

# Bioinformatics for structural biologists

Prof Dan Rigden



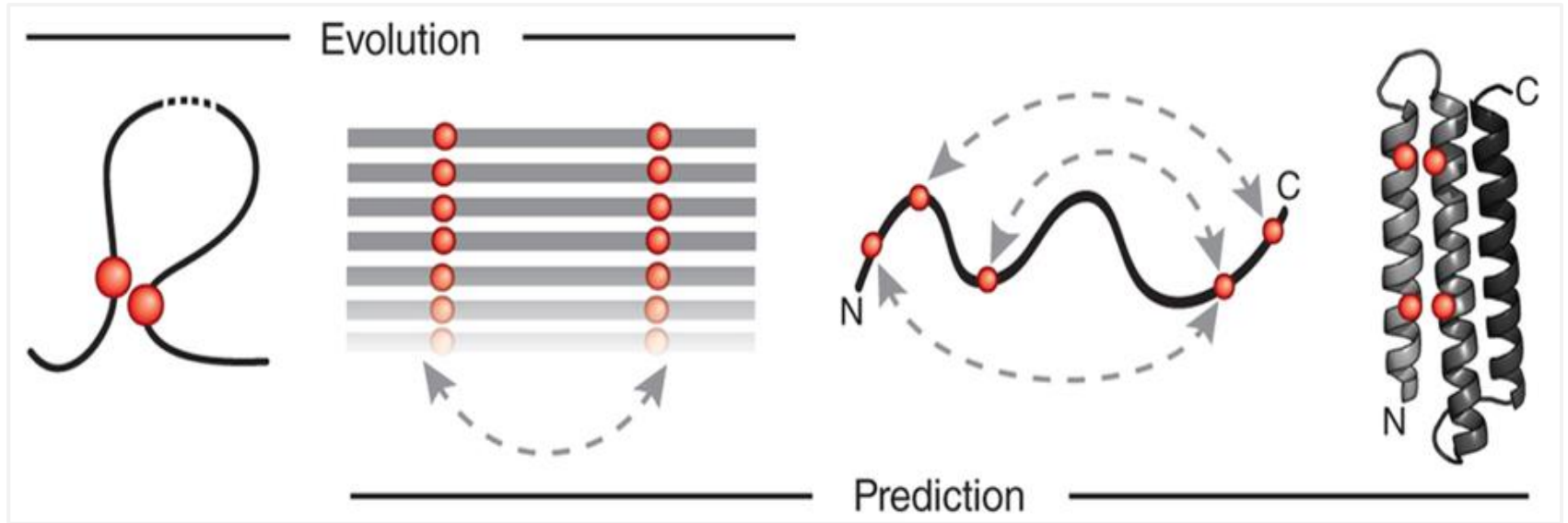
- Introduction
  - Predicting contacts from sequences using evolutionary covariance
  - Deep Learning-based structure prediction eg AlphaFold 2
- Bioinformatics throughout structure determination
  - Predicting domain structure
    - Construct design, experimental strategy
    - Protein engineering
  - ~~Predicting tertiary structure *ab initio* for MR~~
  - Quaternary structure and protein interactions
  - Finalising the structure, validation
  - Structure-based function interpretation
    - Majoring on easily available servers/predictions
    - New Deep Learning-based methods
    - Case study from Structural Genomics (if time!)
  - The sequence alignment in your paper...
- Cross-cutting messages
  - Using multiple methods for a task is good
  - CCP4 has many useful options

# Introduction: Evolutionary covariance and AF2/RF

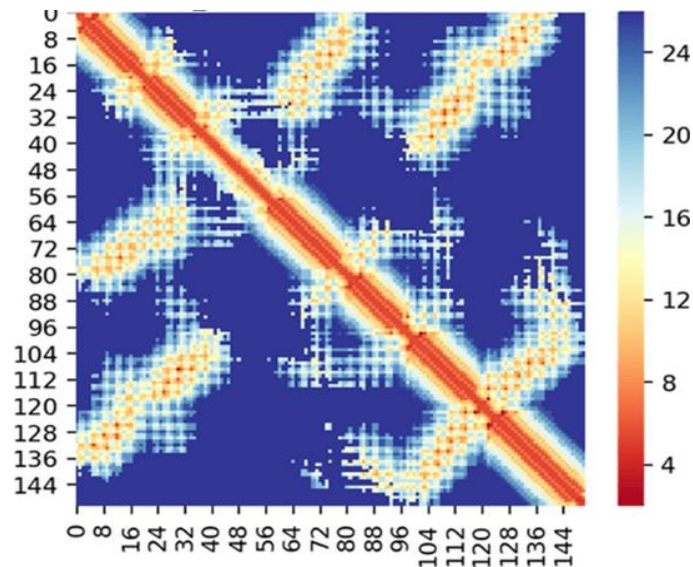
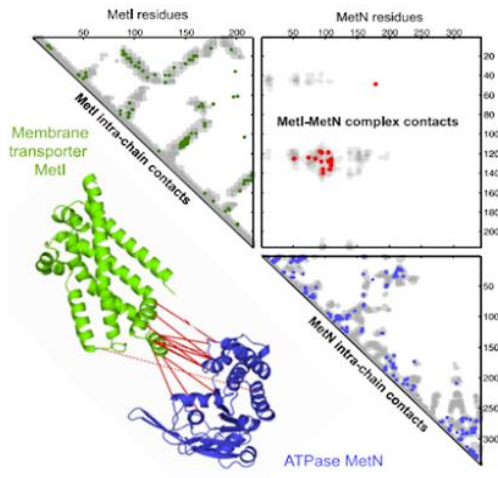
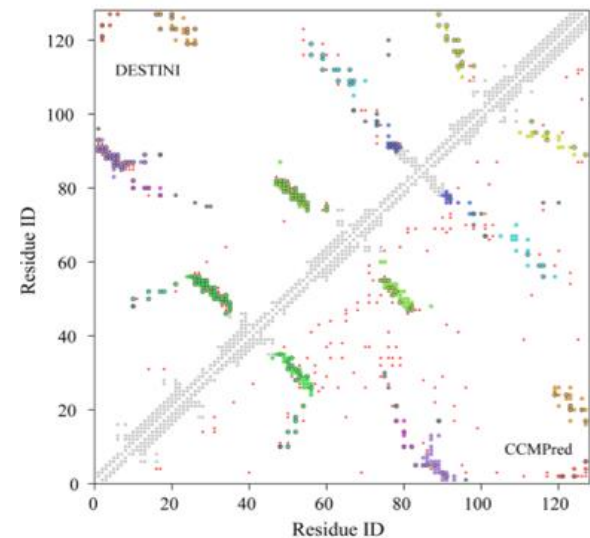
Predicting contacts and distances between residues

Deep Learning-based structure prediction methods

# Evolutionary covariance



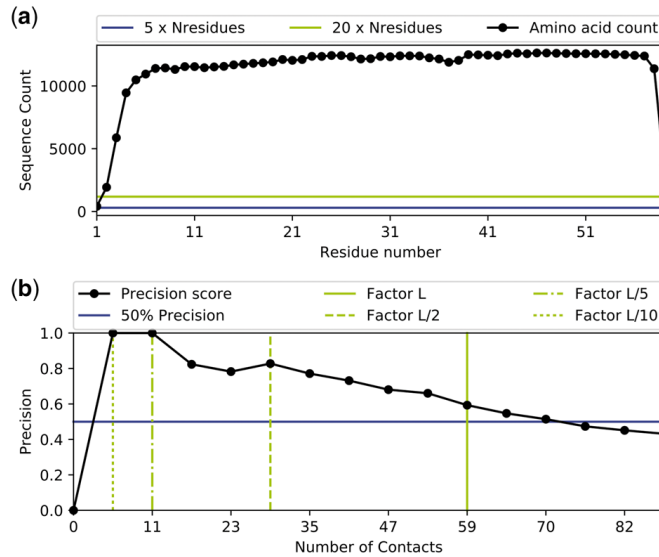
# Predicting contact maps and distograms



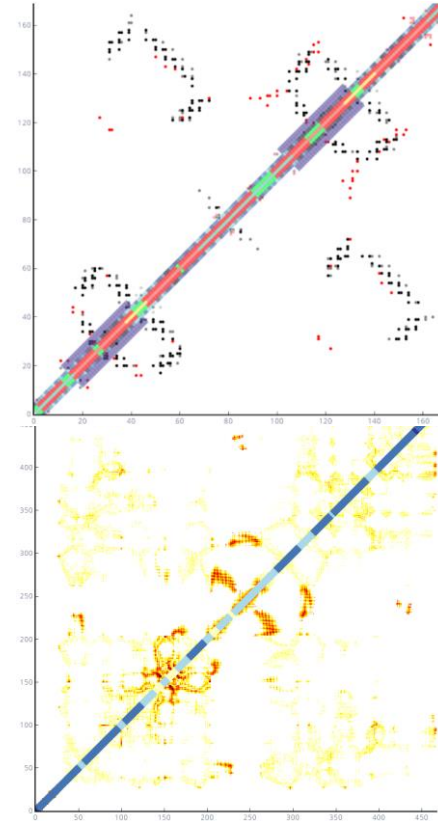
Drove modelling by: EVFold, DMPfold etc

trRosetta, AlphaFold (1)

# ConKit and ConPlot.org



Simkovic *et al.* (2017) Bioinformatics **33** 2209

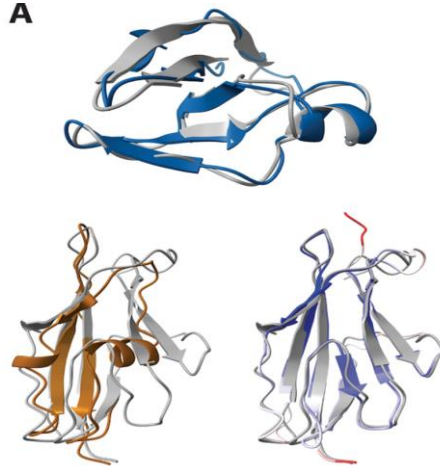


Sánchez Rodríguez *et al.* (2021) Bioinformatics btab049

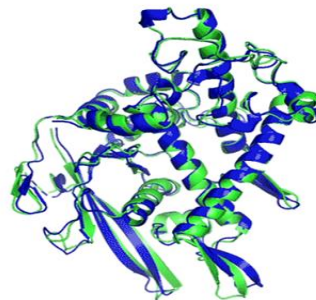
# Multiple methods: RoseTTAFold and AlphaFold 2

- Still use information including covariance from MSAs but networks learn to extract information without imposition of a particular model.
- End-to-end networks produce models directly rather than two separate steps

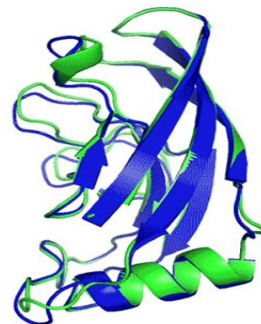
A



Baek et al (2021)  
Science 373,871



T1037 / 6vr4  
90.7 GDT  
(RNA polymerase domain)

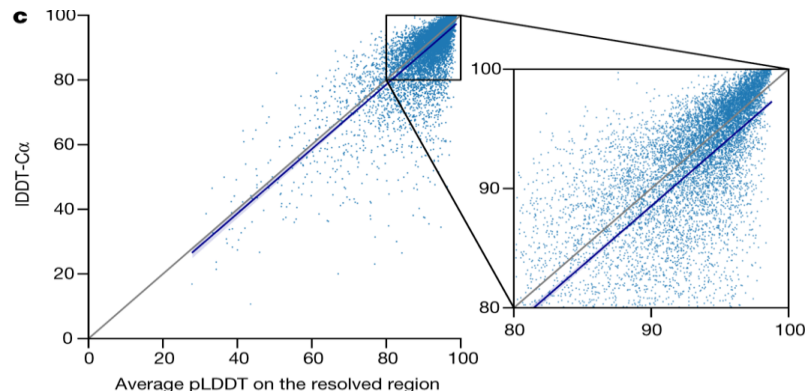


T1049 / 6y4f  
93.3 GDT  
(adhesin tip)

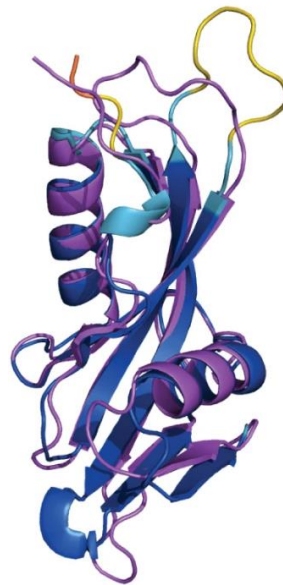
Jumper et al (2021) Nature 596,583

# Multiple methods: RoseTTAFold and AlphaFold 2

- Often amazing quality but not always... Need good MSA or template!

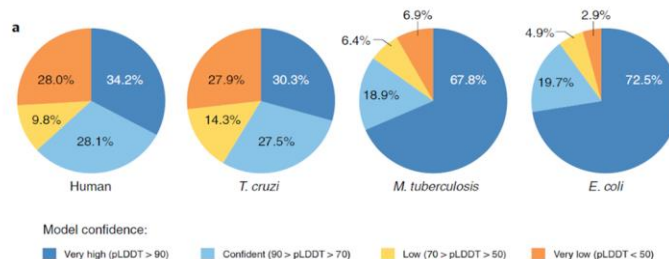


Jumper *et al.* (2021) Nature 596, 583



Phosphatase crystal  
structure vs AF2 model

Insulin crystal structure  
vs AF2 model



Thornton *et al.* (2021) Nature Medicine 27, 1666



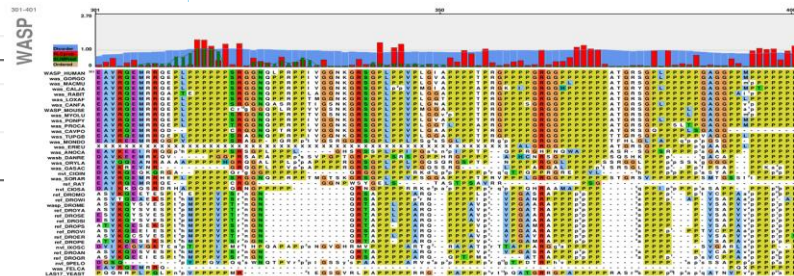
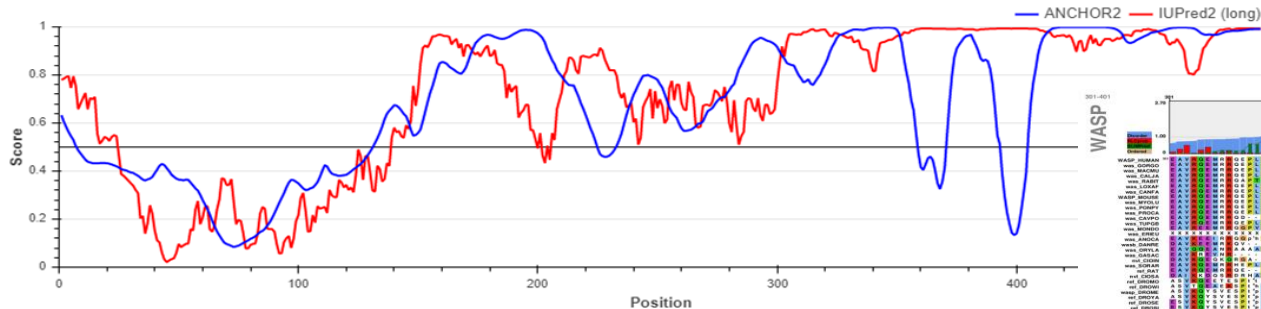
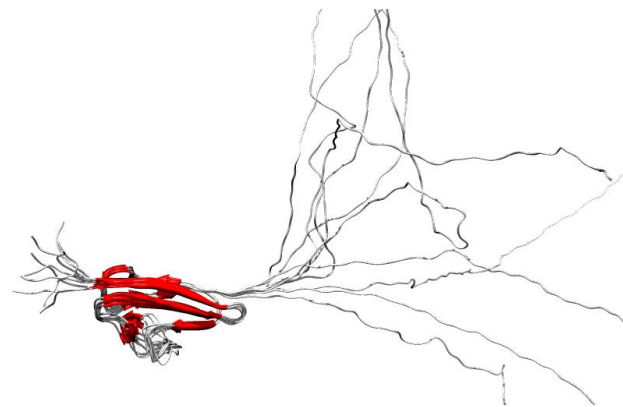
# Domains, construct design and solubility

Which part to express for crystallisation?

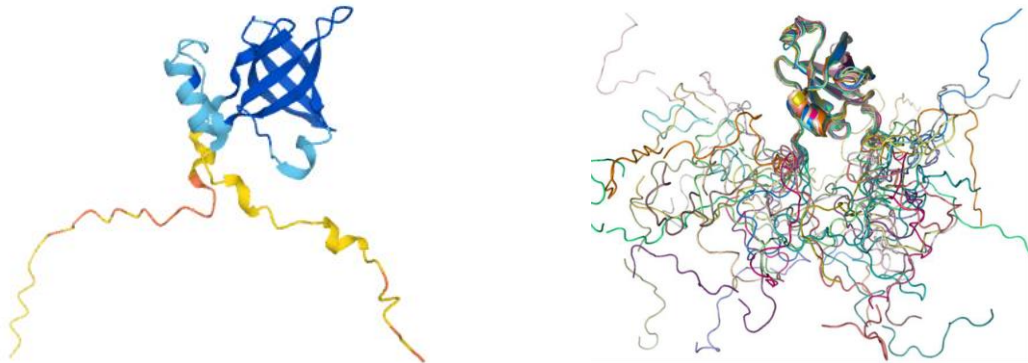
If necessary, can solubility etc be improved?

# Intrinsic disorder prediction

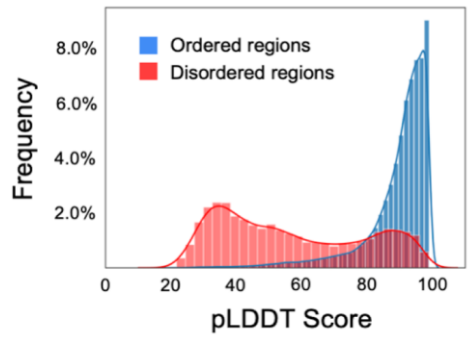
- Not all proteins and protein regions fold into stable structured domains. ID proteins and regions will not crystallise (alone)
- There are many predictors, all performing roughly equally well
- I recommend AIUPred (fast) and MetaDisorder (slow but good)
- Can also look for short interaction motifs in ID regions (ANCHOR, SlimPred)



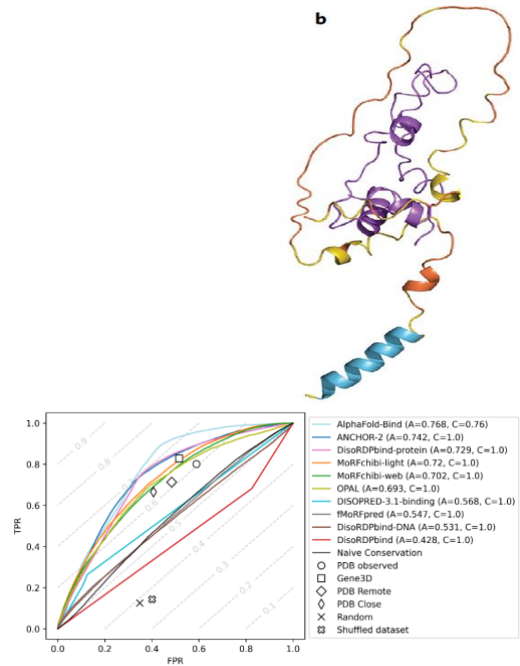
Most low-confidence AF2 regions are disordered but they may just be plain wrong



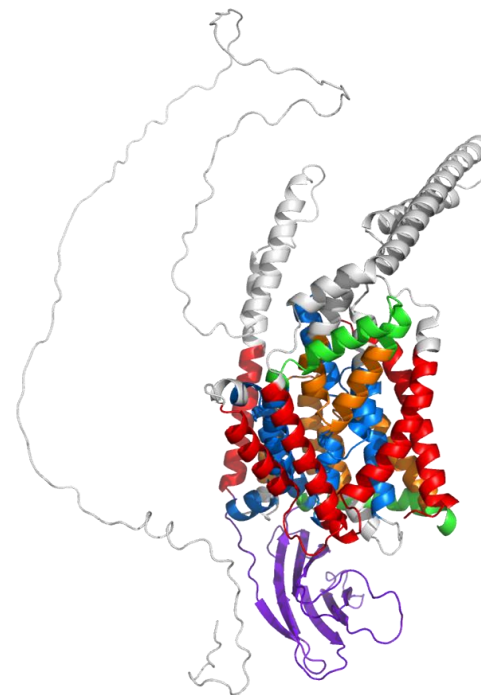
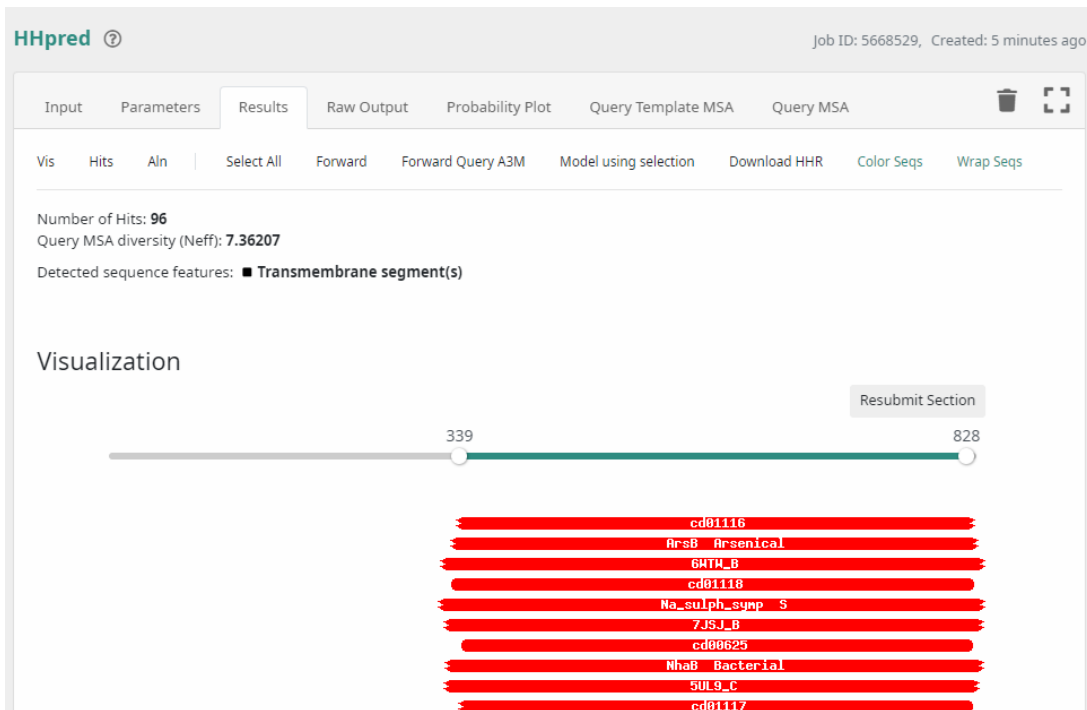
Relatively high pLDDT regions in AF2 disordered regions may predict interaction motifs



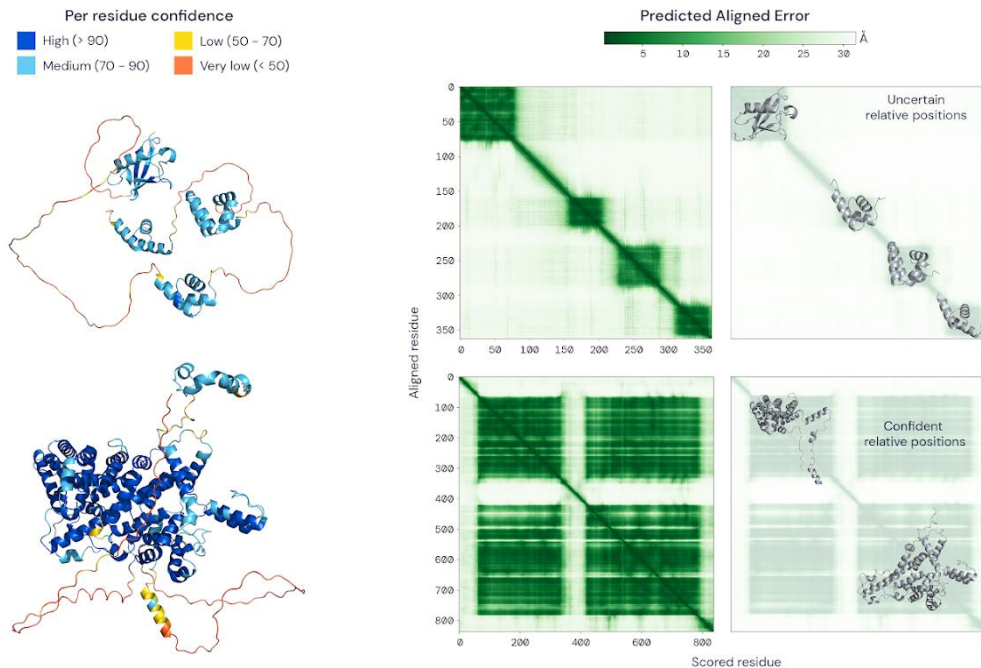
Binder et al (2022) COSB 74, 102372



# Beyond sequence matching: AF2 for domain discovery



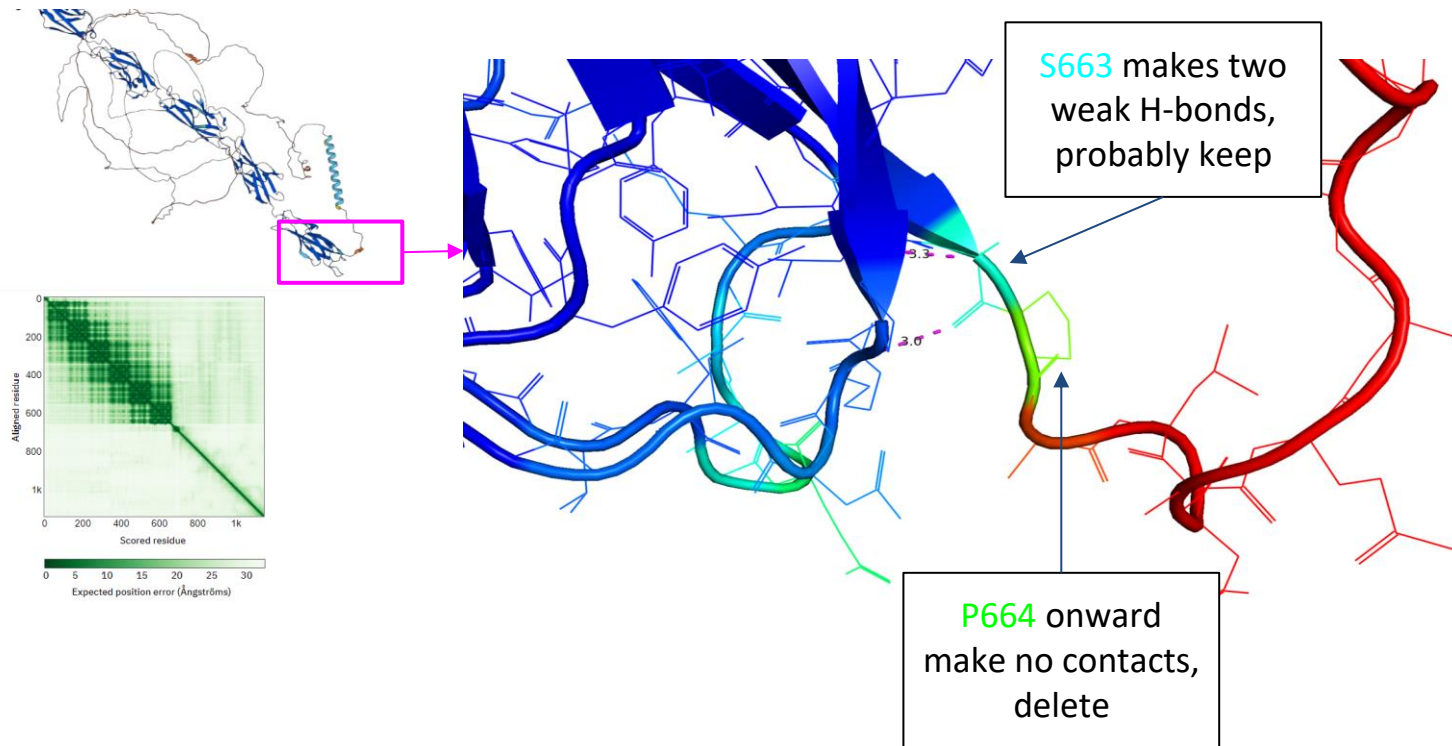
# AF2 for domain boundary prediction



**Slice'n'Dice** uses the PAE or other kinds of clustering to split structures into domains for MR

<https://deepmind.com/research/publications/2021/enabling-high-accuracy-protein-structure-prediction-at-the-proteome-scale>

# Fine details too, if reliability allows



# Recognising homology using a structure

FoldSeek can search PDB, AFDB, ESMAtlas, BFVD using a structure in seconds

### Search Settings

Databases

- ☒ AlphaFold/UniProt50 v3
- ☒ AlphaFold/UniProt50-best v3
- ☒ AlphaFold/Swiss-Prot v2
- ☒ AlphaFold/Proteome v2
- ☒ PDB100 220722
- ☒ GMGCL 2204

Mode

☒ 3D/AA

☐ TM-align

[Taxonomic filter](#)

**mamm**

- Mammalia
- Mammantavirinae
- Lassa mammarenavirus
- Mammarenavirus
- Mammalian orthoreovirus
- Mammalicoccus
- Mammalicoccus sciuri

[illegible]

<https://search.foldseek.com/>

Van Kempen et al. (2022) Nature Biotech  
<https://doi.org/10.1038/s41587-023-01773-0>

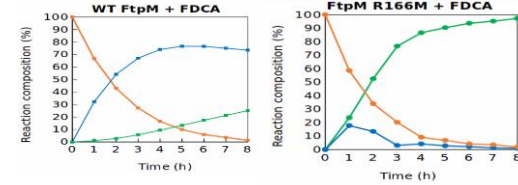
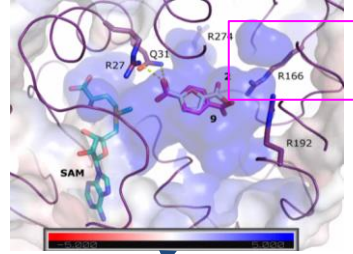
# Protein engineering

Where you have confidently interpreted a model as reliable then you can treat it almost like a crystal structure...

Rational enzyme engineering using an AF2 prediction

Combining AF2 predictions with other structure-based methods eg for stability prediction. AlphaMissense is another option

Predicting a stable fusion protein based on analysis of a complex structure prediction

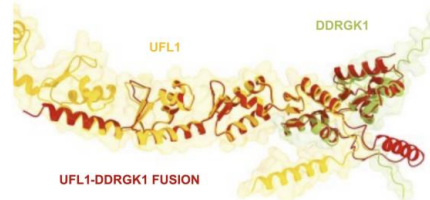
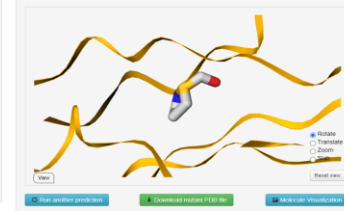


Ward et al (2022) Angewandte Chemie 61,e202117324

## DUET - Protein Stability Change Upon Mutation

mCSM Predicted Stability Change ( $\Delta\Delta G$ ):  
-0.779 kcal/mol (Destabilizing)  
SDM Predicted Stability Change ( $\Delta\Delta G$ ):  
-0.38 kcal/mol (Destabilizing)  
DUET Predicted Stability Change ( $\Delta\Delta G$ ):  
-0.559 kcal/mol (Destabilizing)

Mutation:  
Wild type: PRG  
Position: 881  
Residue type: ARG  
Chain: A  
Secondary structure: Loop or Irregular

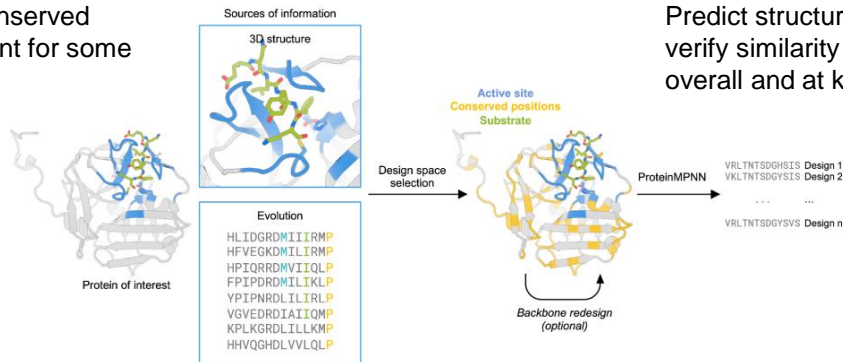


Banerjee et al (2022)  
BioRxiv  
<https://doi.org/10.1101/2022.09.15.508077>

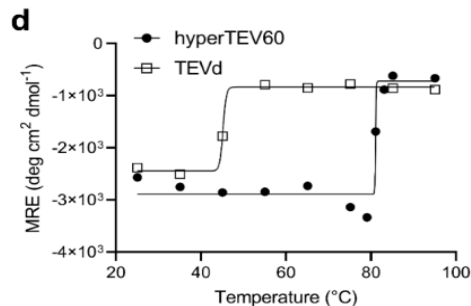
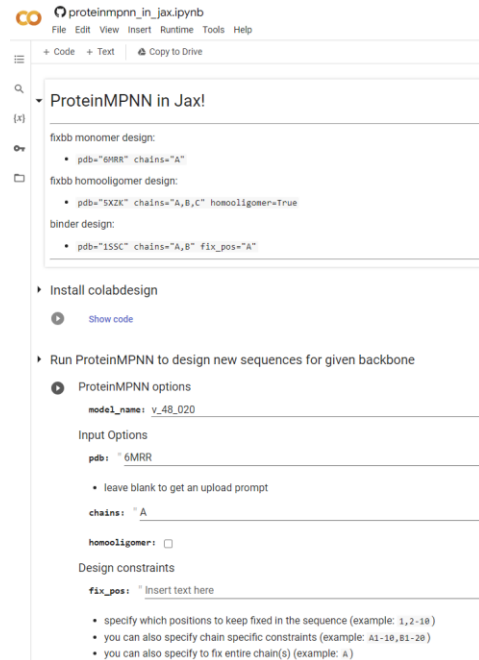


# Protein engineering with deep learning

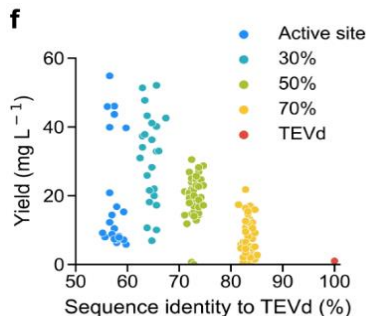
Identify and retain residues near ligand (functionally important) and conserved positions (important for some reason)



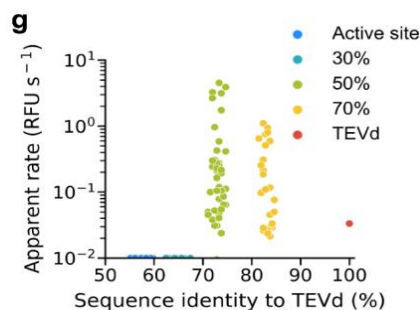
Allow ProteinMPNN to fill in the gaps and design variants.  
Predict structures with AF2 and verify similarity to starting point, overall and at key positions



Better thermostability



Better expression



Better activity

# Tertiary structure

AF2 and RF output as search models for MR

# AF2/RF and Molecular Replacement

MR *is* a kind of structure prediction so the availability of accurate models of most proteins has impacted structure solution hugely

Similarly, accurate models can be used to interpret cryo-EM maps

See other talks for how to find and deploy AF2 models with [MrParse](#), [MrBUMP](#), [Slice'n'Dice](#), [ARCIMBOLDO\\_SHREDDER](#) etc

# Getting diversity in your models

This will be needed for **hard cases** and for cases where **multiple conformations** are accessible or sought

## Ways to sample conformation more broadly

- **Network dropout** (eg `num_samples` and/or `is_training` on the advanced colabfold page)  
[https://colab.research.google.com/github/sokrypton/ColabFold/blob/main/beta/AlphaFold2\\_advanced.ipynb](https://colab.research.google.com/github/sokrypton/ColabFold/blob/main/beta/AlphaFold2_advanced.ipynb)
- Feed AF2 **templates** in the 'right' conformation (and maybe ignore MSA features)

de Alamo et al (2022) *elife* 11:e75751

- Deliberately make the input MSA more shallow

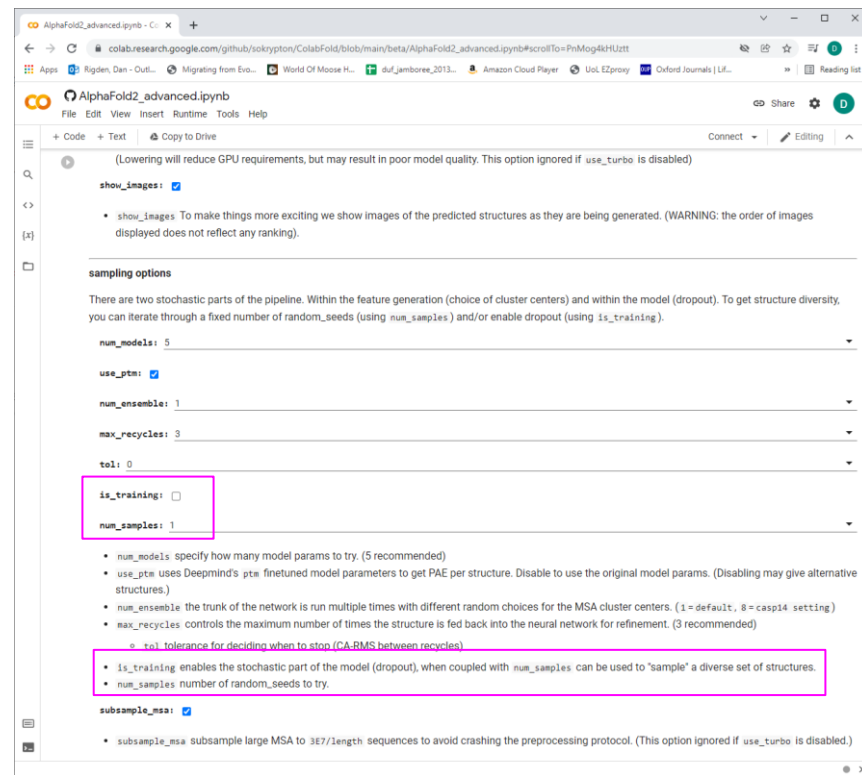
Heo &amp; Feig (2022) PSFB DOI: 10.1002/prot.26382

- Cluster the input MSA and try individual sub-clusters

Wayment-Steele et al (2024) Nature 625, 832

- **Edit the input MSA** to mutate to Ala residue pairs that are driving the 'wrong' conformation

Stein and Mchaourab (2022) PLoS CB 18: e1010483.

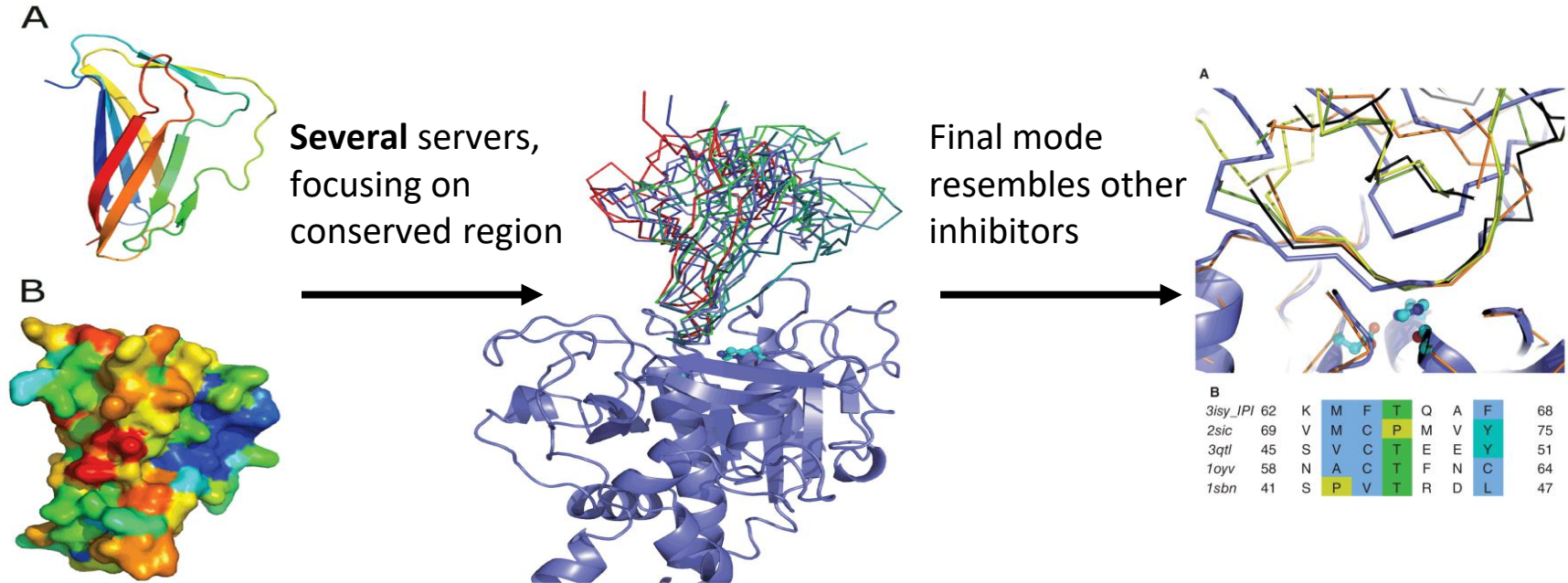


# Quaternary structure and ligand interaction

# Predicting protein-protein interactions

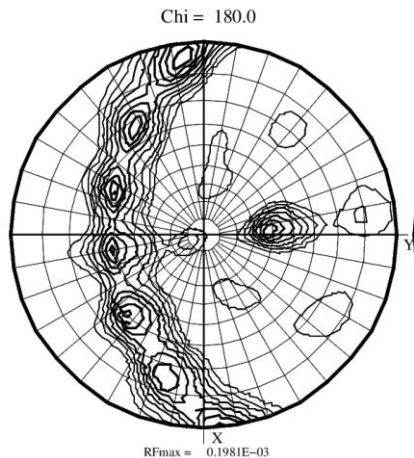
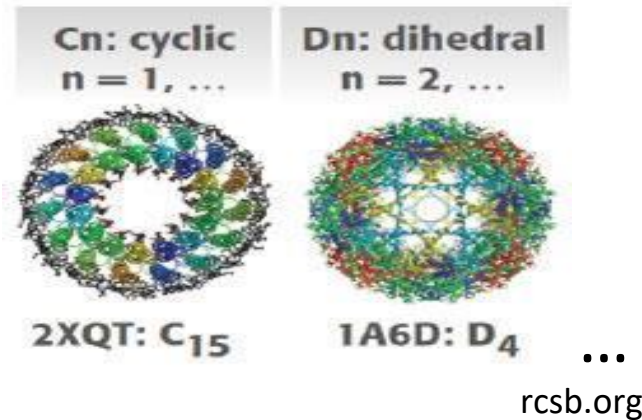
- Relevant to MR eg proteins A and B are cocrystallised but neither alone solves. An accurately predicted complex, being larger, might solve
- Many docking methods predict complexes based on steric complementarity plus other scoring functions
- Recommendable servers include
  - ClusPro, the best performing docking method
  - Haddock, which has a good server with different modes
  - Each allows inclusion of other information eg known interface residues. RF/AF2 do not (easily, yet)
  - Symmetric docking at ROSIE server. Also unavailable in RF/AF2

# Multiple methods in bioinformatics: *B. subtilis* IPI docking to protease

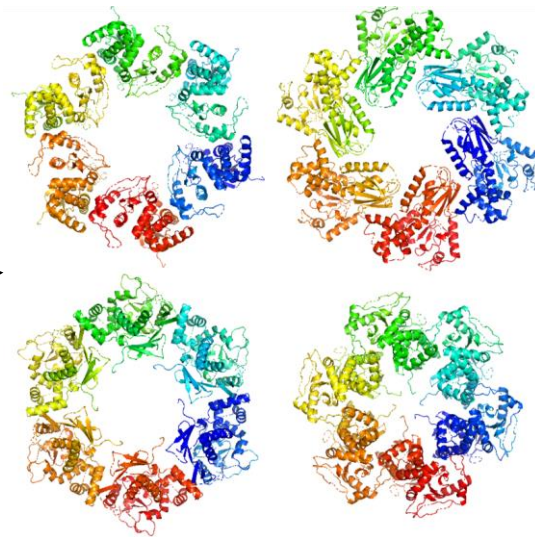


# ROSIE symmetric docking for oligomers

- Only cyclic ( $C_n$ ) or dihedral ( $D_n$ ) symmetry at server
- Clues from self-rotation function may be available
- AF2 cannot use this information!

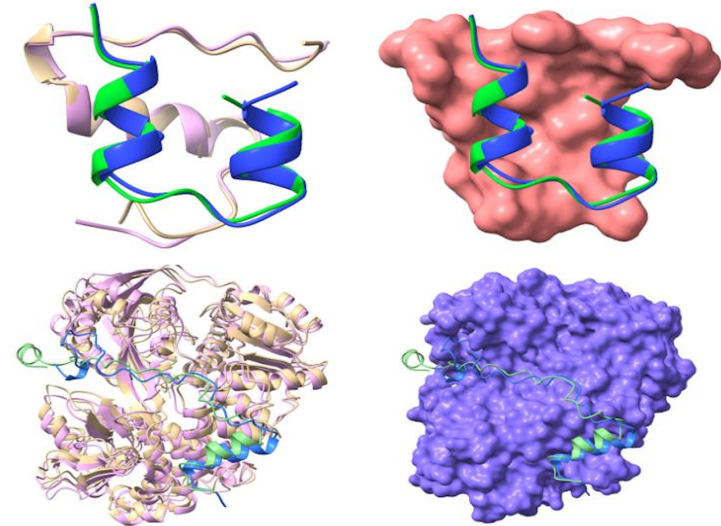
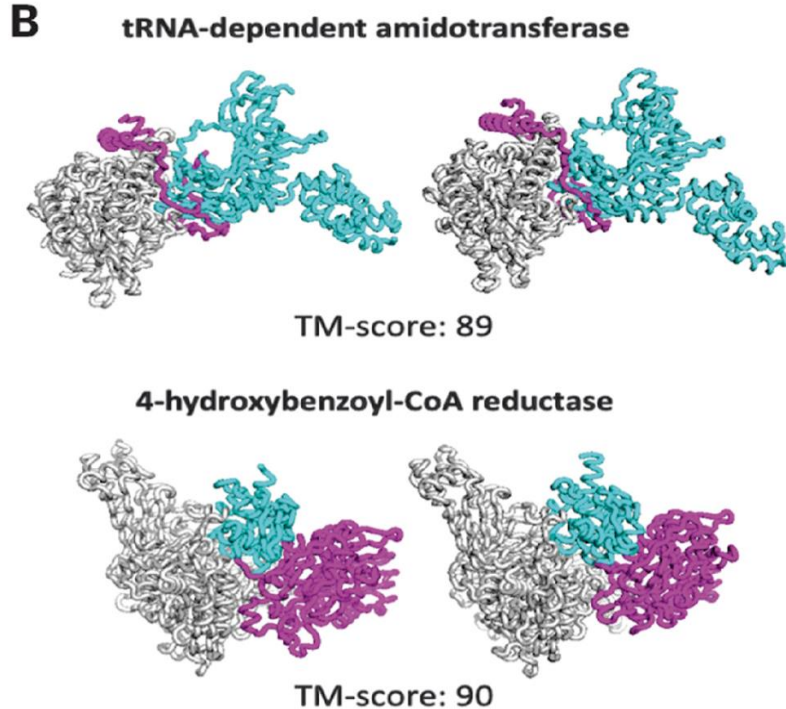


Generate hexamers



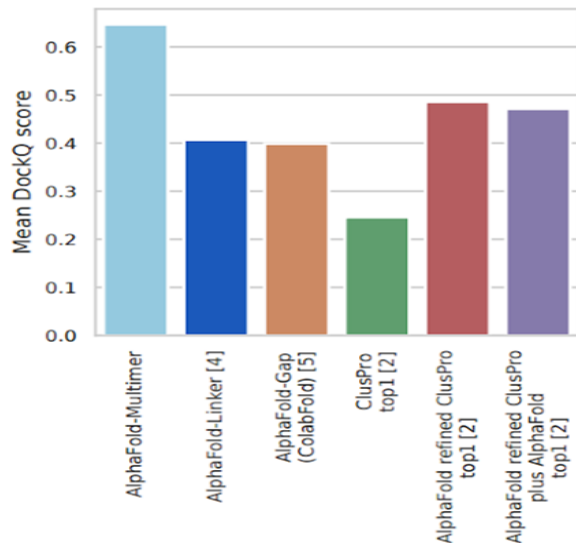


# RF/AF2 to predict complexes

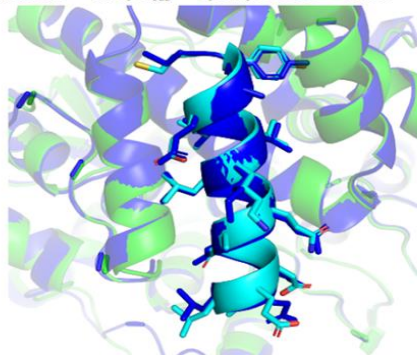


Modelling protein-peptide interactions as separate chains or linked by polyAla are complementary approaches.

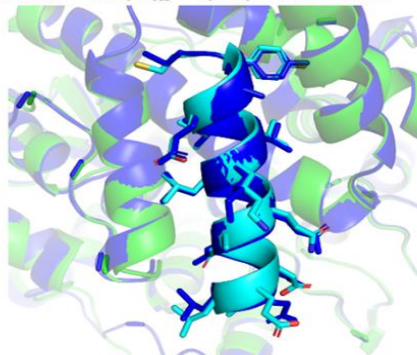
# New multimer-trained AF2



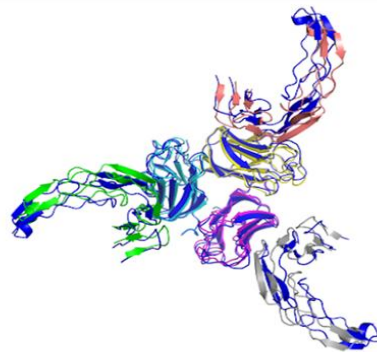
(a) A2B2C2 heteromer  
TM-score = 98.0,  $N_{\text{res}} = 1,246$ , PDB ID = 6E3K



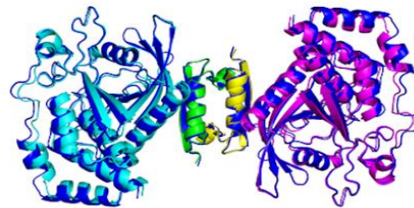
(c) Protein-peptide complex  
TM-score = 96.0, DockQ = 0.948,  
 $N_{\text{res}} = 385$ , PDB ID = 6JMT



(b) A3B3 heteromer  
TM-score = 89.3,  $N_{\text{res}} = 795$ , PDB ID = 7KHD



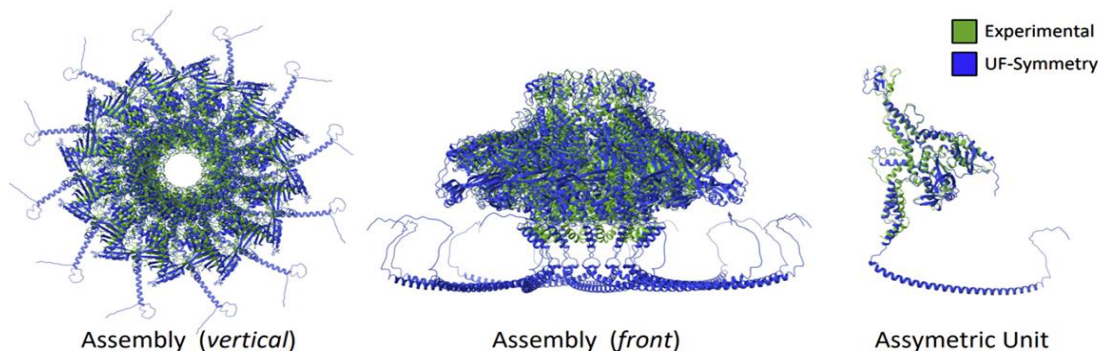
(d) A2B2 heteromer  
TM-score = 98.3,  $N_{\text{res}} = 716$ , PDB ID = 6IWD



AF2 can predict oligomeric state eg asking for 5 copies may produce the natural tetramer + one left over, rather than a pentamer

# Uni-Fold symmetry

- Models a single chain with known symmetry to generate oligomer
- Much quicker and slightly better than other methods on oligomers



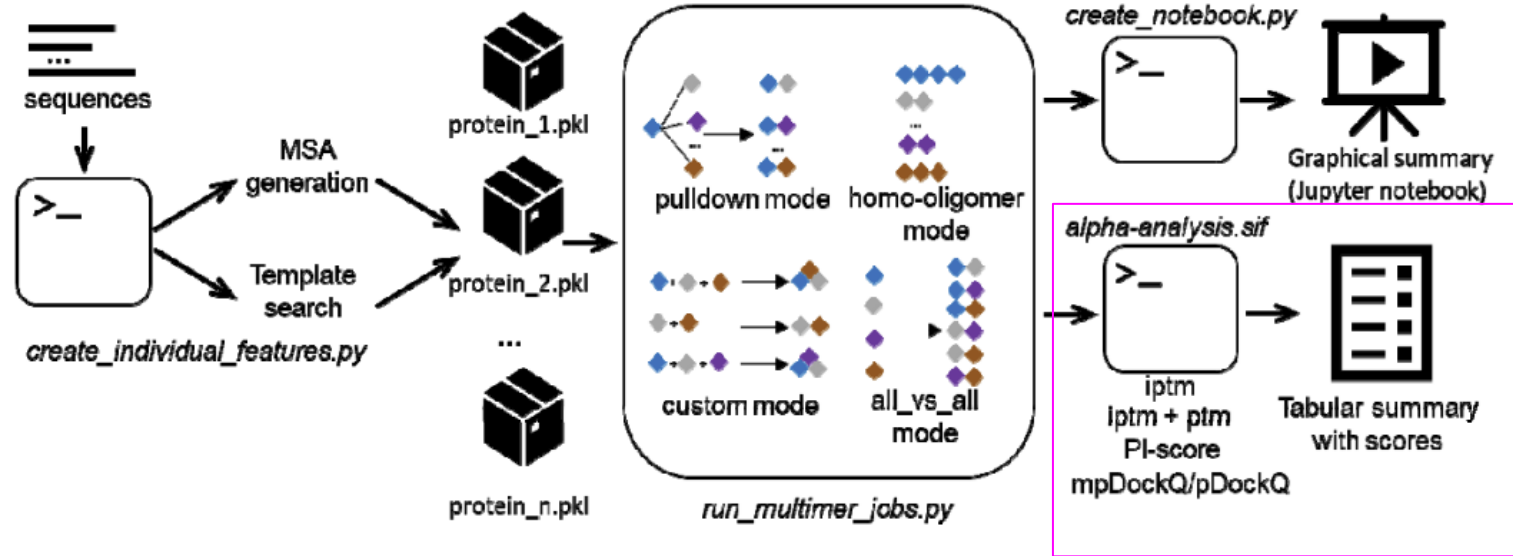
Type	Symmetry	Number of structures
Monomers	–	83,392
Multimers	Asymmetric	27,470
	C2	45,496
	C3	6,037
	C4	1,736
	C5	893
	C6	639
	Larger cyclic	587
	D2	8,571
	D3	2,577
	D4	815
	D5	302
	D6	191
	Larger dihedral	239
	Icosahedral	1,182
	Octahedral	544
	Tetrahedral	475
	Helical	581
	All	98,335
Total	–	181,727

C12 symmetry. AF2 and regular Uni-Fold fail

Li et al (2022) BioRxiv <https://doi.org/10.1101/2022.08.30.505833>

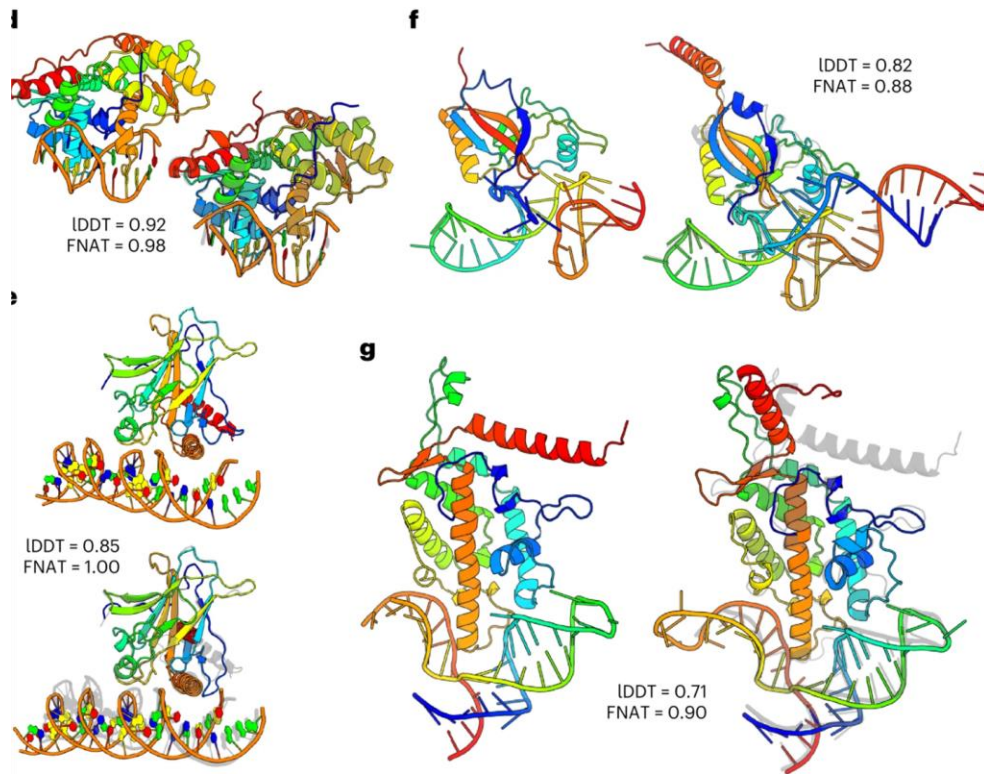
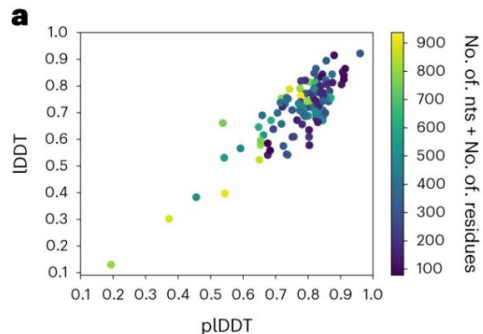
<https://colab.research.google.com/github/dptech-corp/Uni-Fold/blob/main/notebooks/unifold.ipynb>

# AlphaPulldown for complicated cases



# Protein-NA complexes with RoseTTAFoldNA

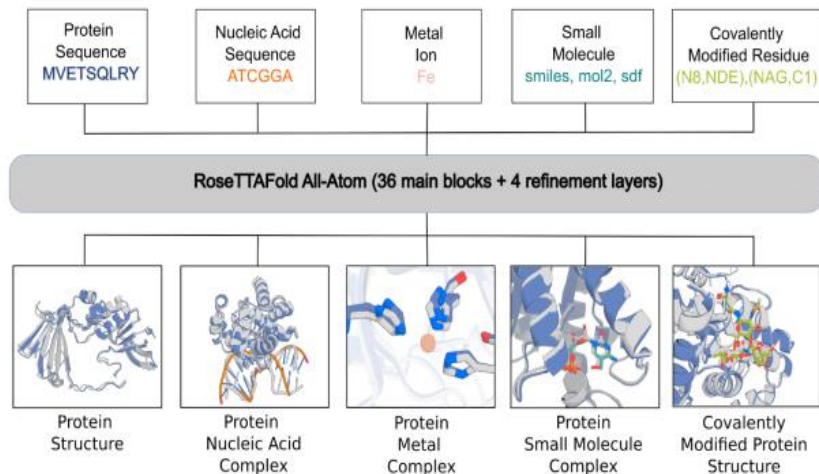
As good as AF2 on proteins; as good as DL-based RNA tools on natural RNAs; USP *was* protein-NA complexes





# Generic protein-ligand prediction

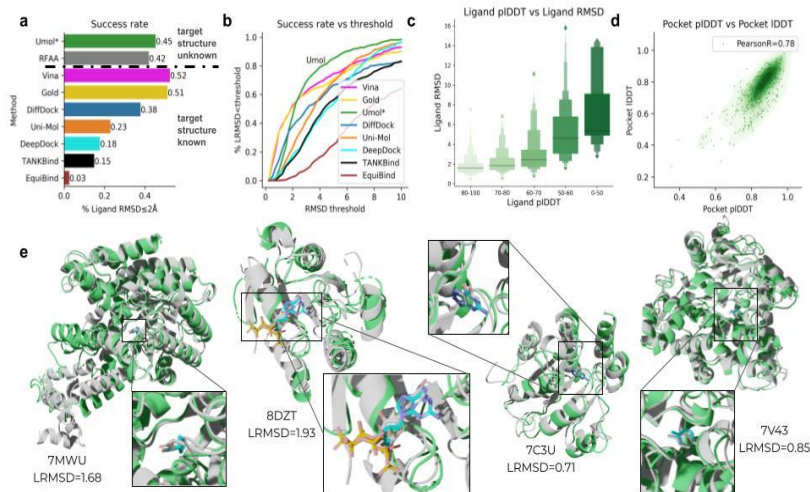
## RoseTTAFold All-Atom



Code

Krishna et al (2024) Science doi: 10.1126/science.adl2528.

## Umol



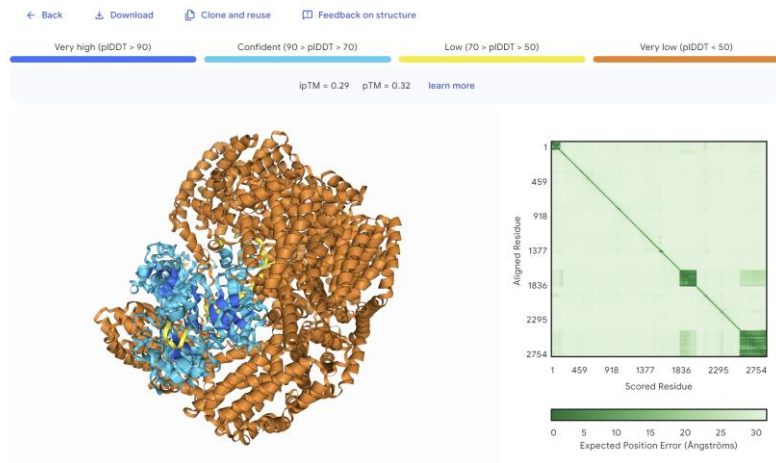
Code, Colab page!

Bryant et al (2024) Nature Communications doi: 10.1038/s41467-024-48837-6

# The latest diffusion methods

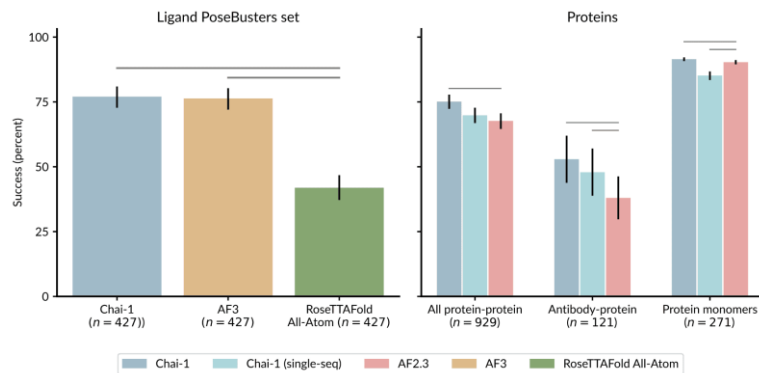
## AlphaFold 3

2024-10-06\_14:05



Not a direct development of AF2. Google's server and now code. Server 20 jobs/day, limited ligand library, template always *on*

## Chai-1



One of five (!) AF3 copycats. Can natively use extra information. Server slow, downloads lack pLDDT. Allows commercial use (Nov 2024)

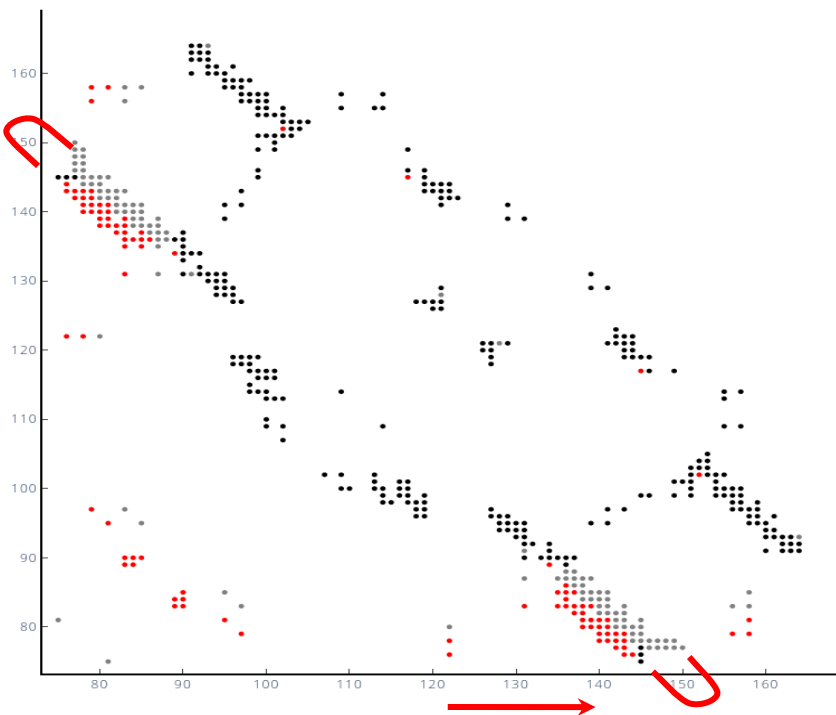
# Finalising the structure

Does it contain any residual errors?

What is the biologically relevant quaternary structure?

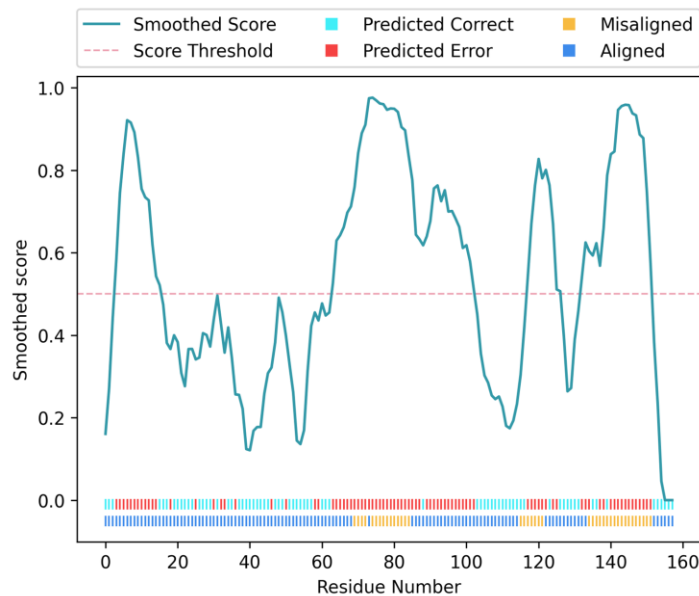


# New covariance-based metrics for model validation



+4 res.

Contact map overlap picks up register errors, SVM detects errors in general

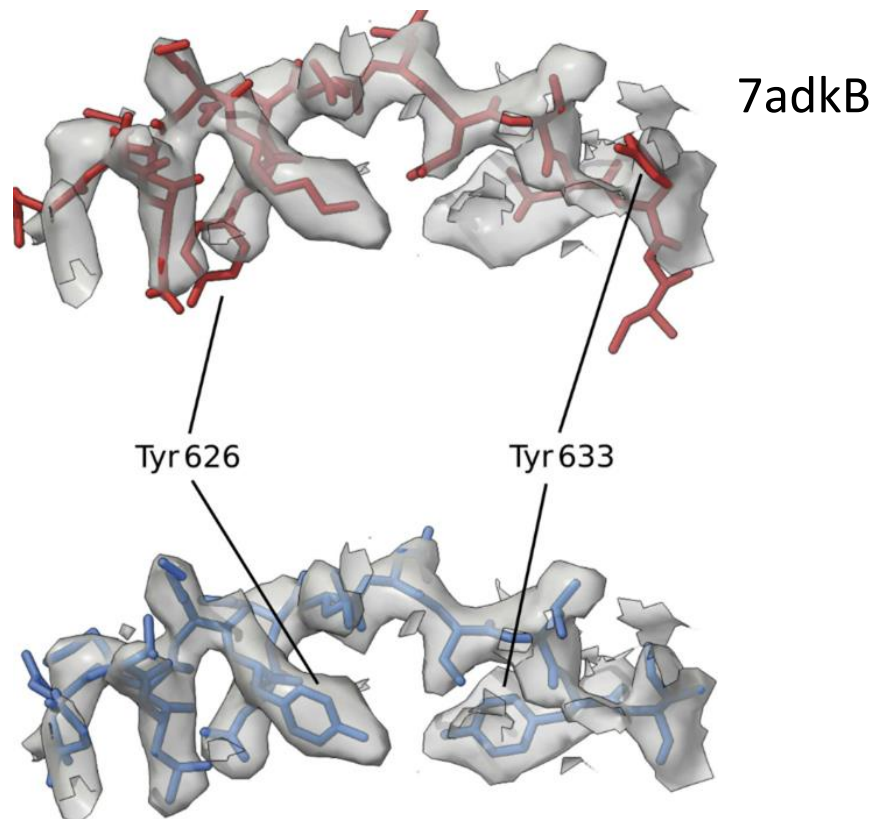


```
$ conkit-validate model.pdb prediction_af2.pkl sequence.fasta
```

# New covariance-based metrics for model validation

## Key points

- Is resolution- and modality-independent
- Suggests the register shift required to correct
- Limits
  - Still needs sufficient sequences for covariance signal
  - Fold-switching proteins
- 3 FP filters
- Errors validated by map, where resolution allows

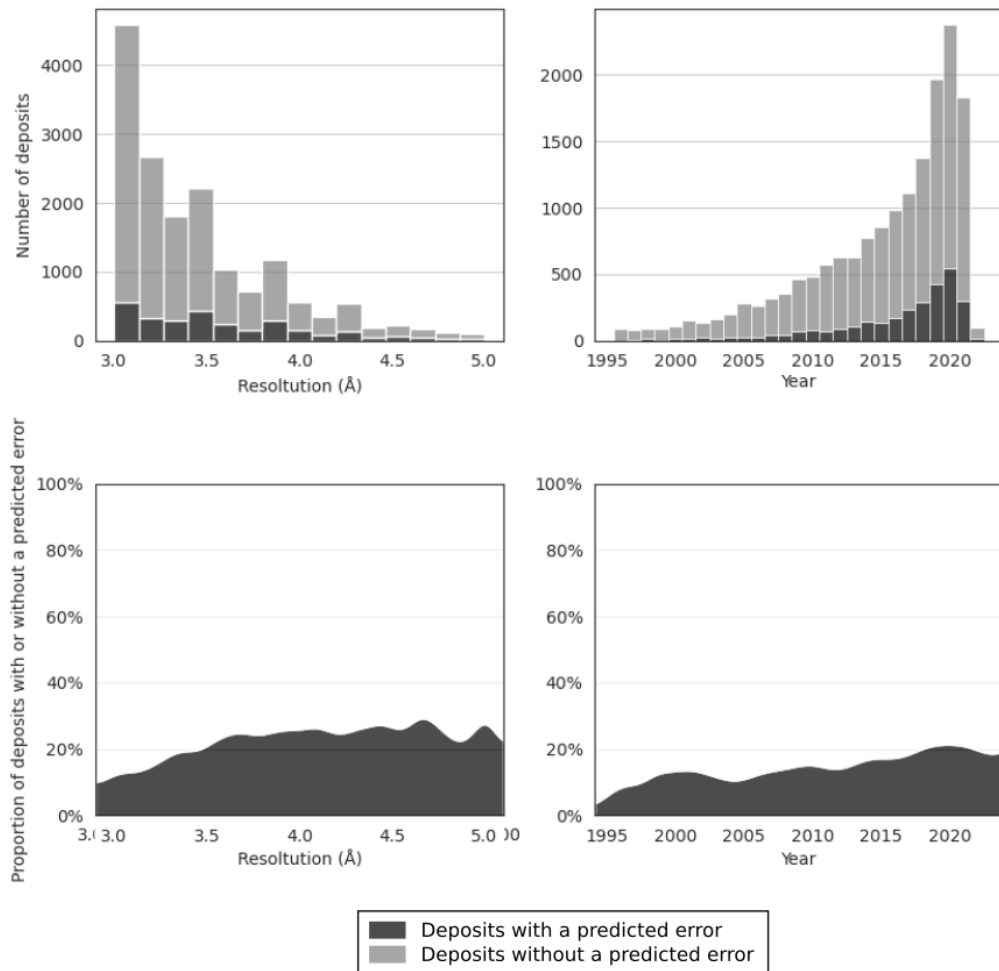


# PDB-wide screen

Lower resolution structures tend to contain more errors, but dependence is quite modest

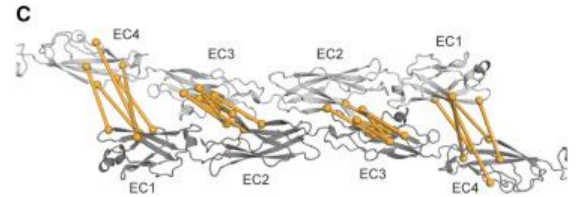
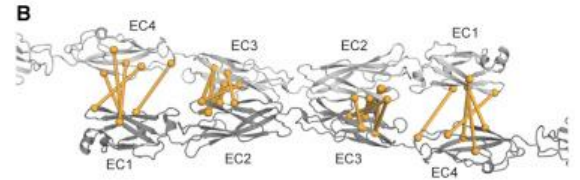
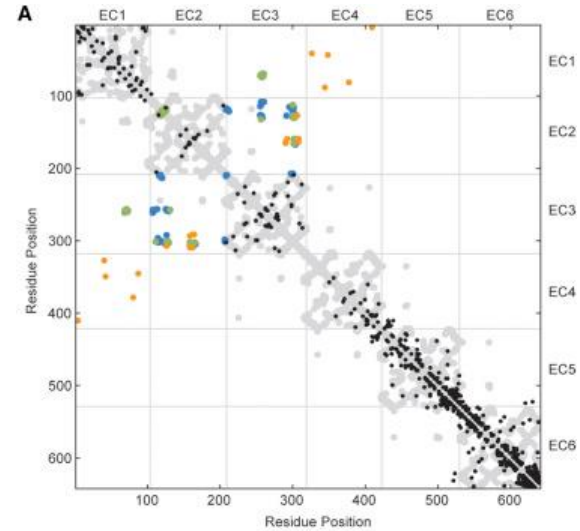
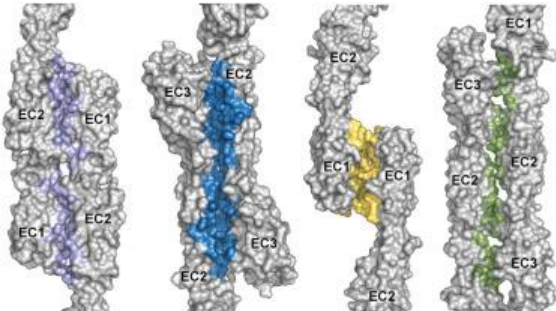
Error rate not declining recently  
- are experimentalists continually tempted to lower resolutions?

Cryo-EM structures contain more errors than crystal. Issue of local resolution, or maybe just because they are bigger?



# Validating crystal structure contents

- **PISA** is an excellent general method, but contact predictions help in some cases
- Crystal showed various ways in which protocadherins could interact
- Predicted contacts supported only some of the modes
- **PISA-cov** is on the way...



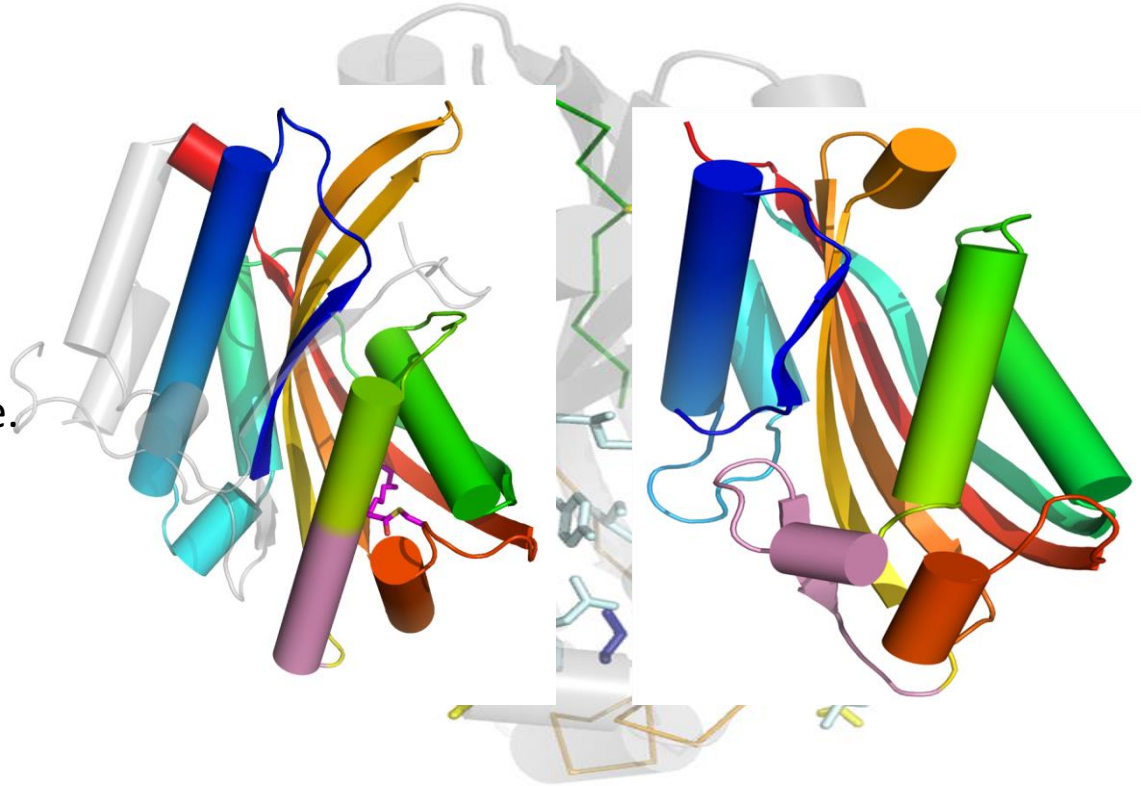
# Structure-based function interpretation

Where are the functional/catalytic sites?

# Multiple methods in bioinformatics:

## Structure comparisons of Evf

- Reported as novel fold...
- ... but in fact related to *Bacillus* toxin structures (DALI)
- Both bind to host insect membranes
- Palmitate seen in Evf structure. Matches conserved region of toxins...
- **GESAMT** is an excellent CCP4 option
- FoldSeek is great for a quick search of the AFDB, ESMAtlas etc



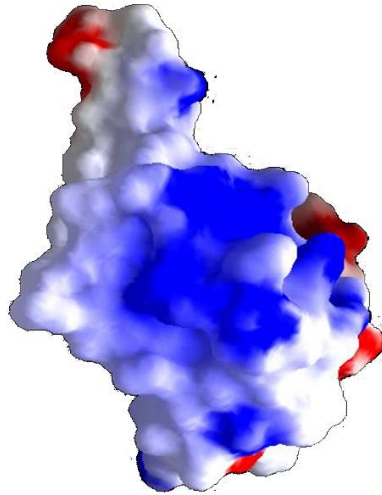
# Structure-based function annotation

- Finding functional sites is based on their being different somehow to the rest of the protein surface. Important general methods are based on
  - Shape (castP, ProFunc, PyMOL)
  - Electrostatics (PyMOL, APBS)
  - Evolutionary conservation (Consurf)
- Less well-known but valuable characteristics are
  - Statistics of surface atom 'triangles' (STP)
  - Probe interaction energetics (ISMBLab)
  - Predicted pKa values (THEMATICS/POOL)
- New generation Deep Learning-based methods

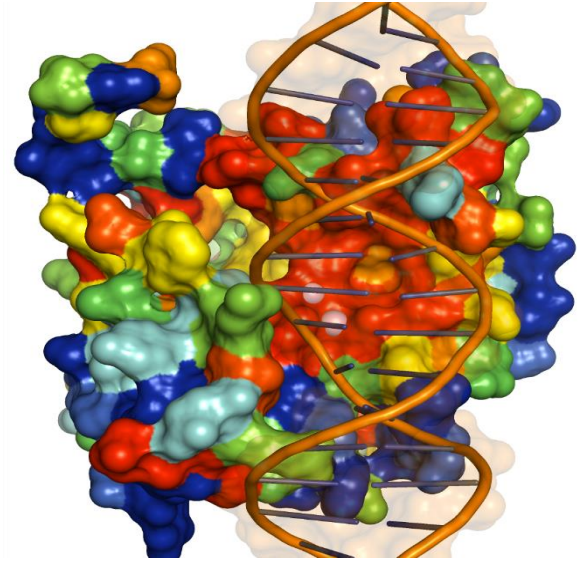
# Important general methods



Shape  
*CastP*



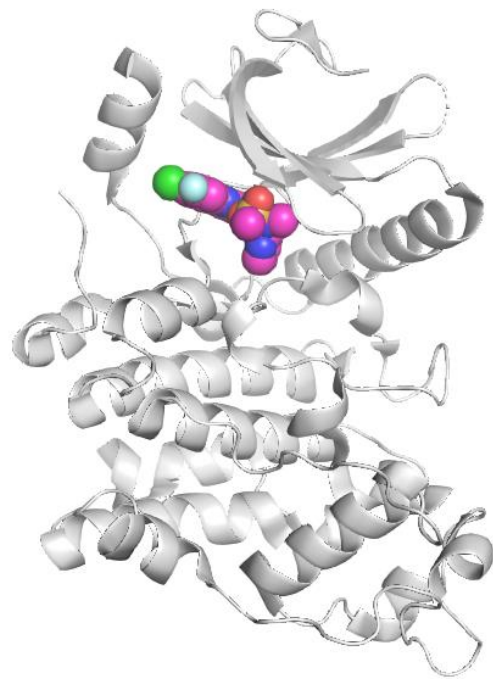
Electrostatics  
*APBS/PyMOL*



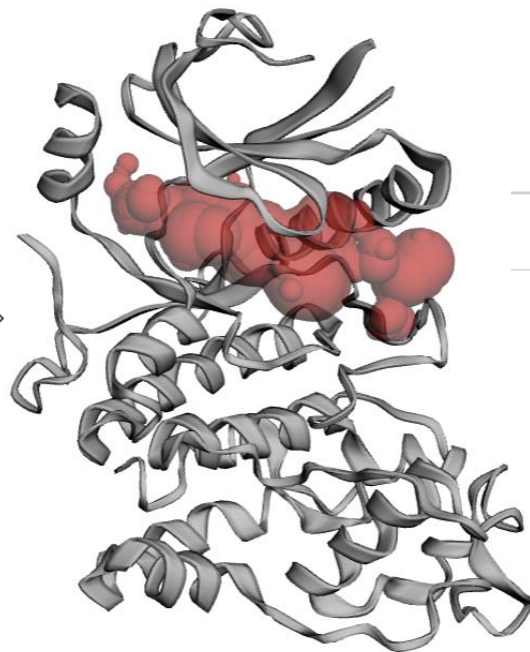
Conservation  
*ConSurf*




# Some servers require thought...

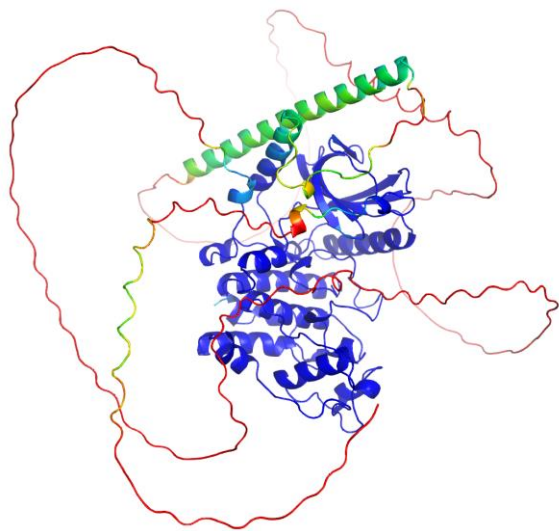


CastP  
→

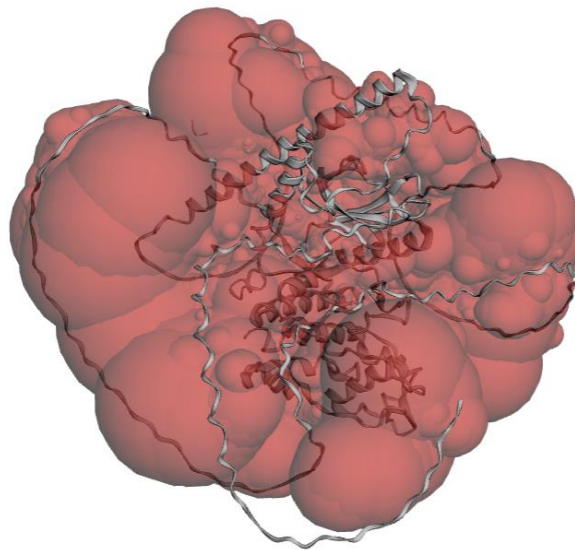


PocID 	Area (SA) Å <sup>2</sup>	Volume (SA) Å <sup>3</sup>
1	762.617	506.394

# Some servers require thought...

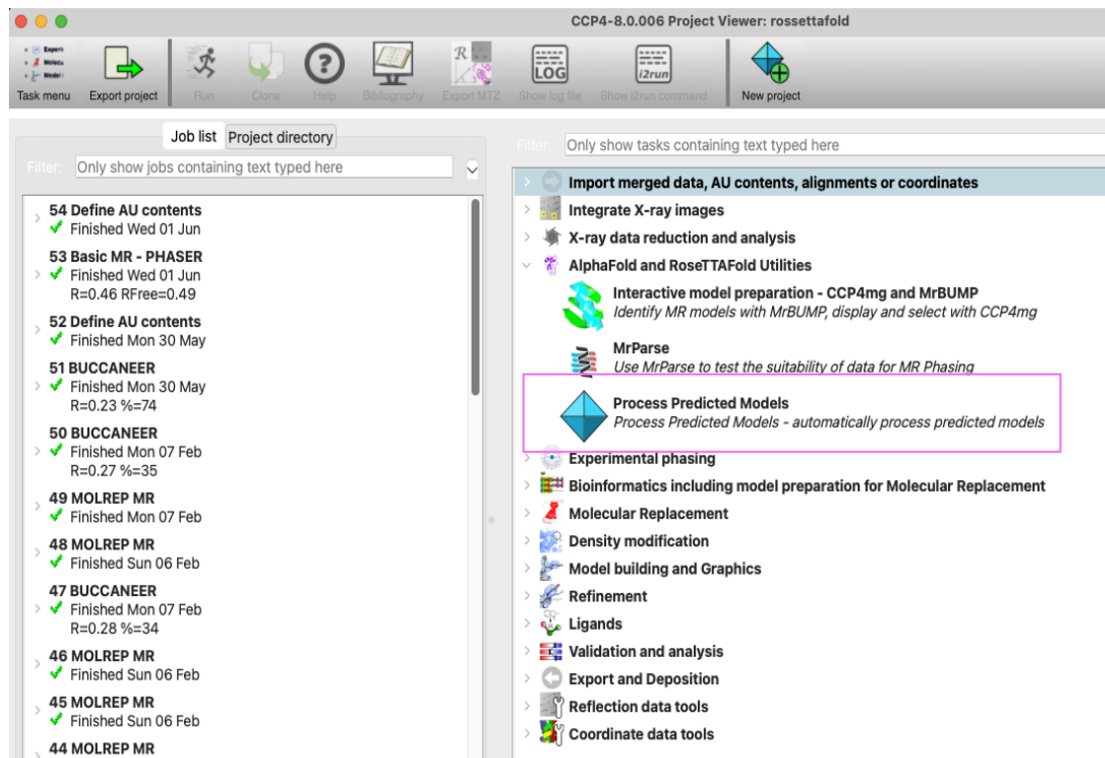


CastP  
→



PoclD 	Area (SA) Å <sup>2</sup>	Volume (SA) Å <sup>3</sup>
1	21186.058	159841.419

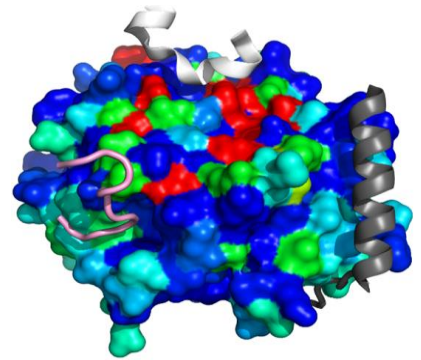
# CCP4 i2 can remove the red spaghetti (and convert pLDDT to B-factor)



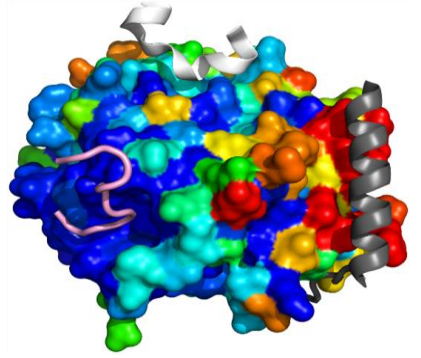
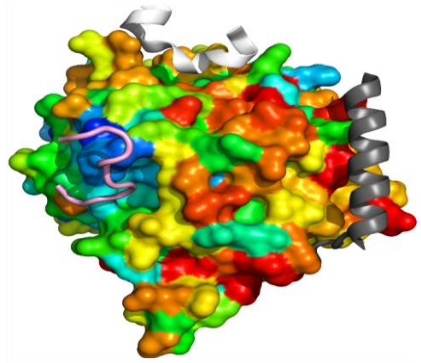
# Some servers require thought...

- Consurf maps sequence conservation onto a structure revealing functional sites
- Excellent, general method, but results depend on sequence set chosen for mapping: selecting all or only near relatives gives different results. Either might be more appropriate for you

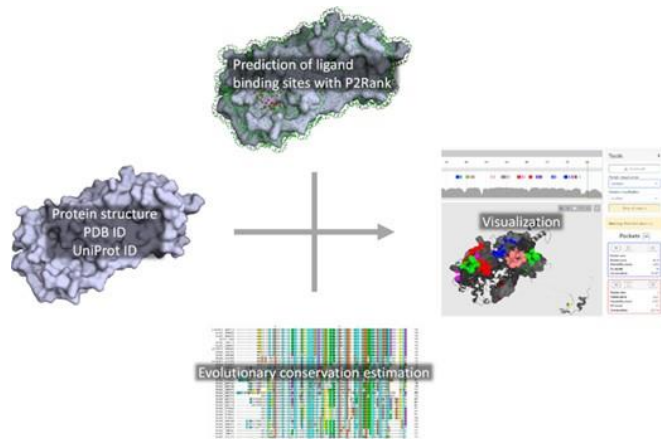
Mapping 300 homologues mixes different activities so no information on binding sites



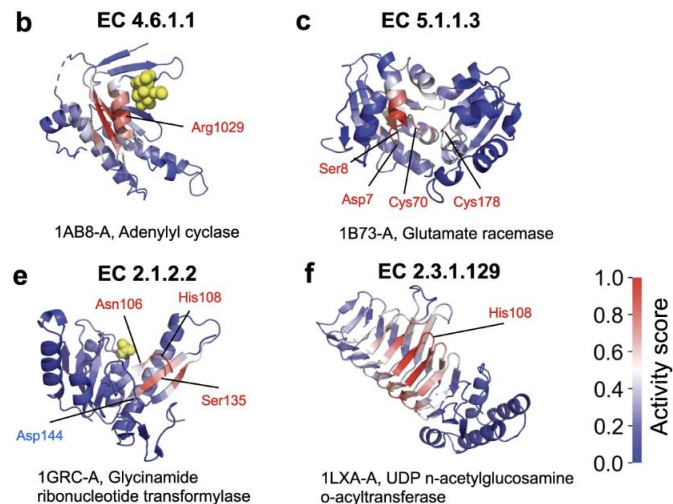
But restricting to a single protein family shows only 'pink' site is function in both Diptera and Lepidoptera



# Advanced methods use Machine Learning and/or multiple signals



PrankWeb uses structural and physicochemical properties then displays pockets with conservation analysis <https://prankweb.cz>

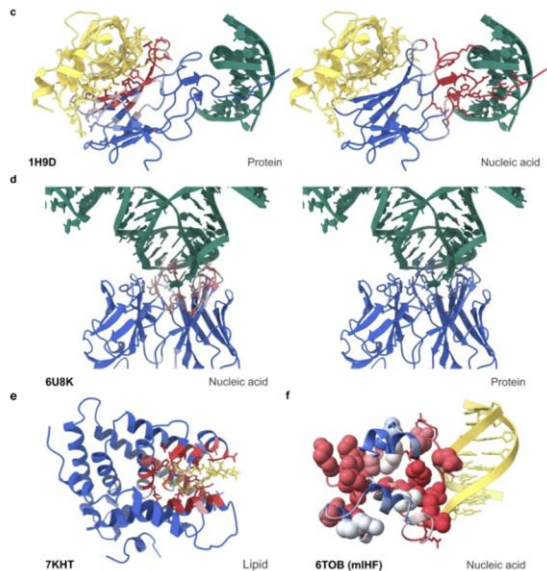


Combines LM and contact map features in CNN to predict GO terms and sites

<https://beta.deepfri.flatironinstitute.org/>

# My favourite Deep Learning-based methods

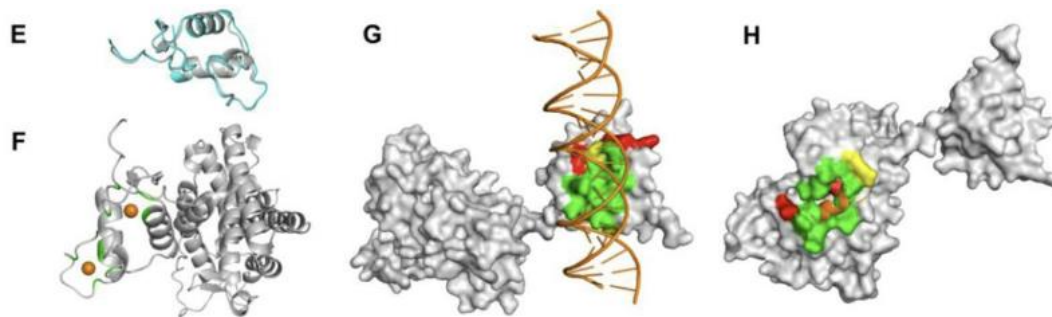
## PESTO



Geometry and contacts of input structure analysed. Interface propensities displayed

Krapp et al (2023) Nature Comms 14, 2175  
<https://pesto.epfl.ch/>

## GPsite



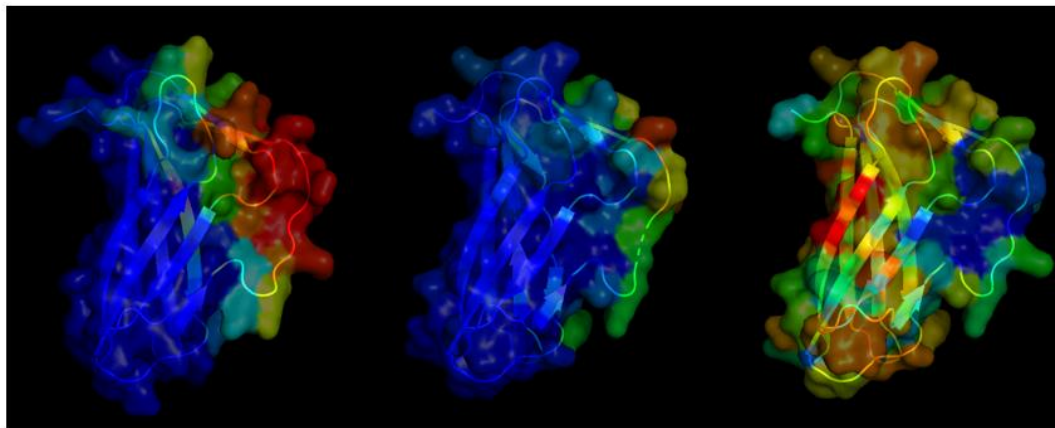
True Positive False Positive False Negative

Sequence modelled with ESMFold. Various binding sites and GO terms predicted

Yuan et al (2024) eLife <https://doi.org/10.7554/eLife.93695.1>  
<https://bio-web1.nscg-gz.cn/app/GPSite>



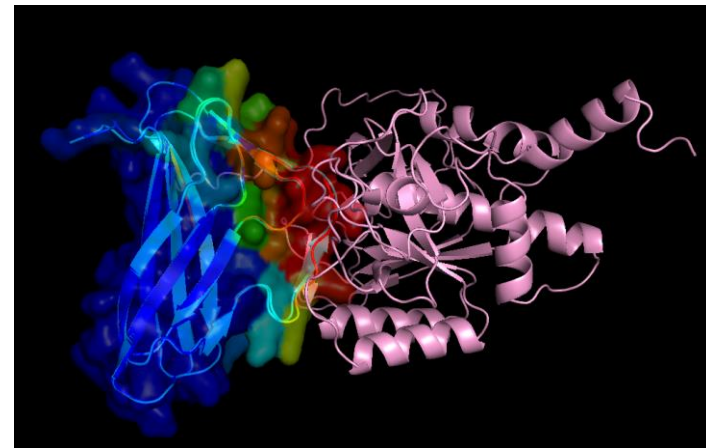
# Multiple methods in bioinformatics: Structure-based function methods



GPsite

PESTO

Consurf



All methods pick out the very probable binding site of this protease inhibitor. DL methods give a somewhat cleaner signal than general conservation

# Case study from Structural Genomics



# GxGYxYP proteins

- Named for a conserved sequence motif. Molecular function unknown
- Over-represented in gut bacteria
- Found in **P**olysaccharide **U**talization **L**oci in *Bacteroides thetaiotaomicron*
- Q: What does the protein do?*



# GxGYxYP proteins

- Domain architectures also predict carbohydrate connection

Q8A5P5 *Bacteroides thetaiotaomicron* (3SGG)



A6LZL0 *Clostridium beijerinckii*



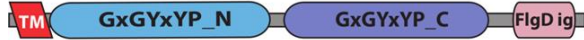
G9S6Q7 *Tannerella* sp.



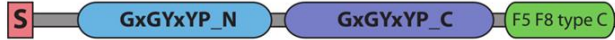
C7PHK0 *Chitinophaga pinensis*



H1XSR2 *Caldithrix abyssi*



B3JFZ1 *Bacteroides coprocola*

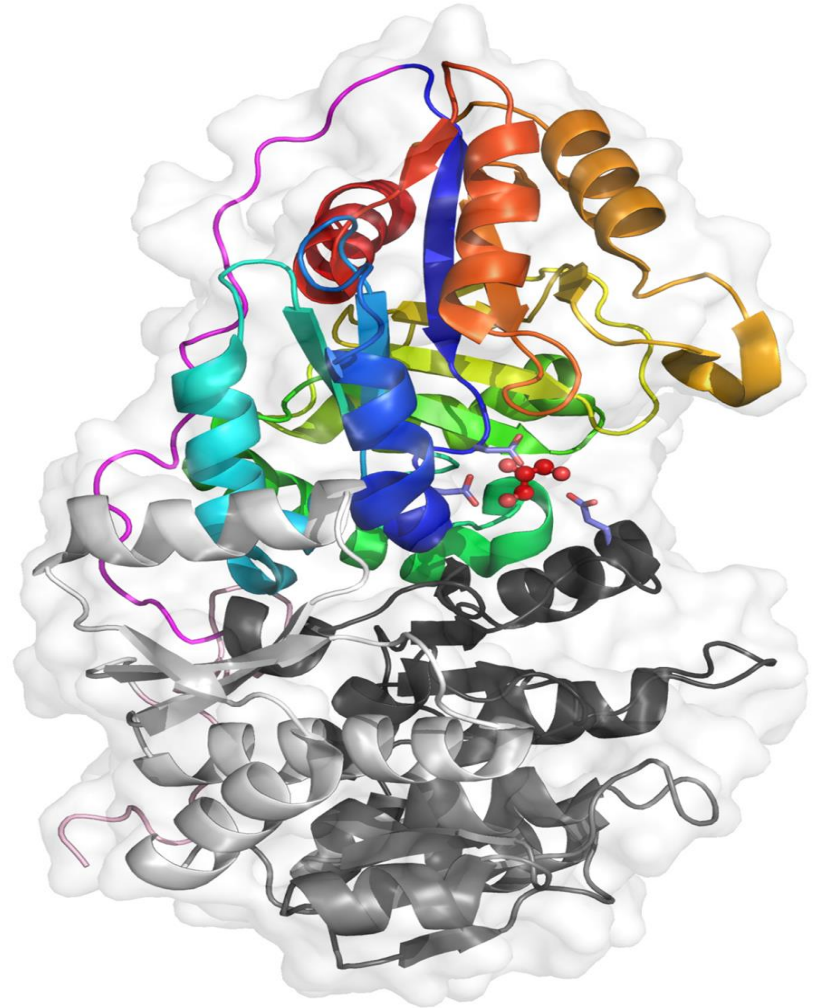


B9XJ10 *Pedospaera parvula*



# GxGYxYP proteins

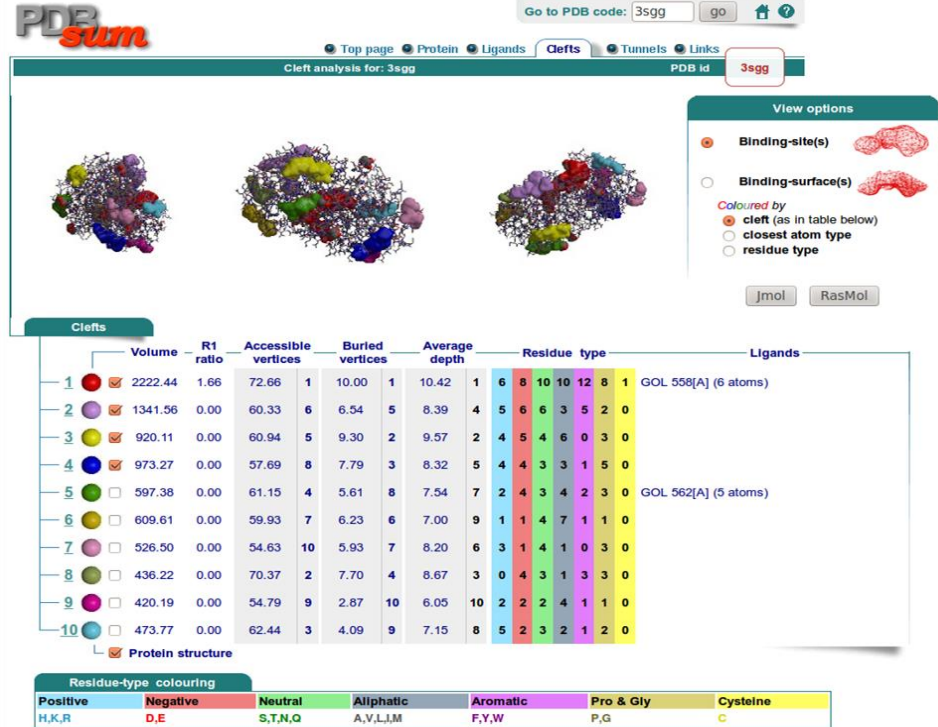
- Overall folds don't help much
- TIM barrel + 3 x novel  $\alpha+\beta$  unit
- TIM barrel DALI Z >13 for
  - Allantoinase
  - Polysaccharide deacetylase
  - Glycosyltransferase



# GxGYxYP proteins

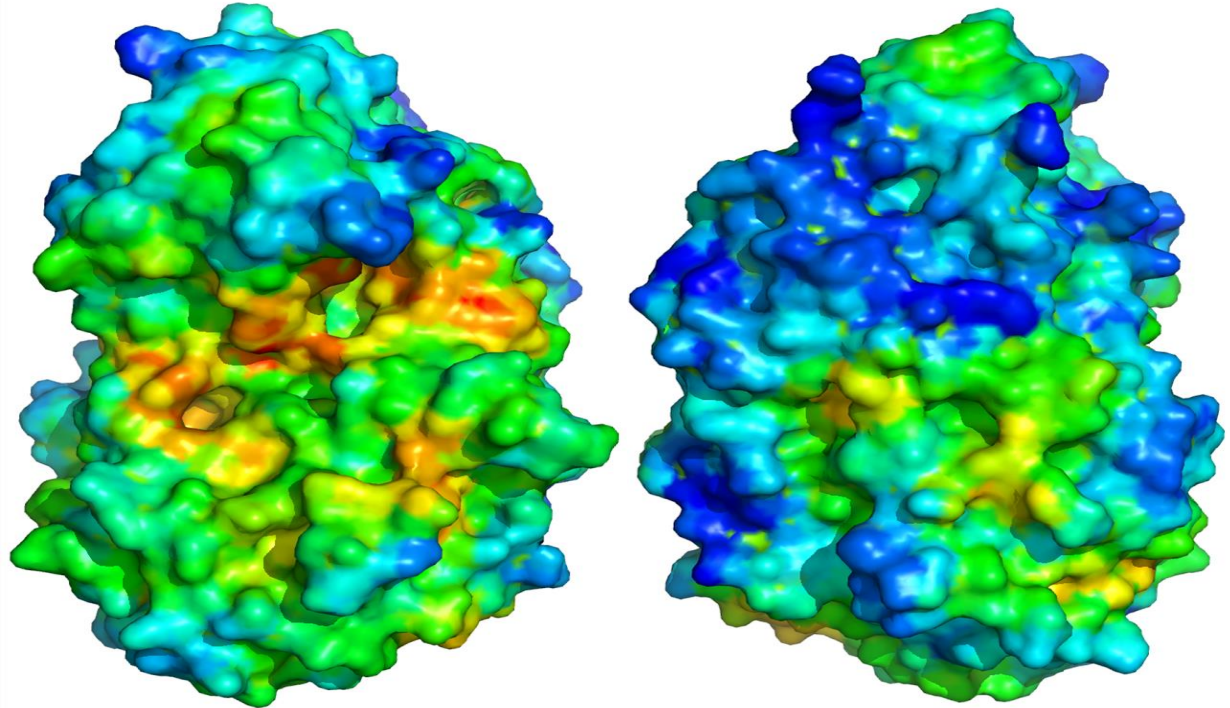
- Largest cavity lies between domains
- Glycerol from crystallisation solution in it

## PDBsum entry 3sgg



# GxGYxYP proteins

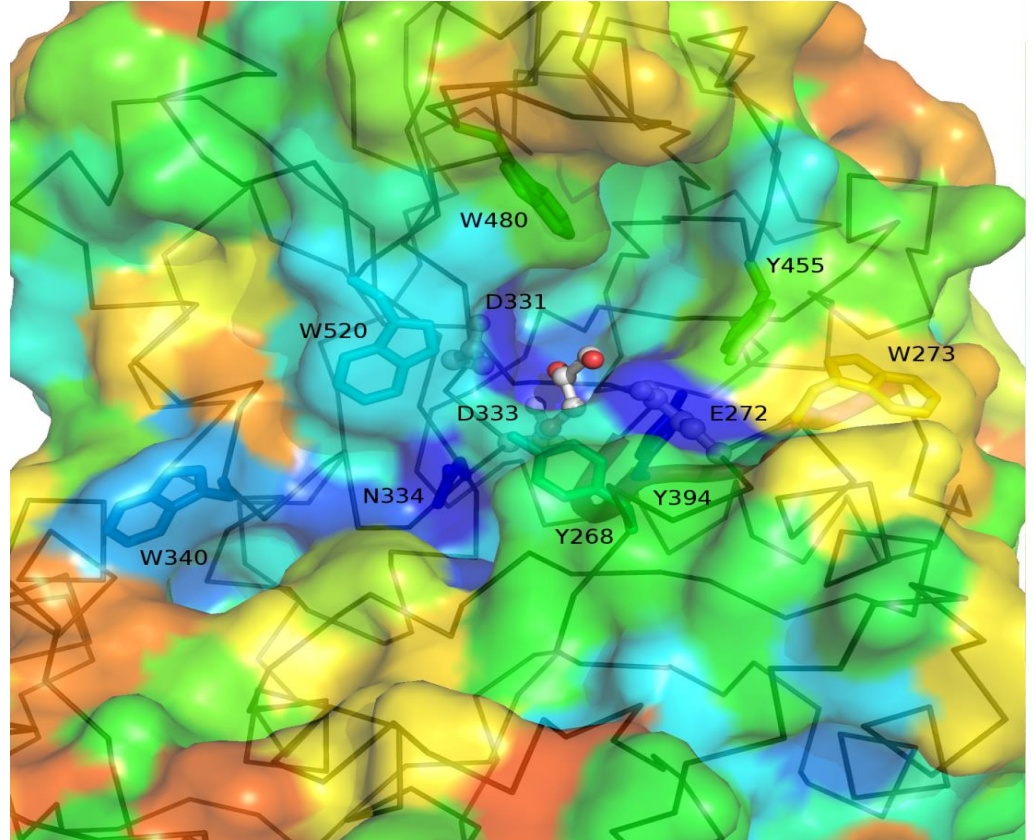
- Largest cavity lies between domains
- Glycerol from crystallisation solution in it
- Picked out by non-geometry based STP (surface triplet propensities)





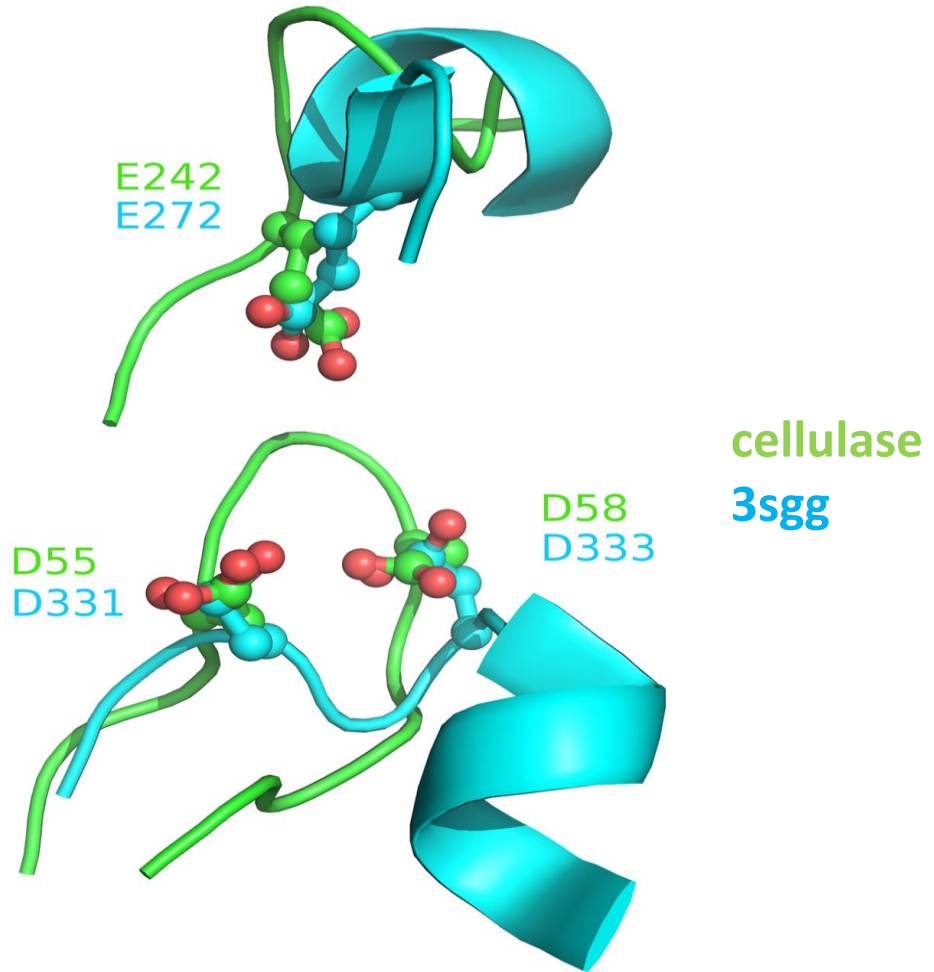
# GxGYxYP proteins

- The patch is conserved
- Contains multiple aromatic residues, often surface lying in carbohydrate binding sites...



# GxGYxYP proteins

- The patch is conserved
- Contains multiple aromatic residues, often surface lying in carbohydrate binding sites...
- ... and acidic residues resembling known glycosidase site... (SPRITE)
- ... and with perturbed pKa values (THEMATICS)



# Multiple methods in bioinformatics

## GxGYxYP conclusion

- GxGYxYP is a novel Glycoside Hydrolase family
  - Genome context
  - Domain composition
  - Cavity
  - Bound glycerol
  - STP
  - Conservation
  - Match to known GH catalytic site
  - pKa perturbation

**Structure-based  
methods**



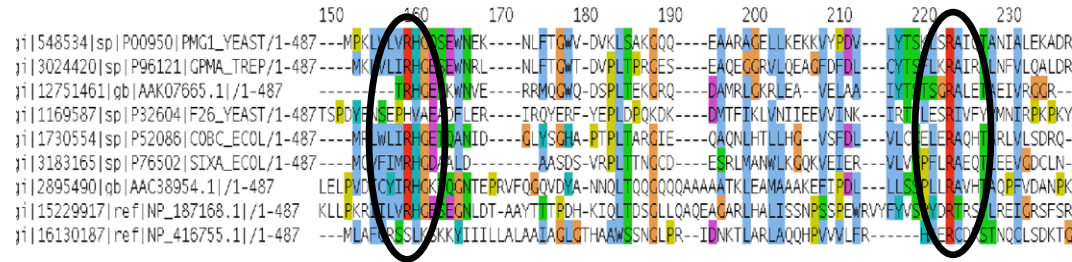
...and finally, you're putting a  
manuscript together

Calculating and presenting sequence alignments

# Your sequence alignment

- Don't use ClustalW! It's 24 years old! Modern methods like MUSCLE, Probcons and MAFFT are much better

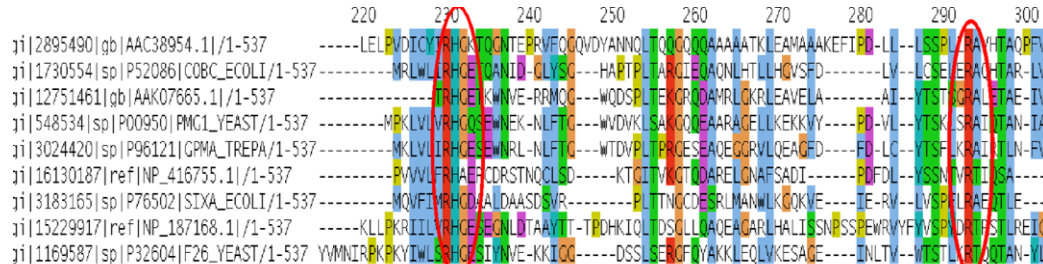
ClustalW misses relatively obvious RHG motif in some of diverse sequence set...



Sequence alignment snippet showing positions 150 to 230. The RHG motif (residues 160-170) is highlighted with a red oval. The alignment shows a gap in the RHG motif for the sequence j1|16130187|ref|NP\_416755.1|/1-487.

```
j1|548534|sp|P00950|PMG1_YEAST/1-487---MPKLVVRHCESEWNEK---NLFTGTV-DVKLSAKGQQ---FAARAGELKKEKKVYFDV---LYTSLSRAITANIALEKADR
j1|3024420|sp|P96121|GPMA_TREP/1-487---MKLVLRHCESEWNEK---NLFTGTV-DVPLTPGES---EAGEGCRVLEAGDFD---CYTSFLKRAIRLNFVLQALDR
j1|12751461|gb|AAK07665.1|/1-487-----TRHCEKWNVE---RRMOGQ-DSPLTEKQRQ---DAMRLCKRLEA--VELAA---LYTSLSRALEAIEIVRGR--
j1|1169587|sp|P32604|F26_YEAST/1-487TSPDYERSEPHVAE-DFLER---IRQYERF-YEPLDRQKDK---DMTFIKLVNITIEEWINK---TRTLESRTVYFMNIRKKBY
j1|1730554|sp|P52086|COBC_ECOL/1-487---MKLVLRHCESEWNEK---GLYSCH-PTPLTARQIE---CAQNLHTLHG--VSFDL---LVLPLEAEQHTLRVLSDRQ-
j1|3183165|sp|P76502|STXA_ECOL/1-487---MGVFIMRHCDALD---AASDS-VRPLTNGCD---ESRLMANWLKQKVEIER---LVLPPLAEQHTLRVLSDRQ-
j1|2895490|gb|AAC38954.1|/1-487LELFDVICYTHGKQDTEBRVFGQGVYANNQLTQQGQQAAAAATKLEAMAAKEFTPD---LLSSPLRAVHHAQEFV
j1|15229917|ref|NP_187168.1|/1-487KLLPKRILVRHCESEWNEKLDYAYTTTPDHKIQLTDSQLLQAEACARLHALISSNPSSPEWRVYFVSPDRITSTLREIC
j1|16130187|ref|NP_416755.1|/1-487---MLAEPSLSLKKKIIILLALAAIAGLTHAAWSSNQLR---IDNKTLARLAQHPVWYLER-----LEKCTQVLTNQSLSDKTC
```

... but MUSCLE gets it



Sequence alignment snippet showing positions 220 to 300. The RHG motif (residues 230-240) is highlighted with a red oval. The alignment shows the RHG motif correctly aligned for the sequence j1|16130187|ref|NP\_416755.1|/1-537.

```
j1|2895490|gb|AAC38954.1|/1-537-----LELFDVICYTHGKQDTEBRVFGQGVYANNQLTQQGQQAAAAATKLEAMAAKEFTPD---LLSSPLRAVHHAQEFV
j1|1730554|sp|P52086|COBC_ECOLI/1-537-----MRLVLRHCESEWNEK---GLYSCH-PTPLTARQIECAQNLHTLHGVSFD-----LVLPLEAEQHTLRVLSDRQ-
j1|12751461|gb|AAK07665.1|/1-537-----TRHCEKWNVE-RRMOG---WQDSPLTEKQPDAMRLCKRLEAVELA---AI---LYTSLSRALEAIEIVRGR--
j1|548534|sp|P00950|PMG1_YEAST/1-537-----MPKLVVRHCESEWNEK-NLFTG---WVDKLSAKGQEAARAGELKKEKKVY---PD-VL---LYTSLSRAITANIA
j1|3024420|sp|P96121|GPMA_TREPA/1-537-----MKLVLRHCESEWNEK-NLFTG---WTDVPLTPGESAEQECRVLEAGDFD---FD-LG---LYTSFLKRAIRLNFV
j1|16130187|ref|NP_416755.1|/1-537-----RVVLFVRHAEFCRSTNQGLSD---KTCTVYKTDARELQNAFSADI---PDVFL---LYSNVITITISA
j1|3183165|sp|P76502|STXA_ECOLI/1-537-----MGVFIMRHCDALDAASDSVR---PLTNGCDSRLMANWLKQKVE---TE-RV---LVSPPLAEQHTLRVLSDRQ-
j1|15229917|ref|NP_187168.1|/1-537-----KLLPKRILVRHCESEWNEKLDYAYTTTPDHKIQLTDSQLLQAEACARLHALISSNPSSPEWRVYFVSPDRITSTLREIC
j1|1169587|sp|P32604|F26_YEAST/1-537YMNIRKKBYIWLSEHCESEWNEK-KKICG---DSSLSEKCFYAKKLEQLVESAGE---INLTV---LYTSLSRAITANIA
```

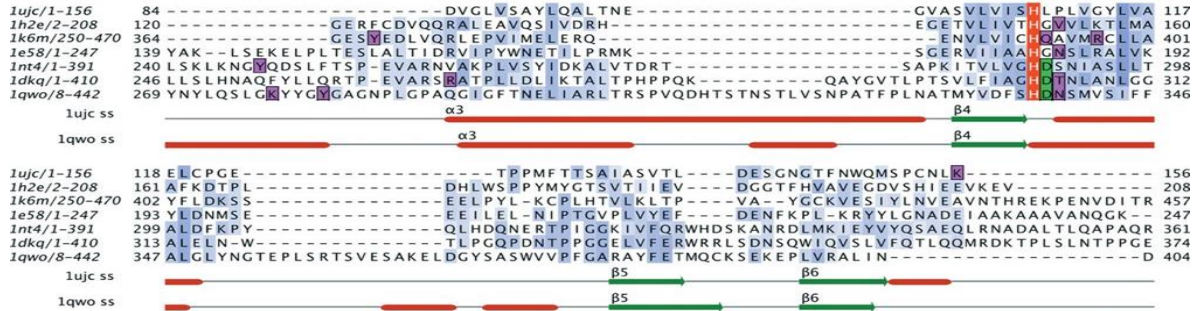
# Multiple methods in bioinformatics : Jalview.org

- All these alignment methods and more are available through Jalview on Dundee servers

The screenshot displays the Jalview 2.9.0b2 interface. The main window shows a multiple sequence alignment of FER1 proteins, including FER\_CAPAA1-97, FER\_CAPAN1-144, FER1\_SOLLO1-144, Q03XIG\_SOLTU1-144, FER1\_FEA1-149, Q7XARL\_TRIPR1-152, FER1\_MESCR1-148, FER1\_SPIOL1-147, FER3\_RAPSA1-96, FER2\_ARATH1-96, FER2\_ARATH1-148, Q03XIG\_ARATH1-118, FER1\_MAIZE1-150, and Q04120\_MAIZE1-140. The alignment is color-coded by residue type. Below the alignment, there are tracks for Secondary Structure, Iron Sulphur Contacts, Conservation, and Quality. A context menu is open over the alignment, listing various analysis tools: Tcoffee with Defaults, Protein Disorder, Analysis, Conservation, Fetch DB References, Muscle with Defaults, MAFFT with Defaults, MSAProbs with Defaults, GLprobs with Defaults, Clustal, and Realign with ClustalO. Other panels show a phylogenetic tree (Average distance) and a 3D protein structure (Jmol view for FER1 S...).

# Jalview

- Also helps you produce figures like this...



- ... rather than like this



Don't forget to cite it (and all your bioinformatics)!

# Questions?

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