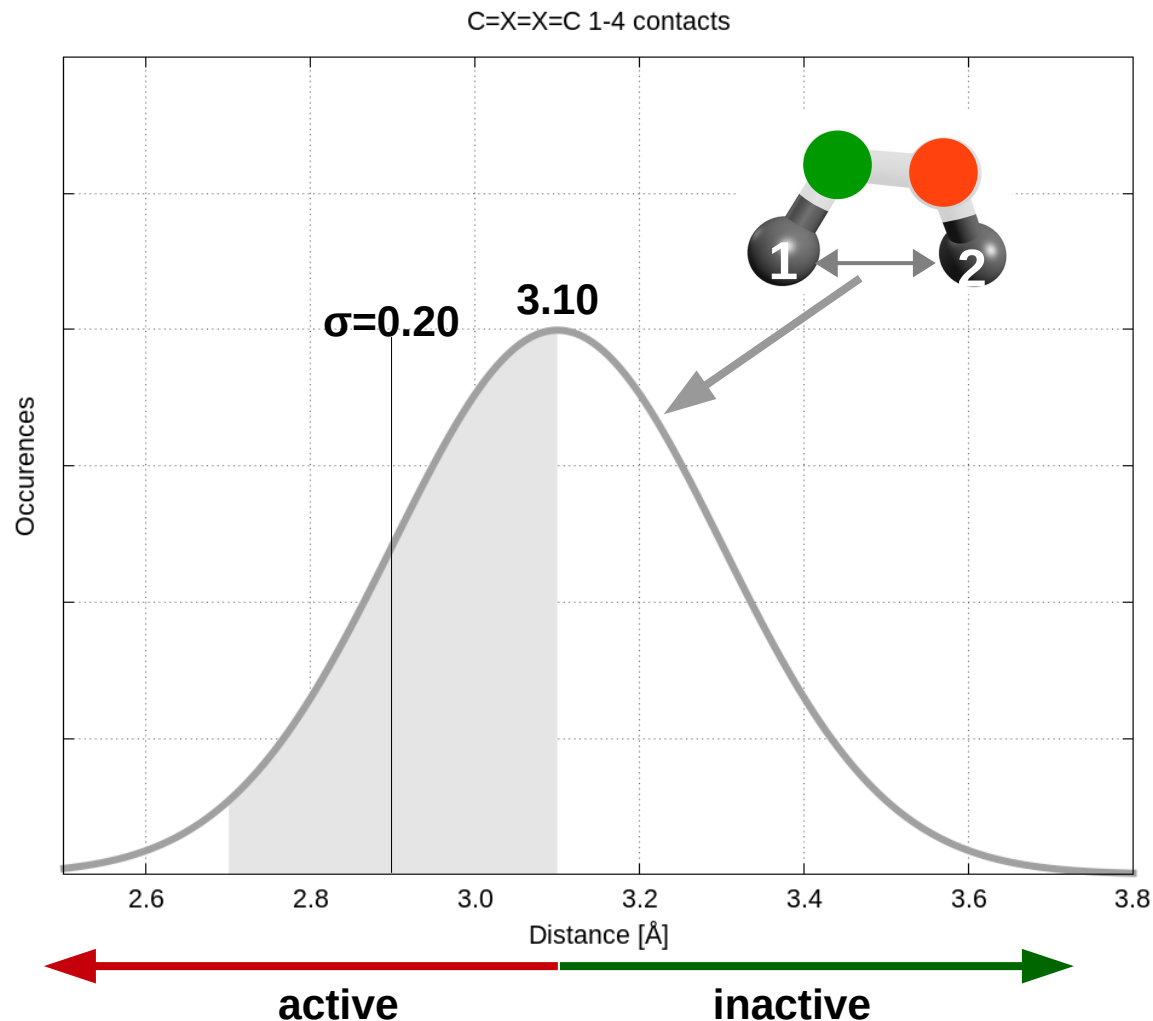
- NCS
- targeting

❑ Conformational restraints

- *Ramachandran*
- *rotamer*



Contact restraints for 1-4 connected atoms - V1



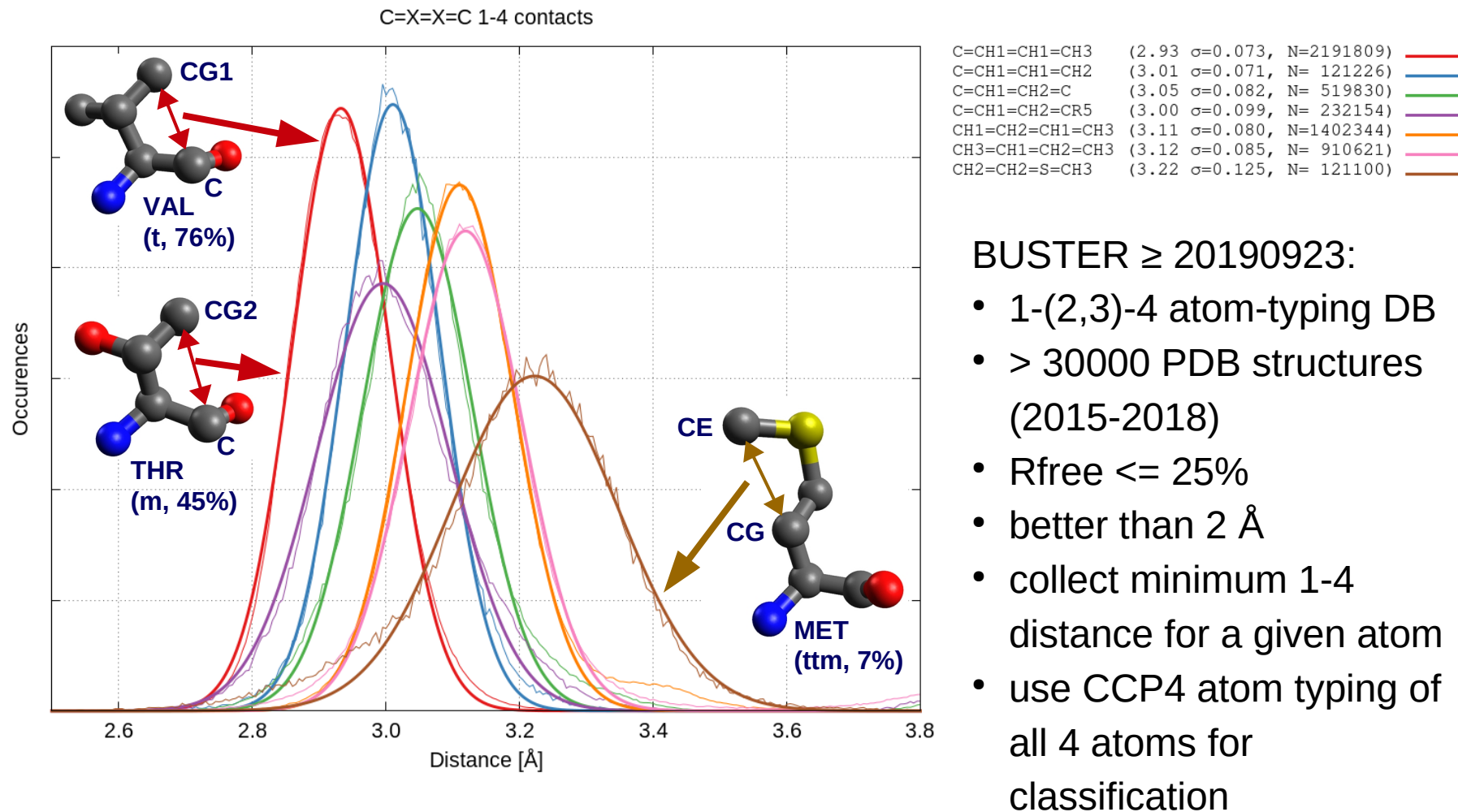
only first and last atoms of
1-4 (torsion) contact taken
into account

BUSTER pre-20190923:
 $\text{vdW}_1 + \text{vdW}_2 + \text{Dinc}_1 + \text{Dinc}_2$

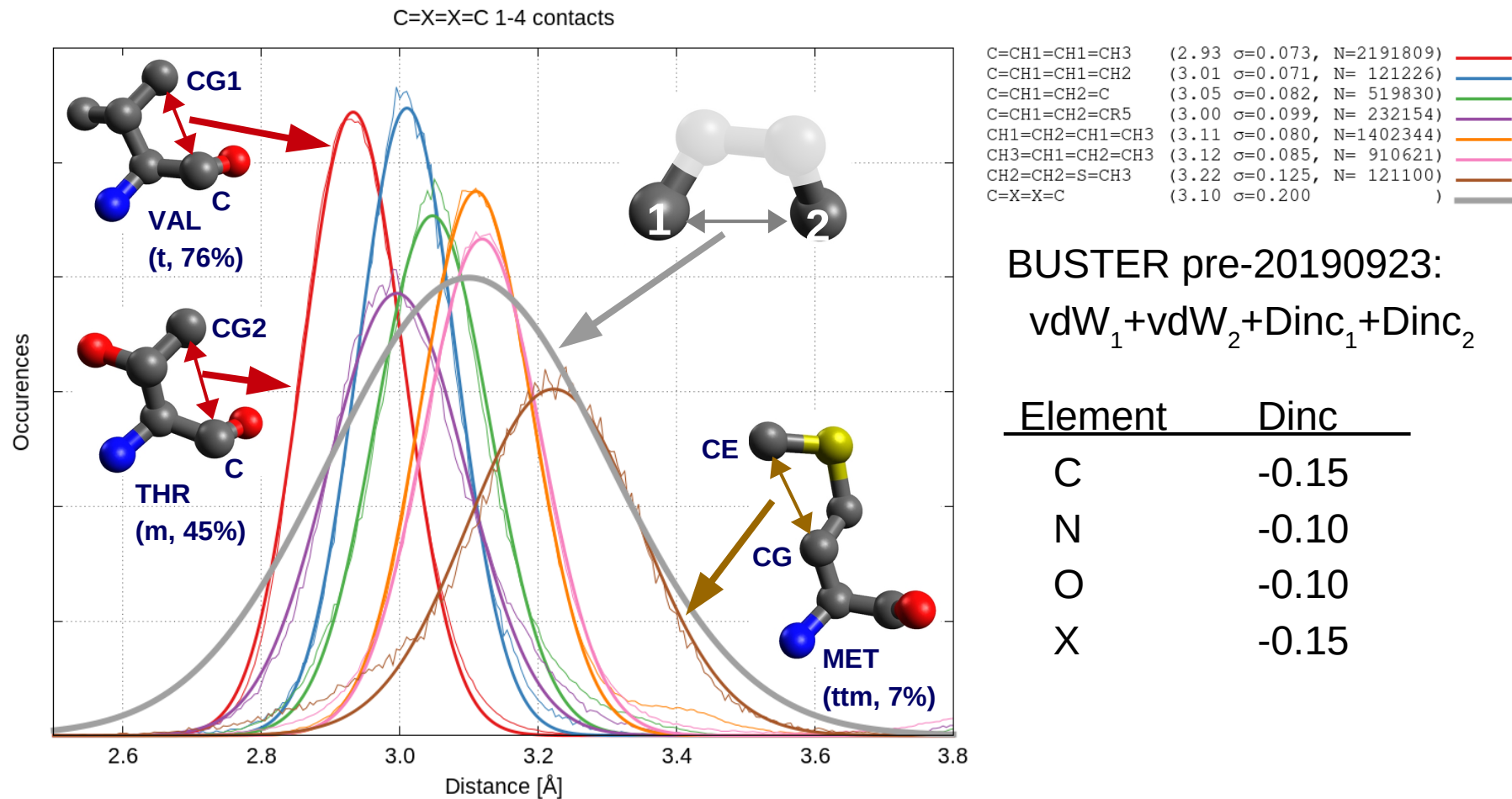
Element	Dinc
C	-0.15
N	-0.10
O	-0.10
X	-0.15

fixed sigma (0.20)

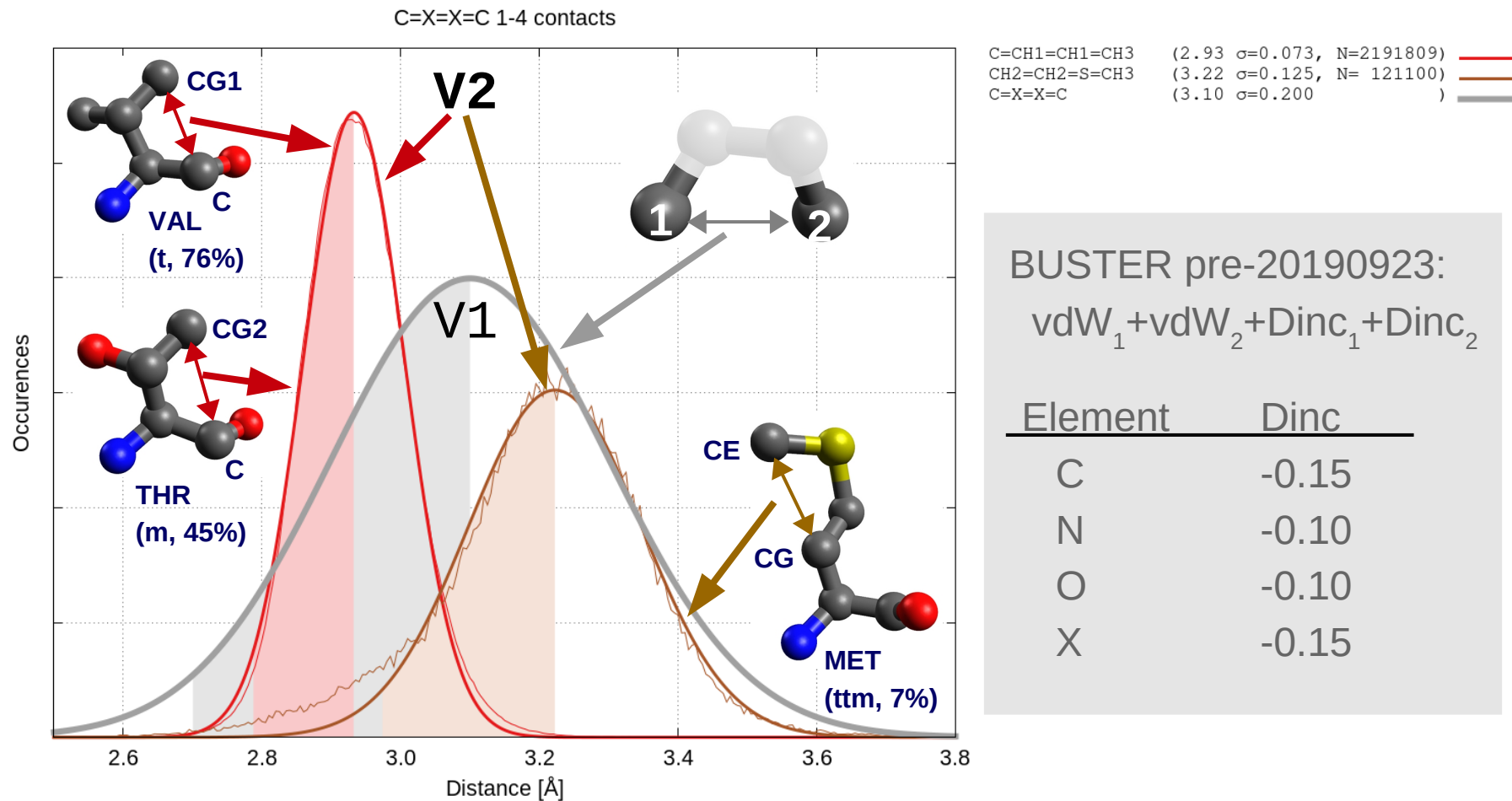
Contact restraints V2



Contact restraints V1 vs V2

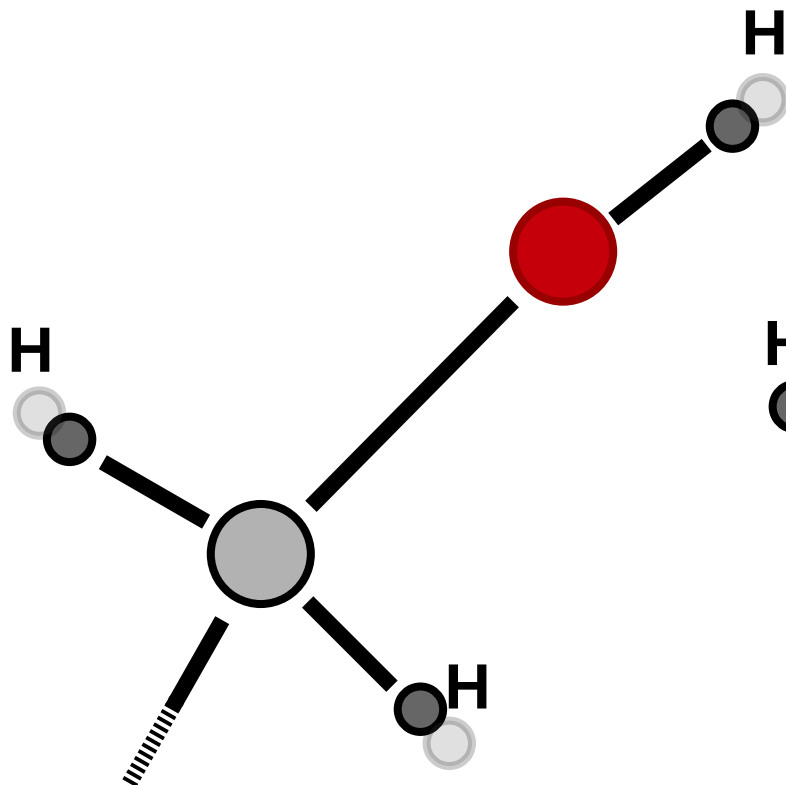


Contact restraints V1 vs V2

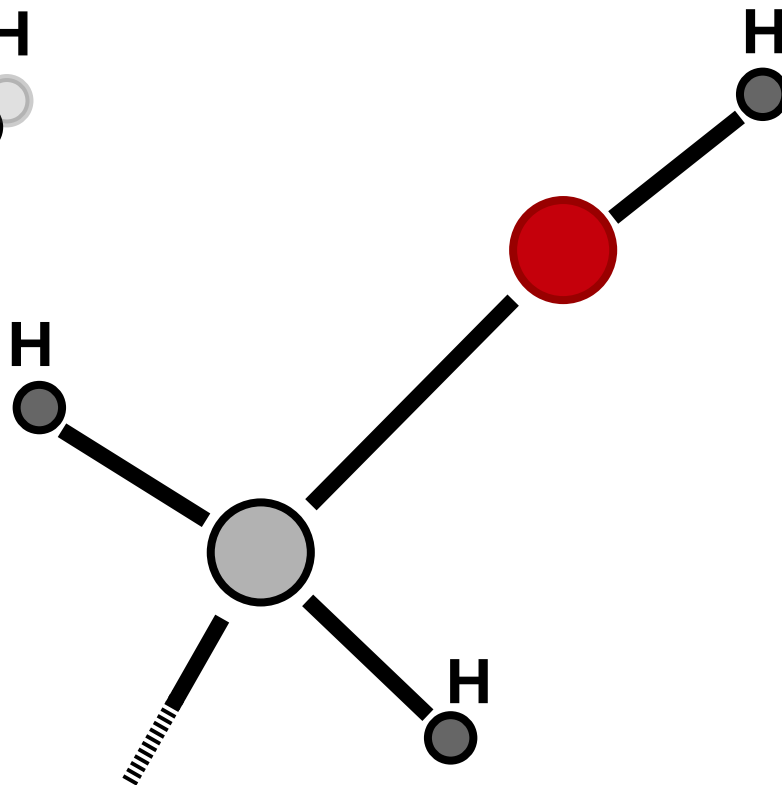


Hydrogen handling in X-Ray refinement

e⁻-cloud position

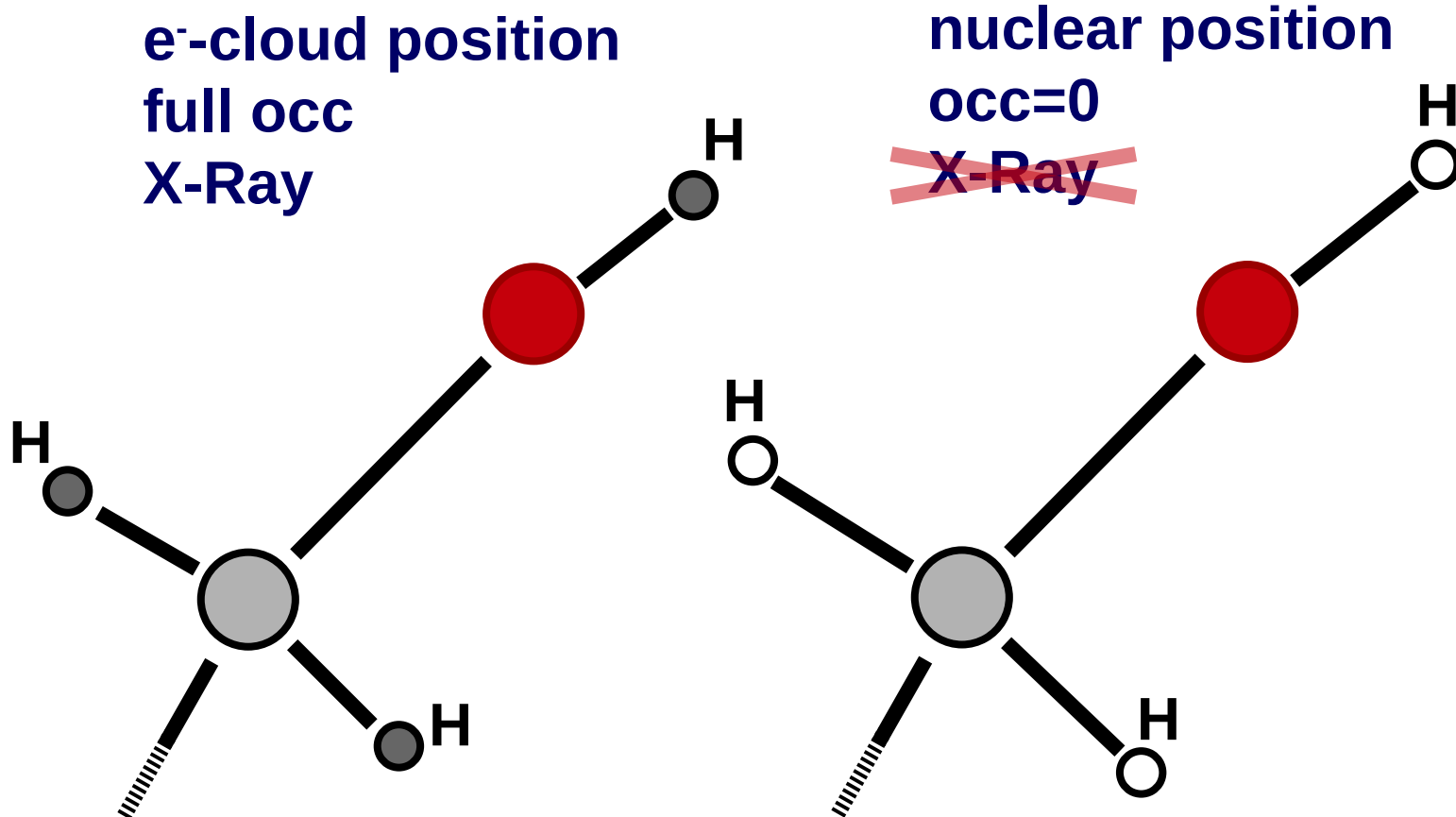


nuclear position



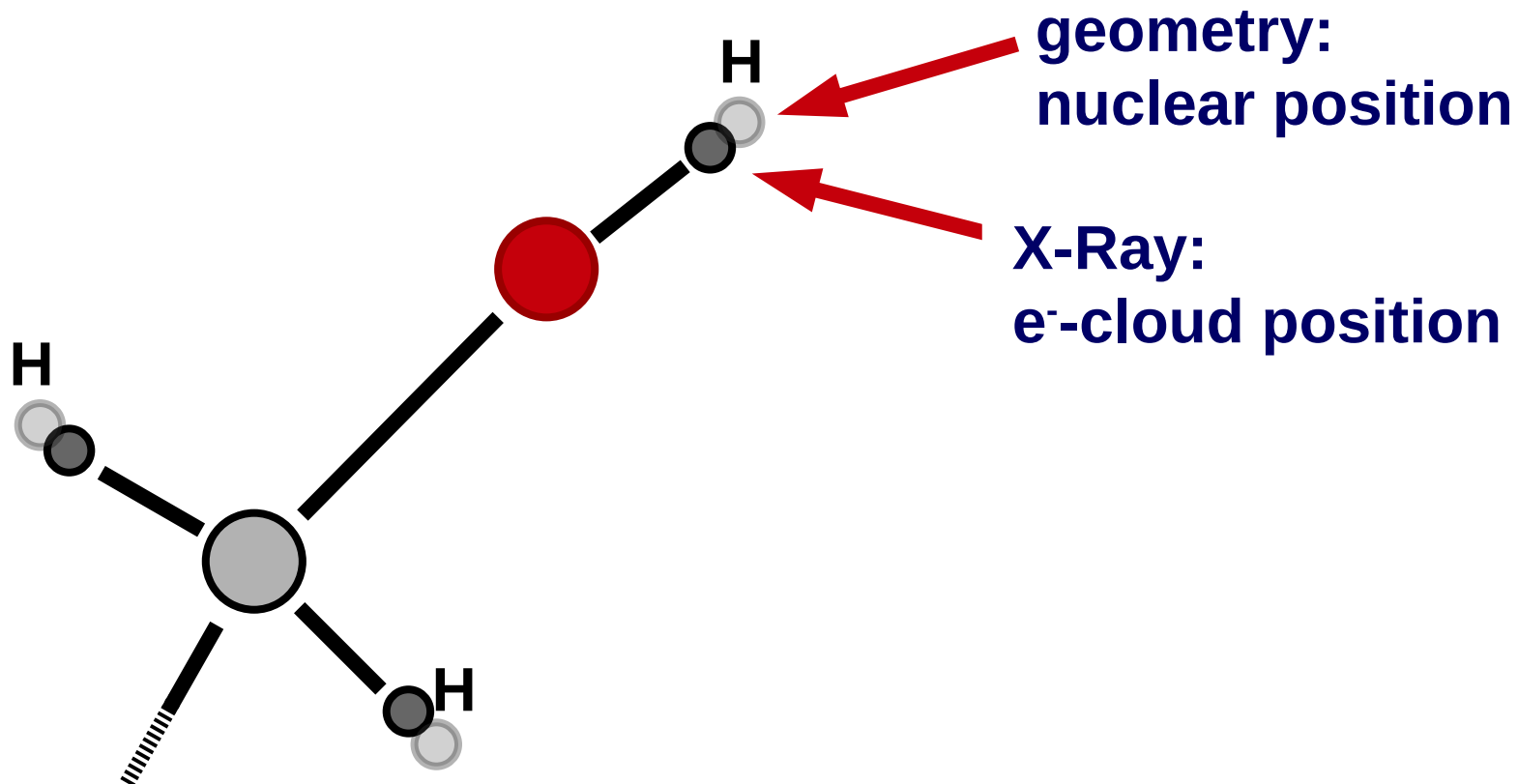
See e.g.: Williams et al (2018). Protein Science, 27, 293-315.

Hydrogen placement has consequences



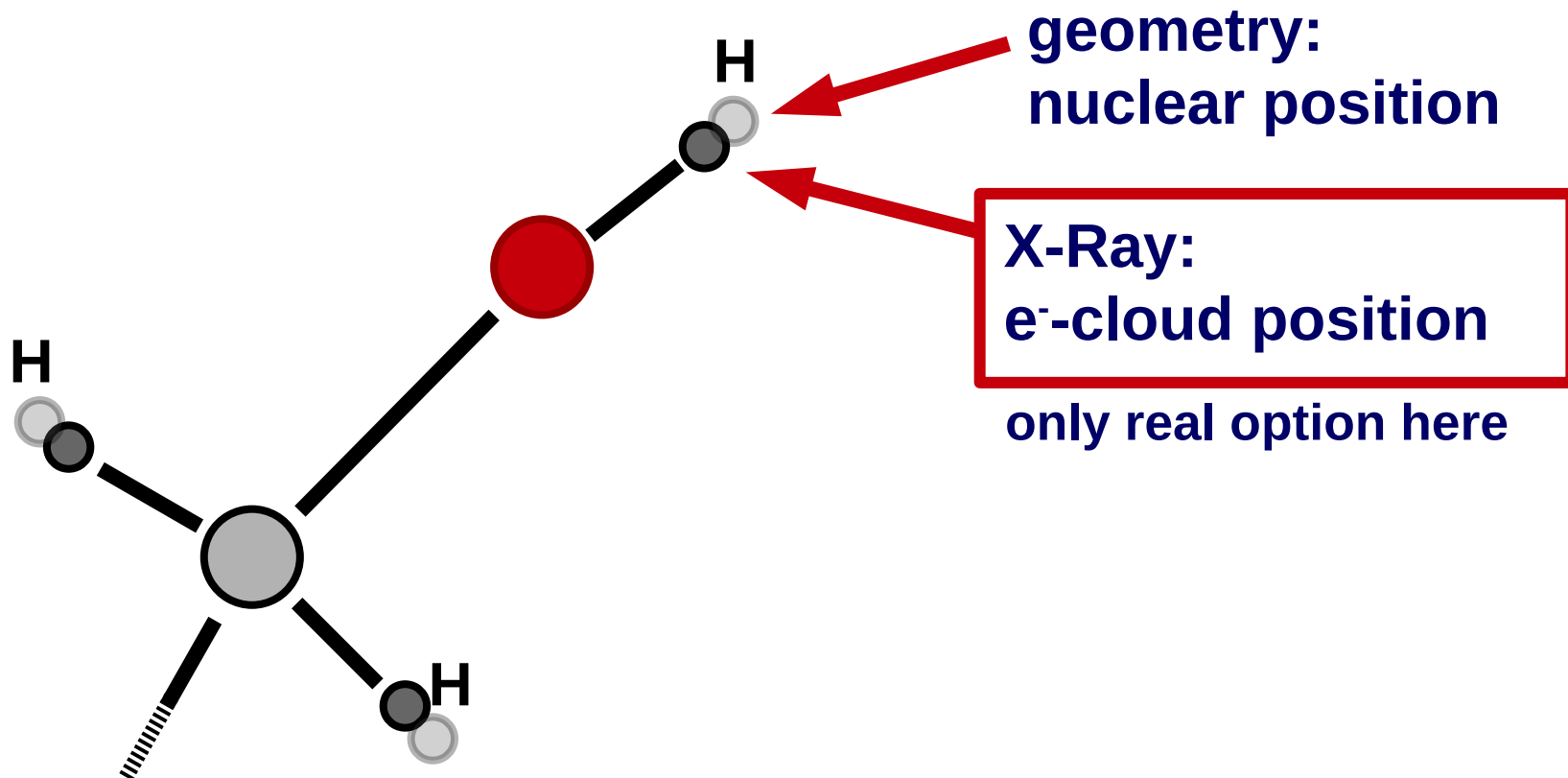
BUSTER: hydrogens refined as any other atom - not “riding hydrogen” model

Mixed-mode (hybrid) hydrogen model

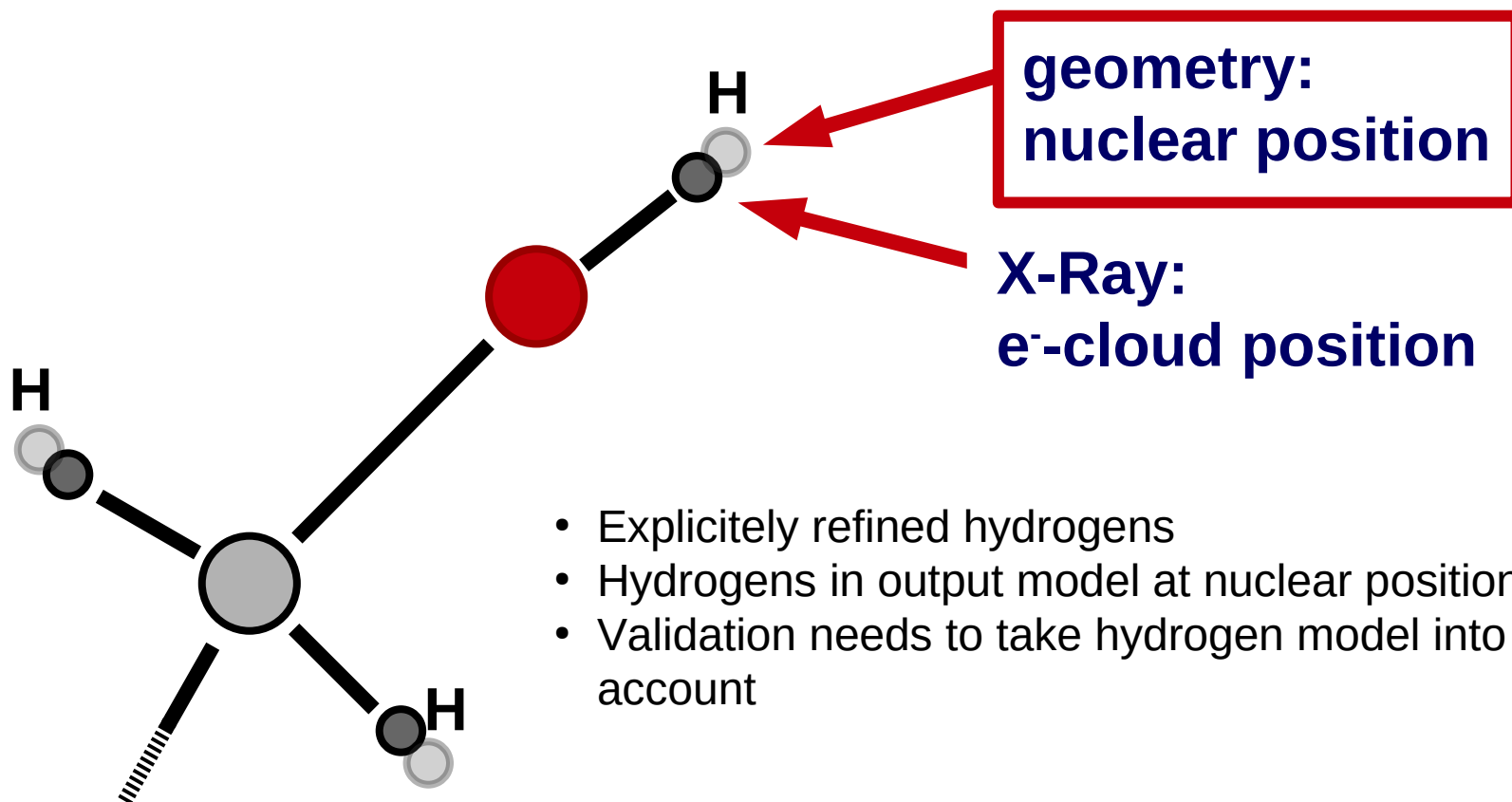


BUSTER: refine -M HydrogenHybridModel ...

Mixed-mode (hybrid) hydrogen model



Mixed-mode (hybrid) hydrogen model

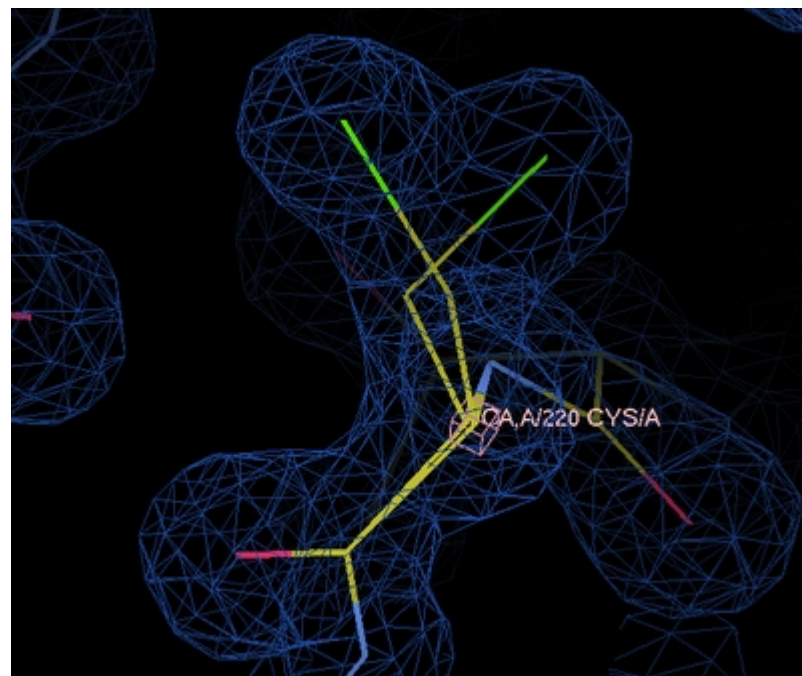


See also discussions in: Liebschner et al (2018). Acta D74, 800-813.

Alternate conformations

Alternate conformation

- ❑ Atoms with altConf “A” see atoms without altConf or with same altConf “A”
- ❑ Atoms with different altConfs don’t see each other
- ❑ Set occupancies you want to refine to something not equal to 1.0, and then run
 - **pdb2occ** -p model.pdb -o occ.gelly
 - **refine** -Gelly occ.gelly ...
- ❑ If $\text{occ}(A) + \text{occ}(B) = 1$ in input, **pdb2occ** will restrain occupancy sum (to 1)



Some notes about rotamers and outliers

- ❑ During validation, MolProbity rotamer analysis often used as a quality criteria for model
- ❑ Outliers are considered problematic und highly unlikely (the stated goal is <1% outliers)

6D0E (deposited)

2%	A	10	LEU:1.00:0.4:280.2:36.0:::Allowed:mp
2%	A	19	LEU:1.00:0.4:273.1:36.4:::Allowed:mp
2%	A	24	LEU:1.00:0.4:260.6:43.2:::Allowed:mp
2%	A	45	LYS:1.00:0.5:199.5:130.2:301.8:173.9:Allowed:ttmt
	A	50	ILE:1.00:0.0:249.5:205.1::: OUTLIER :OUTLIER
20%	A	82	AVAL:0.33:2.1:311.2::: Favored :m
	A	97	LEU:1.00:0.0:235.3:15.2::: OUTLIER :OUTLIER
50%	B	23	LEU:1.00:0.4:247.2:198.3:::Allowed:mt
?	B	41	ARG:1.00:0.4:142.9:262.7:191.0:181.3:Allowed:tmt170
2%	B	45	LYS:1.00:0.3:203.2:129.9:301.1:170.3:Allowed:ttmt
8%	B	50	ILE:1.00:0.4:215.5:155.0:::Allowed:tt
20%	B	82	VAL:1.00:1.2:312.5:::Allowed:m
	B	97	LEU:1.00:0.0:236.7:13.8::: OUTLIER :OUTLIER

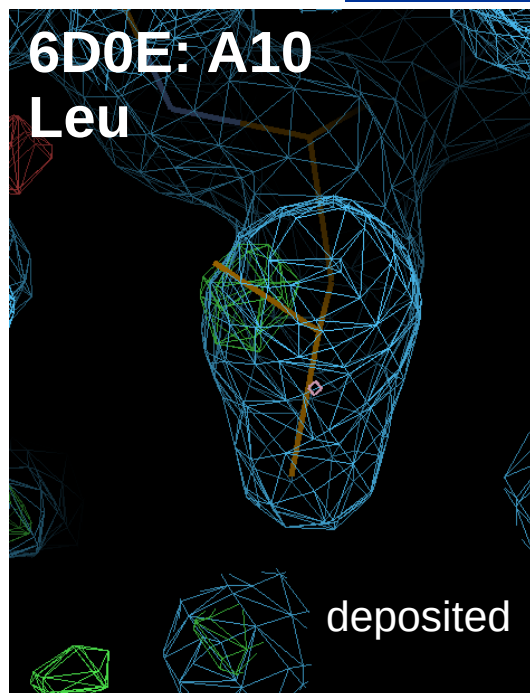
BUSTER

A	10	LEU:1.00:0.0:259.1:21.3::: OUTLIER :OUTLIER
A	19	LEU:1.00:0.1:309.3:303.1::: OUTLIER :OUTLIER
A	24	LEU:1.00:0.2:297.4:296.7::: OUTLIER :OUTLIER
A	45	LYS:1.00:0.1:197.6:122.2:308.4:165.6: OUTLIER :OUTLIER
A	50	ILE:1.00:0.0:239.0:174.6::: OUTLIER :OUTLIER
A	82	AVAL:0.33:0.2:320.6::: OUTLIER :OUTLIER
A	97	LEU:1.00:0.0:238.9:5.1::: OUTLIER :OUTLIER
B	23	LEU:1.00:0.0:180.7:193.4::: OUTLIER :OUTLIER
B	41	ARG:1.00:0.1:138.1:266.5:172.2:188.0: OUTLIER :OUTLIER
B	45	LYS:1.00:0.1:197.5:123.2:307.8:166.2: OUTLIER :OUTLIER
B	50	ILE:1.00:0.0:238.2:171.6::: OUTLIER :OUTLIER
B	82	VAL:1.00:0.1:326.1::: OUTLIER :OUTLIER
B	97	LEU:1.00:0.0:238.8:5.0::: OUTLIER :OUTLIER

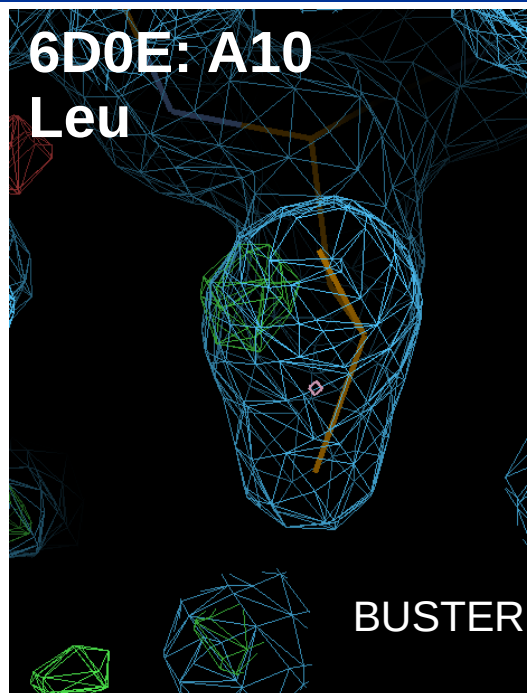
Unlikely – but not
classified as “outlier”

- ❑ Do we have a problem with rotamers in BUSTER refinements?

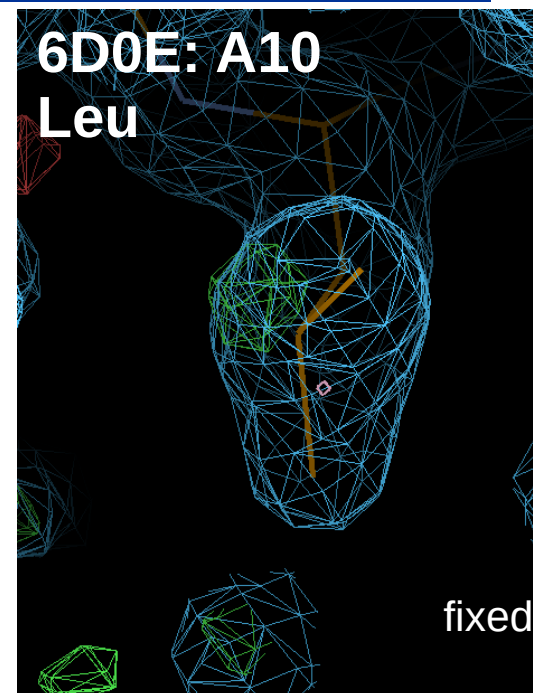
Rotamer has to fit density (for X-Ray crystallographic models)



mp (2%): Allowed



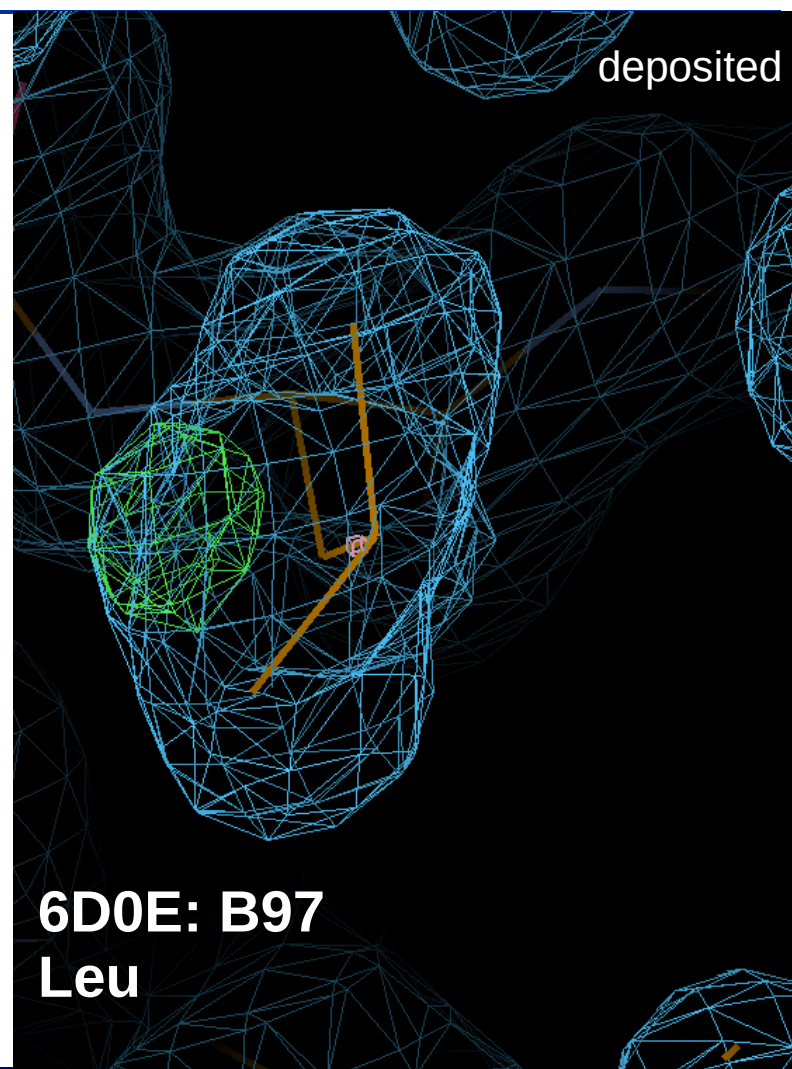
OUTLIER



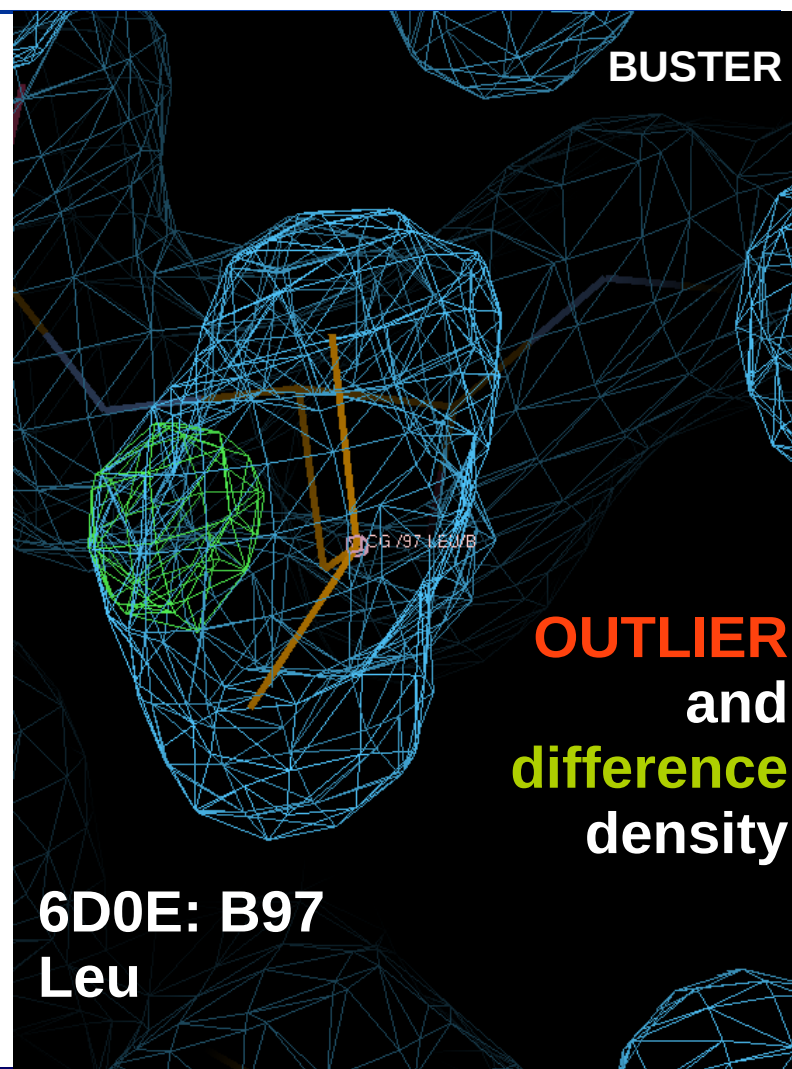
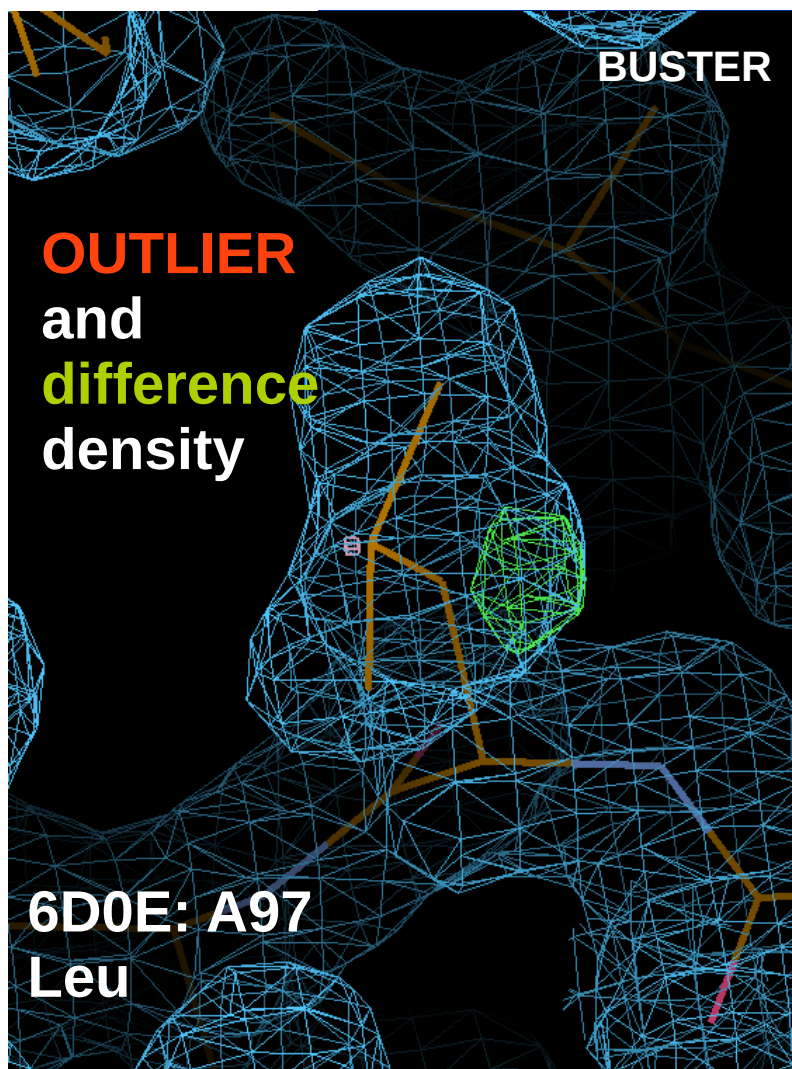
mt (59%): favoured

- small, but significant difference between mt (59%) and mp (2%)
- BUSTER refines model away from poor/wrong rotamer and gives clear indications:
 - positive difference density
 - rotamer now highlighted as outlier
- enforcing rotamer restraints can mask incorrect side-chain conformations

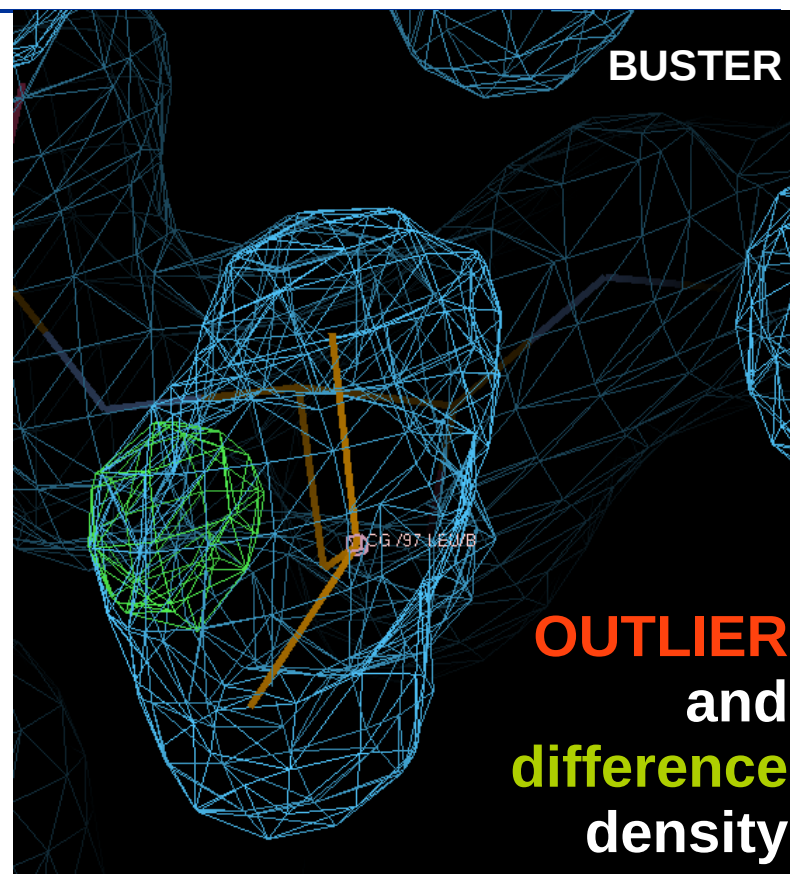
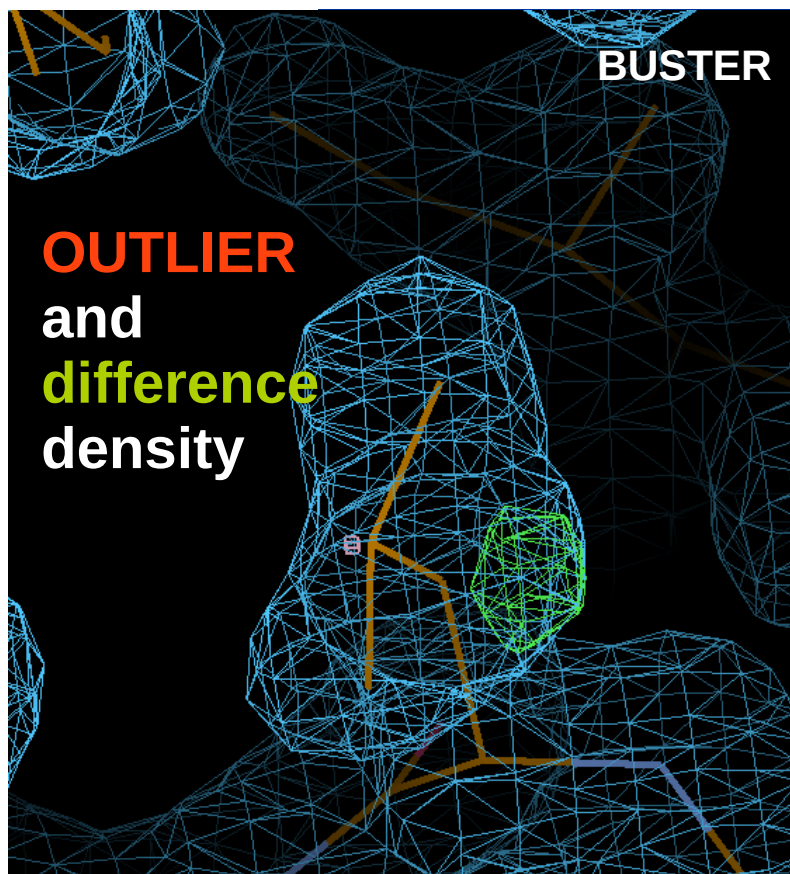
Rotamer outliers are useful markers



Rotamer outliers are useful markers

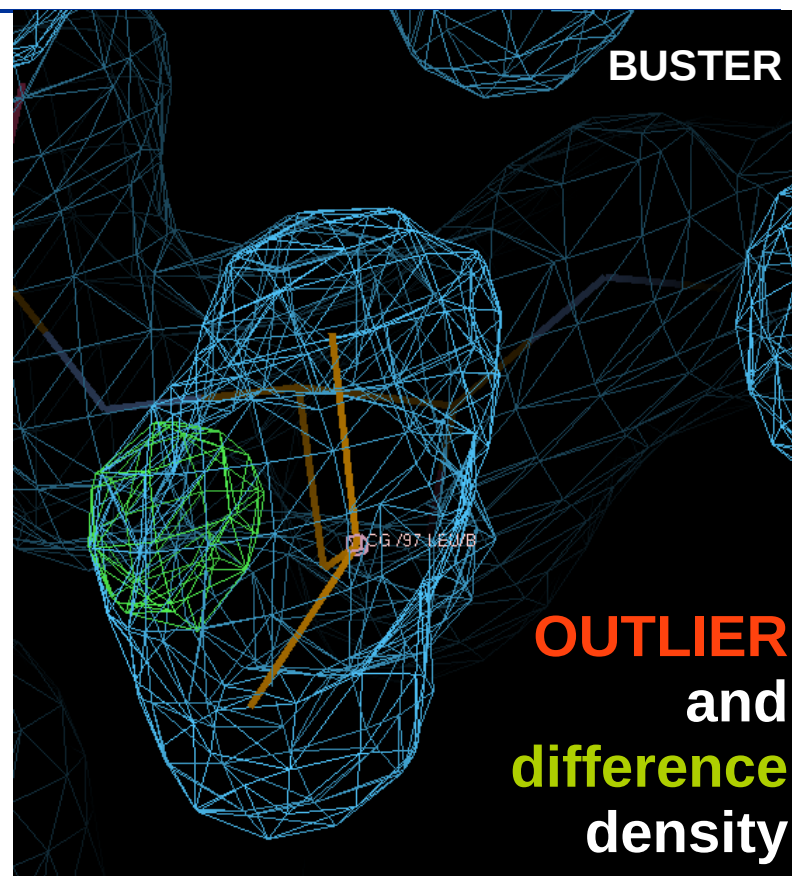
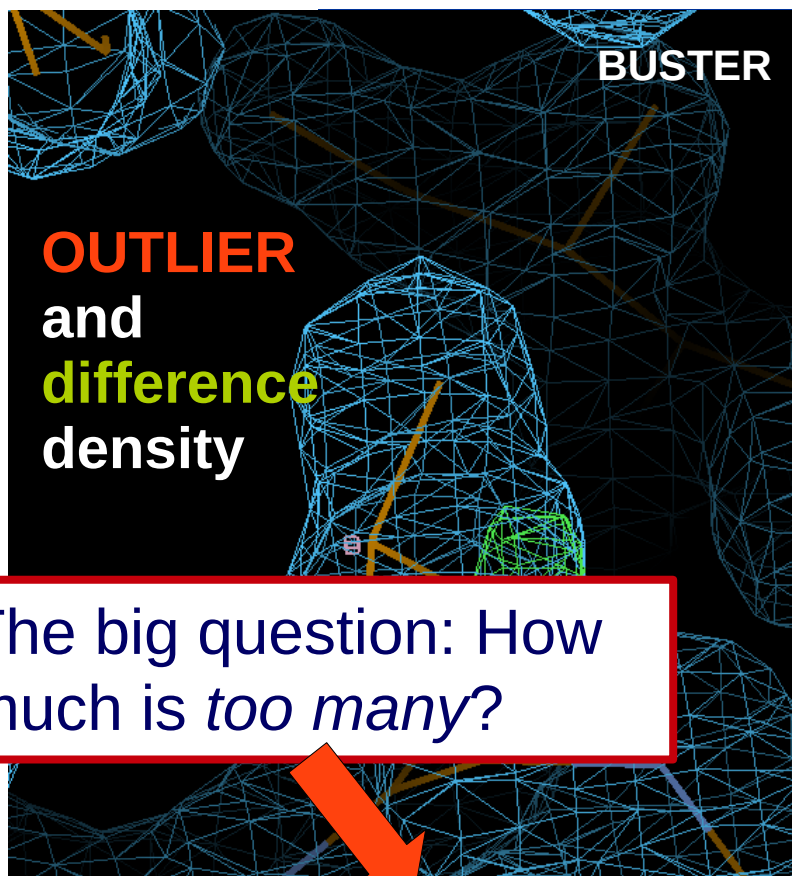


Rotamer outliers are useful markers



Do not use *too many* additional restraints to keep geometry under control if underlying problem is incorrect atomic model - as visible in density!

Rotamer outliers are useful markers



Do not use *too many* additional restraints to keep geometry under control if underlying problem is incorrect atomic model - as visible in density!

BUSTER: Is there any any ligand bound?

Acta Cryst. (2005). **A61**, C248

Automated Structure Refinement for High-throughput Ligand Detection with BUSTER-TNT

Clemens Vornrhein, Gerard Bricogne, *Global Phasing Ltd., Cambridge, UK.* E-mail: vornrhein@GlobalPhasing.com

The use of crystallography for the discovery of lead compounds often involves a large number of experiments with different soaking or co-crystallization trials. The subsequent refinement and analysis of the resulting datasets can be time-consuming and tedious. Since the crystallographic parameters (resolution, space group, cell parameters) are quite often similar, this task is ideally suited for automation.

We present a method (**autoBUSTER**) that automates the refinement, solvent model update, ligand detection and analysis. Centered around the BUSTER-TNT program [1,2], it requires a minimal amount of user input. Although it can be used at any resolution and for any kind of macromolecular structure, it is tuned to the refinement of protein structures at better than 2.8 Å resolution.

The knowledge of any (possibly) bound ligand can be given (a) explicitly by supplying a PDB file of dummy atoms that describes the assumed binding site, or (b) by letting the system automatically analyze the residual density of difference Fourier maps. A unique feature of BUSTER-TNT is used, where the various masks describing the known fragment, the bulk solvent and the missing part can be given independently from each other. The results show that this can greatly enhance the capability of uniquely defining any bound ligand.

[1] Bricogne G., Irwin J., *Macromolecular Refinement: Proceedings of the CCP4 Study Weekend*, Warrington: Daresbury Laboratory, 1996, 85-92. [2] Blanc E., Roversi P., Vornrhein C., Flensburg C., Lea S. M., Bricogne G., *Acta Cryst.*, 2004, **D60**, 2210-2221.

- ❑ BUSTER's '-L' feature tries to take an **unbiased look** at data & model to decide **if, where and how** something ***might*** have bound.
- ❑ No prior knowledge (bias?) required/used.
- ❑ Used since 2003, with first academic release in 2009

Liebschner et al (2017): "Polder maps: improving OMIT maps by excluding bulk solvent.", *Acta D* 73, 148

BUSTER: Is there any any ligand bound?

GΦL LigandDetectionModes

☐ Favorite?

 Search


Attachments



(You are *AnonymousGnome*)

Content:

- [Introduction](#)
- [Unknown location](#)
- [Known location](#)
- [Caveats](#)
- [Summary](#)

Introduction

Apart from the standard procedures for detecting ligands (difference Fourier maps), BUSTER has one particular feature that needs a bit further explanation - to explain what it can do, what it can't do and what potential bias it might introduce.

This feature is triggered by the `-L` and `-Lpdb` flags to the refine command:

```
% refine -p some.pdb -m some.mtz -L ...
- or -
% refine -p some.pdb -m some.mtz -Lpdb bindingsite.pdb ...
```

It will treat a certain region of the model differently during the last big cycle of refinement: that region will neither contain an atomic model nor a contribution from the bulk solvent. However, if there is some electron density present in this region, it should show up in difference (Fo-Fc) maps as strong positive density. The interpretation, what this electron density might represent (atomic model, bulk solvent or a mixture) is up to the user.

Here we're going to explain the typical usage of this BUSTER feature, their assumptions and caveats.

Unknown location

The least biased assumption is that there might be some ligand bound, but its location is unknown. In that case one would use the

- ☐ Unbiased towards “expected” binding (just because the crystal was soaked/co-crystallised with compound doesn't mean it actually is bound)
- ☐ Unbiased towards binding site (there can be unexpected or new binding sites)
- ☐ Unbiased towards binding pose

BUSTER wiki

www.globalphasing.com/buster/wiki/

Difference Fourier maps - simple & useful

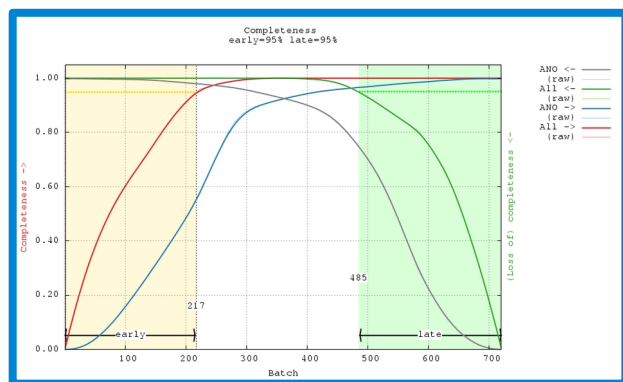
- **Anomalous:**
 - distinguish ion from solvent
 - help difficult sequencing (Cys/Met marker)
 - help placing of compound
 - automatically generated and analysed in BUSTER
- **Early-Late:**
 - show radiation damage effects
 - automatically prepared in autoPROC - then generated and analysed in BUSTER
- **Fo-Fo:**
 - (Compound - Apo) to show compound

diff_fourier -h

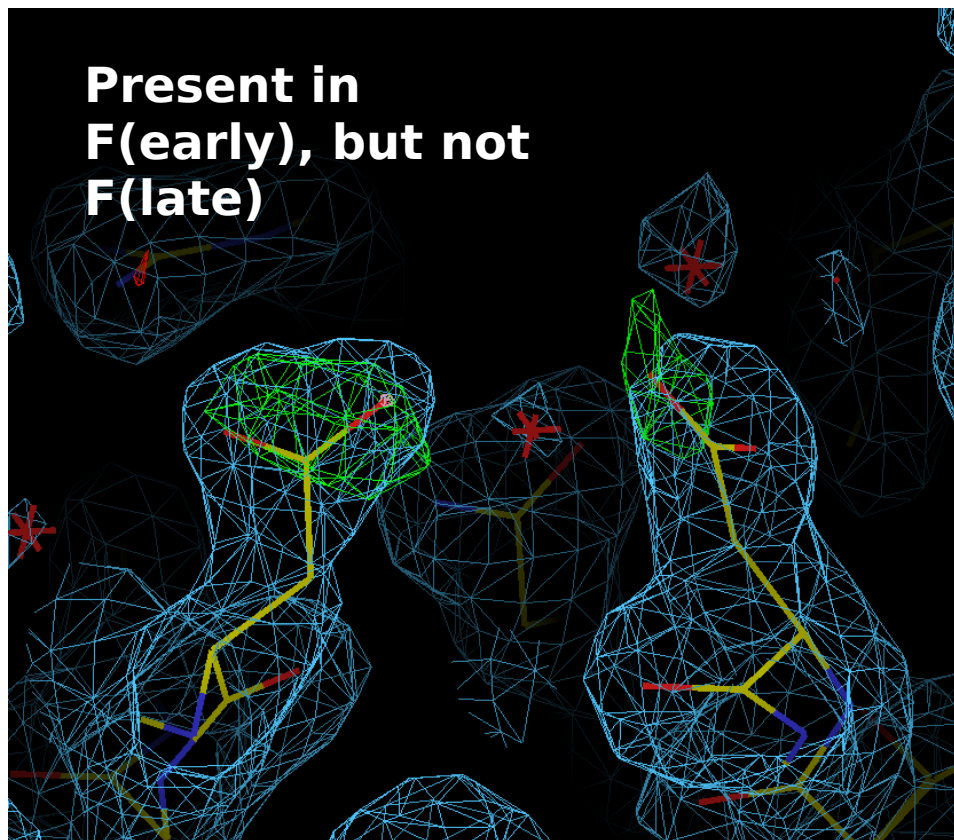
Difference Fourier maps: $F(\text{early}) - F(\text{late})$ detecting/describing radiation damage

Typical decarboxylation of
ASP/GLU residues

Normal mFo-DFc maps will
show negative peaks.

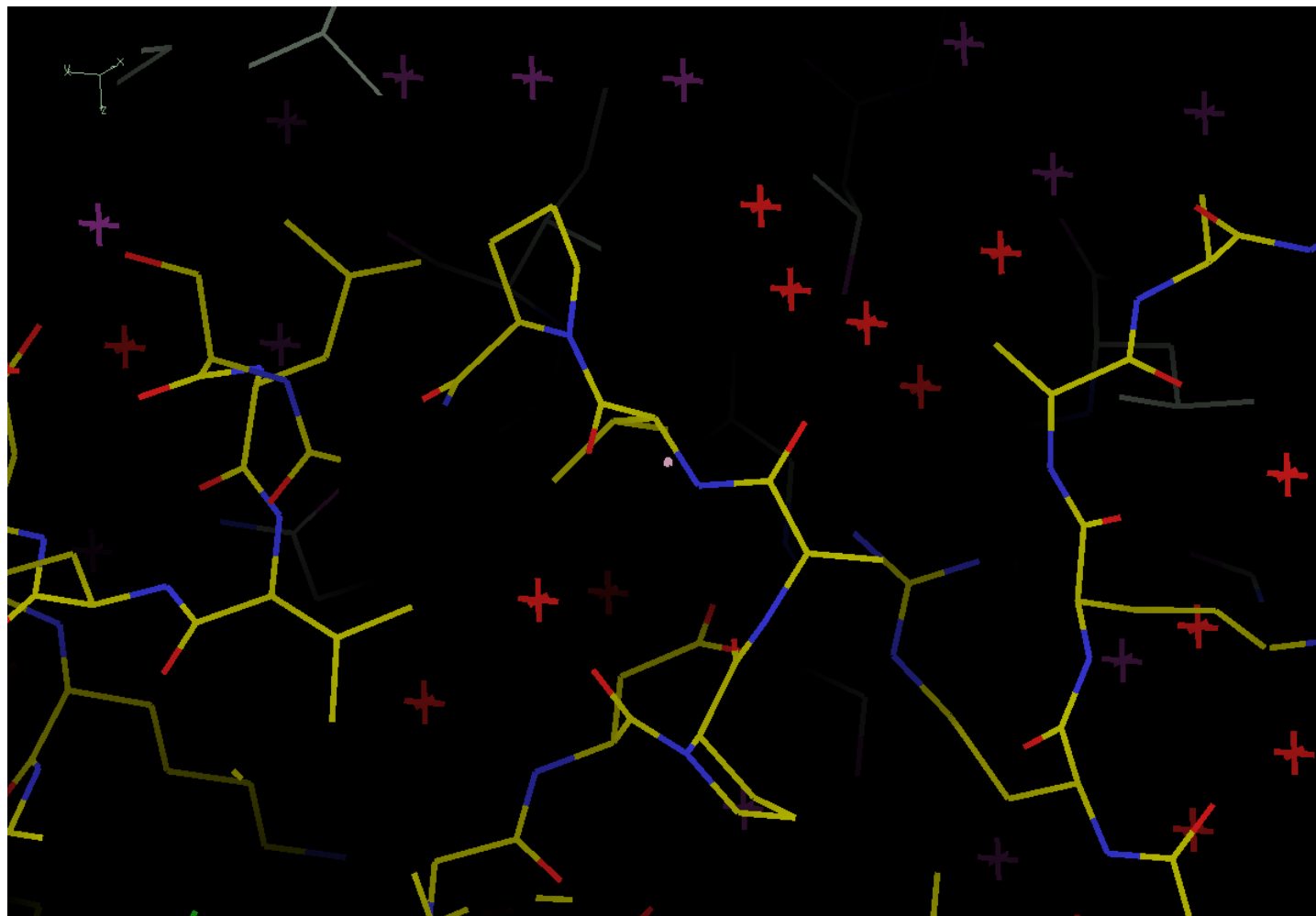


low dose, high-multiplicity

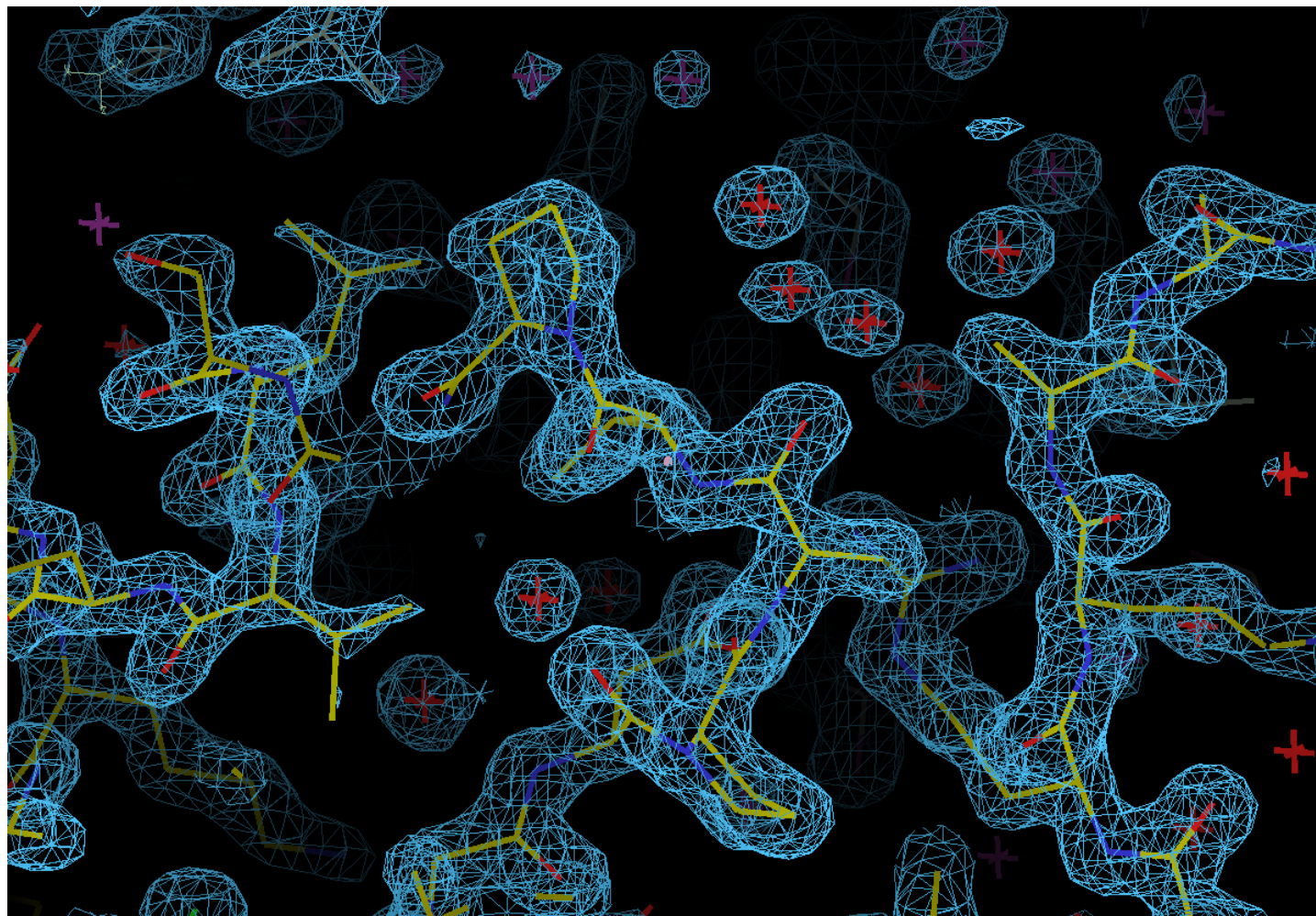


A novel mode of Gleevec binding is revealed by the structure of spleen tyrosine kinase. Atwell, S. et al.(2004) J.Biol.Chem. 279, 55827-55832.

F(early)-F(late): water in crystal contact



F(early)-F(late): water in crystal contact

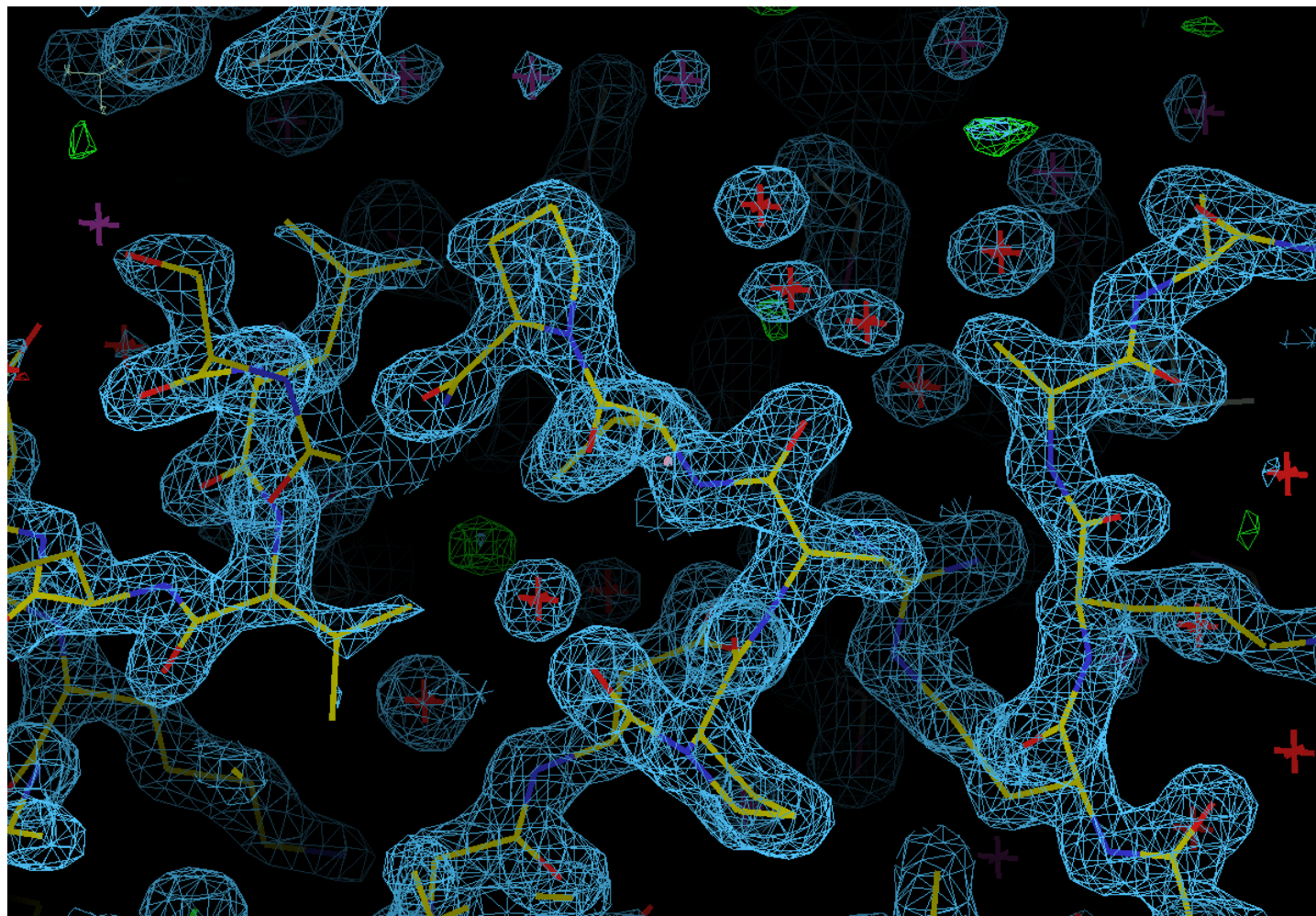


4LER

2Fo-FC

1.5 rms

F(early)-F(late): water in crystal contact



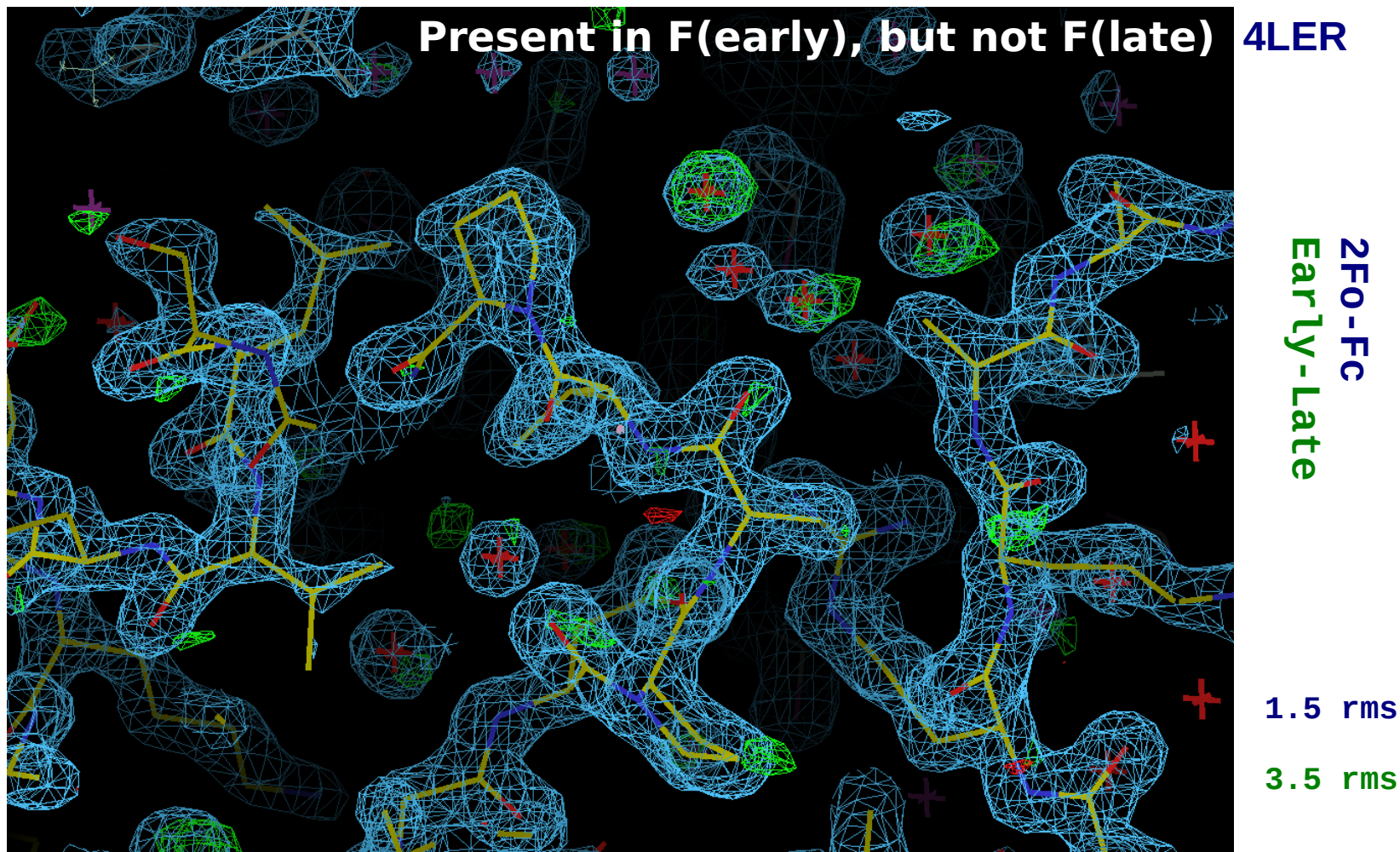
4LER

2Fo-FC
Fo-FC

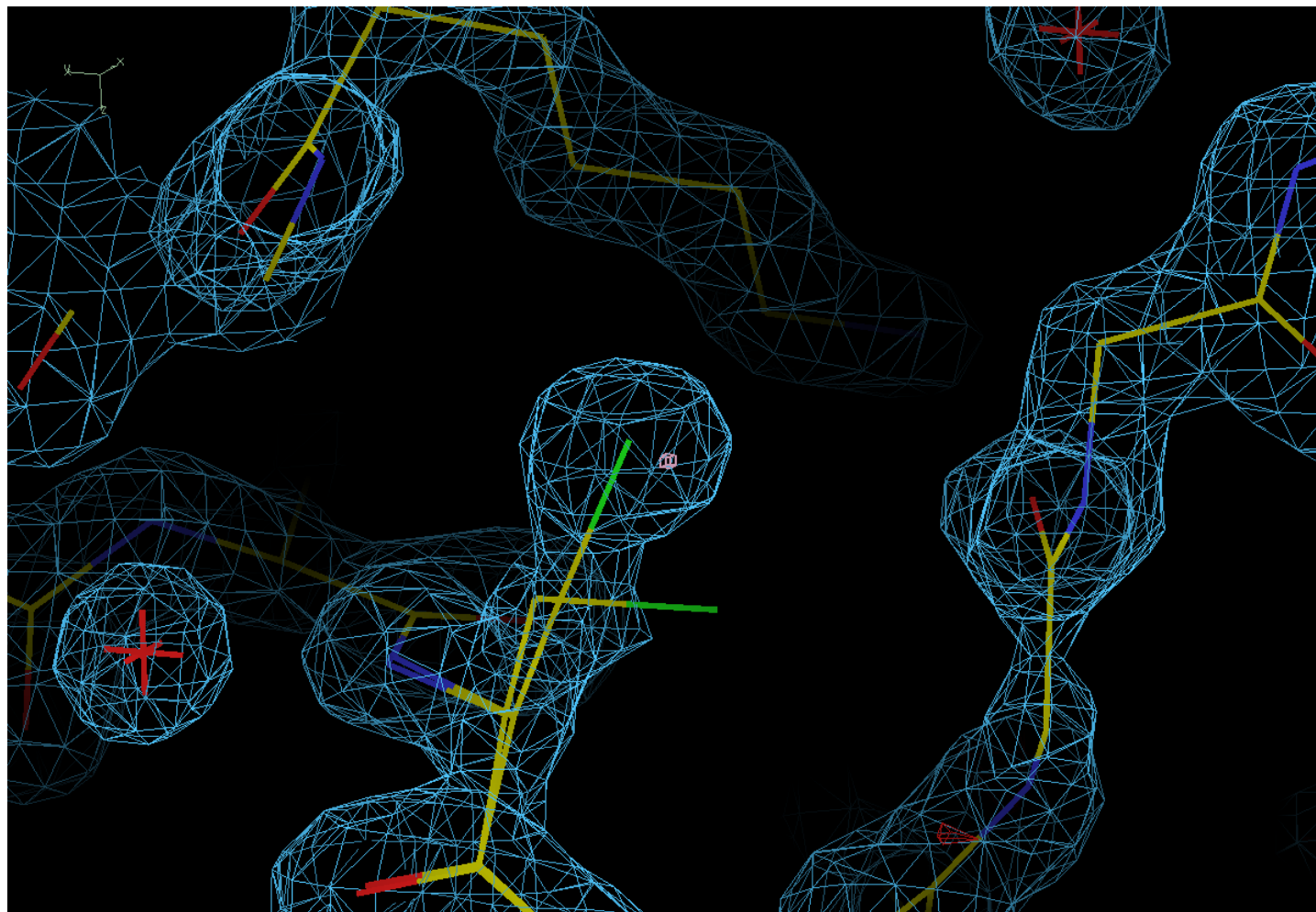
1.5 rms

3.5 rms

F(early)-F(late): water in crystal contact



F(early)-F(late): alternate conformation?



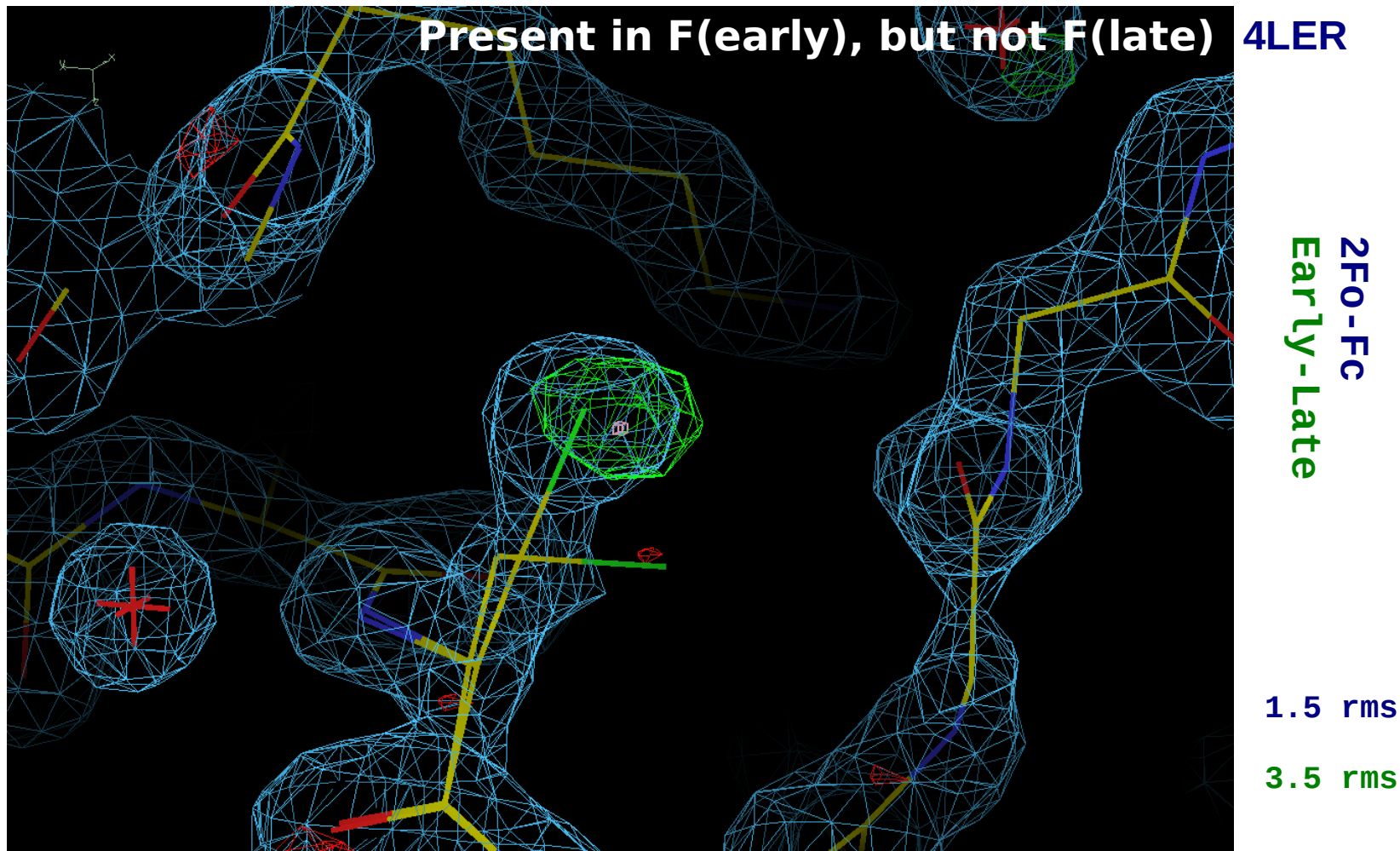
4LER

2Fo-FC
Fo-FC

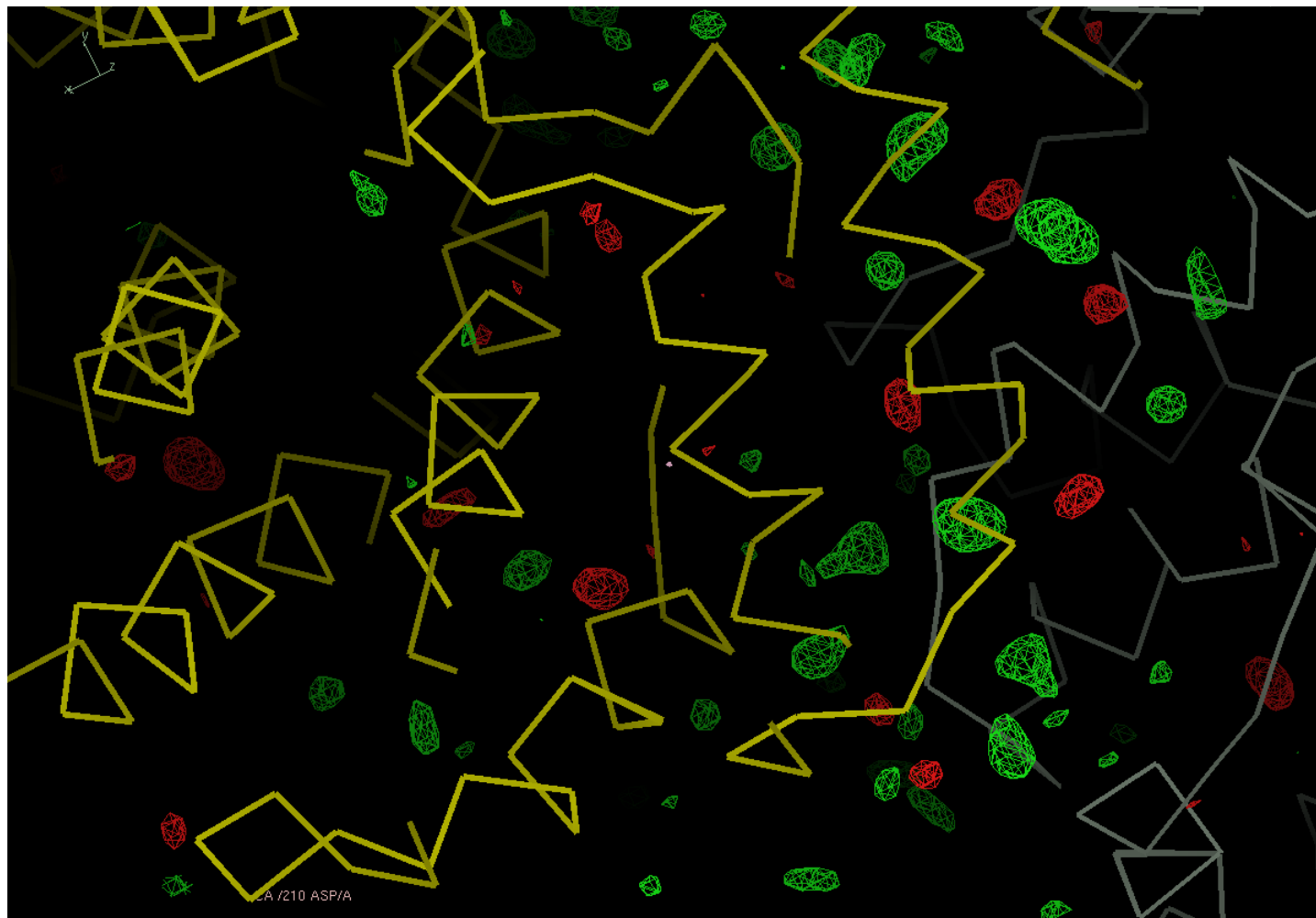
1.5 rms

3.5 rms

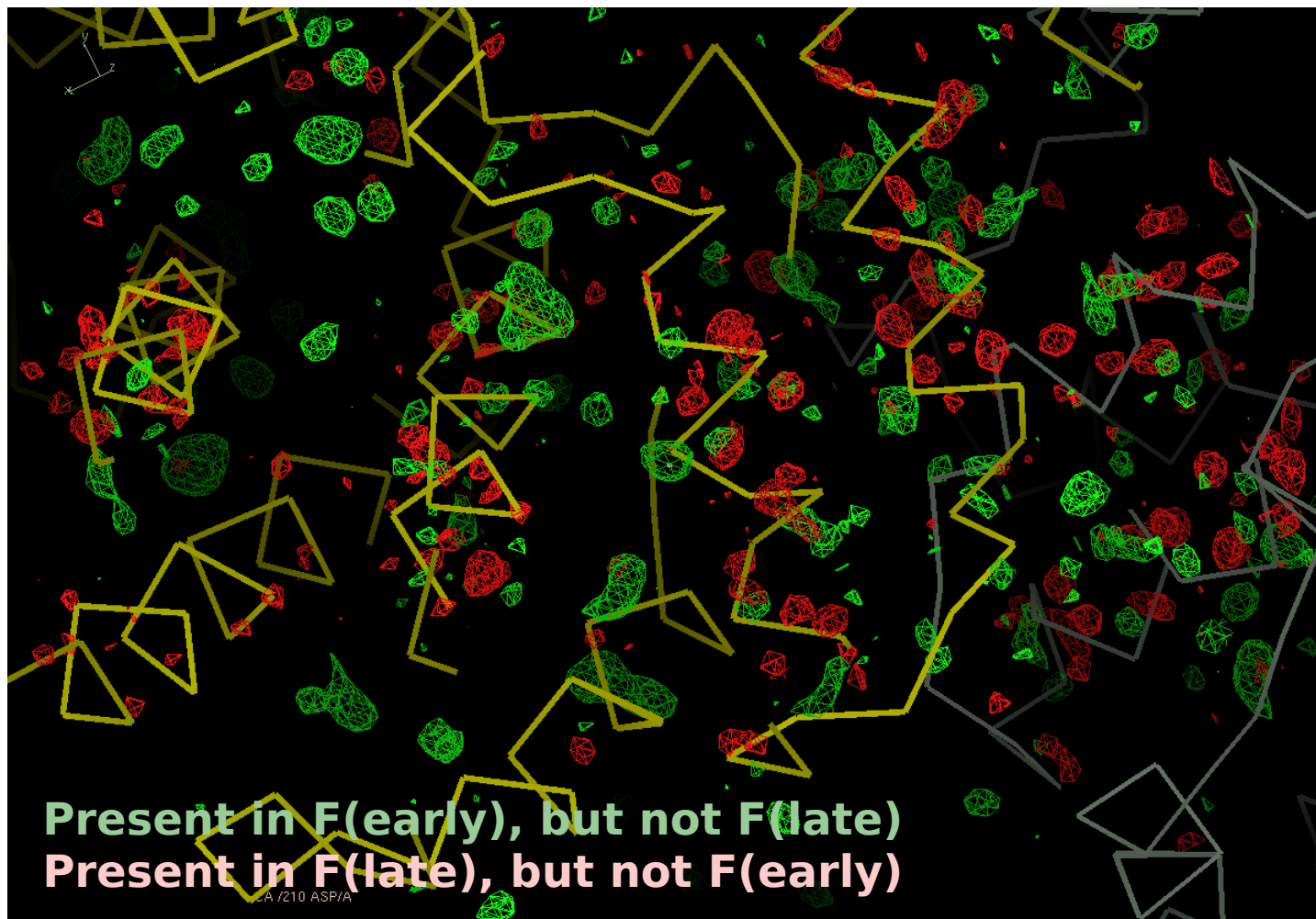
F(early)-F(late): alternate conformation?



F(early)-F(late): large-scale movements



F(early)-F(late): large-scale movements

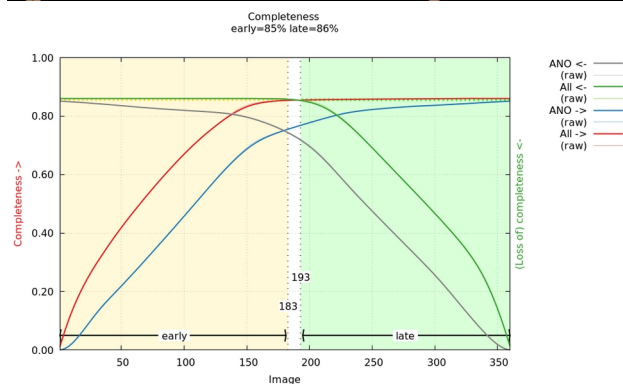
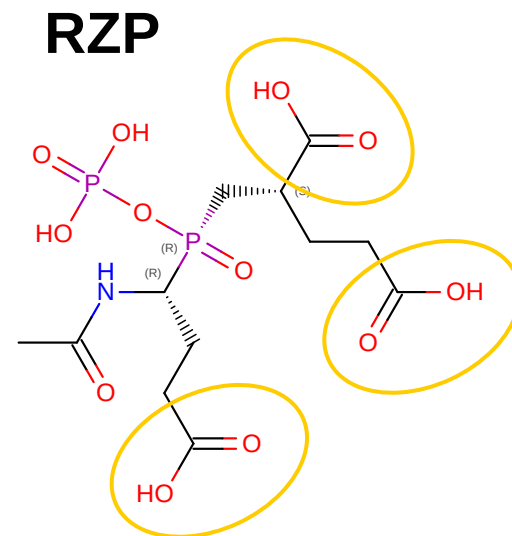
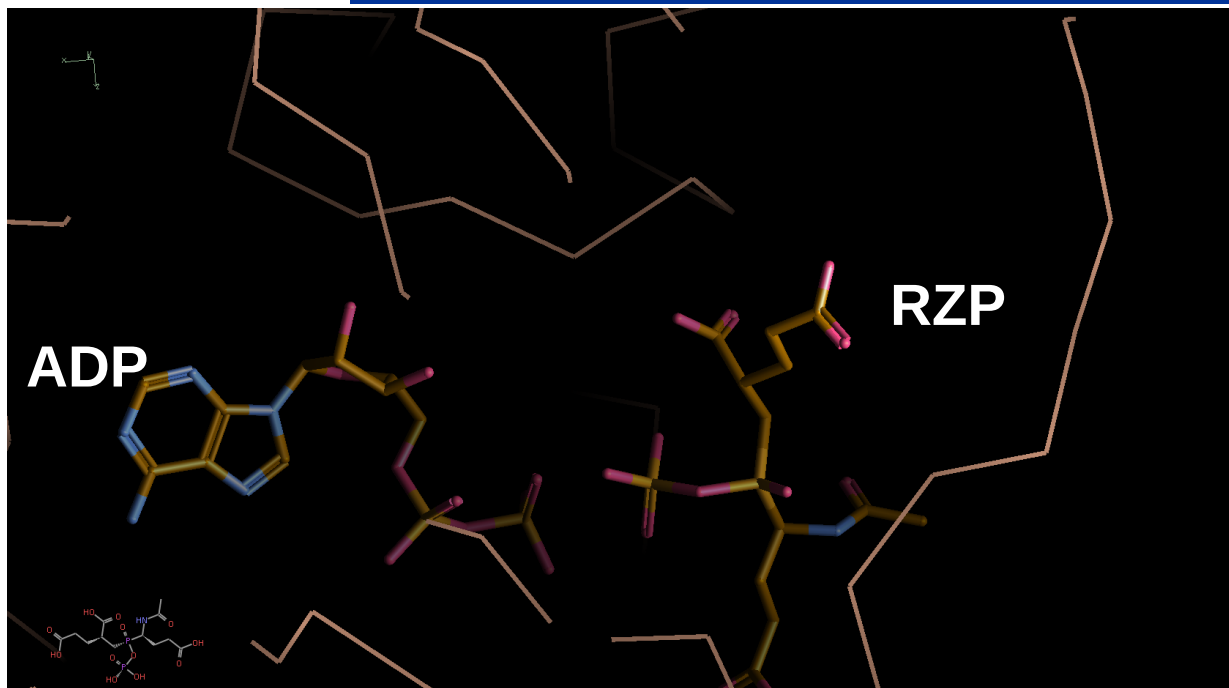


4LER

Early-Late

3.5 rms

Radiation damage - decarboxylation



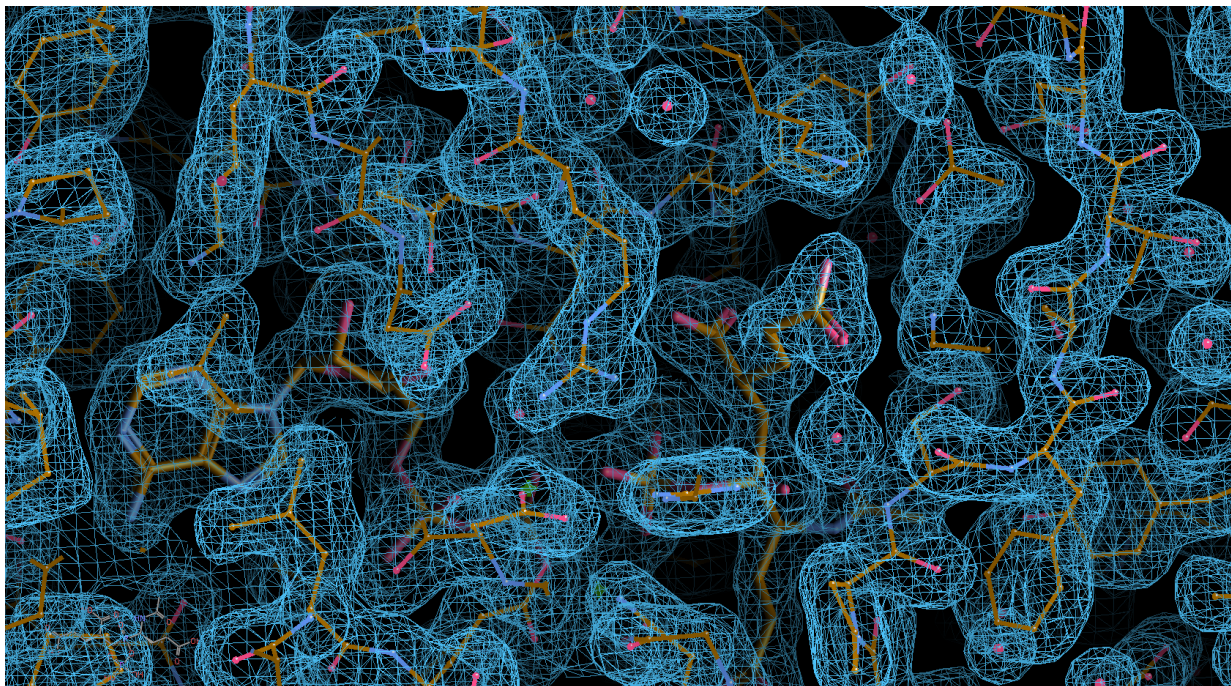
6VZU

Structural basis for polyglutamate chain initiation and elongation by TTL family enzymes.

[Mahalingan, K.K.](#), [Keith Keenan, E.](#), [Strickland, M.](#), [Li, Y.](#), [Liu, Y.](#), [Ball, H.L.](#), [Tanner, M.E.](#), [Tjandra, N.](#), [Roll-Mecak, A.](#)

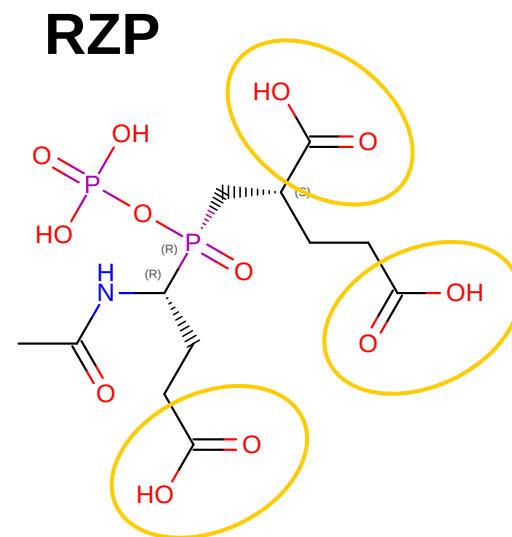
(2020) Nat Struct Mol Biol **27**: 802-813

Radiation damage - decarboxylation

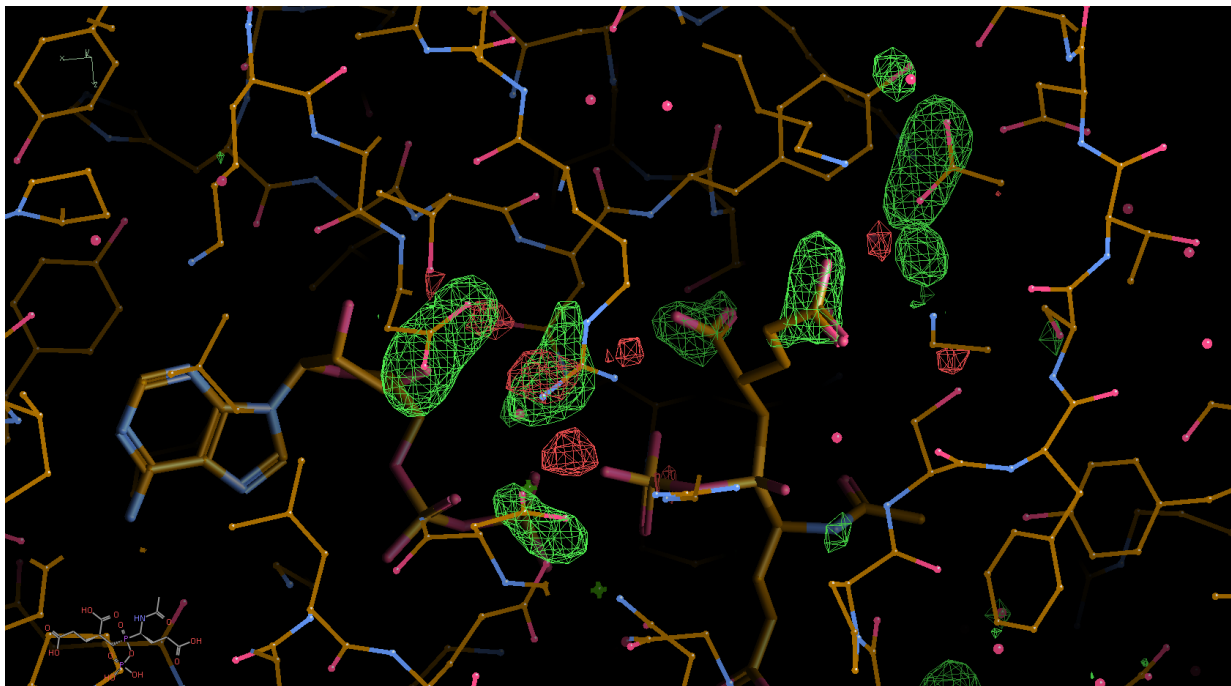


2mFo-DFc @ 1.0 rms, 2 Å
resolution.

6VZU



Radiation damage - decarboxylation



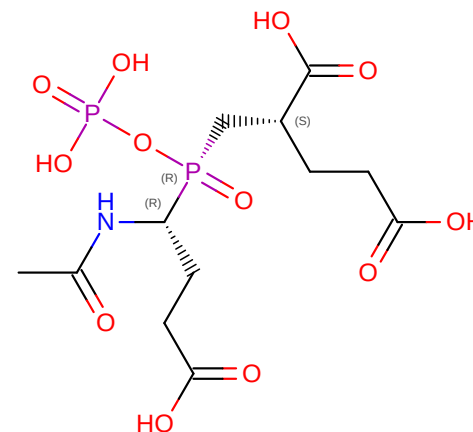
F(early)-F(late) @ 6.0 rms

Showing loss of density due to

- reduction in occupancy
- increase in temperature factor
- loss of atoms/groups
- movement of atoms/groups

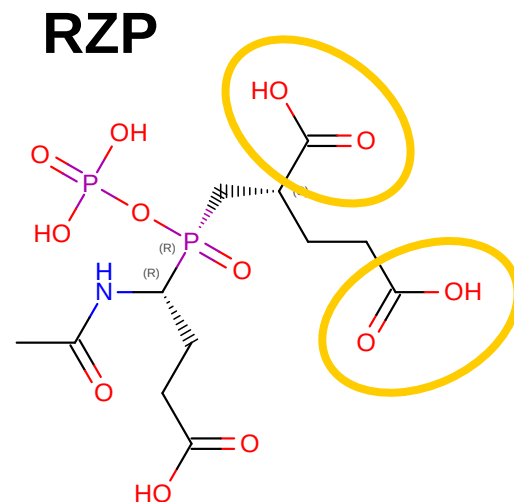
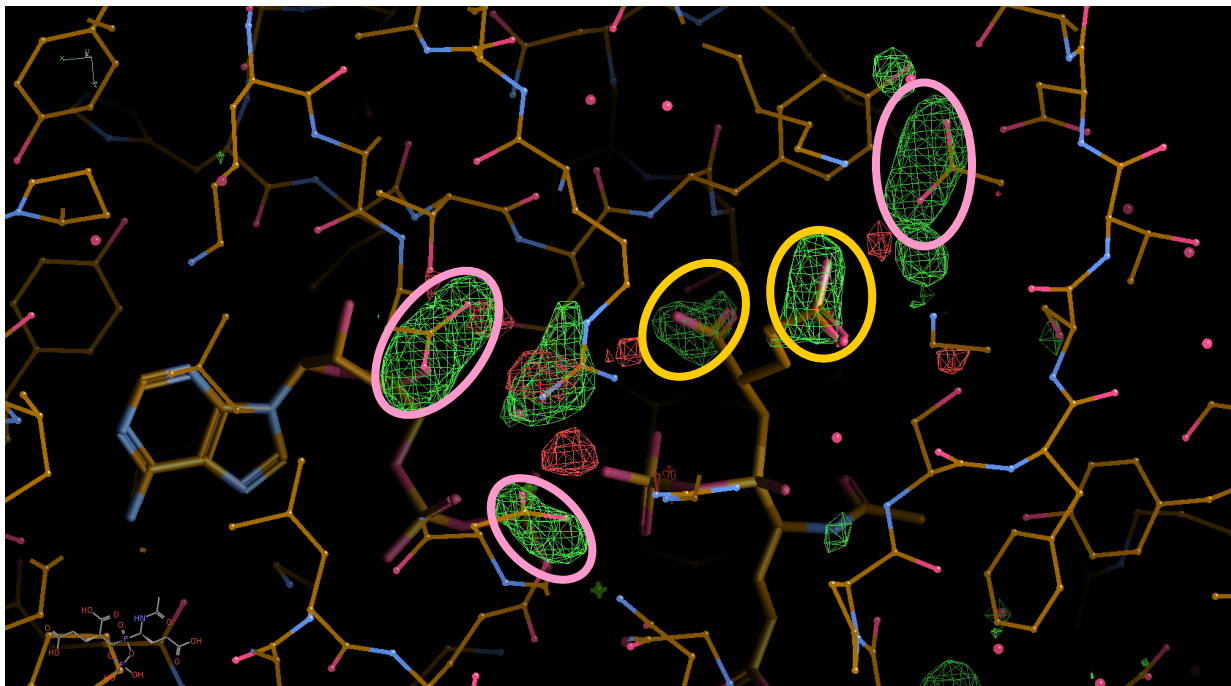
6VZU

RZP



- **early and late data** automatically generated by **autoPROC**
- **F(early)-F(late)** maps automatically generated and analysed by **BUSTER**

Radiation damage - decarboxylation



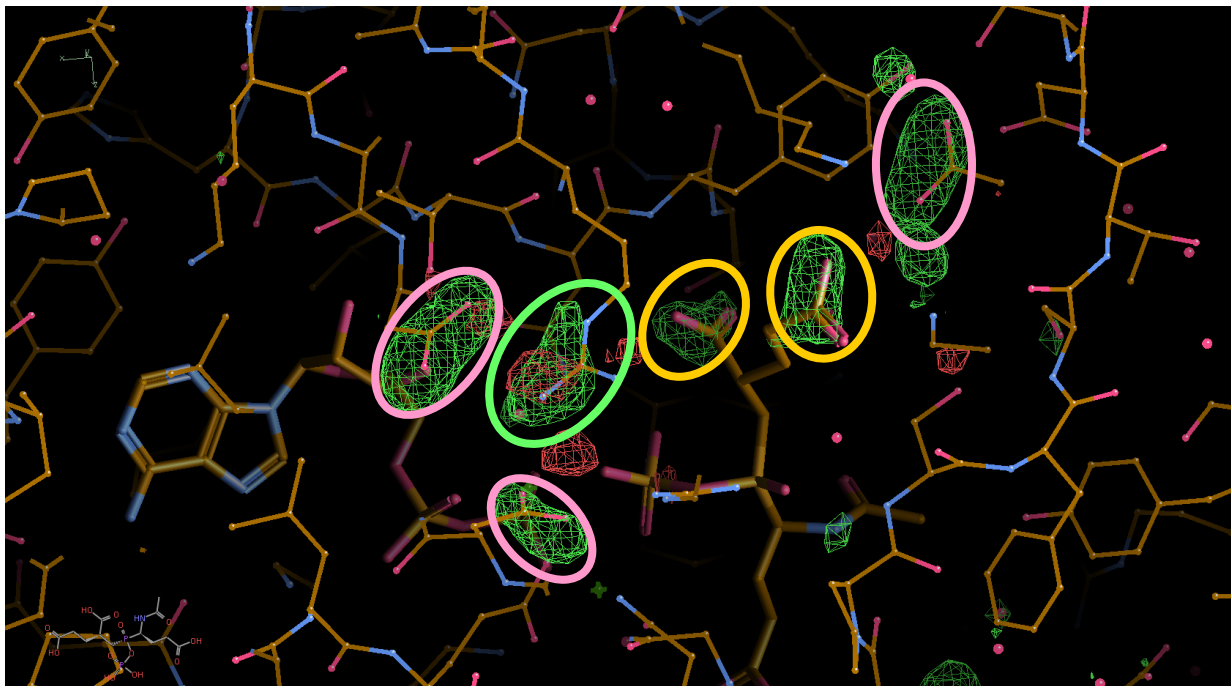
F(early)-F(late) @ 6.0 rms

- Showing loss of density due to
- reduction in occupancy
 - increase in temperature factor
 - loss of atoms/groups
 - movement of atoms/groups

6VZU

de-carboxylation of ASP/GLU and compound

Radiation damage - decarboxylation



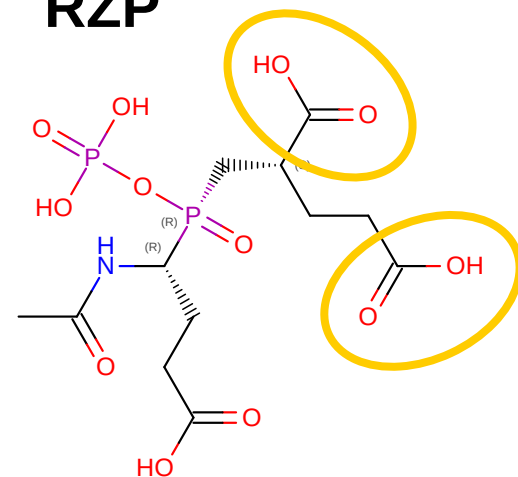
F(early)-F(late) @ 6.0 rms

- Showing loss of density due to
- reduction in occupancy
 - increase in temperature factor
 - loss of atoms/groups
 - movement of atoms/groups

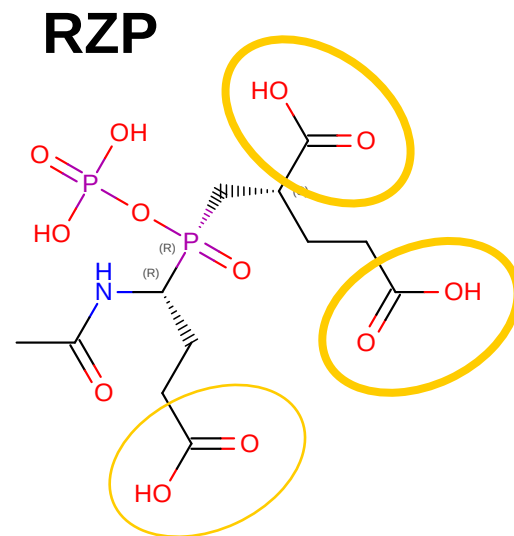
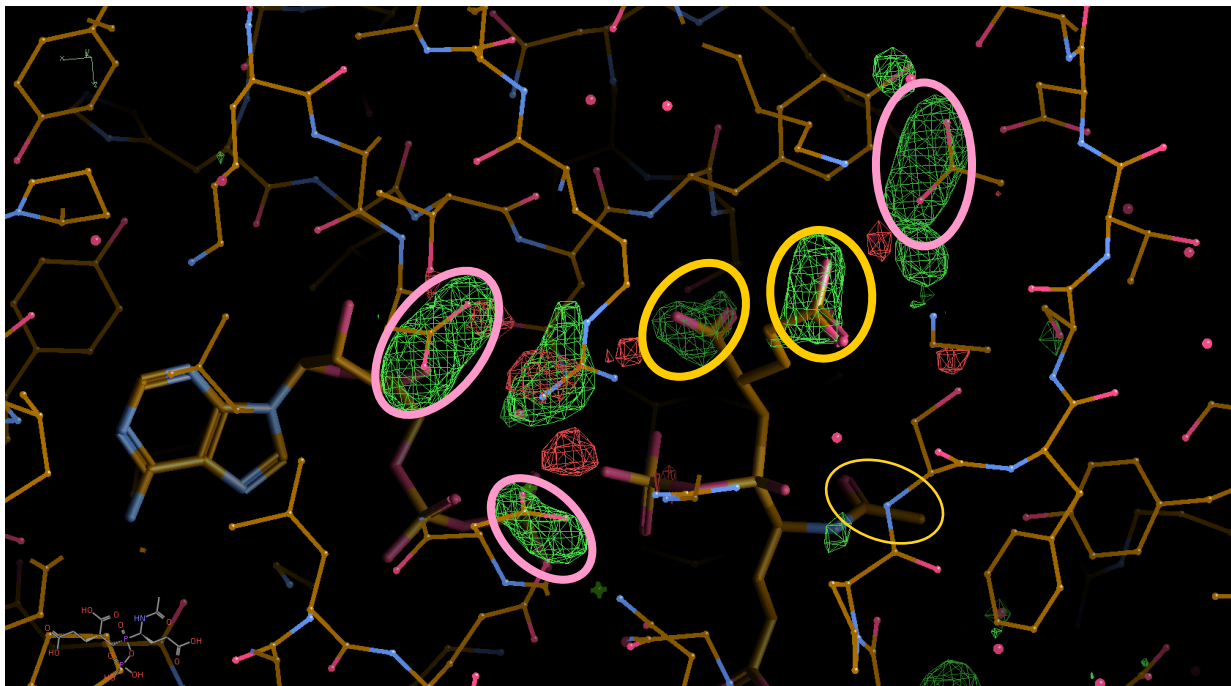
6VZU

de-carboxylation of ASP/GLU and compound

RZP



Radiation damage - decarboxylation



F(early)-F(late) @ 6.0 rms

- Showing loss of density due to
- reduction in occupancy
 - increase in temperature factor
 - loss of atoms/groups
 - movement of atoms/groups

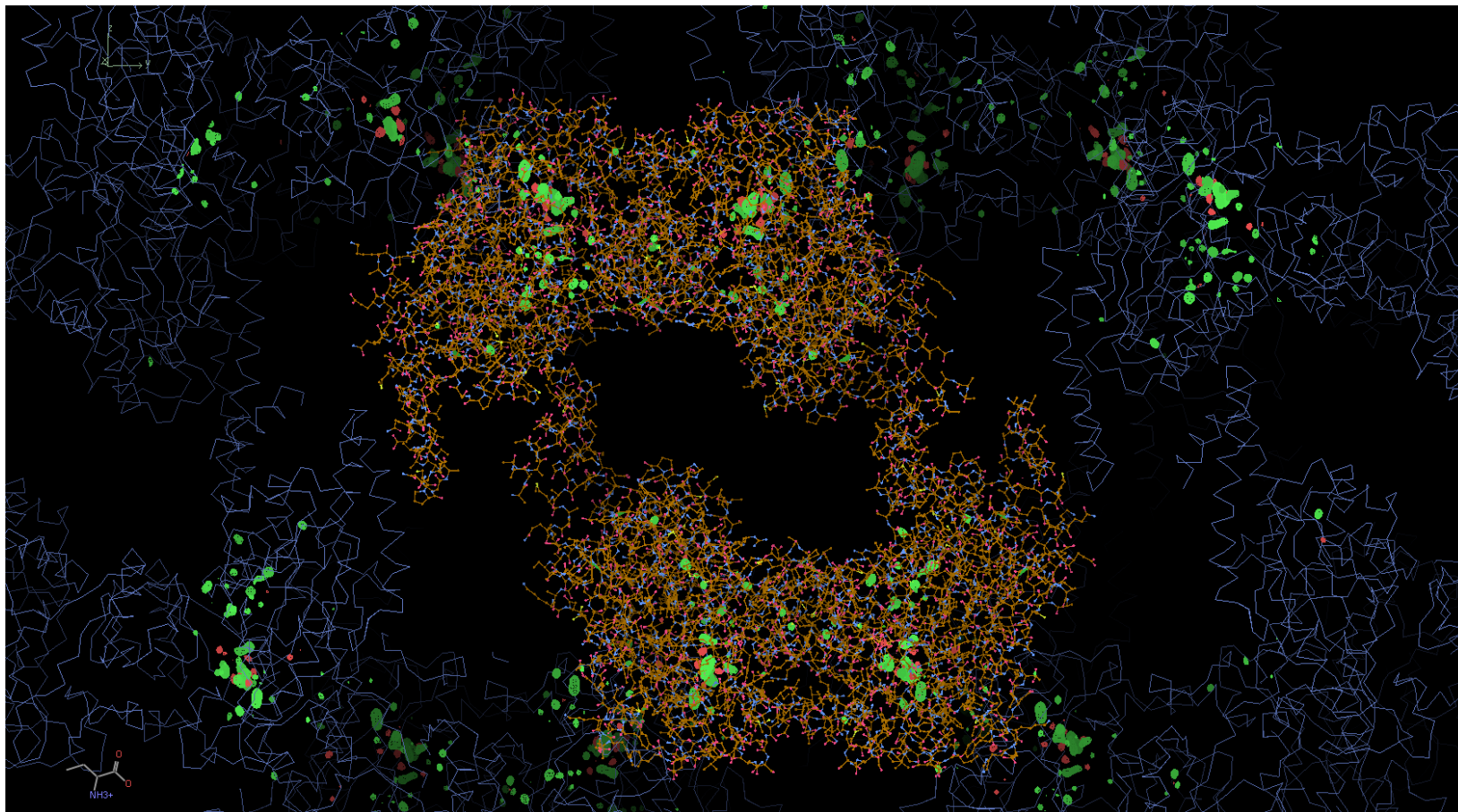
6VZU

de-carboxylation of ASP/GLU and compound

not every carboxy group suffers radiation damage to the same extent

Radiation damage - decarboxylation

6VZU

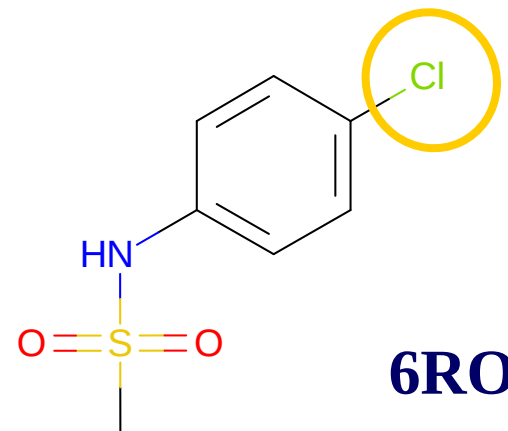
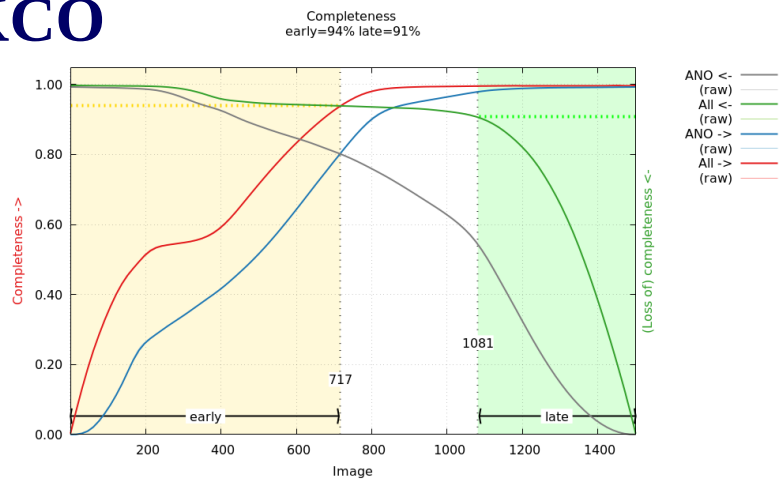


F(early)-F(late) @ 6.0 rms

showing hotspots and “connected” regions

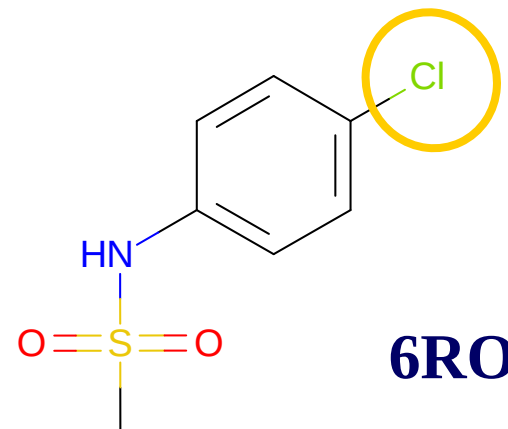
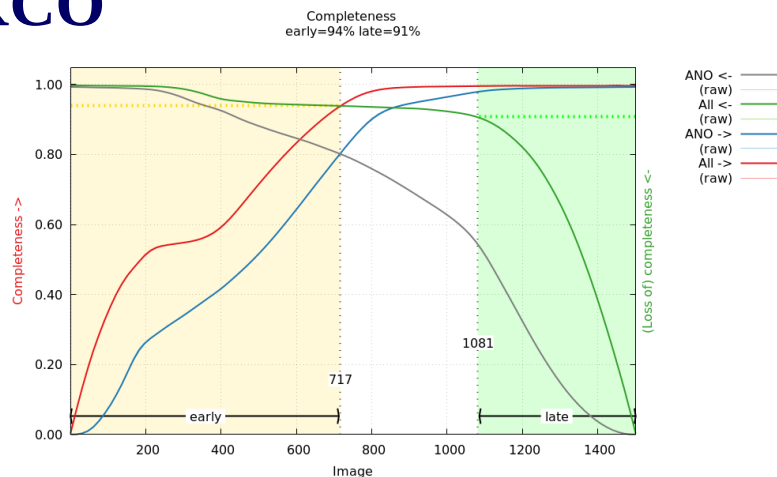
Radiation Damage - improved model parametrisation

5KCO



Radiation Damage - improved model parametrisation

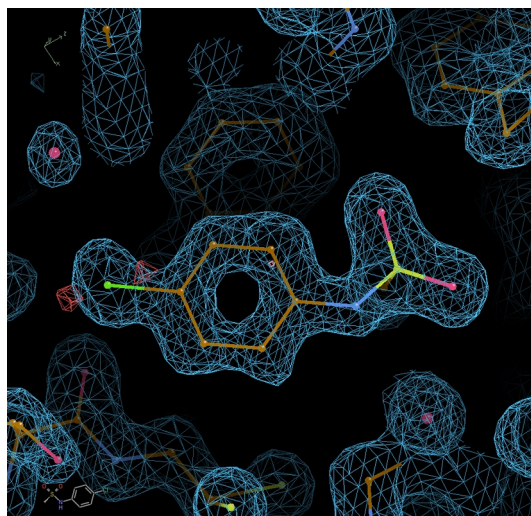
5KCO



2mFo-DFc @ 1.0
rms

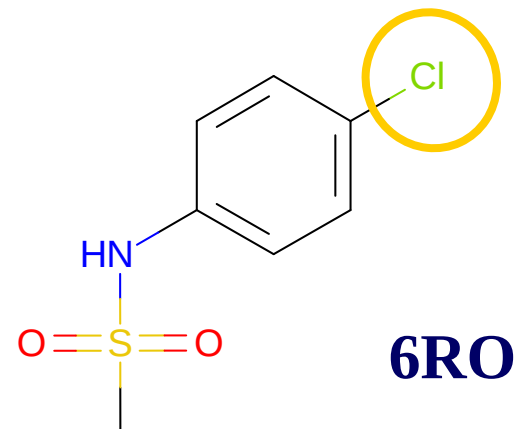
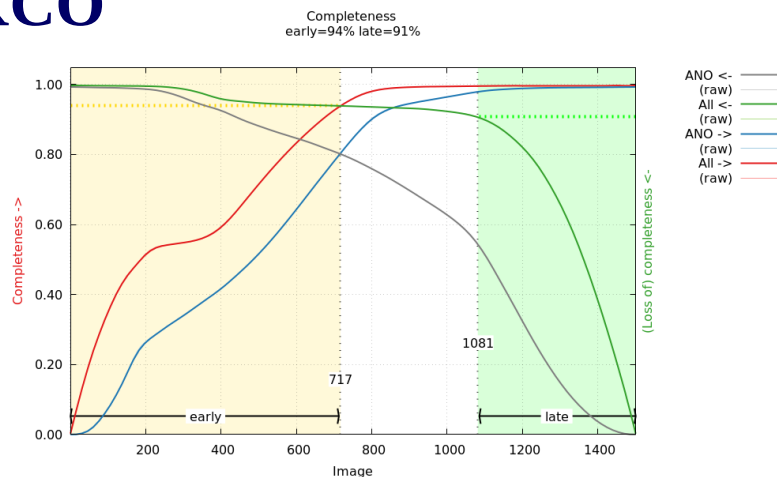
mFo-DFc @ 3.5
rms

very
clean →



Radiation Damage - improved model parametrisation

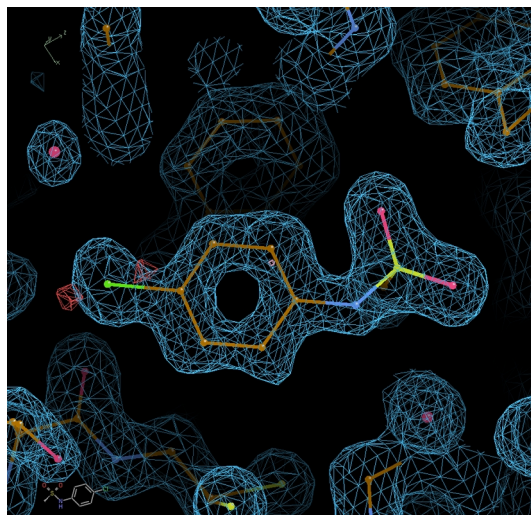
5KCO



2mFo-DFc @ 1.0
rms

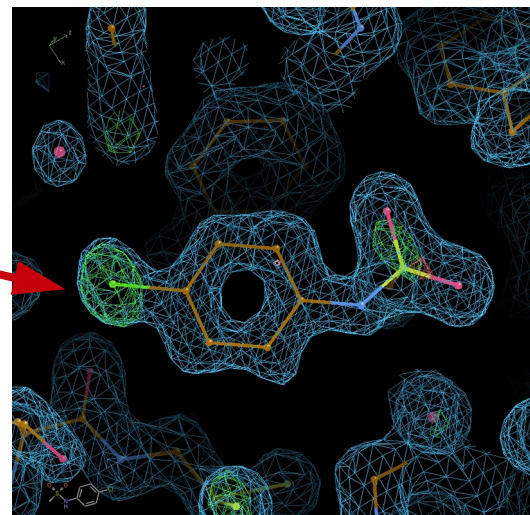
mFo-DFc @ 3.5
rms

very
clean

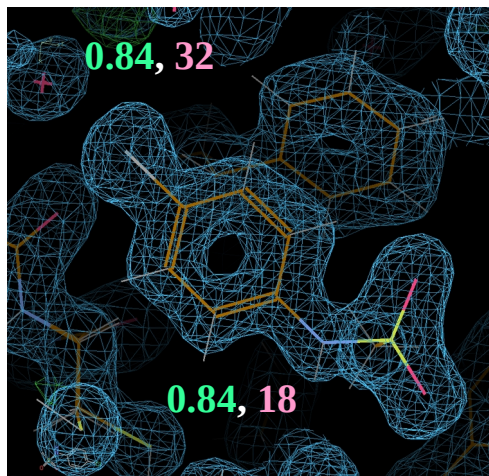
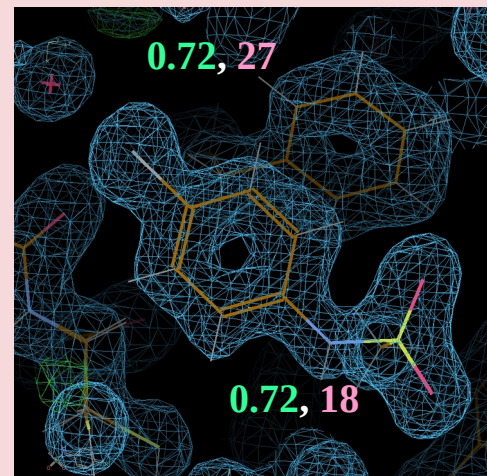
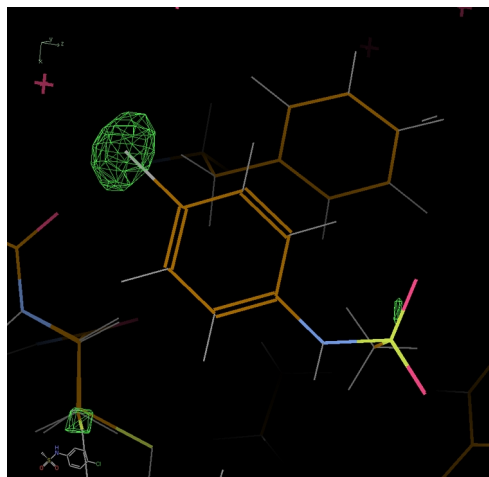
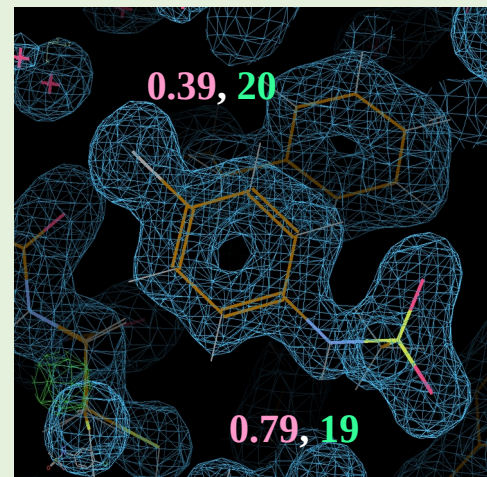


F(early)-F(late)
@ 4.0 rms

strong
peak on
chlorine



Deposited

Re-processed
single OCCF(early)-F(late)
@ 5.0 rmsRe-processed
split OCC

-
- ❑ **Careful processing of data** can be crucial (not everything is Lysozyme)
 - A **good collection strategy** (that adapts to crystal quality, SG, orientation, detector, beamline, goniostat) will result in better data, better density and better models
 - ❑ Collect all **prior knowledge**
 - Geometric restraints (good ligand dictionaries)
 - Similarity restraints (NCS, targeting)
 - Occupancy
 - Correct formfactors (fluorescence scan)
 - ❑ Refine to **convergence**
 - Otherwise difference maps can be hard to interpret
 - And statistics like R/Rfree are (fairly) meaningless
 - ❑ What happened to crystal during data collection is still (very) important at this stage
 - **Radiation damage**: F(early)-F(late) maps
 - **Anisotropy** (STARANISO)
 - ❑ Often the defaults (quick click) work well ... but to get the correct structure interpretation the **best/correct parametrisation and refinement is important**
-

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 - ❑ CCDC
 - ❑ CCP4

 - ❑ Global Phasing Consortium members
 - ❑ ... many, many users & collaborators

<https://www.globalphasing.com/>

<https://www.globalphasing.com/buster/wiki/> (Tutorials etc)