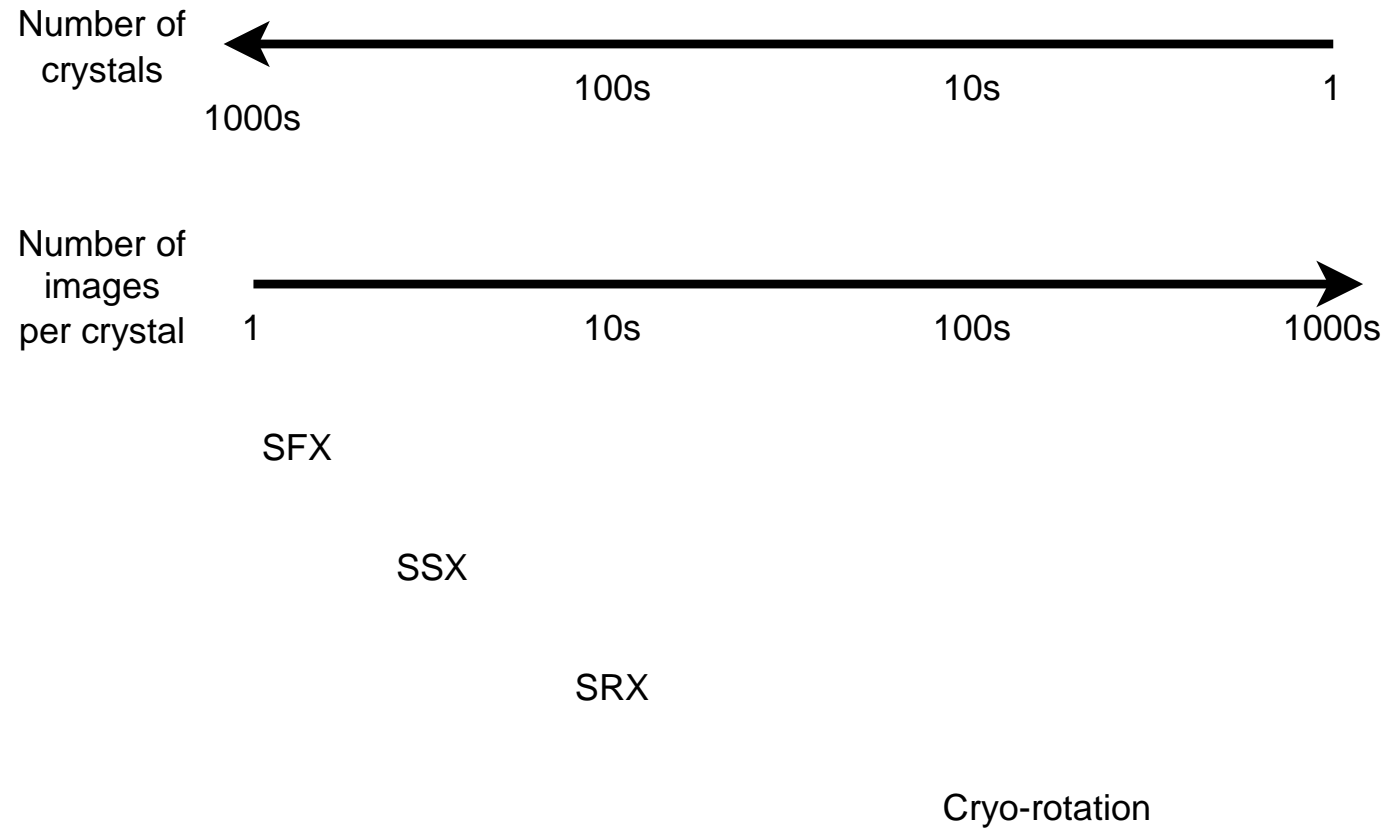


Multi-Crystal Approaches

Xia2.Multimorph and Xia2.SSX

Amy Thompson (PDRA on VMXi)

Multi-Crystal Approaches

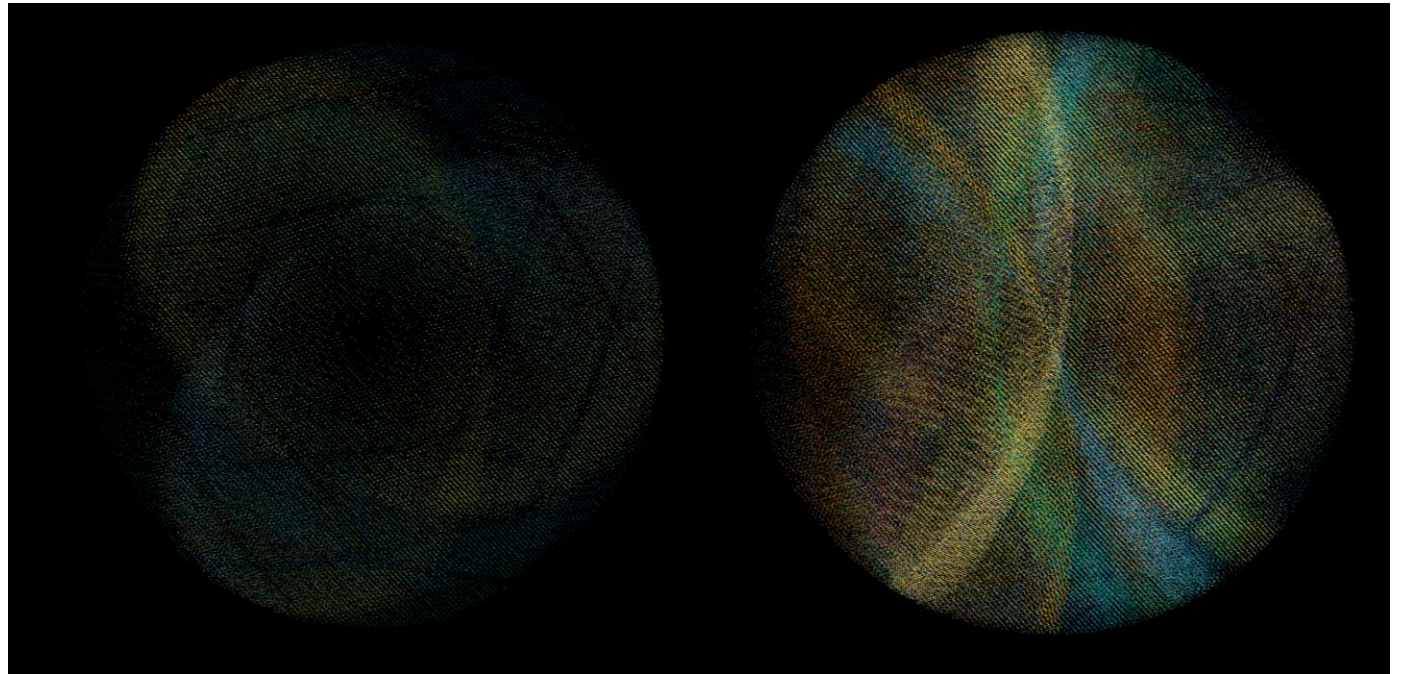


Why Use Multi-Crystal Strategies?

In some cases, it is only possible to collect incomplete data from a single crystal due to radiation damage: obtain a complete dataset by combining data from multiple crystals

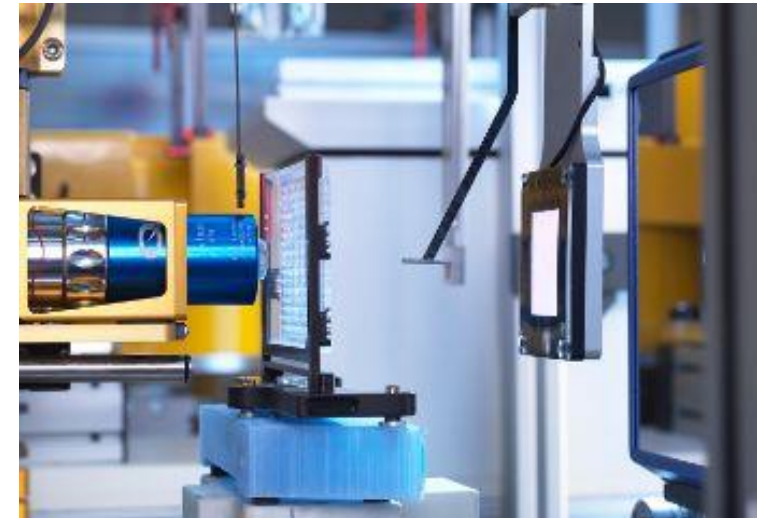
This strategy allows for the use of:

- small crystals
- room-temperature data collection



Room Temperature Advantages

- Cryo-cooling may hide biologically significant structural features
- Virus crystals often suffer on cryo-cooling
- *In situ* data collection



Fraser, J., Clarkson, M., Degnan, S. et al. (2009) Nature 462, 669–673

Sanchez-Weatherby, J., Sandy, J., Mikolajek, H. et al. (2019) J. Synchrotron Rad. 26, 291–301

Nave, C. & Garman, E. F. (2005) J. Synchrotron Rad. 12, 257–260

Merging Multiple Datasets Increases Signal-to-Noise

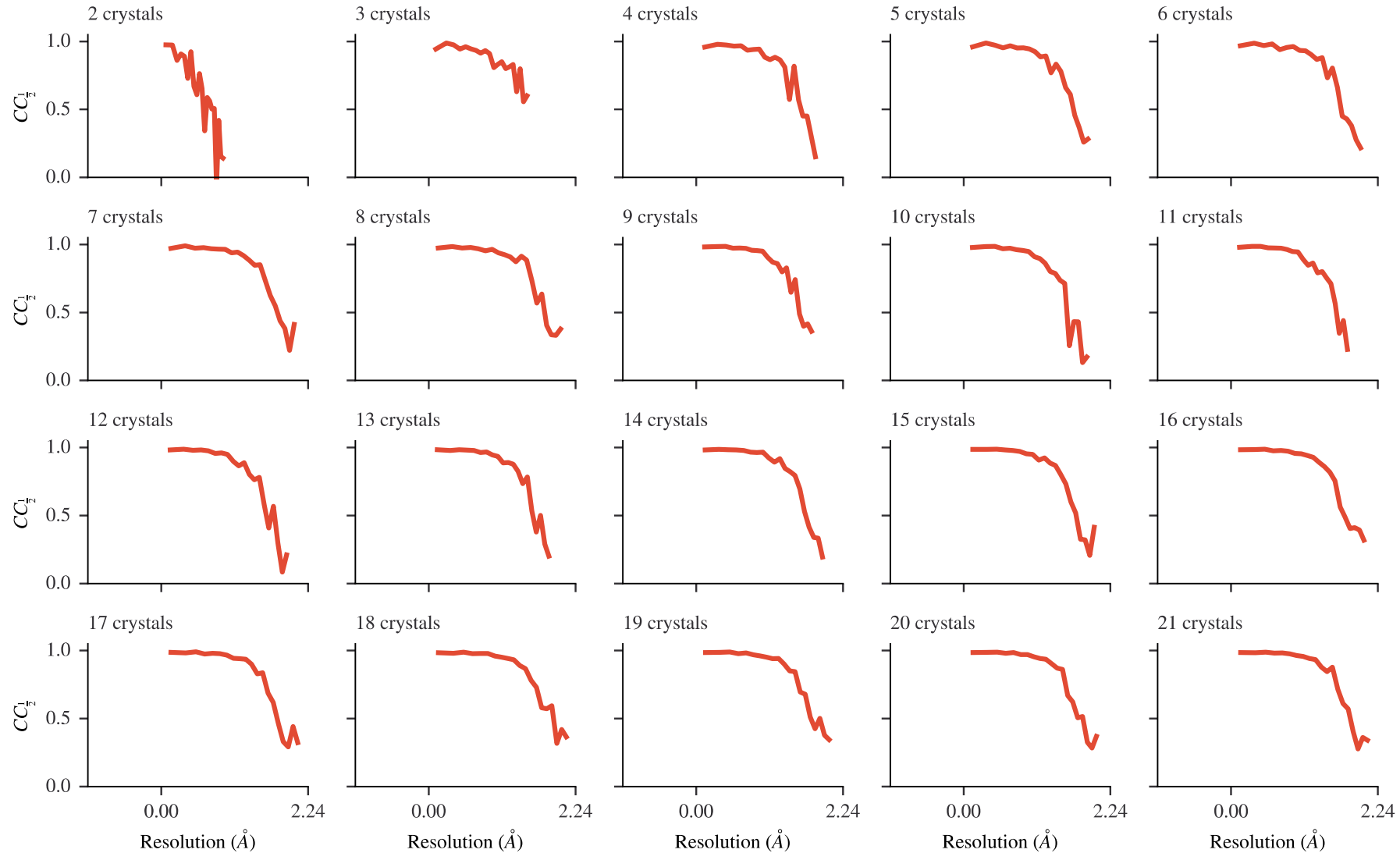
- Given only random errors, the standard error σ in a measurement is reduced by \sqrt{n} if the measurement is repeated n times
- Beware R_{merge} and R_{meas} !
- Use $\text{CC}_{1/2}$ and $\langle I/\sigma \rangle_{\text{mrgd}}$

Dataset	<i>Big</i>	<i>Tiny</i>	<i>T100</i>	<i>Big+T100</i>	<i>Big2</i>
Multiplicity	2	2	200	202	4
$\langle I/\sigma \rangle_{\text{ind}}$	2.0	0.2	0.2	0.22	2.0
R_{merge}	28%	280%	399%	395%	35%
R_{meas}	40%	400%	400%	396%	40%
R_{pim}	28%	280%	28%	28%	20%
$\langle I/\sigma \rangle_{\text{mrgd}}$	2.8	0.28	2.8	4.0	4.0
$\text{CC}_{1/2}$	0.66	0.04	0.66	0.80	0.80

Karplus, P. A., & Diederichs, K. (2015) Current opinion in structural biology, 34, 60-68

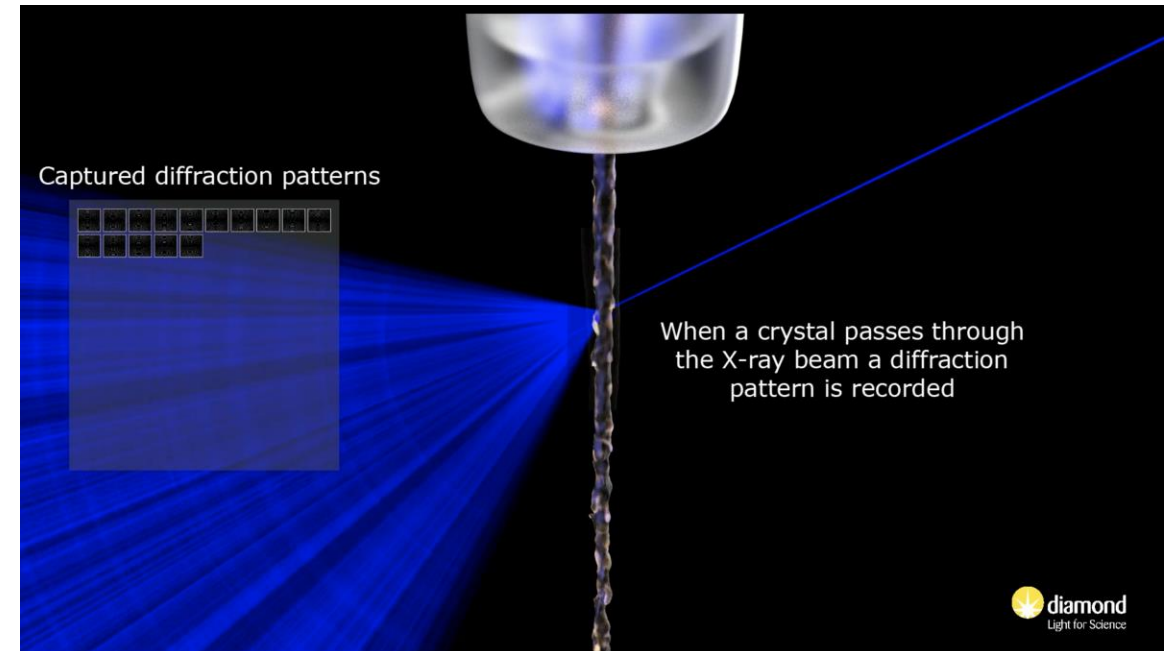
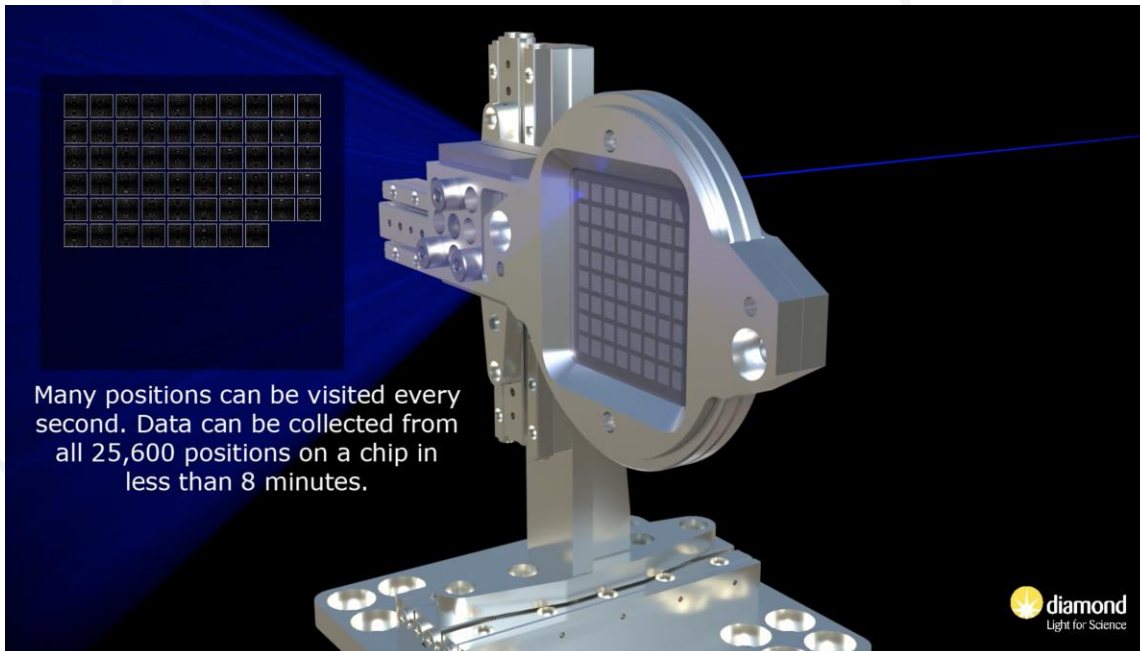
Better CC1/2 at higher resolution

```
xia2.compare_merging_stats unmerged_1.mtz  
unmerged_2.mtz [...] small_multiples=True
```



Time-Resolved Crystallography

- Study dynamics of light- or chemical-induced reactions using serial femtosecond (SFX) or synchrotron (SSX) crystallography

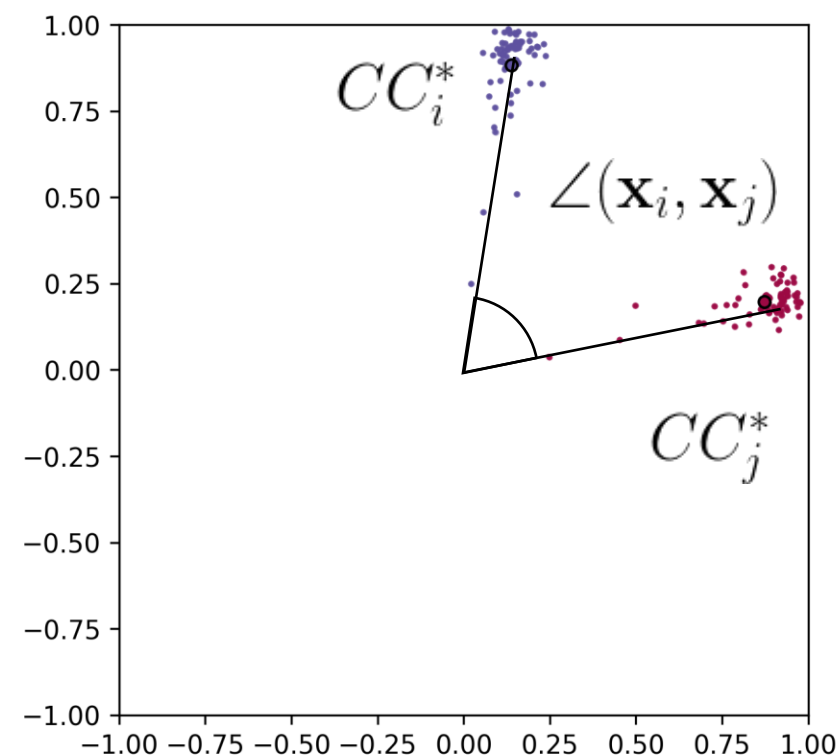


Challenges

- Symmetry determination
- Non-isomorphism
- Preferential orientation

Symmetry Determination

- Identification of consensus symmetry from narrow wedges or stills can be challenging
- Complicated by presence of potential indexing ambiguity
- New algorithms have been developed to help in symmetry determination from narrow wedges and stills (dials.cosym)

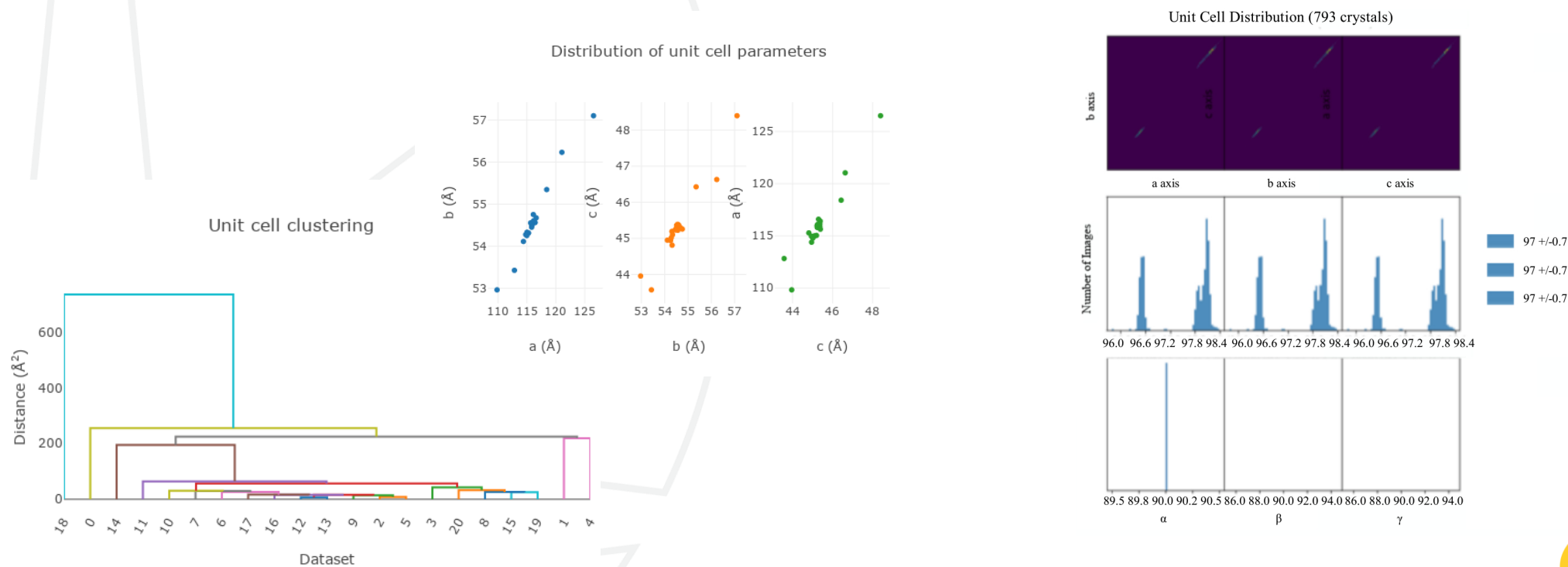


Brehm, W. & Diederichs, K. (2014). *Acta Cryst.* D70, 101–109

Gildea, R. J. & Winter, G. (2018). *Acta Cryst.* D74, 405–410

Non-Isomorphism

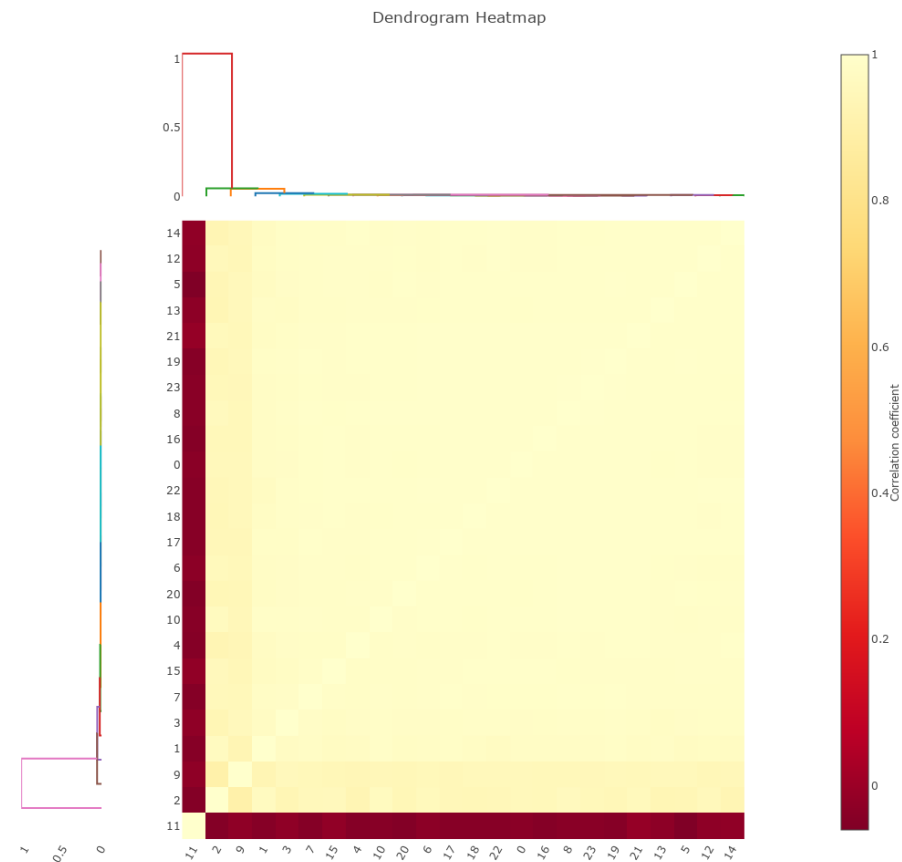
- Inclusion of non-isomorphous crystals may degrade the final data set
- Unit cell clustering may help identify outliers or different populations



Non-Isomorphism

Clustering on pairwise correlation coefficients may help identify outliers

$$r_{i,j} = \frac{\sum_h [I_i(h) - \bar{I}_i] [I_j(h) - \bar{I}_j]}{\left\{ \sum_h [I_i(h) - \bar{I}_i]^2 \sum_h [I_j(h) - \bar{I}_j]^2 \right\}^{1/2}}$$

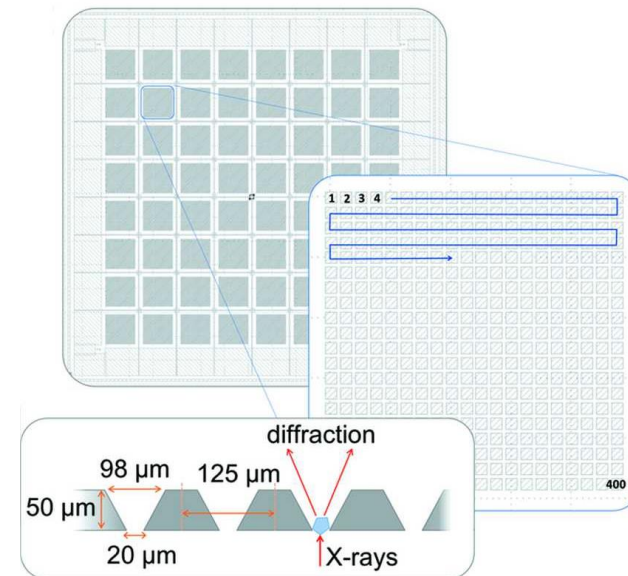
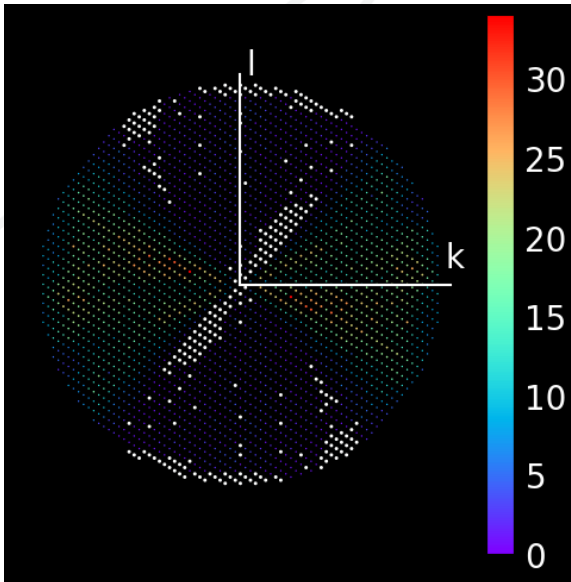


Giordano, R., Leal, R. M. F., Bourenkov, G. P. et al. (2012). Acta Cryst. D68, 649–658

Santoni, G., Zander, U., Mueller-Dieckmann, C. et al. (2017). J. Appl. Cryst. 50, 1844–1851

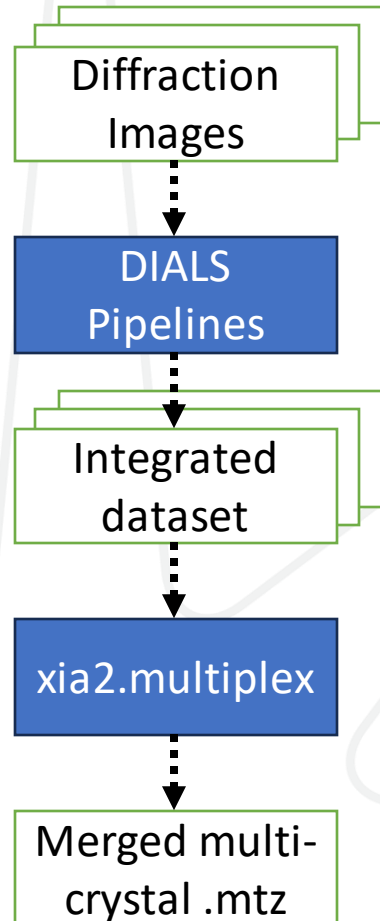
Preferential Orientation

- Crystal symmetry and morphology combined with data collection conditions may lead to preferential crystal orientation
- May result in under-sampled regions of reciprocal space
- Check stereographic projection and multiplicity plots

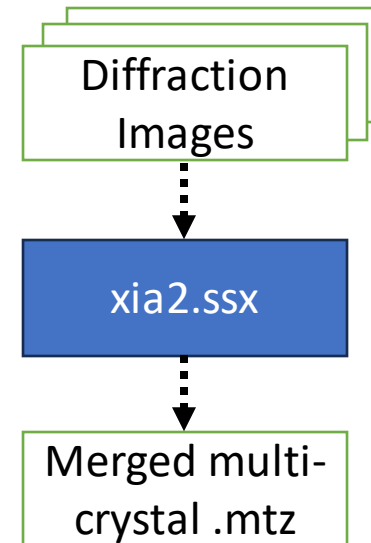


Processing Multi-Crystal Data at Diamond

xia2.multiplex – for rotation data



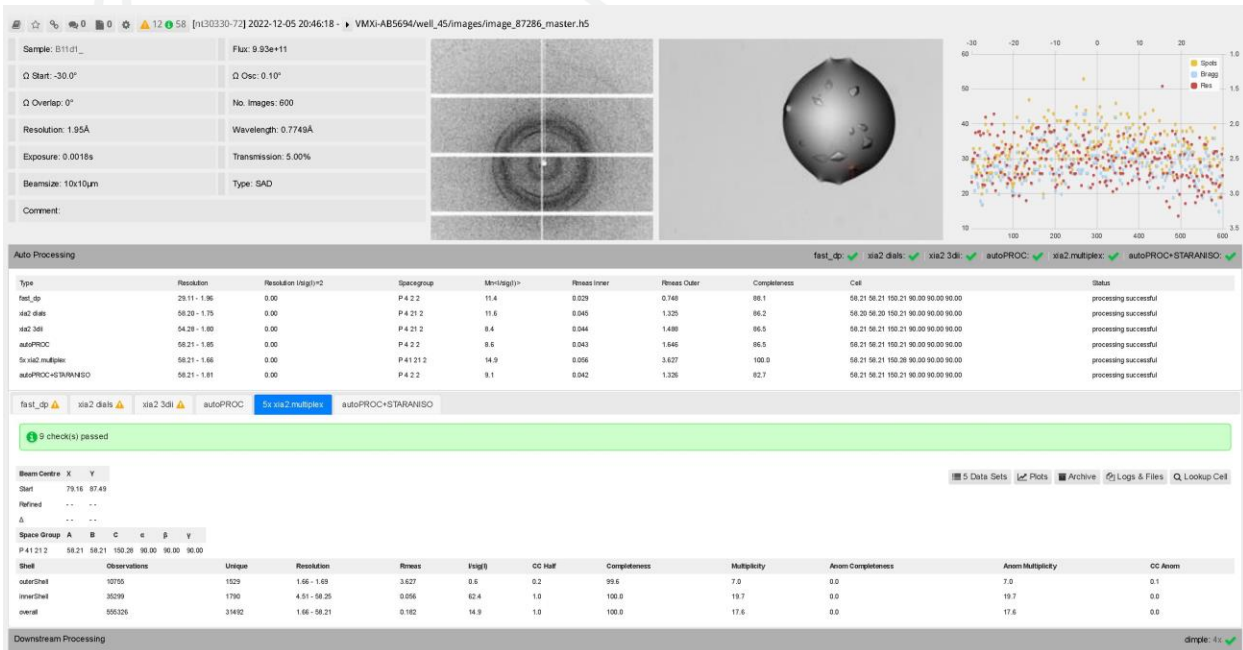
xia2.ssx – for still data



Runs automatically on I24 – otherwise need to run manually via command line

Run automatically when collected at Diamond

Availability through ISPyB at Diamond



Sample Group Management

Create Sample Group

Group Name	Container Barcode	Number of Samples	Action
cell_2	Thaum_1722_Thaum_1721	89	
cell_4	Thaum_1721	43	
1722_rowA	Thaum_1722	24	
1722_rowG	Thaum_1722	24	
1722_rowEandB	Thaum_1721_Thaum_1722	48	
1780_all	H58_1780_20	182	
ACHR_1	2012_ACHR	99	
2018_rowA	P400-VM09-AB0216	12	
2018_rowB	P400-VM09-AB0216	12	
2018_rowC	P400-VM09-AB0216	12	
2018_rowD	P400-VM09-AB0216	12	
h58		0	
100percentH58	kangaroo_AB02173_lys_kroka_AB02172_lys_kroka_AB02171_lys_walaby_AB02174_lys	128	
75percentH58PercentH58	kangaroo_AB02173_lys_kroka_AB02172_lys_kroka_AB02171_lys_walaby_AB02174_lys	128	
50percentH58PercentH58	kangaroo_AB02173_lys_kroka_AB02172_lys_kroka_AB02171_lys_walaby_AB02174_lys	128	

1722_rowF

Edit Sample Group

Container: VMXi-AB1722



Summary of last multiplex jobs from group 1722_rowF

Sample Group Data Collection

Type	Resolution	Spacegroup	Mn<1/sig(I)>	Rmeas Inner	Rmeas Outer	Completeness	Cell	Status
1 23x xia2 multiplex	1.59 - 75.19	P 4 1 2 1	16.4	0.077	7.880	100.0	58.25 58.25 100.32 90.00 90.00 90.00	processing successful
2 20x xia2 multiplex	1.57 - 75.19	P 4 1 2 1	21.9	0.075	10.507	100.0	58.25 58.25 100.32 90.00 90.00 90.00	processing successful
3 25x xia2 multiplex	1.60 - 75.19	P 4 1 2 1	20.0	0.077	8.951	100.0	58.25 58.25 100.32 90.00 90.00 90.00	processing successful

23x xia2 multiplex processing job details



Beam Centre X Y

Start 79.19 87.49

Refined -- --

Δ -- --

Space Group A B C α β γ

P 4 1 2 1 58.25 58.25 100.32 90.00 90.00 90.00

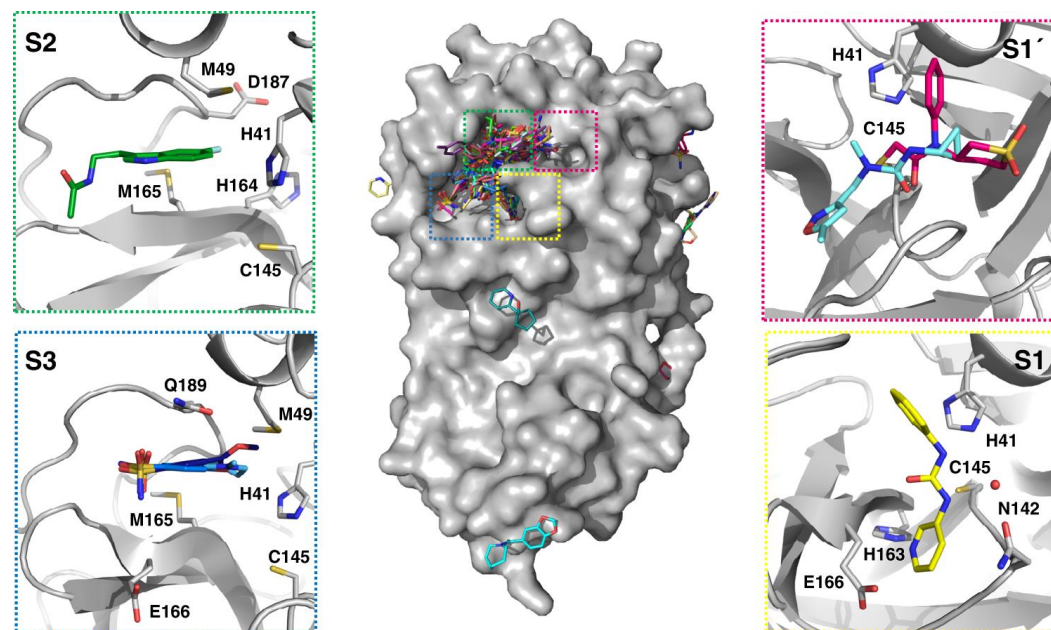
Shell	Observations	Unique	Resolution	Rmeas	1/sig(I)	CC Half	Completeness	Multiplicity	Anom Completeness	Anom Multiplicity	CC Anom
outerShell	31477	1765	1.58 - 1.62	7.880	0.4	0.2	100.0	17.8	0.0	17.8	-0.0
innerShell	157308	2029	4.32 - 75.24	0.077	92.2	1.0	100.0	77.6	0.0	77.6	-0.0
overall	2254804	35840	1.59 - 75.16	0.281	18.4	1.0	100.0	62.9	0.0	62.9	0.0



Example: SARS-CoV-2 M^{pro} ligand screening

Conventional Collection at CT

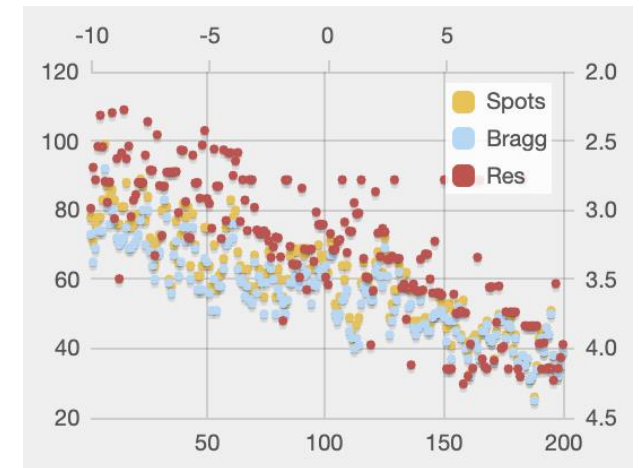
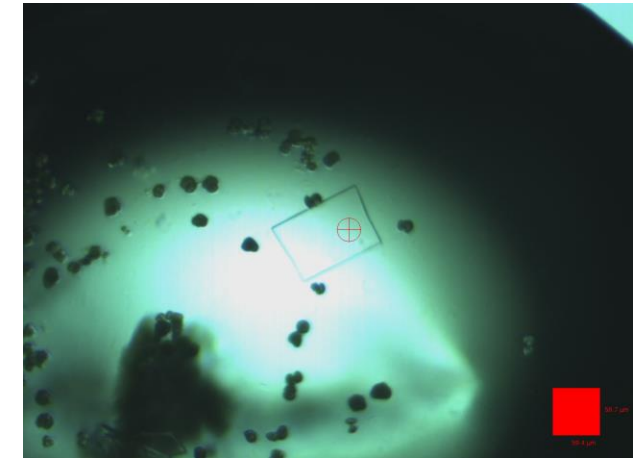
- SARS-CoV-2 main protease
- Central role in viral replication
- Key antiviral drug target
- Conventional fragment-screening campaign performed on I04-1
- Over 1250 unique fragments, identifying 74 high-value fragment hits



Example: SARS-CoV-2 M^{pro} ligand screening

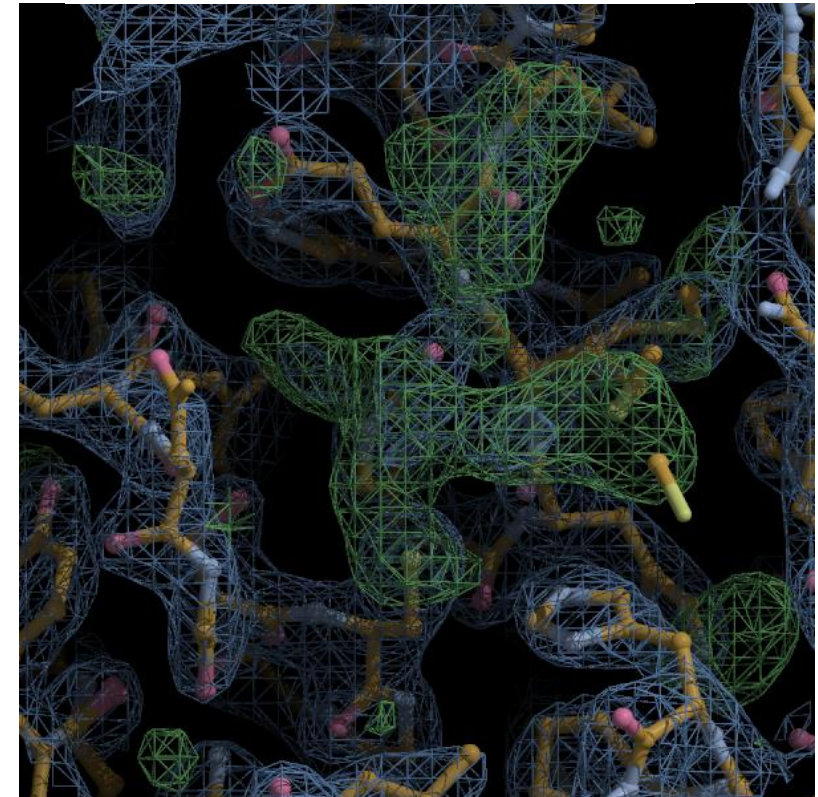
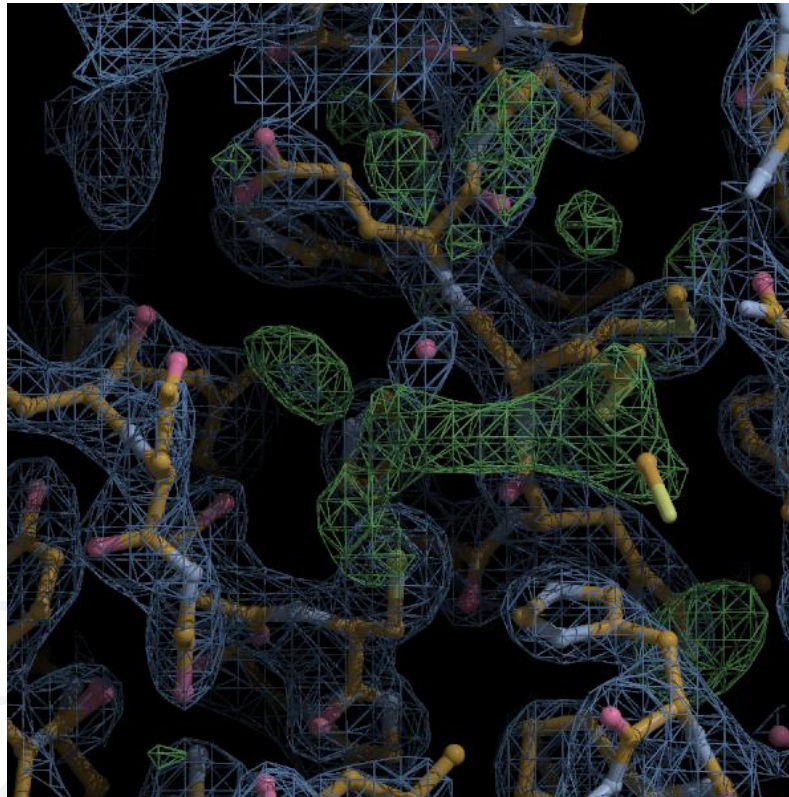
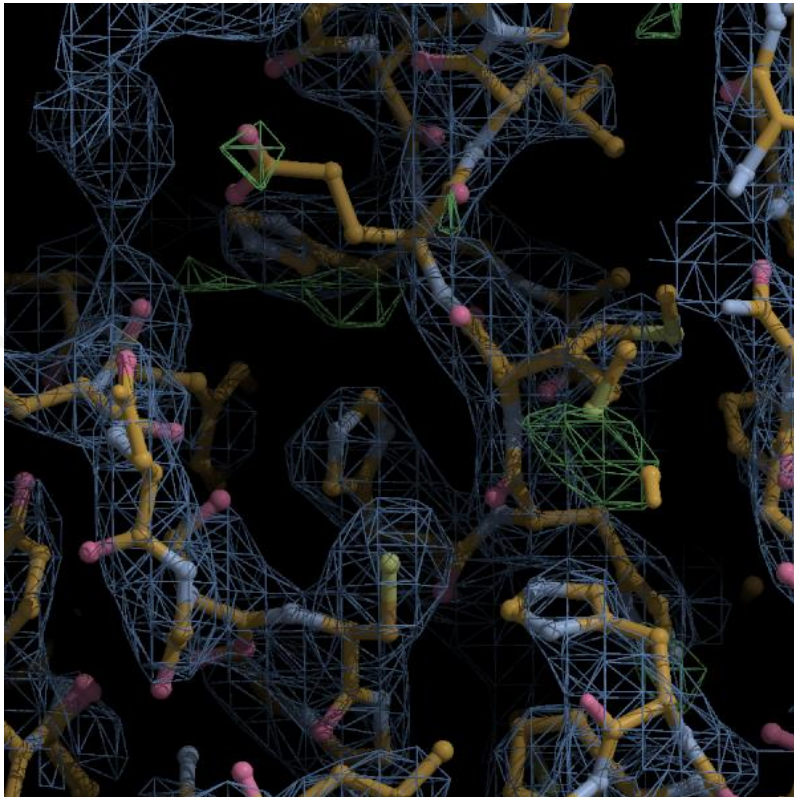
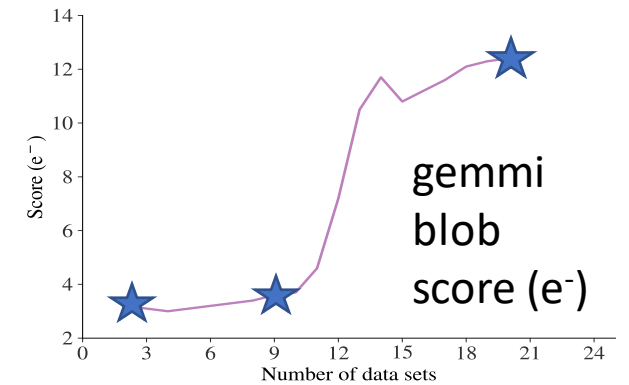
Multi-Crystal Collection at RT

- Initial fragment screening performed at cryo-temperatures (100 K)
- Are room temperature structures identical?
- RT in situ data collections on known ligand hits performed on I24 and VMXi
- Preferred orientation (plate-like crystals): vary starting angle
- xia2.multiplex provided near real time feedback during the experiment



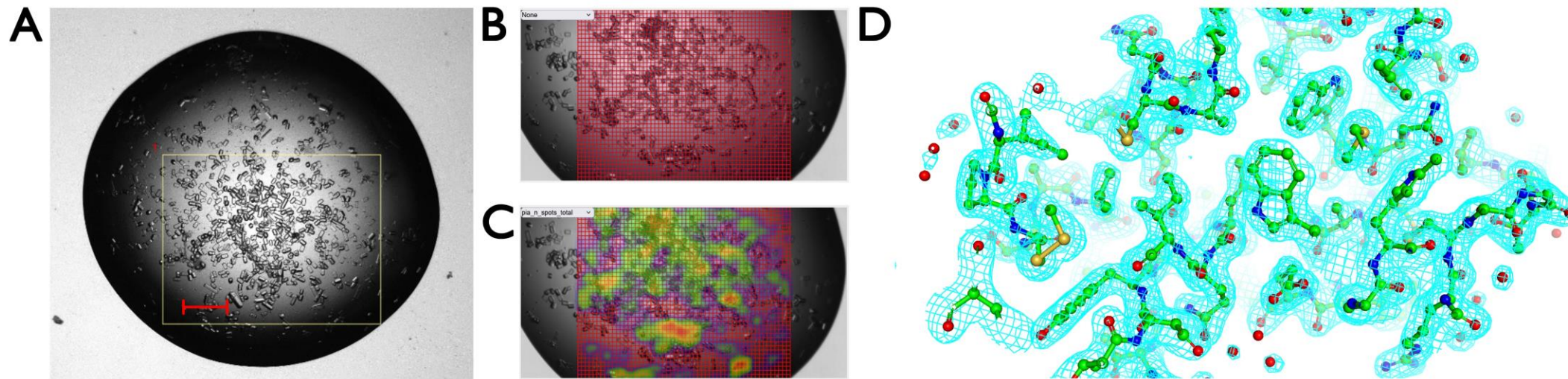
Example: SARS-CoV-2 M^{pro} ligand screening

Automatic dimple maps



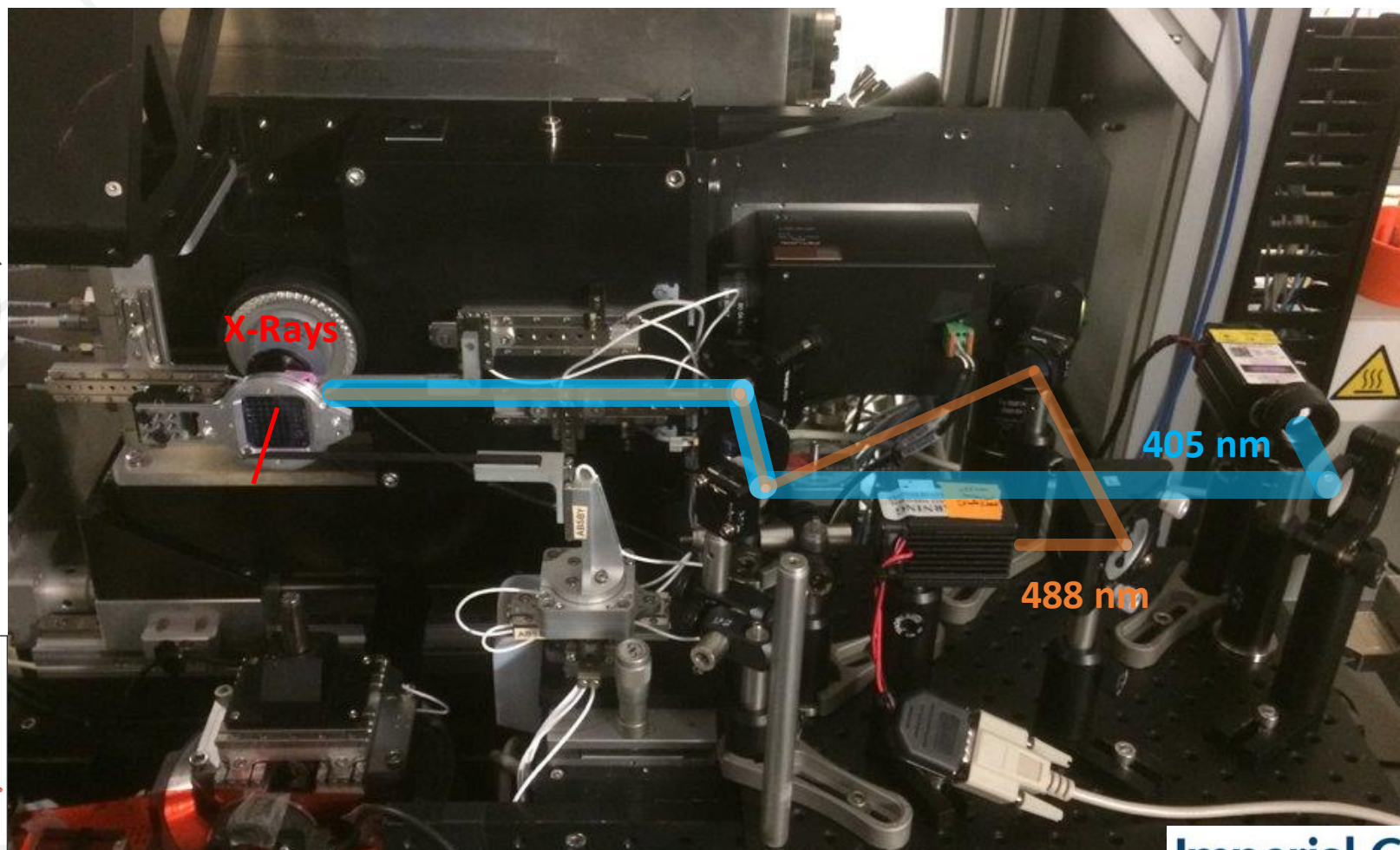
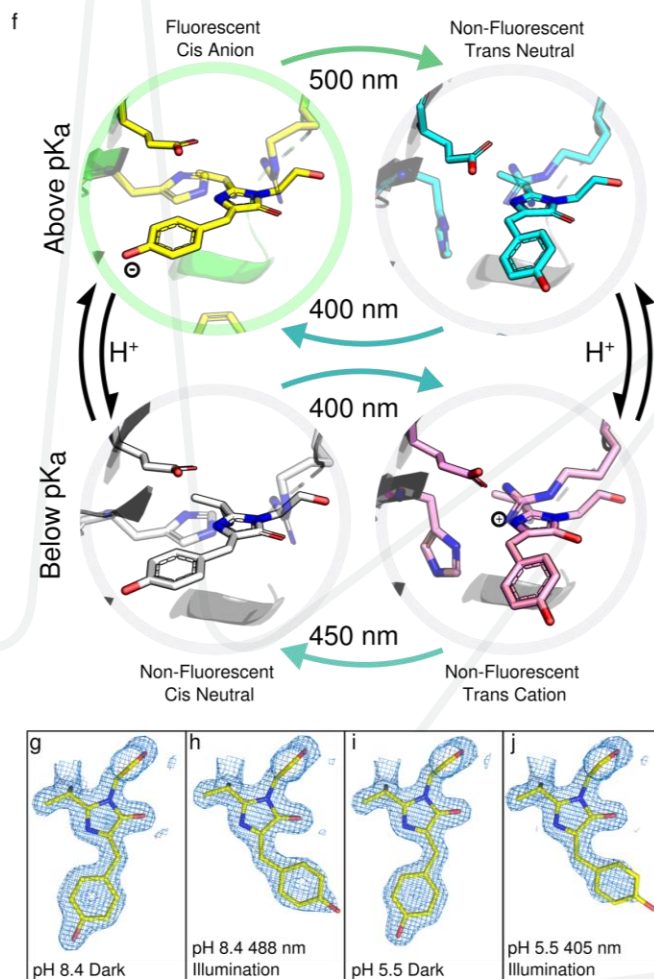
Example: Using VMXi to screen for serial experiments

You can get decent quality serial data from grid scans on VMXi – example: lysozyme



Statistics for eight wells merged together	Completeness (%)	Multiplicity	$I/\sigma(I)$	R_{split}	$CC_{1/2}$	Unique Observations	Indexed Images
Overall	100	95.5	20.8	0.063	0.998	8422	9891 / 25906 (38.18 %)
Low (55.58 – 5.43)	100	147.1	81.7	0.028	0.999	488	
High (2.03 – 2.00)	100	75.3	1.2	1.092	0.410	411	

Example: I24 SSX (Van Thor Group – Imperial College London)



Baxter, J. M., Hutchison, C. D., Maghlaoui, K. et al. (2022) J. Phys. Chem. B, 126, 45, 9288–9296

Furthering Output of Multi-Crystal Collections

Can we do more than just structure solution with multi-crystal data?

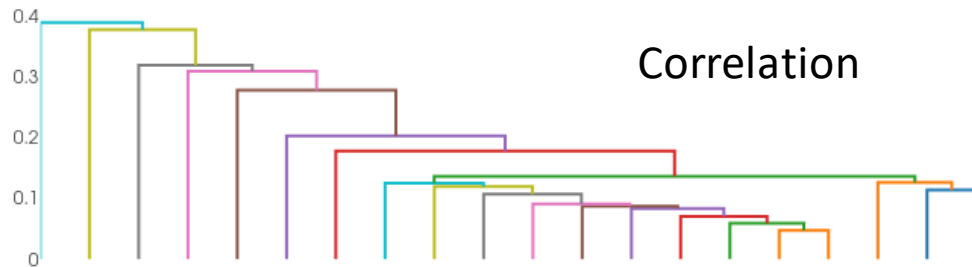
Clustering of Multi-Crystal Data

When merging many datasets together, there are going to be differences: especially when collected at room temperature!

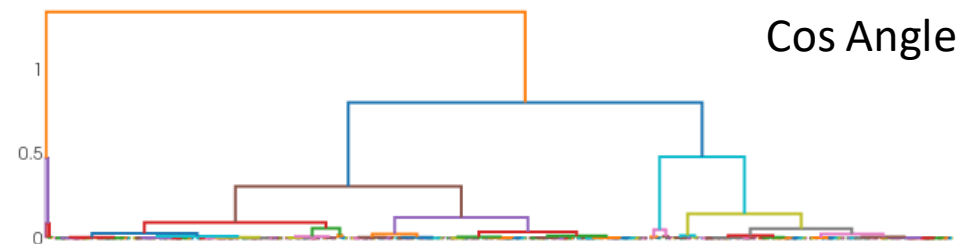
Multiplex includes two different intensity-based clustering tools to group datasets:

- Correlation Coefficient
- Cos Angle

Dendrogram Heatmap



Dendrogram Heatmap

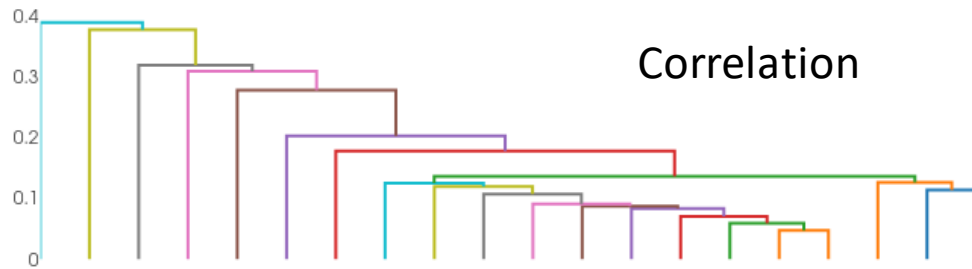


Clustering of Multi-Crystal Data

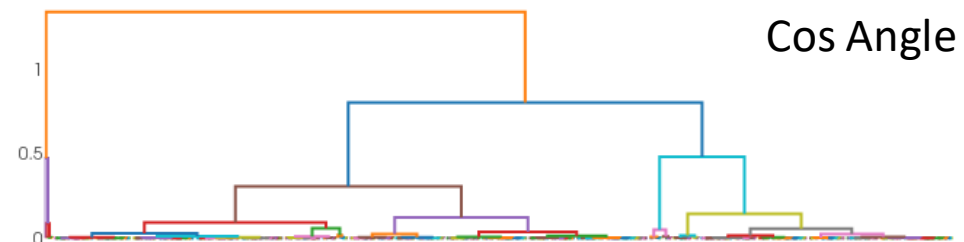
By analysing differences between clusters, can we extract more scientific outcomes from multi-crystal datasets?

- Structural dynamics?
- Ligand binding dynamics?

Dendrogram Heatmap



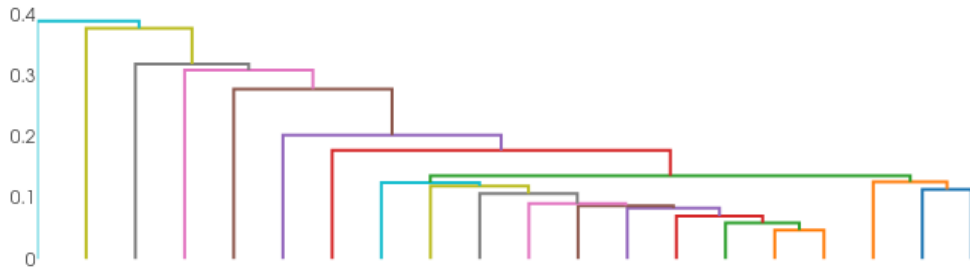
Dendrogram Heatmap



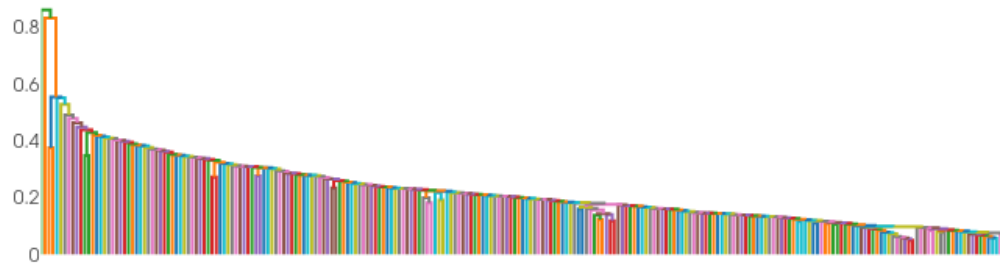
Correlation Clustering with xia2.multiplex

Clustering based on correlation coefficients can improve data quality for small datasets – but could it identify distinct states for large datasets?

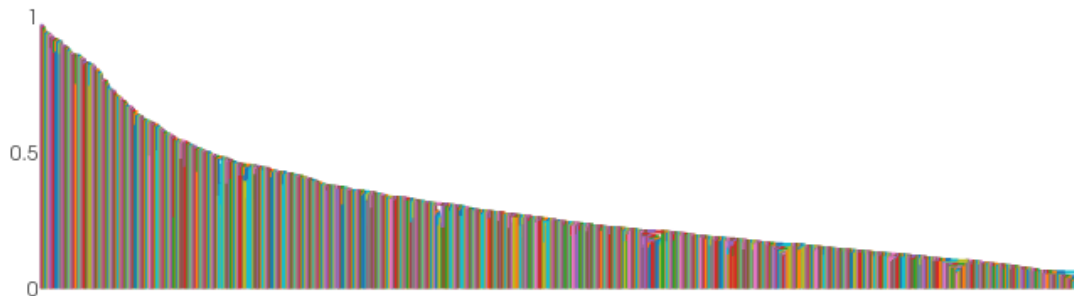
Dendrogram Heatmap



Dendrogram Heatmap



Dendrogram Heatmap



Issues with current method of cluster output!

Which ones are scaled and merged is based on completeness – for cubic crystals, 100% can be achieved with as little as 2 datasets. If you have thousands of datasets, this is thousands of clusters!

Computationally expensive and time wasting when you may only want to analyse a couple of clusters!

1500 x 20° wedges of insulin on VMXi

Improvements to Multiplex Towards Cluster Analysis

- Inclusion of a cluster picking algorithm to automatically identify clusters that may be of interest

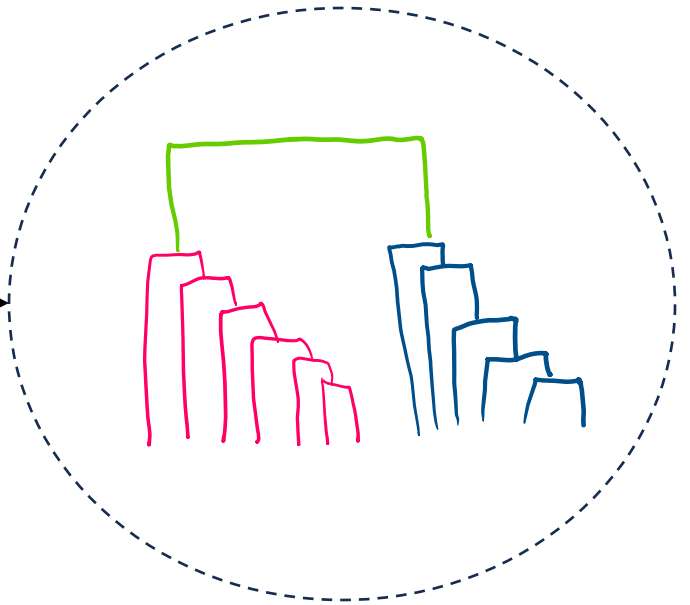
Run multiplex with
reference and
cluster analysis



Data is reindexed
to reference



Identify potential
clusters of interest



Output full merge
and interesting
clusters (as .mtz)

Consistent Rfree flags too!

Initial Findings – Insulin

Do notice some subtle, but distinct conformational changes between pairs of clusters, despite all clusters starting from the same initial model



Main example: flip of a well-defined HIS side-chain and corresponding water movement

From the merging of 500 datasets

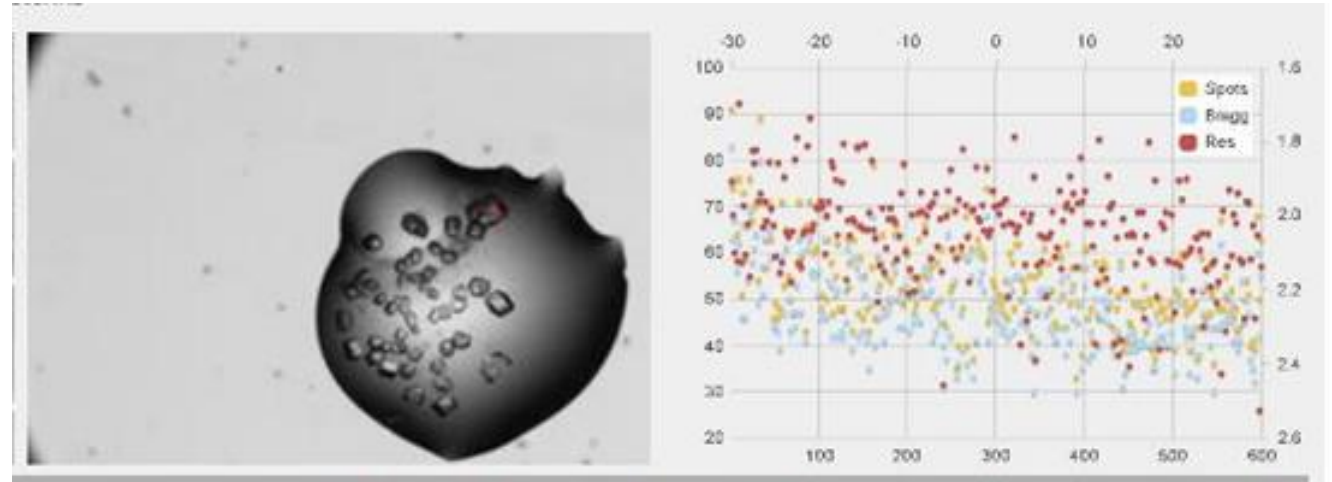
Applications to Ligand Binding?

Hypothetical situation: drop with multiple crystals, all datasets merged together, see a ligand binding. Great right?

But what if:

- Not all crystals actually had a ligand in them?
- Different crystals had different binding states?
- Different crystals had different ligand occupancies?

How much information can multiplex clustering provide towards answering these questions?



Initial Test Case - Lysozyme

Regular Lysozyme

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Measured whole plate on VMXi (LYS_Cl)
50 ° wedges

Drop of NaBr_(aq) on top of each position

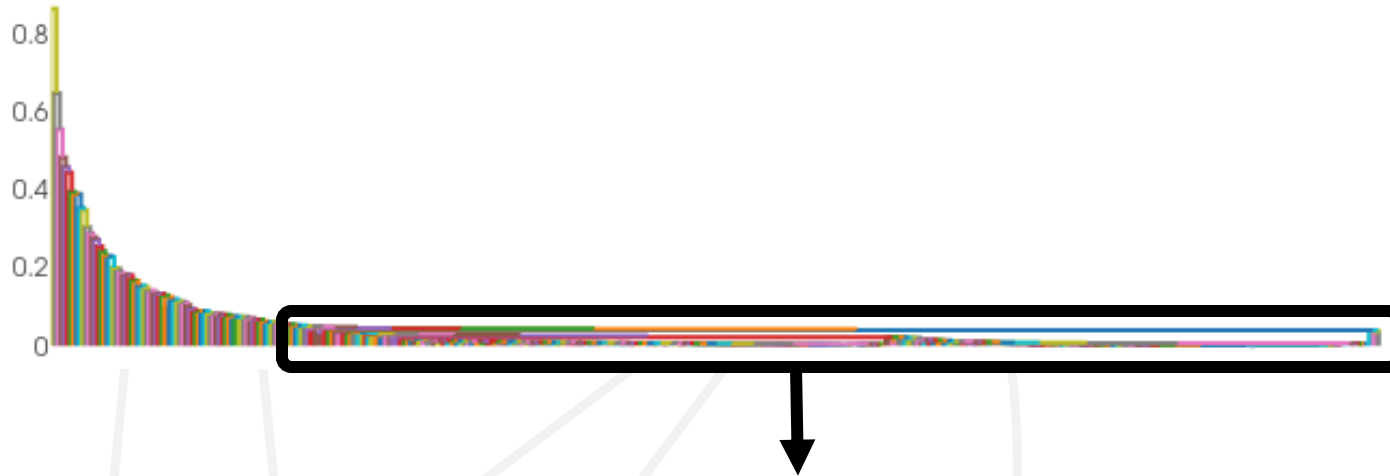
	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Measured whole plate on VMXi (LYS_Br)
50 ° wedges

LYS_Cl + LYS_Br through xia2.multiplex
(438 crystals merged together)

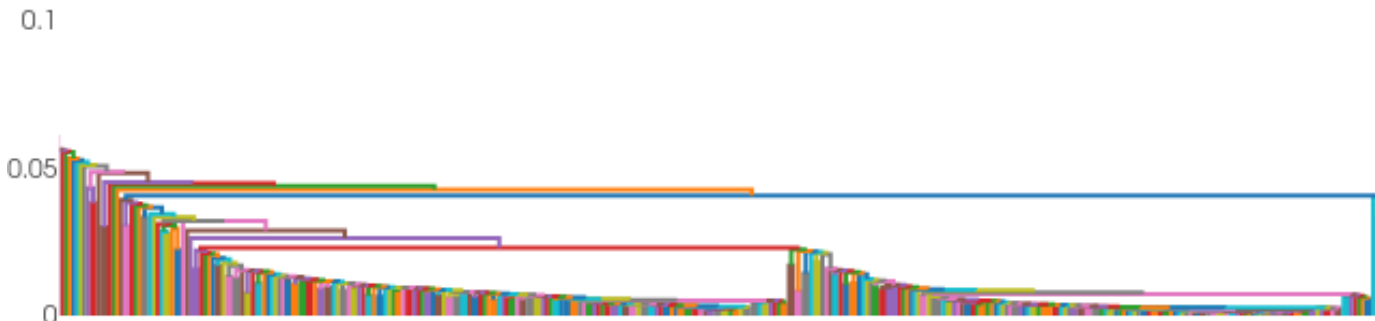
Correlation Clustering of Combined Lysozyme

Dendrogram Heatmap



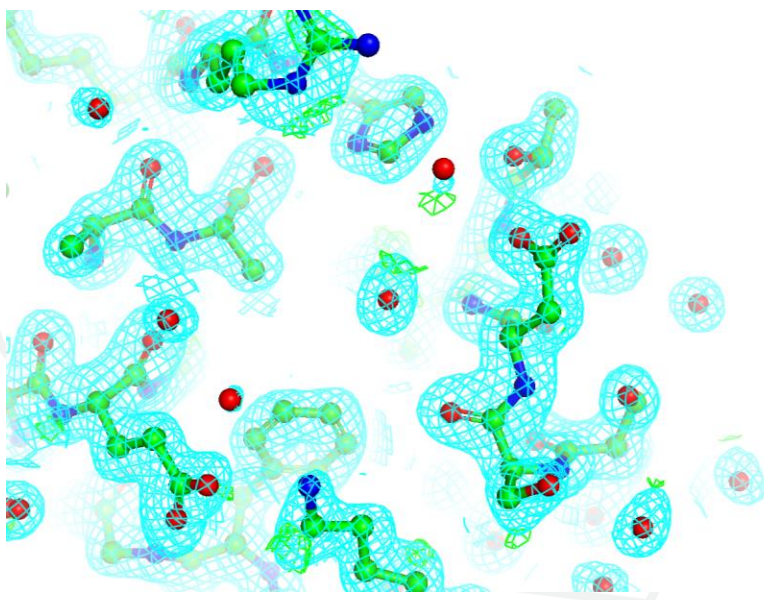
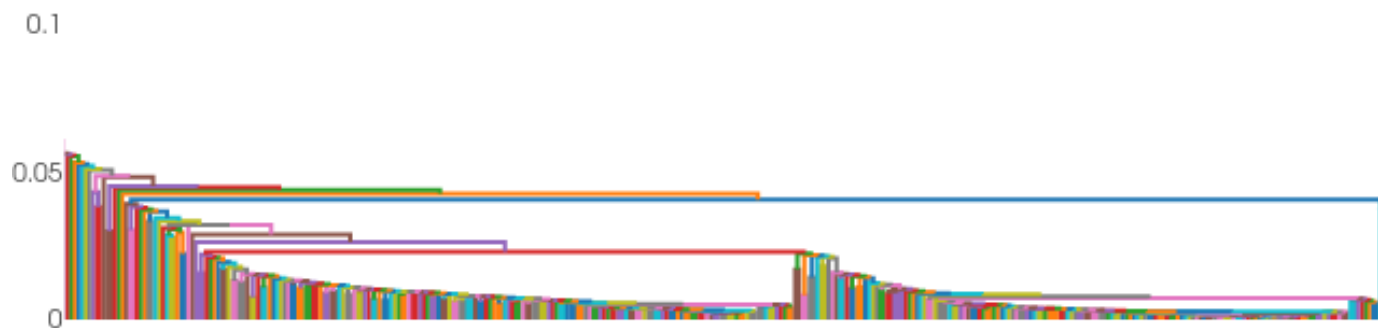
Datasets corresponding to each cluster indicates that they correspond to the two plates!

Dendrogram Heatmap

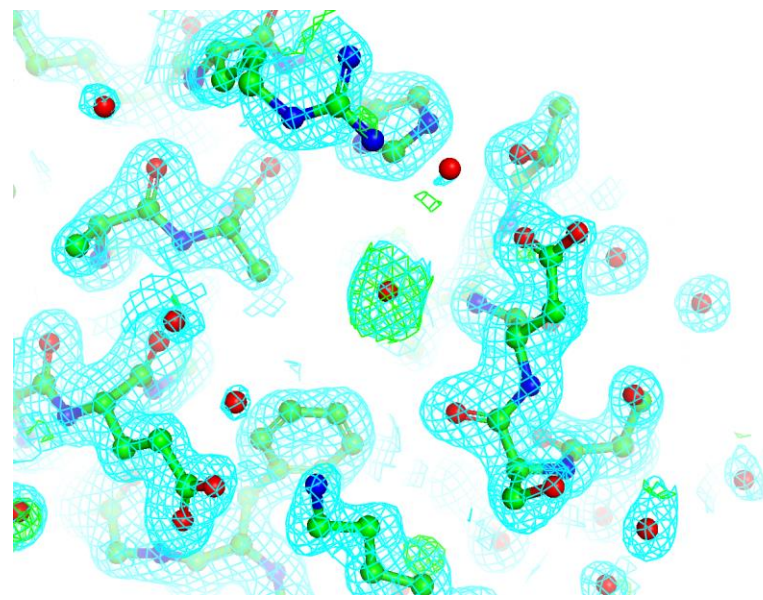


Correlation Clustering with xia2.multiplex

Dendrogram Heatmap



First cluster – only datasets
from NaCl plate



Second cluster – only datasets
from NaBr plate

Acknowledgements

VMXi:

- Michael Hough
- Juan Sanchez-Weatherby
- James Sandy
- Halina Mikolajek
- Megan Lambert
- Hans Pfalzgraf
- Sotaro Fujii
- Cicely Tam

Software Team:

- Graeme Winter
- James Beilsten-Edmands
- Richard Gildea



Questions?