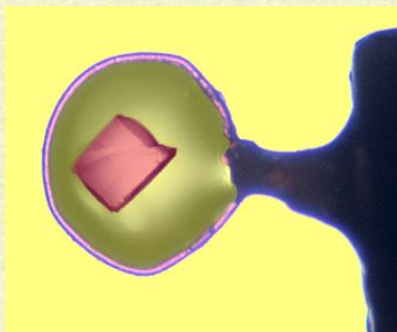
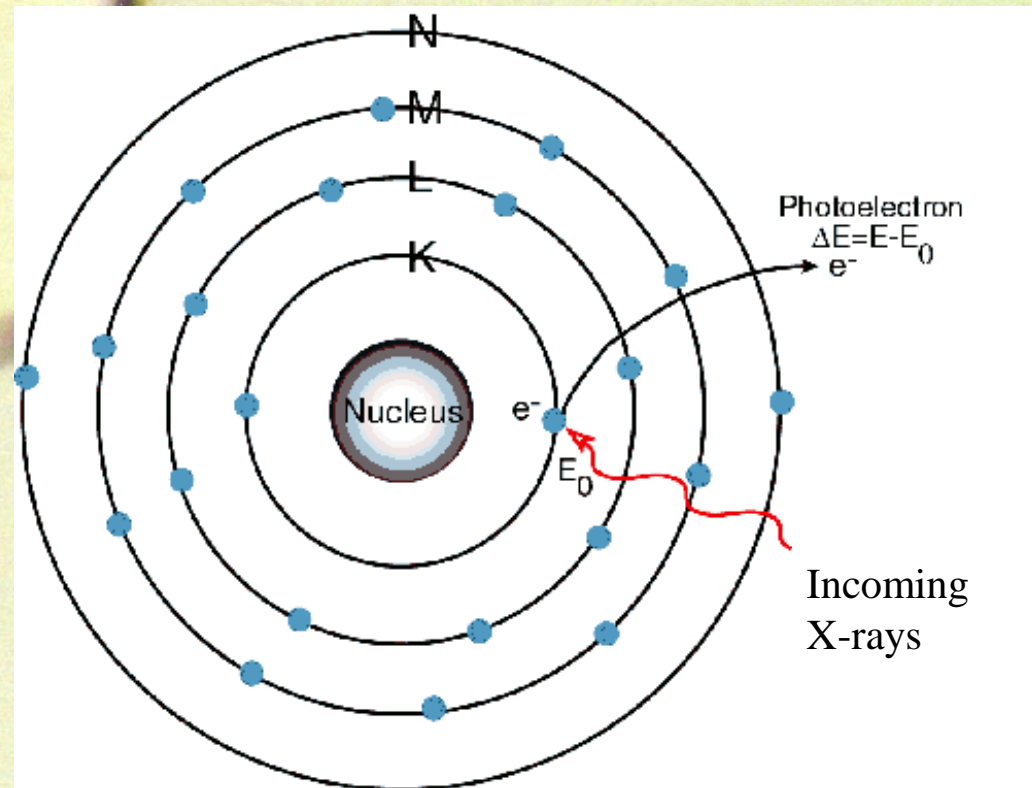
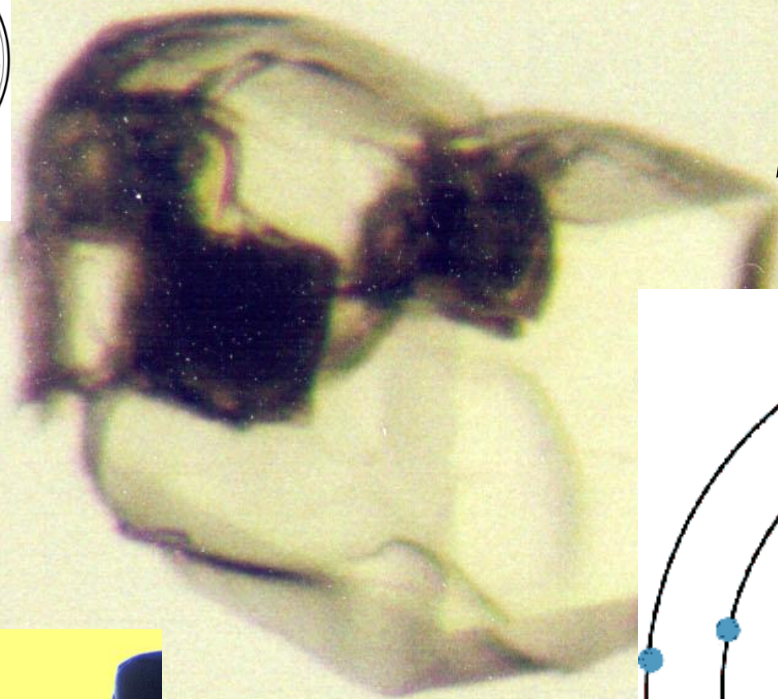
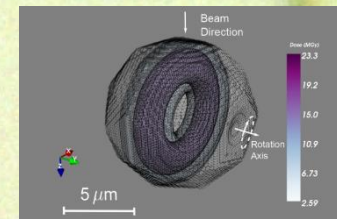
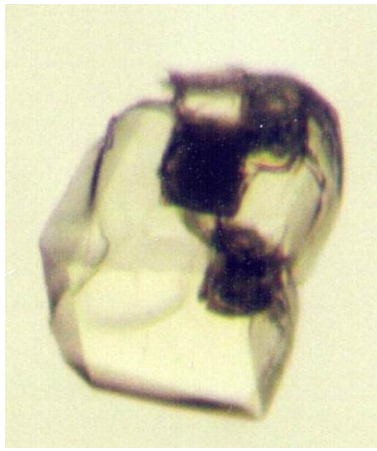


RADIATION DAMAGE:

why do we care?

*DLS/CCP4 Diamond/CCP4
Data Collection and
Structure Solution Workshop
21st November 2023*





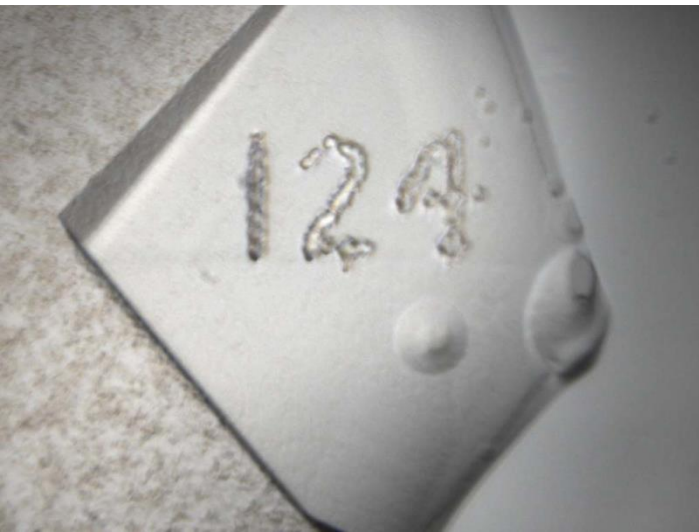
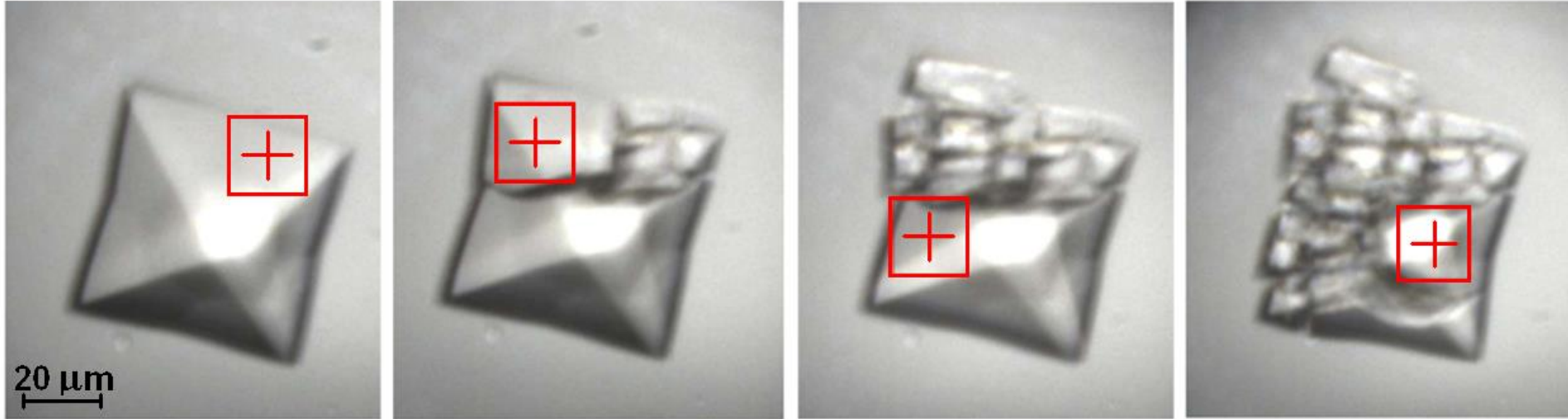
Radiation damage:

The Plan:



- **What are the symptoms?**
- What is it?
- Why do we care? Effect on MAD/SAD.
- How do we estimate the Dose?
- What do we know/would like to know?
- A new RD metric

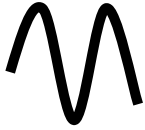
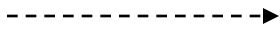
I24, Diamond, *in situ* data collection from a
Bovine Enterovirus 2 crystal, room temperature, 0.5 s
20 μm x 20 μm beam

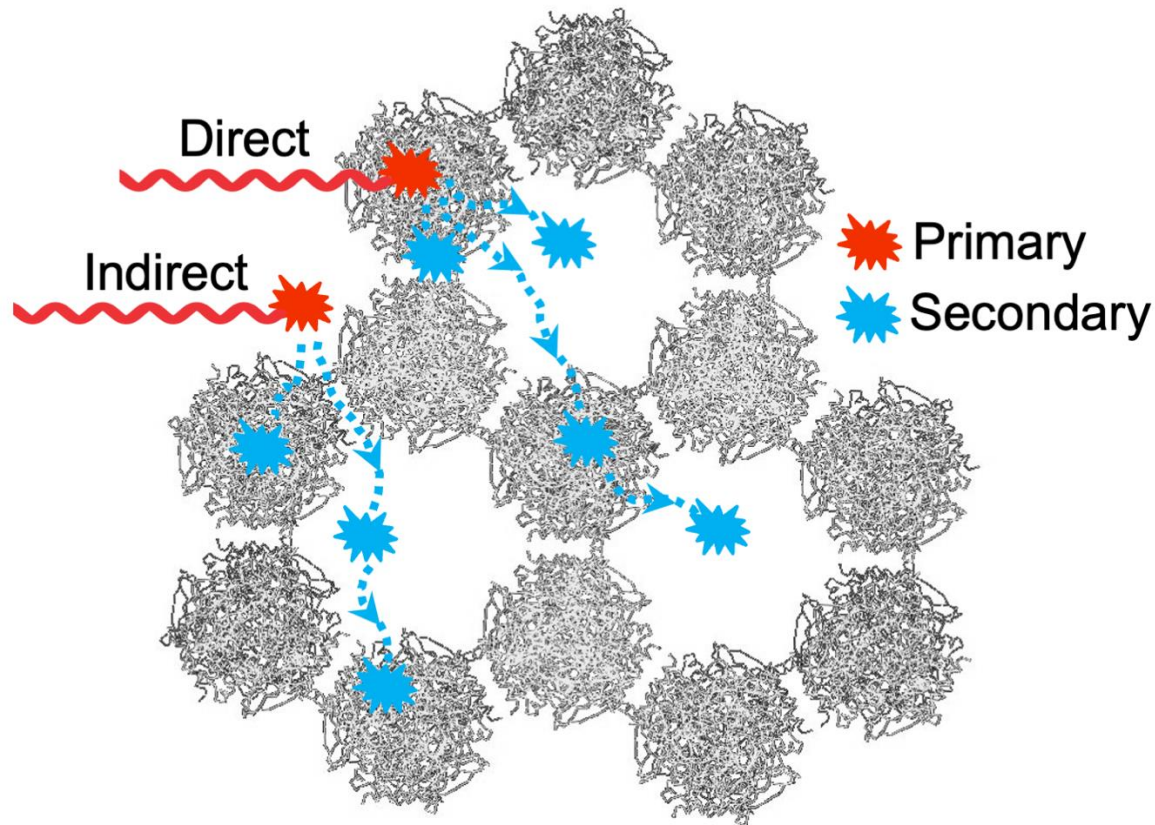


Axford *et al.*,
Acta Cryst D (2012) 592

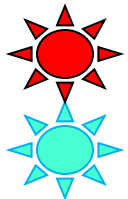
Beamline logo I24
(Gwyndaf Evans *et al.*)

Radiation damage

Primary 
Secondary 



Protein: direct
Solvent: indirect

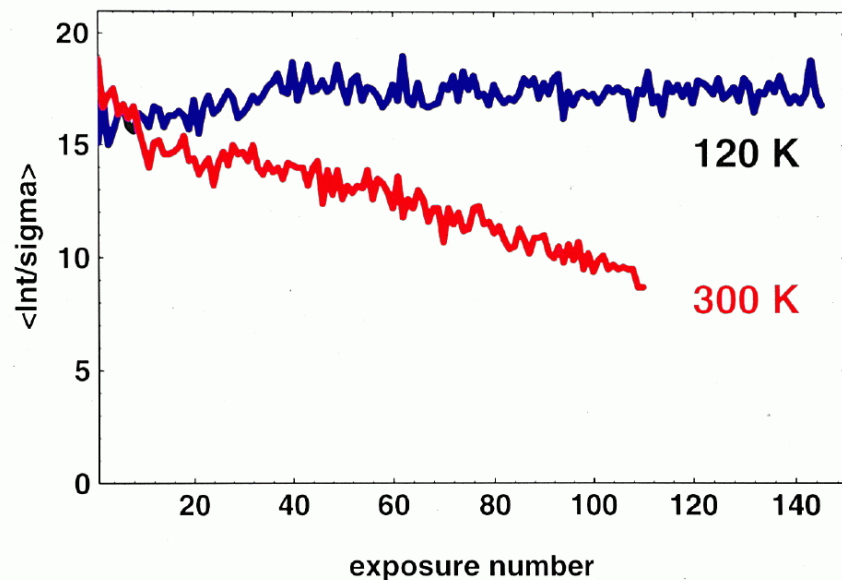
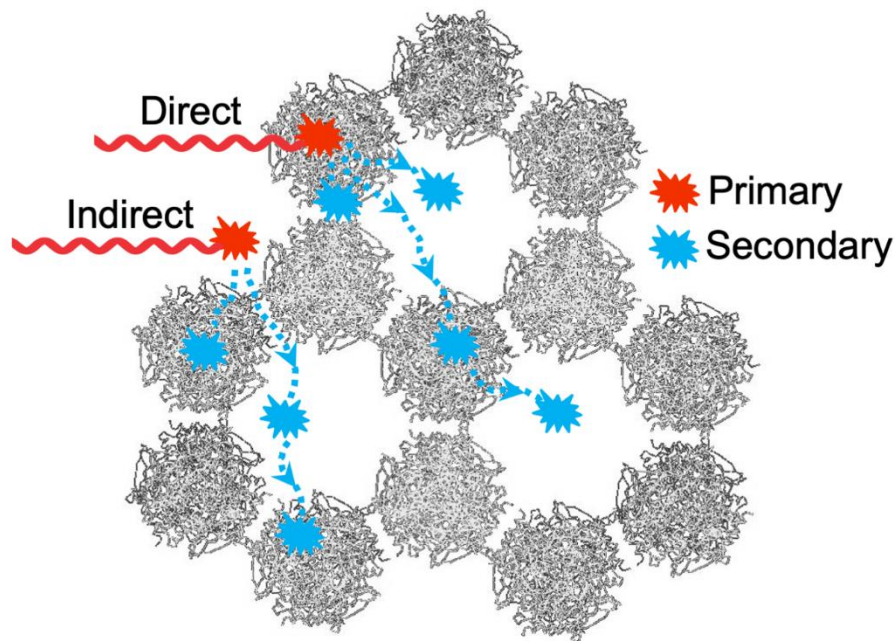


PRIMARY; inevitable, a fact of physics! Neutralise it?



SECONDARY, can we control it?

Radiation damage

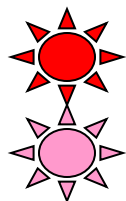
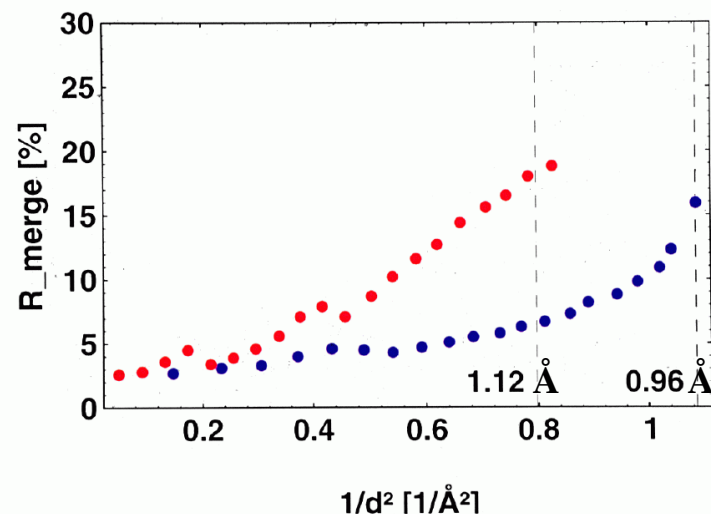


Significantly reduced at
100 K: time factor of ~ 70

[Nave and Garman *JSR* (2005), **12**, 257-260].

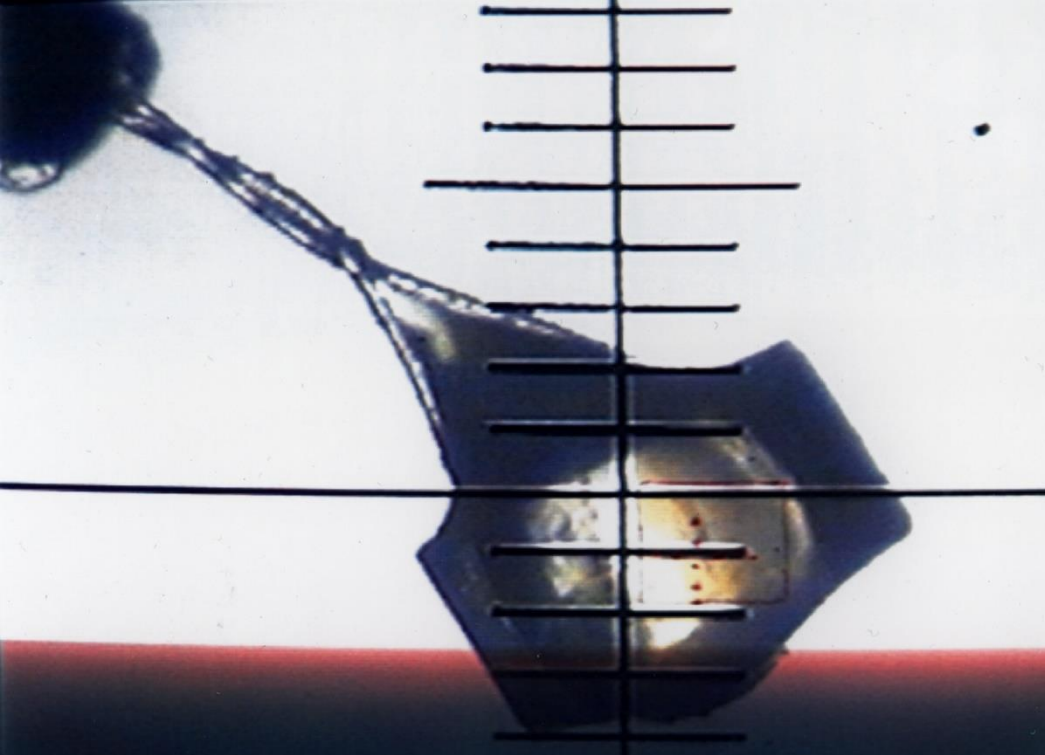
SP445: Data Quality

[T.Schneider]



PRIMARY; inevitable, a fact of physics! Proportions?

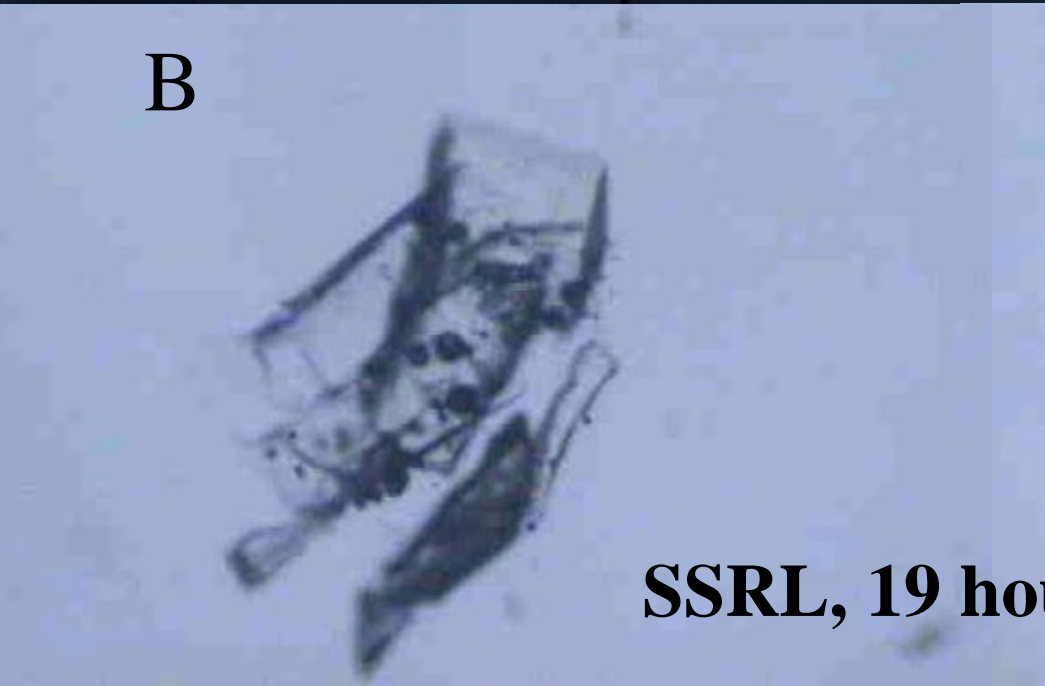
SECONDARY, can we control it?



A



B

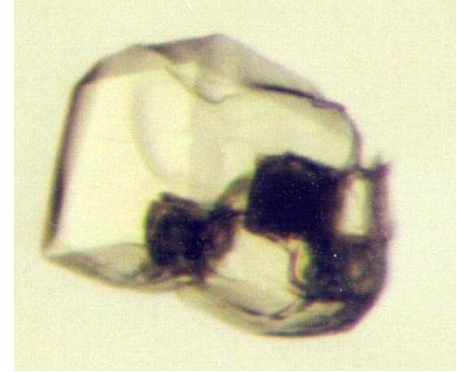
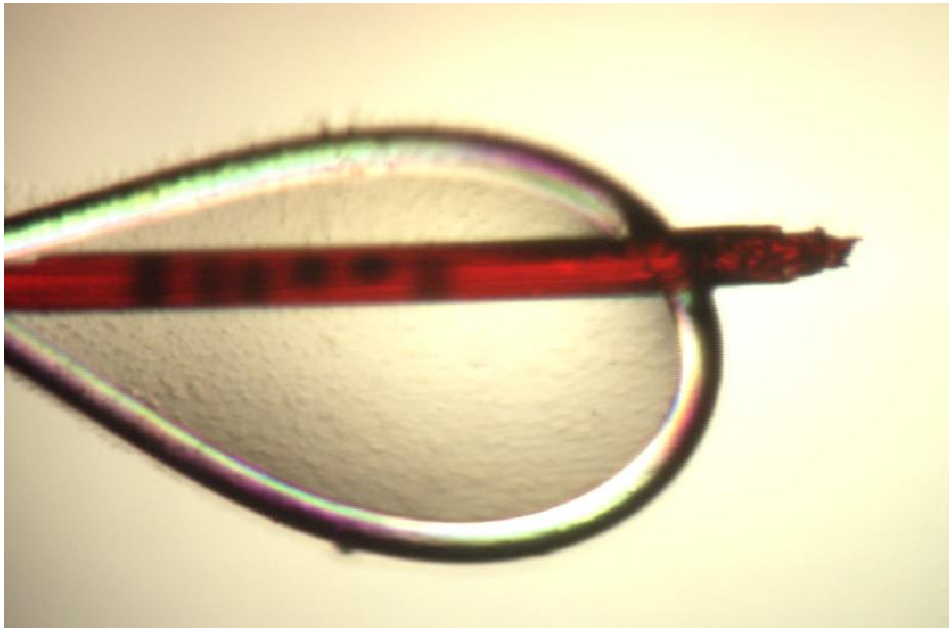


C

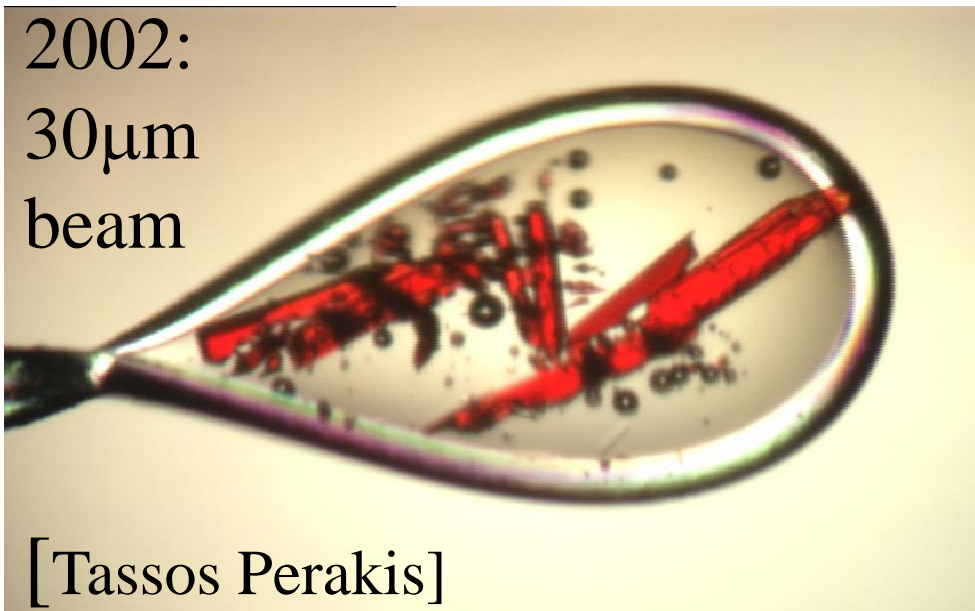


SSRL, 19 hours, 9.1, 1998

1995 onwards: 100 K
BUT THEN, 1999:



2002:
30 μ m
beam



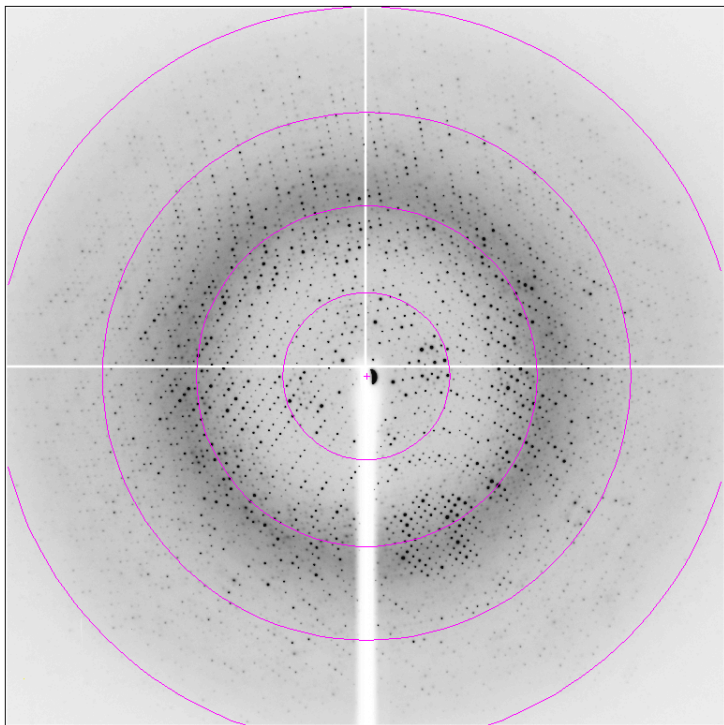
[Tassos Perakis]



Also observe
spectral changes

Iron containing protein, ESRF

Garman & Owen (2006), Acta D62



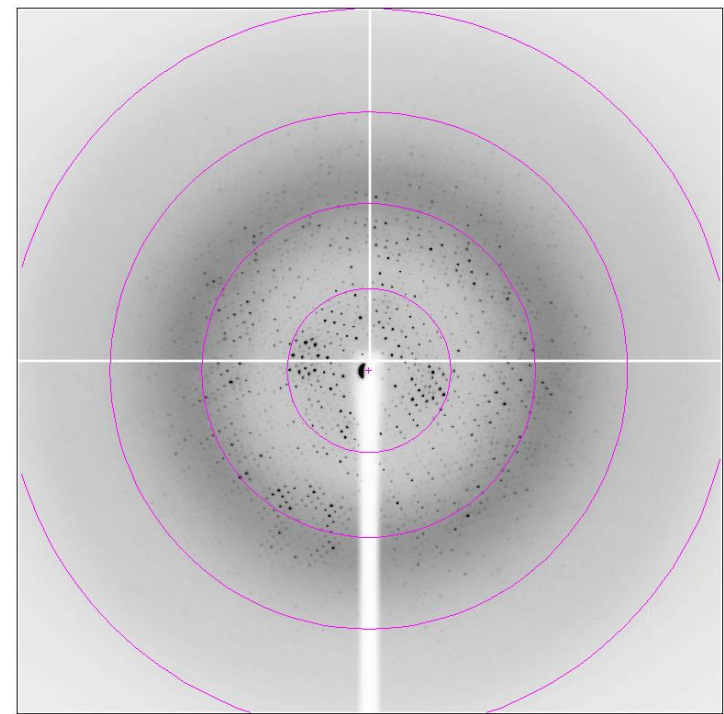
At 100 K

Intensity
decrease

Loss of
diffraction



Incomplete data
from crystals



Happens during 1 dataset at 100K for many crystals

Unit cell volume expansion

Increase in Wilson B factors, Rmerge

Increase in mosaicity

'GLOBAL' damage

ESRF 2000:

1×10^{12} ph s⁻¹ into
100μm square slits



**Diamond Light
Source:**

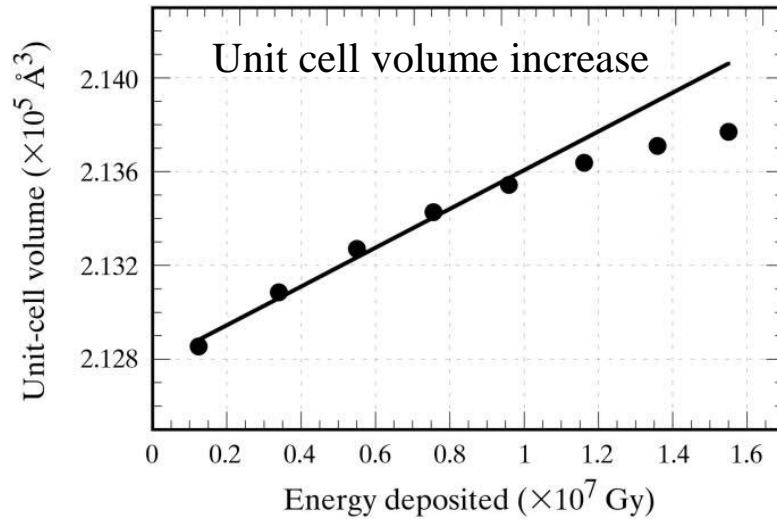
3×10^{12} ph s⁻¹ into
7μm × 6μm
[7 × 10¹⁴]

Australian synch.

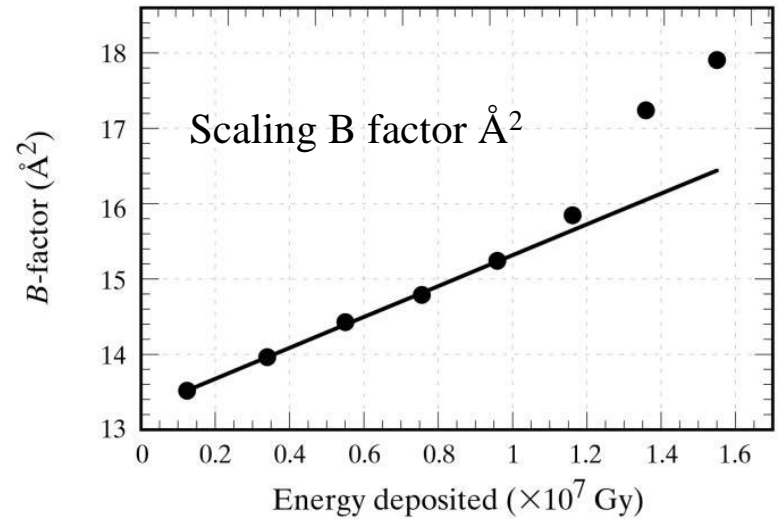
3×10^{13} ph s⁻¹ into
50μm × 70μm [10¹⁴]



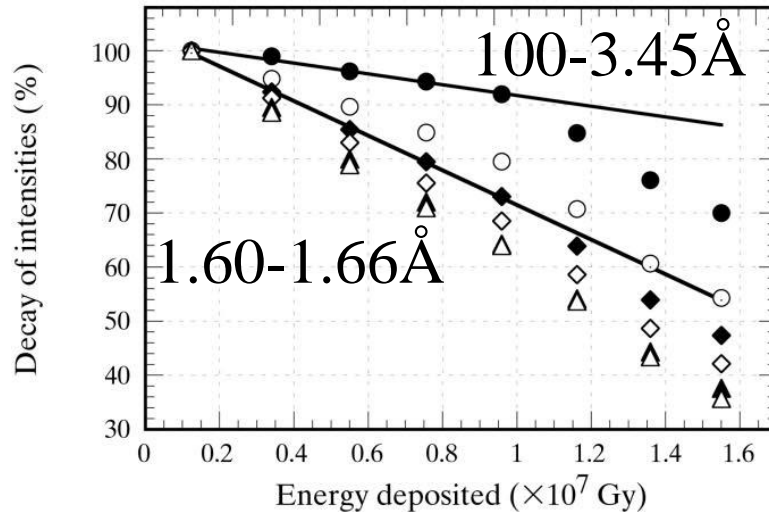
RadDam signatures in reciprocal space 100 K



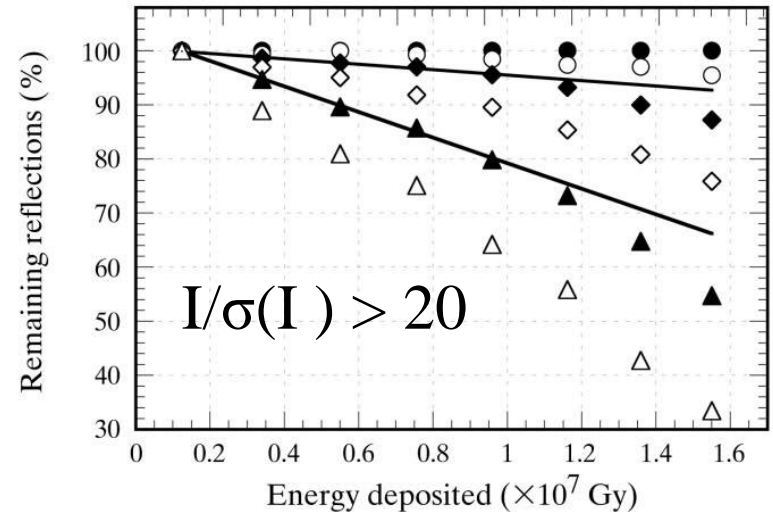
(a)



(b)



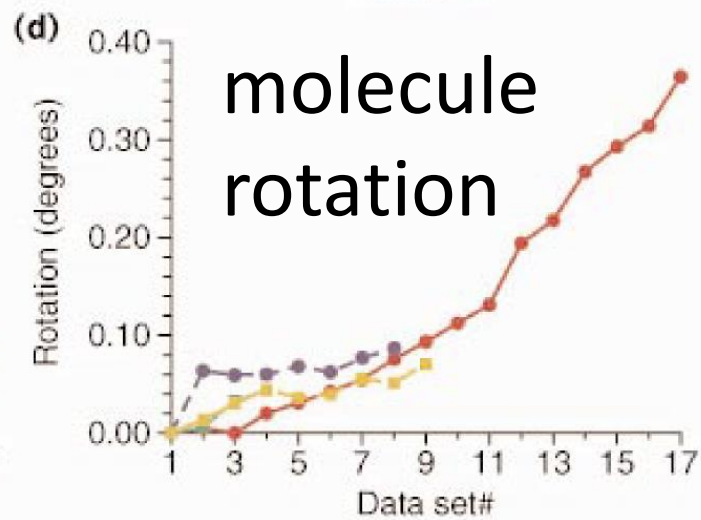
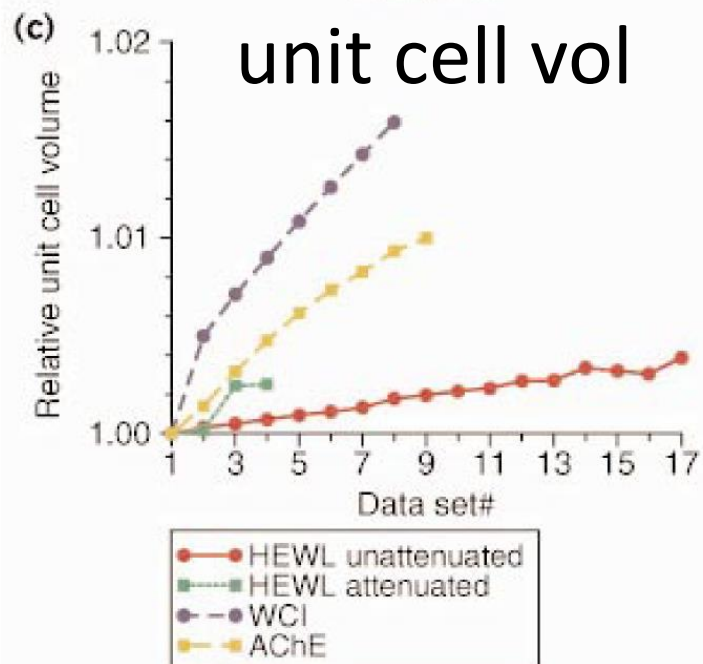
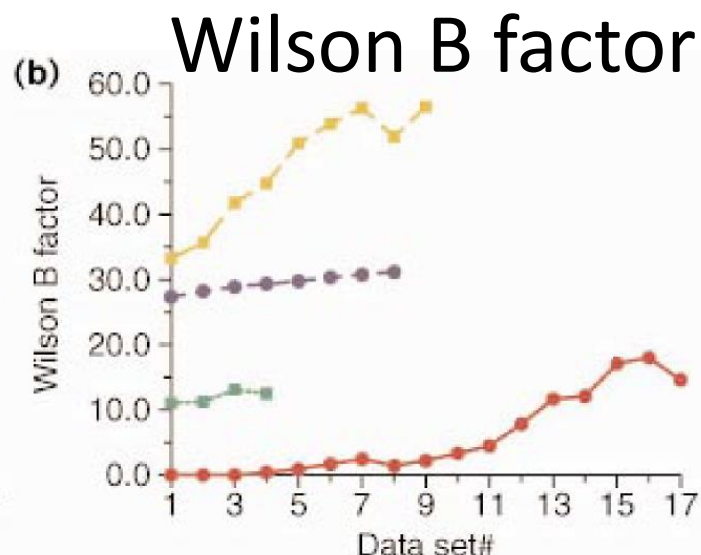
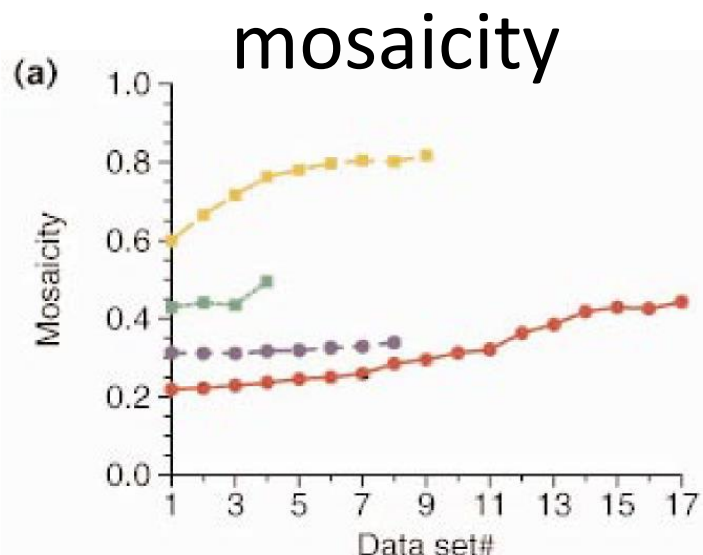
(c)



(d)

Intensity decay

Effects at 100 K



Raimond
Ravelli
1968-2023

RadDam signatures in reciprocal space

Characteristic 'U' type shape in scale factors plotted against image number:

ID14-4, ESRF
Ed Lowe

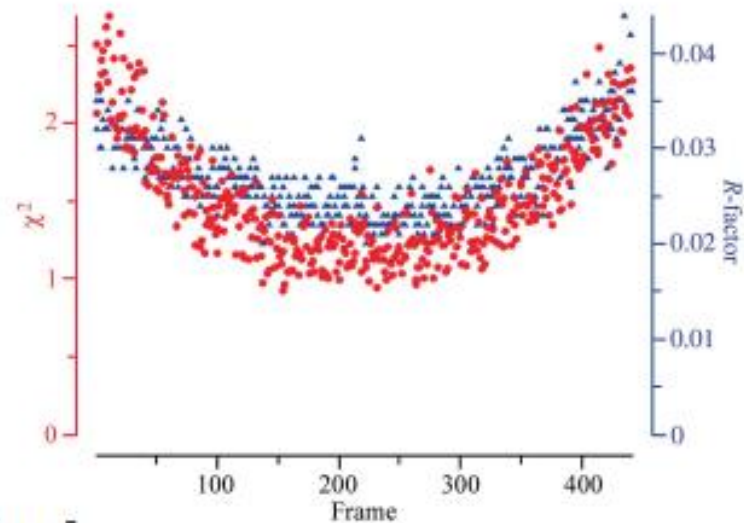
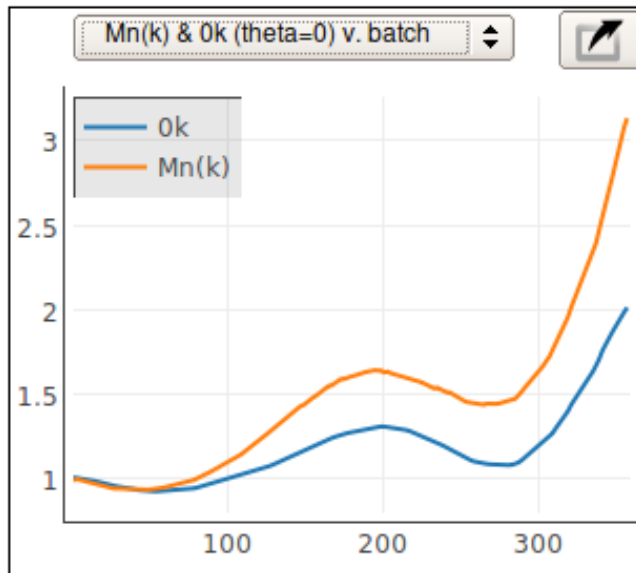








Figure 5

Statistics of scaling the individual images with the average of the data. The presence of radiation change creates a characteristic U-shape in the scaling statistics. Circles indicate $\chi^2 = \langle (I_n - \langle I \rangle)^2 / \sigma_n^2 \rangle$ values for individual diffraction images, triangles show the traditional scaling R -factor $R = \Sigma_n |I_n - \langle I \rangle| / \Sigma_n \langle I \rangle$, where in both cases sums are evaluated for all measurements and where the centroid of the diffraction spot is in a particular image frame.

Data Parameters affected by Radiation Damage

- $I / \sigma(I)$ or resolution limit 
- R_{merge} 
- Scaling B factors 
- Mosaicity 
- Unit Cell expansion a) function of dose 
b) function of cryogen temperature 

Could this be an on-line damage metric?

[Ravelli and McSweeney, (2000) Structure]

No!

[Murray & Garman (2002) JSR, Ravelli et al (2002) JSR]

What global damage
metric should we use and against
what should we plot it?

- I_n/I_1
- Not $I/\sigma(I)$
- Scaling B factors?
- An R_{meas} type measure?

To monitor the damage we need an x -axis!

for comparisons across synchrotrons &
even between beamlines at the same synchrotron:
we **can't** use time or image number.

DOSE estimation

$$\text{Dose} = \frac{\text{energy absorbed}}{\text{unit mass}} = \frac{\text{J}}{\text{kg}} = \text{Gy (gray)}$$

Fundamental metric against which to plot damage.

Cannot be measured, can only be estimated.

Takes care of the physics but NOT the chemistry.

RADDOSE-3D: www.raddo.se

MX experiment

1 MGy/s absorbed by a 100 μm cubed
metal free crystal in a

100 \times 100 μm^2 beam of

12.4 keV (1 \AA) X-rays

flux: 10^{13} photons s^{-1}

MX at 100 K: 30 MGy experimental
dose ‘limit’ reached in ~ 30 s:

4th generations sources $\ll 1$ s,

XFELs: < 80 fs

3 Gy

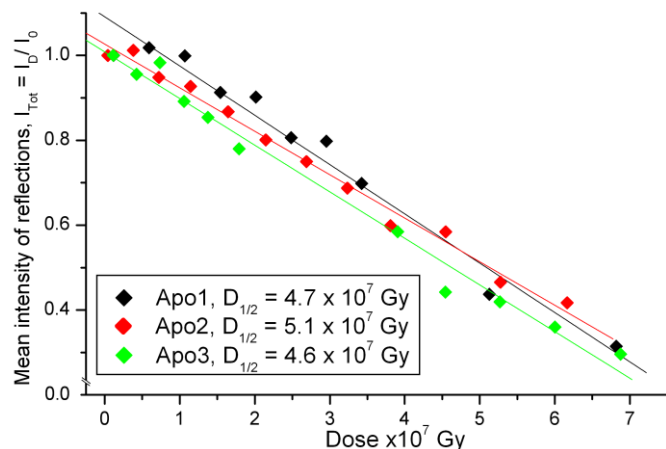


Intensity Decay at 100K

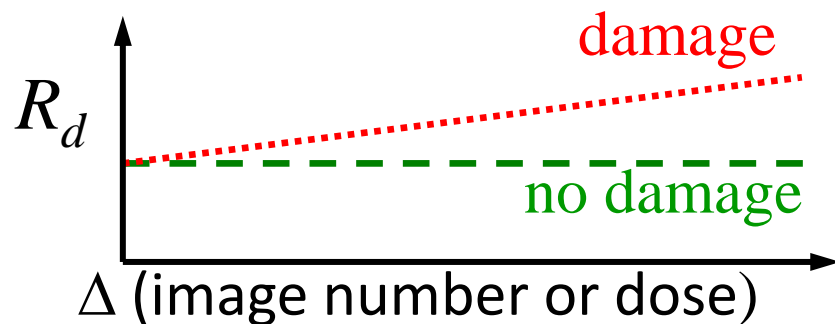
Normalised Intensity vs Dose:

apoferritin

[Owen et al 2006, PNAS]



[Deiderichs 2006 Acta D59, 903]



R_d : pair wise R factor between identical and

symmetry-related reflections occurring on different diffraction images

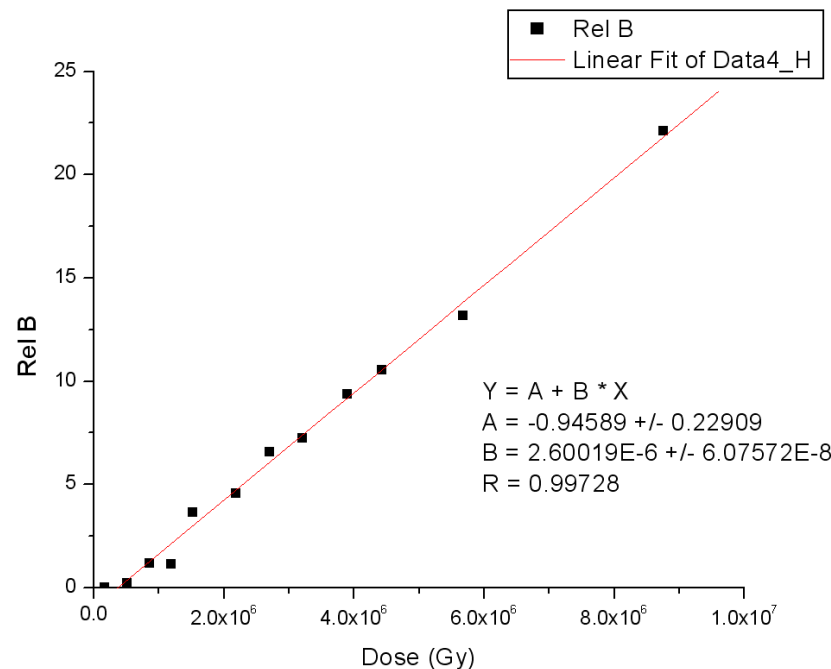
Coefficient of sensitivity α

change in relative isotropic B factor:

$$s_{AD} = \Delta B_{rel} / 8\pi^2 \Delta D$$

(e.g. HEWL@100 K = 0.012 Å²/Gy)

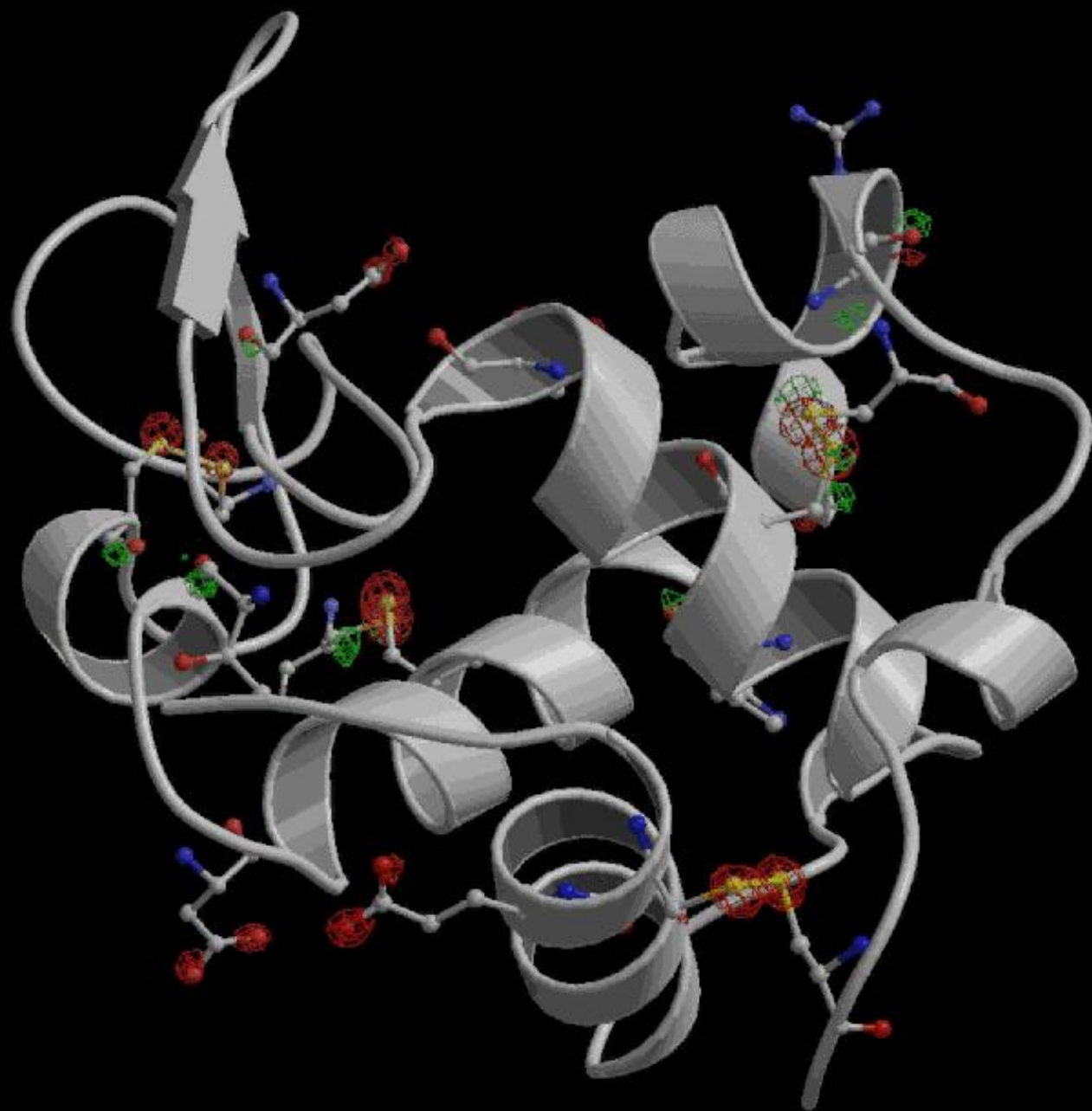
[Kmetko et al 2006, Acta D62, 1030]



Global damage: summary

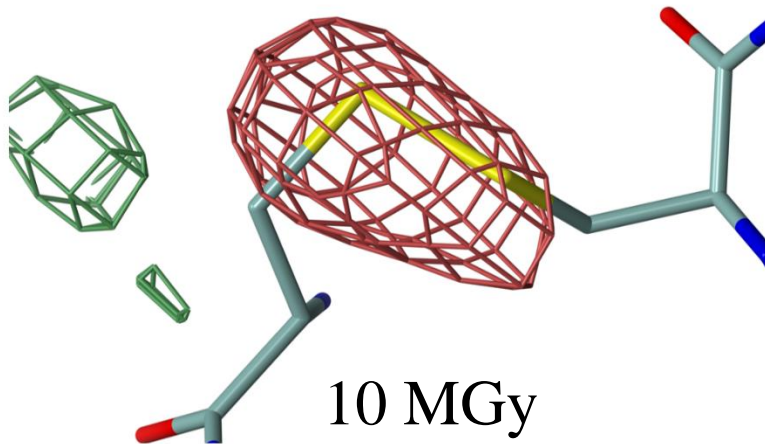
- Incomplete data/lost resolution/scaling 'dip'
- Causes non-isomorphism within a dataset (unit cell grows)
- No significant* ($< \times 2$) dose rate effect at 100 K at current flux densities (10^{15} ph/s/mm²).
- No significant ($< \times 2$) temperature dependence below 100 K, but weak minimum at around 50 K.
- Damage to lattice due to hydrogen abstraction and then build up?
- Heating not significant at current flux densities.
- For a particular system is predictable/can be modelled (using a sacrificial crystal)

* James Holton criterion for a significant result

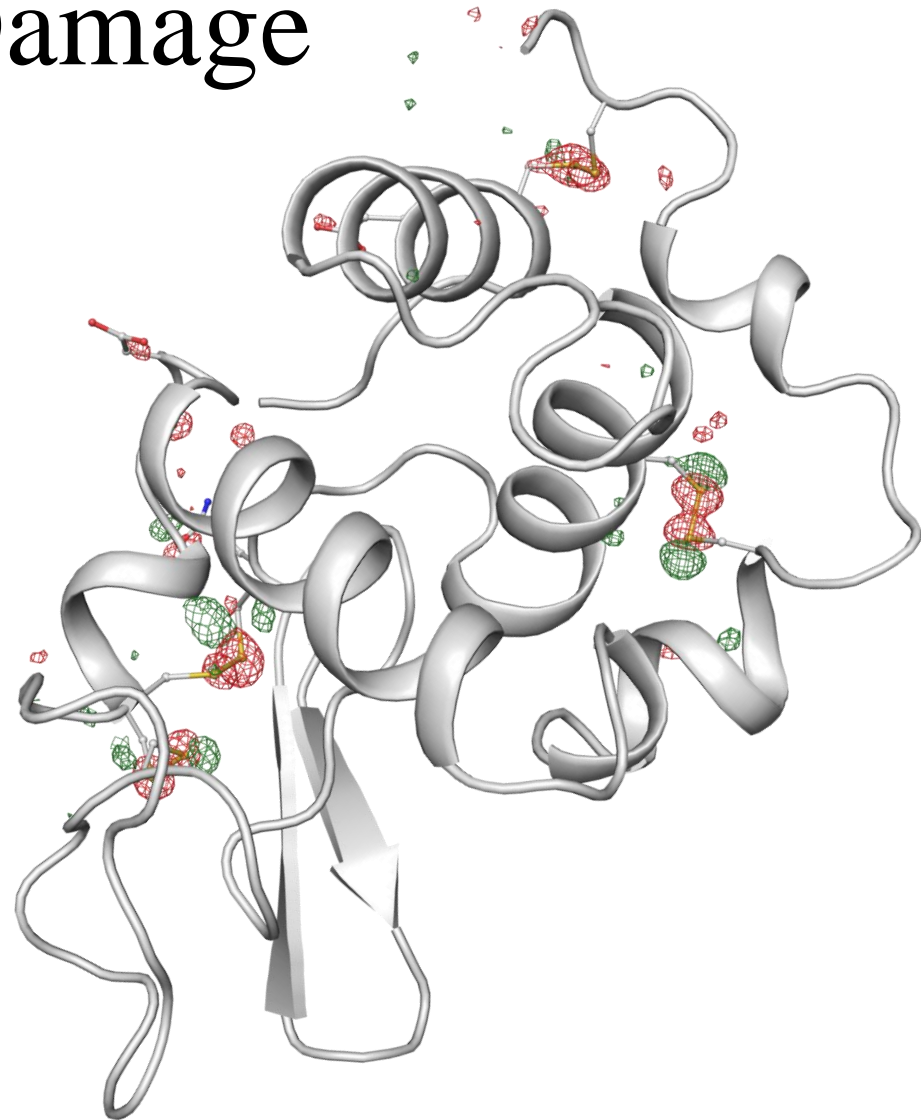


Specific Damage

- Specific damage occurs in a predictable hierarchy in proteins
- Reduction of metallocentres
- Breakage of disulphide bonds
- Asp and Glu decarboxylation



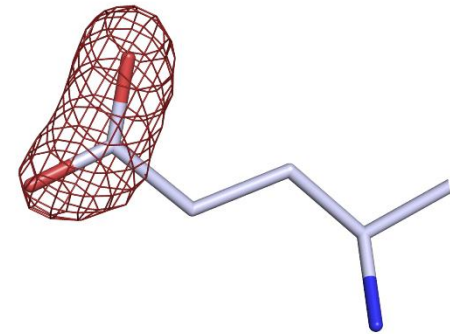
Difference map Fo_4-Fo_1



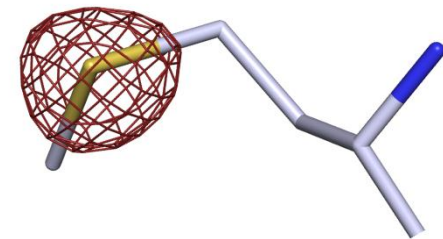
Weik et al. 2000, Burmeister 2000
Ravelli and McSweeney 2000,

Specific Damage

- Specific damage occurs in a predictable hierarchy in proteins
 - Reduction of metallocentres
 - Breakage of disulphide bonds
 - Asp and Glu decarboxylation
 - Cleavage of S—C bond in MET
 - Rupture of covalent bonds to heavier atoms:
C-Br, C-I, S-Hg
- **Note** that if this were due to primary damage alone, damage would be in order of absorption cross sections of atoms, which it is not.

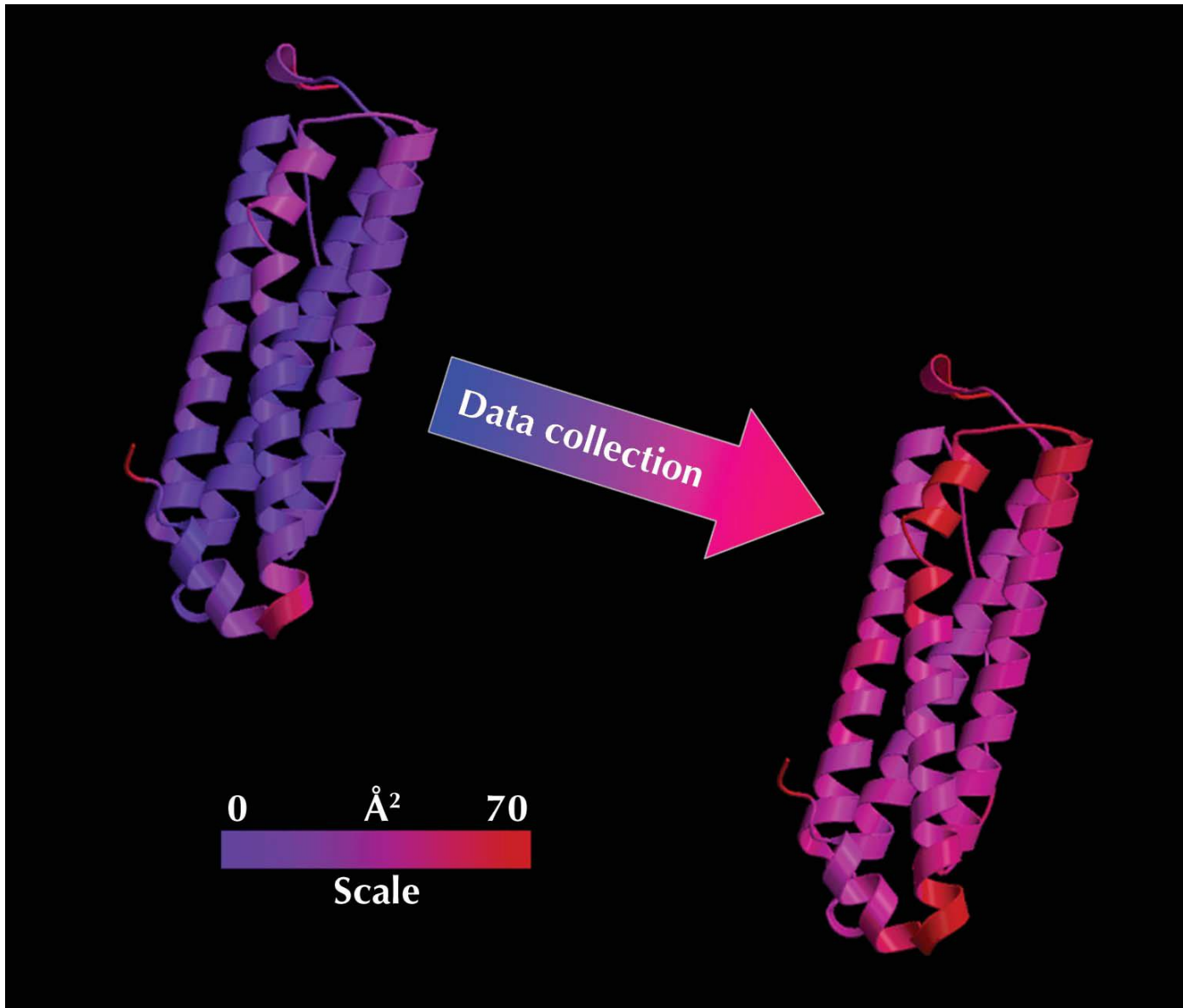


*GLU $\pm 4\sigma$ -level
difference map:
Red = negative density*



*MET $\pm 4\sigma$ -level
difference map:
Red = negative density*

ALSO:
Atomic B-factors increase:

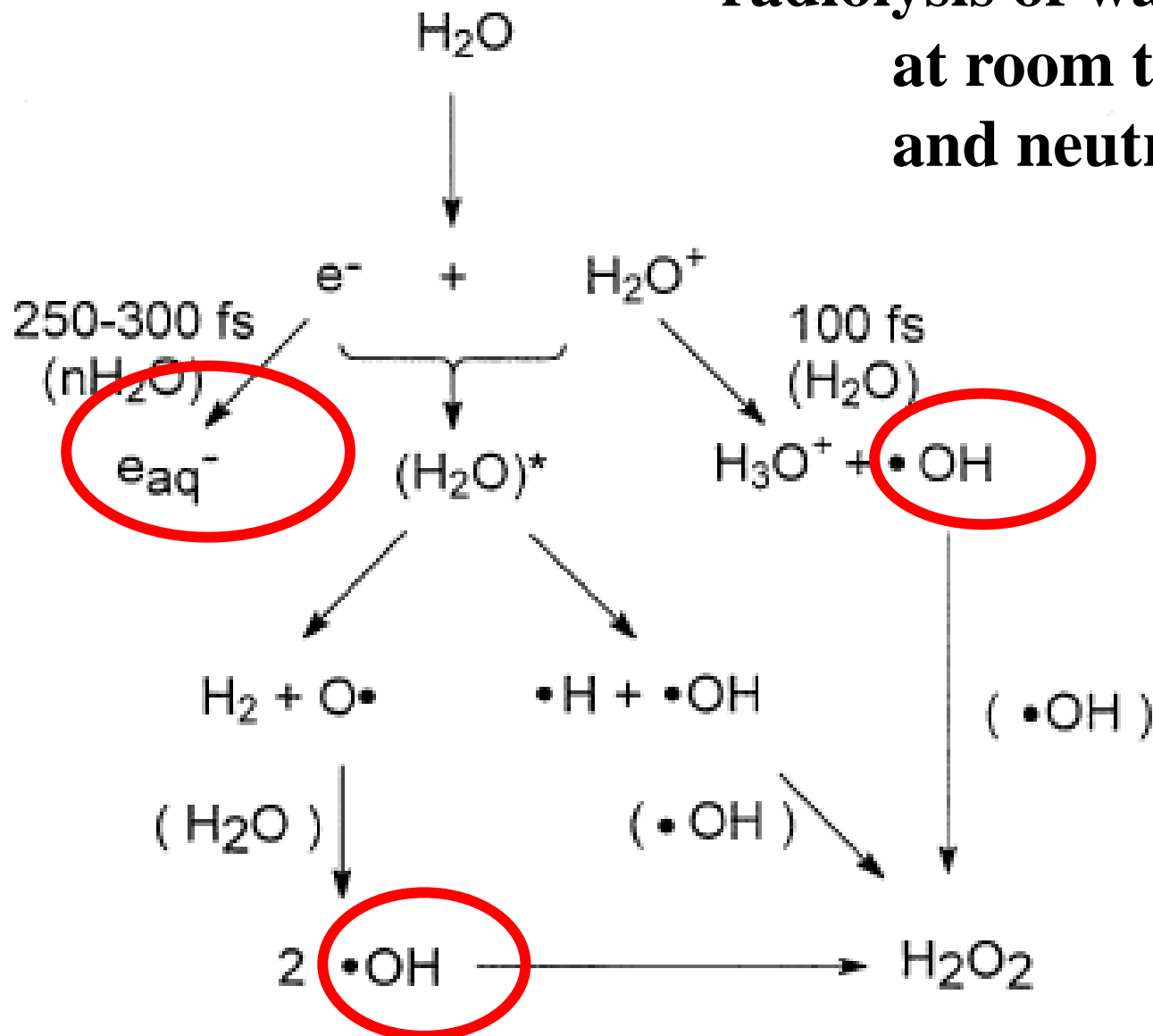


Damage: the Radiation Chemistry

1) INDIRECT RADIATION DAMAGE :

radiolysis of water

at room temperature
and neutral pH:

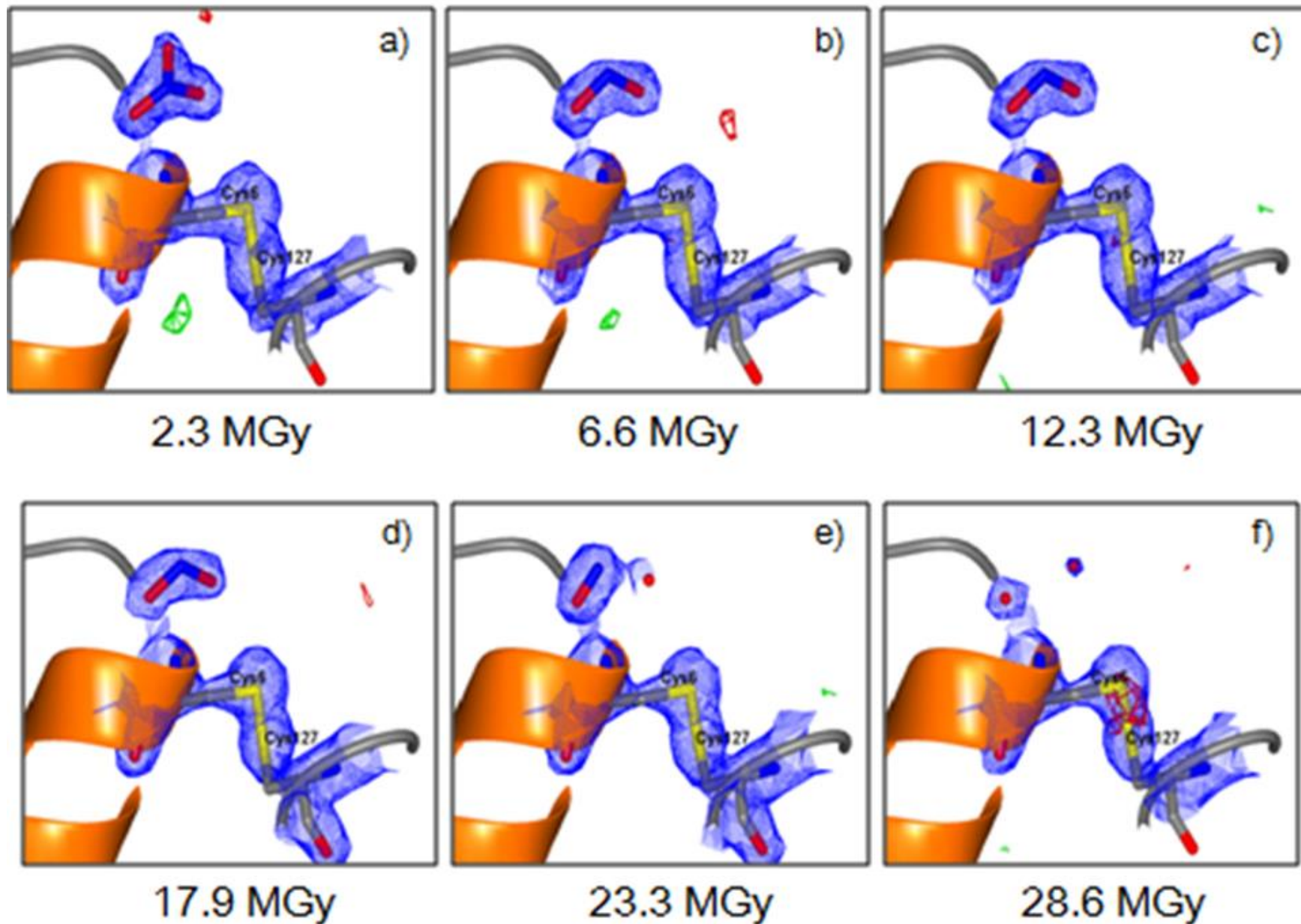


OH thought not to be
mobile in glasses
below 110K

(Owen et al Acta D
2012)

Hiroki, A. Pimblott, S. M.
LaVerne, J. A. (2002)
J Phys Chem A **106**,
9352-9358

Radiation Chemistry in action: Nitrate scavenger



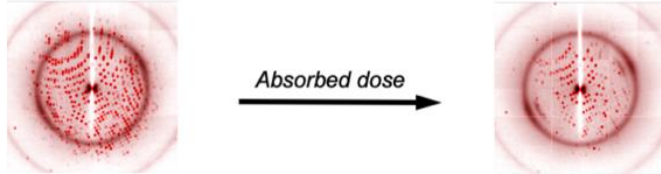
Specific Damage: summary.

- Can compromise biologically relevant observations (e.g. damage enzymatically important glutamates).
- Metallo-enzymes are reduced by X-ray beam.
- Perhaps weakly dose rate dependent ($< \times 2$)
- Perhaps weakly wavelength dependent ($< \times 2$)
- Weakly temperature dependent (varying results) ($< \times 2$)
- Can be reduced with certain scavengers: but very conflicting results (mainly $< \times 2$, benzoquinone RT $\times 9$)
- We DON'T understand pecking order of damage within an amino acid group pH? Solvent accessibility?
Neighbouring amino acids?

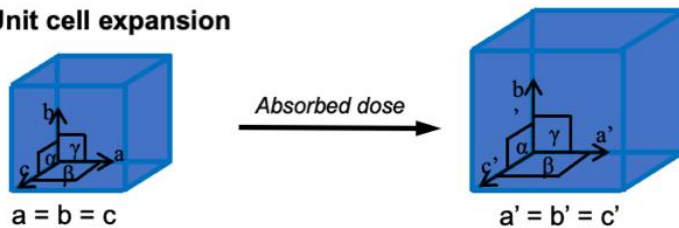
Summary

Global radiation damage

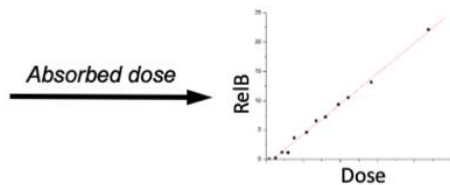
Decreased reflection intensities



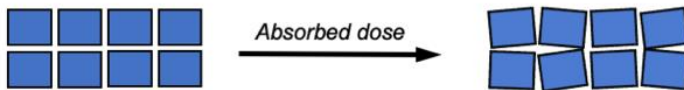
Unit cell expansion



Scaling B factors



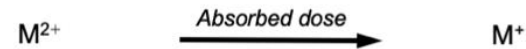
Increased mosaicity



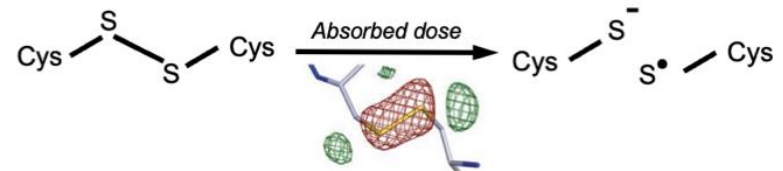
Specific radiation damage

Chemical changes

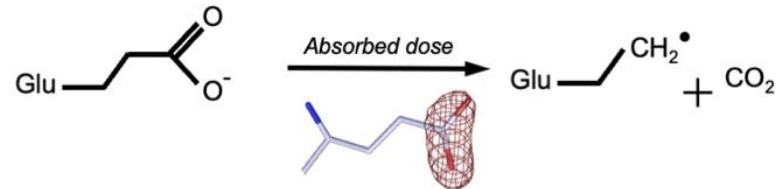
Reduction of metal ions



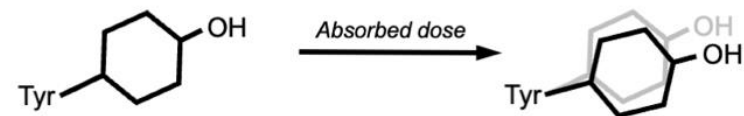
Reduction of disulphide bonds



Glutamate / aspartate decarboxylation

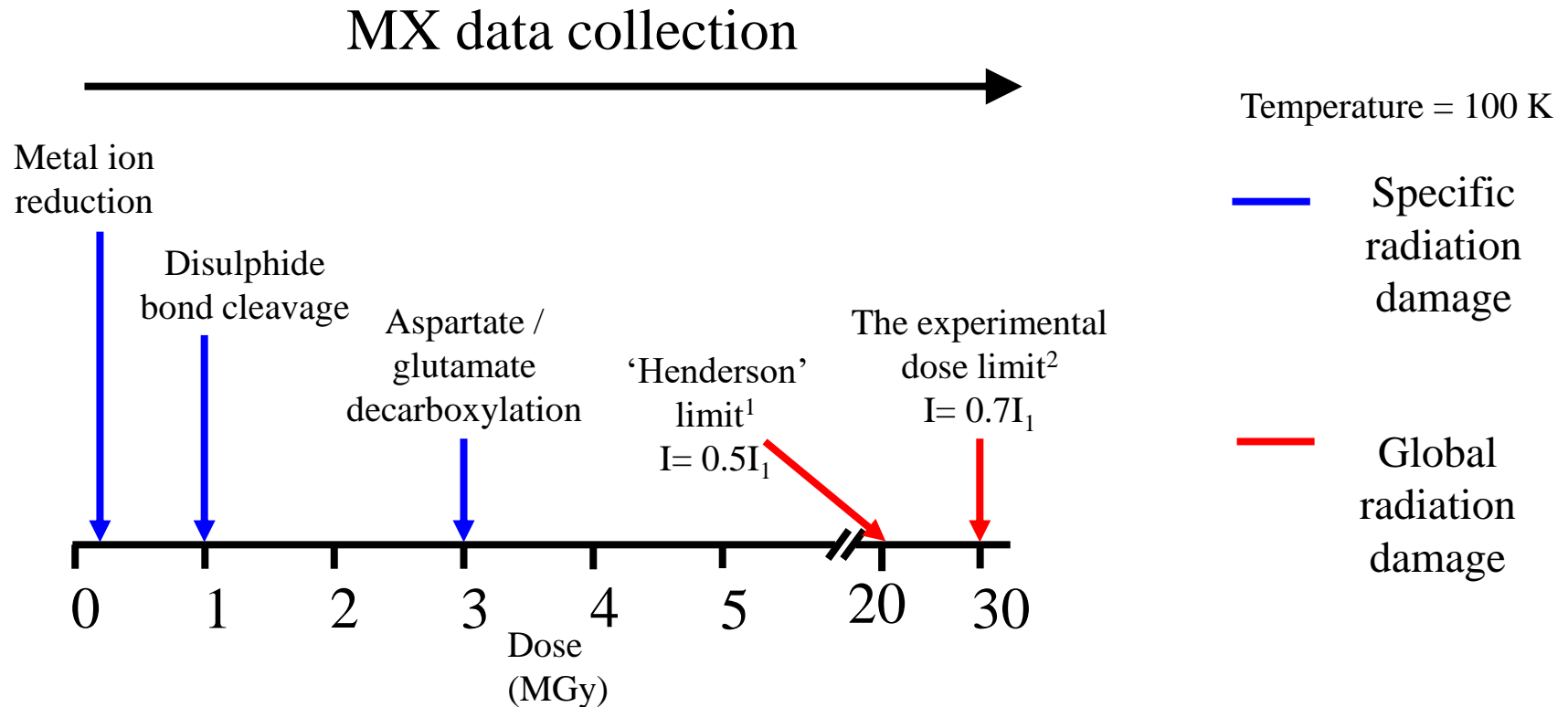


Side / main chain motion



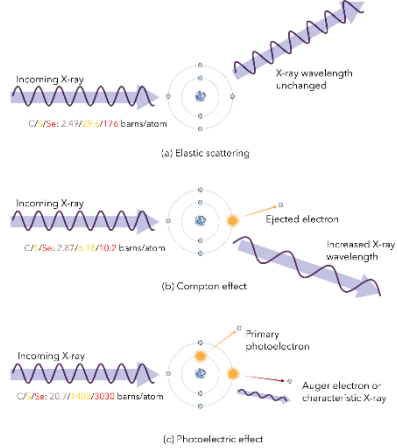
At T=100 K:

- Specific damage effects onset before global damage
- Both classes of damage common in protein crystallography (MX)



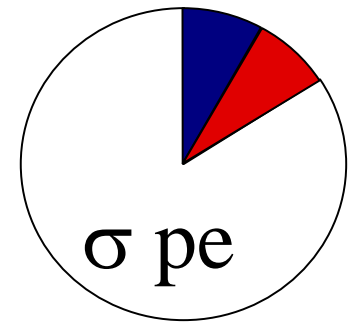
[1. Henderson Phil Trans RS B 1990

2. Owen *et al* PNAS 2006]



Radiation damage:

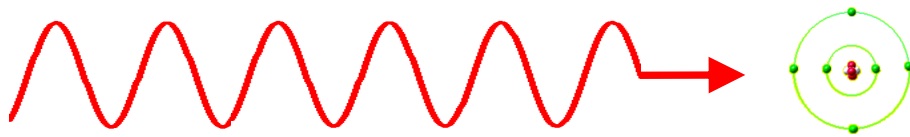
The Plan:



- What are the symptoms?
- **What is it?**
- Why do we care? Effect on MAD/SAD.
- How do we estimate the Dose?
- What do we know/would like to know?
- A new RD metric

A) Primary X-ray interaction processes with crystal and solvent.

Thomson (Rayleigh, coherent) scattering

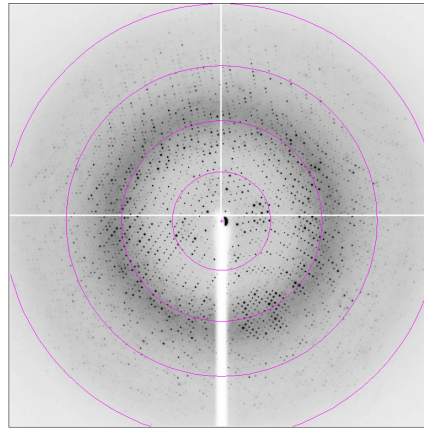


[8% at 1Å]

ELASTIC - no energy loss.

PHYSICS of the interaction of X-rays with crystals.

A) Diffraction



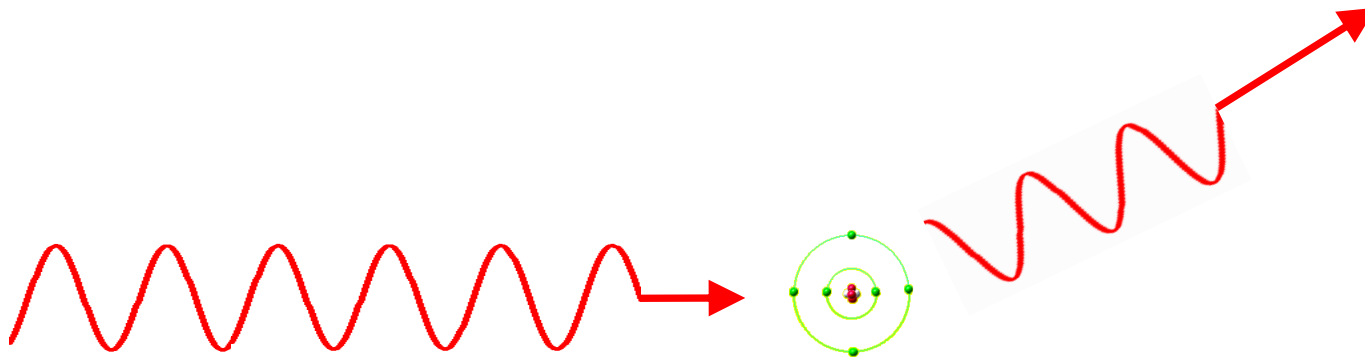
B) Absorption = Energy loss



N.B. $> 90\%$ of the beam does not interact at all,
but goes straight through.

Primary X-ray interaction processes with crystal and solvent.

Thomson (Rayleigh, coherent) scattering

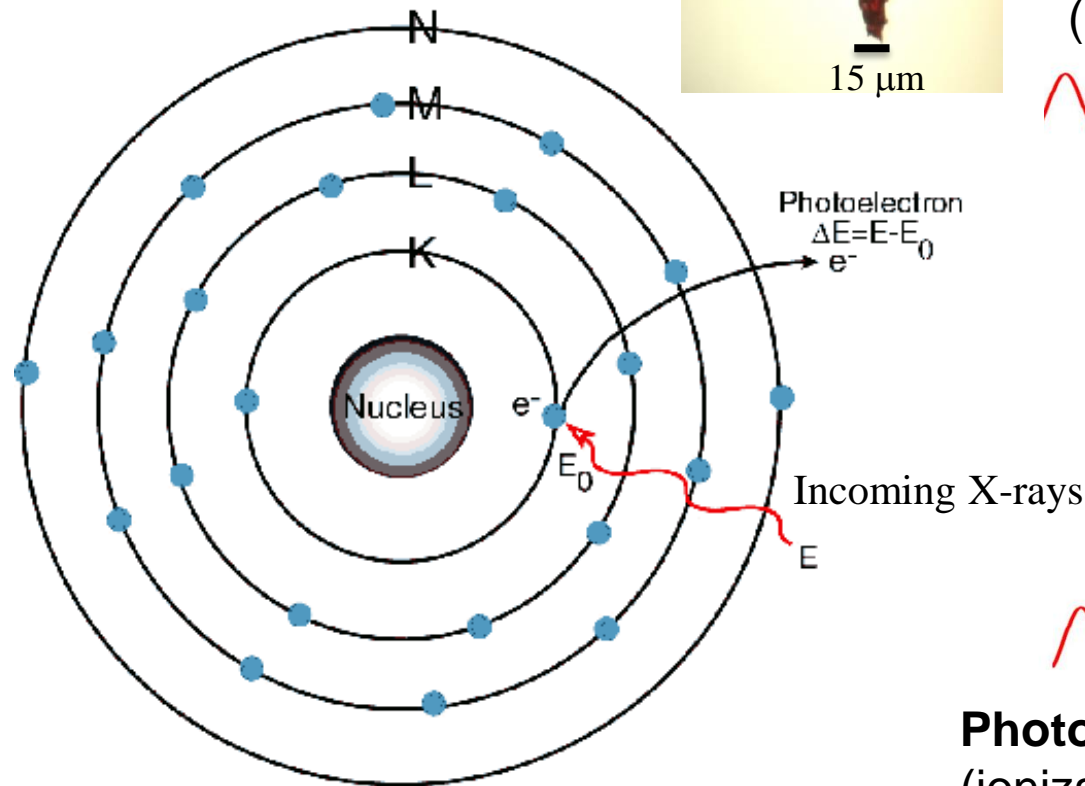
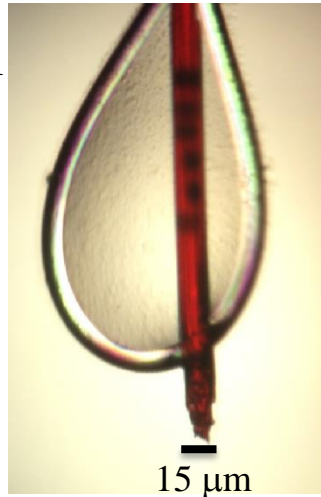


[8% at 1Å]

ELASTIC - no energy loss.

Interaction of X-rays with biological samples

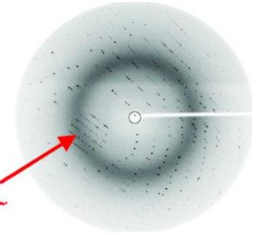
At $E_x = 12.4 \text{ keV}$, $100 \mu\text{m}$ protein crystal, only 2% of beam interacts.



Elastic scattering
(Thomson, coherent)

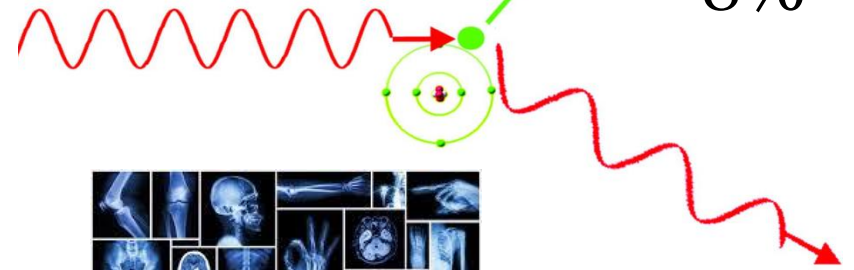


8%



**BUT IT IS THE
BIT WE WANT!!**

Compton scattering
(incoherent)



8%



Primary photoelectron

84%

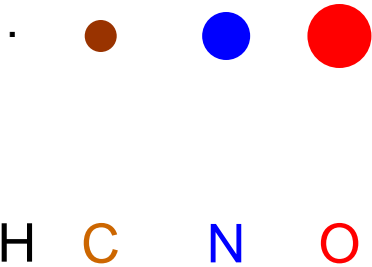
Photoelectric absorption
(ionization of up to 500 other atoms)



Photoelectric Cross Sections (barns/atom) at 13.1 keV

[1 barn= 10^{-28}m^2]

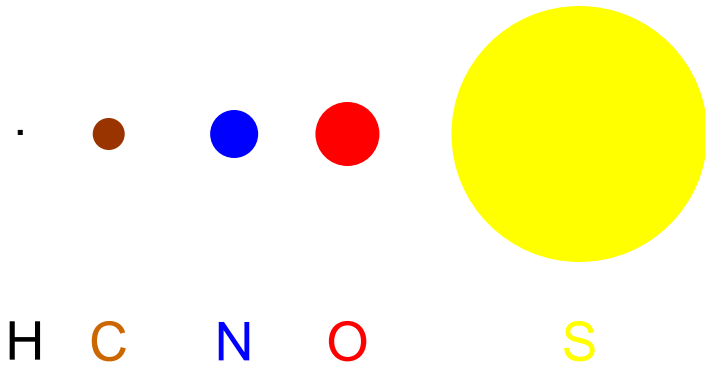
A few heavy atoms can
make a big difference.



Photoelectric Cross Sections (barns/atom) at 13.1 keV

[1 barn = 10^{-28}m^2]

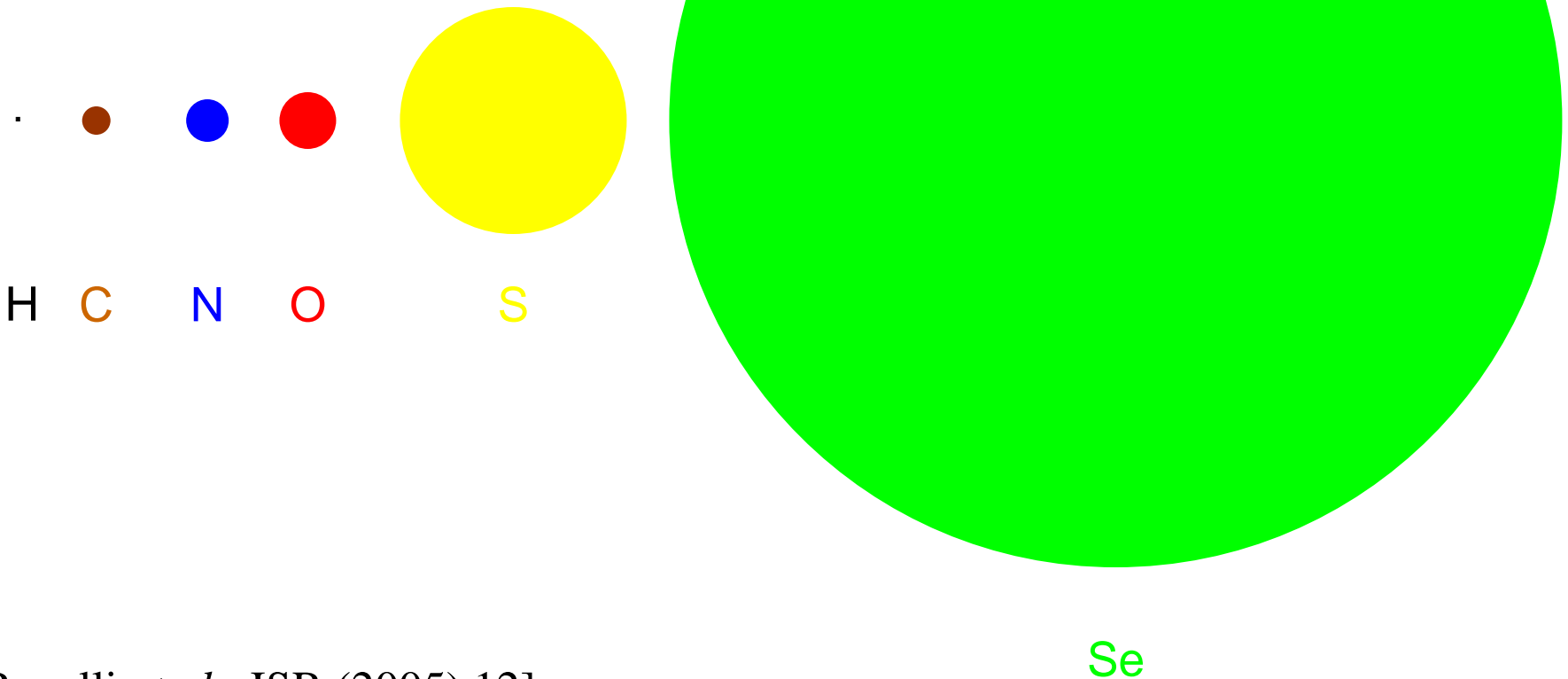
A few heavy atoms can
make a big difference.



Photoelectric Cross Sections (barns/atom) at 13.1 keV

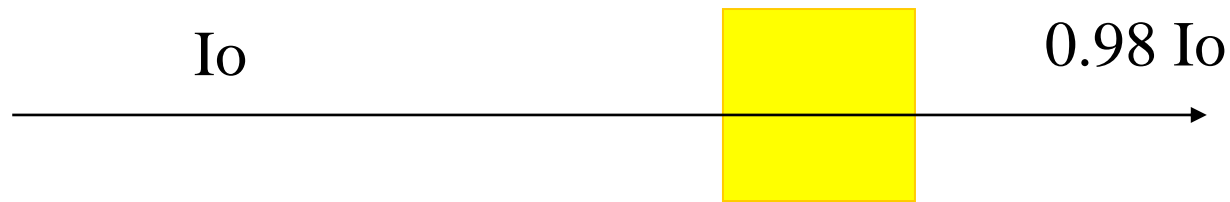
[1 barn=10⁻²⁸m²]

A few heavy atoms can
make a big difference.

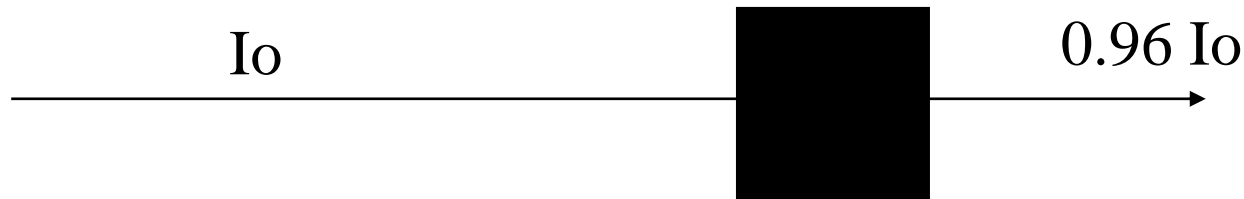


Beam absorption ($\lambda=1\text{\AA}$) by a protein crystal

Native HEWL 100 μm thick



Platinum derivatised (1/molecule)
HEWL 100 μm thick



N.B. INCIDENT FLUX is the SAME but the absorbed dose is DOUBLE

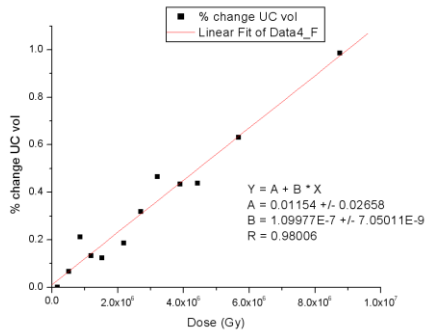
A few heavy atoms in the solvent can make a BIG difference to the absorption cross section and this the dose rate for the SAME flux.

e.g. Cacodylate buffer (arsenic, mass 75 cf selenium = 79)

BACK SOAKING to REMOVE

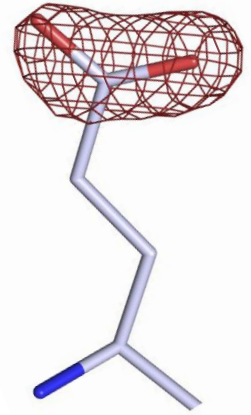
Non-specifically bound heavy atoms

e.g. a brominated DNA-protein complex will radiation damage much faster than a native crystal, and will de-brominate during data collection [Ennfar et al, Acta Cryst D (2002) 1263-1268].



Radiation damage:

The Plan:



- What are the symptoms?
- What is it?
- **Why do we care? Effect on MAD/SAD.**
- How do we estimate the Dose?
- What do we know/would like to know?

Why do we care?

A) We don't get all the data we need!

B) Effect on MAD/SAD phasing methods.

- Failure of structure determination

(Multi-wavelength anomalous dispersion MAD, SAD)

due to creeping non-isomorphism –

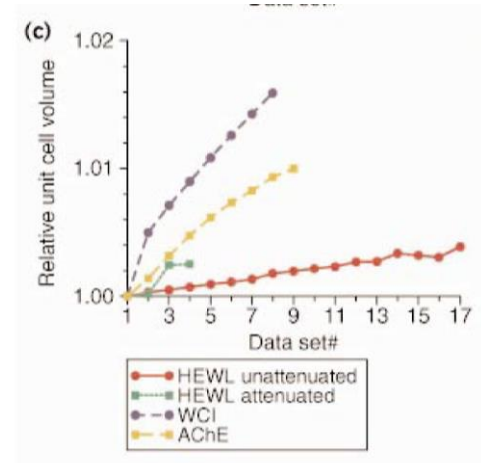
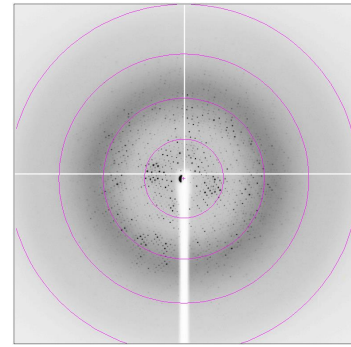
a) cell expansion and

b) movement of molecule in unit cell

c) structural changes DURING experiment.

i.e. **MAD/SAD phasing** signals (<5%) washed out completely

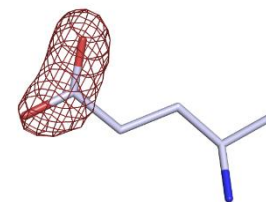
e.g. for a 0.5% change in all 3 unit cell dimensions of 100Å, reflection intensity changes by **15% at 3Å**



Why care?

C) Radiation damage can affect our biological results

- Metallo-proteins often photo-reduced during the experiment [e.g. PSII, Yano et al, PNAS (2005)]
- Decarboxylation of Glu is sometimes part of the protein mechanism, but is indistinguishable from radiation damage at the synchrotron
- X-ray induced structural changes were initially misleading in studies of intermediates
e.g. Bacteriorhodopsin: orange species

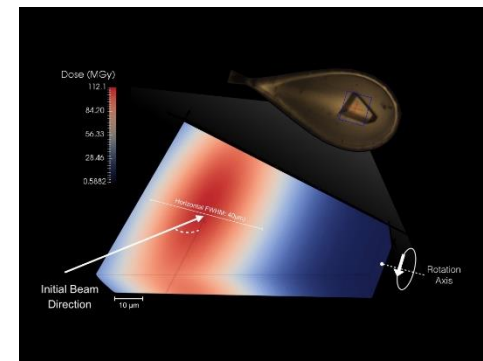


Takeda et al, Crystal structure of the M intermediate of bacteriorhodopsin...JMB (2004),
Wickstrand, et al., Bacteriorhodopsin: Would the real structural intermediates please stand up?,
Biochim. Biophys. Acta (2014)

Radiation damage:

The Plan:

- What are the symptoms?
- Why do we care? Effect on MAD/SAD.
- What is it?
- **How do we estimate the Dose?**
- What do we know/would like to know?
- A new RD metric



DOSE Postulate (Henderson 1990):

- There is a MAXIMUM dose for MX postulated as 20 MGy
(Energy absorbed/unit mass: Joules/kg = Gy)
which protein crystals can tolerate: depends only on the PHYSICS
of the situation.

Henderson: 20 MGy

Experimental: 43 MGy to $I_{0.5}$ – but don't go lower than $I_{0.7}$, 30 MGy*

- Crystal might not reach that limit due to chemical factors, but it is unlikely to last BEYOND the limit.
- Need to be able to calculate the DOSE conveniently: **RADDOSE-3D**

RADDOSE-3D

V1: Zeldin, Gerstel, Garman (2013) *J.Appl.Cryst*,

V2: Bury et al (2018) *Protein Science*

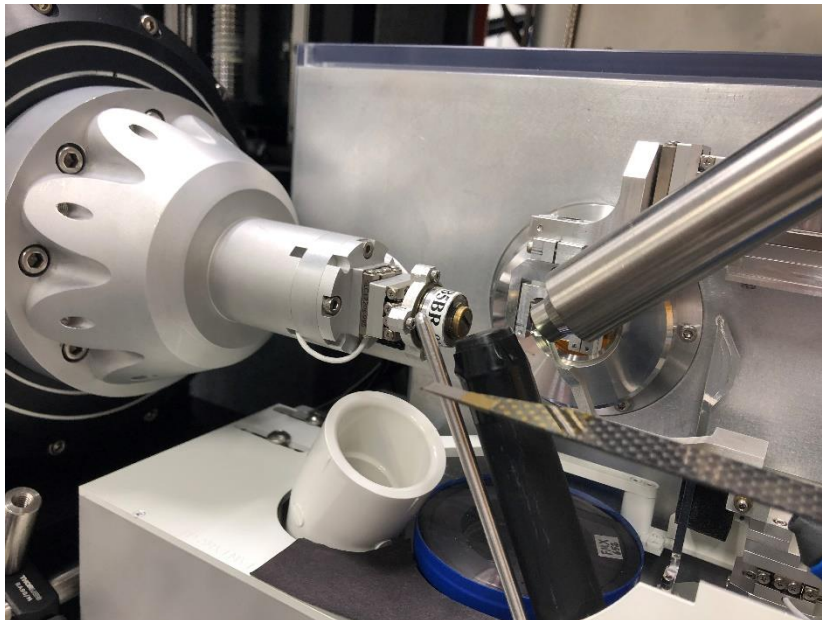
V3: Dickerson and Garman (2021) *Protein Science*

*Owen, Rudino-Pinera, Garman (2006), PNAS

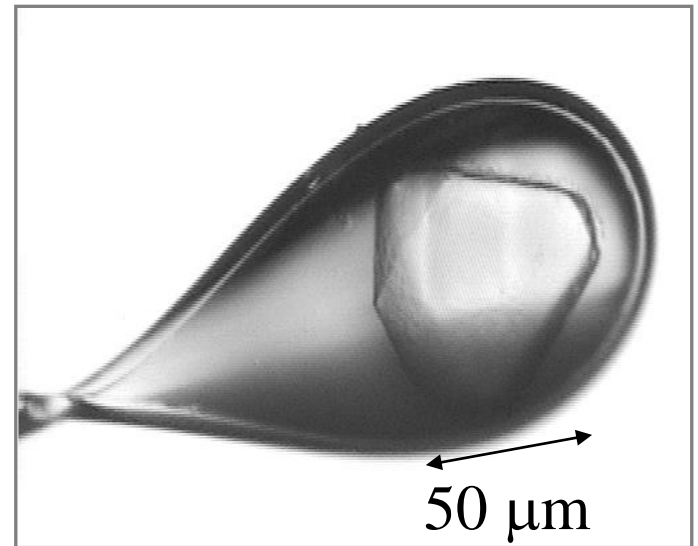
Make the dose calculation convenient for MX (include solvent contribution in mM and heavy atoms explicitly)

To find the energy deposited per unit mass in the crystal, need to characterise two things:

The beam

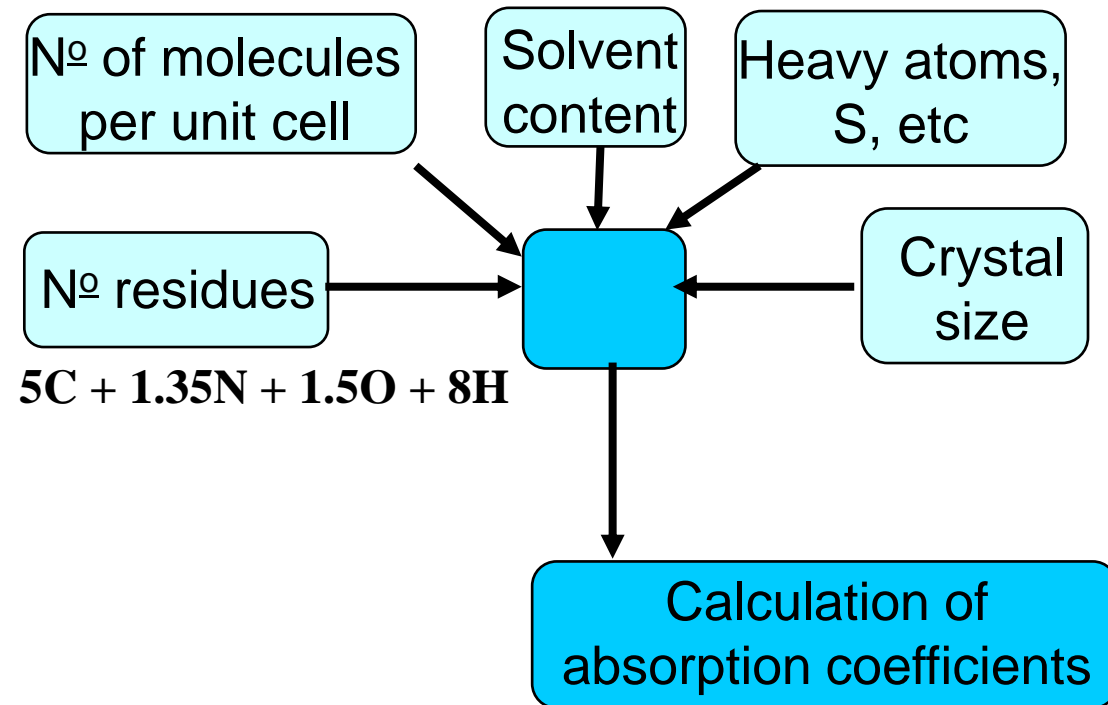


The crystal

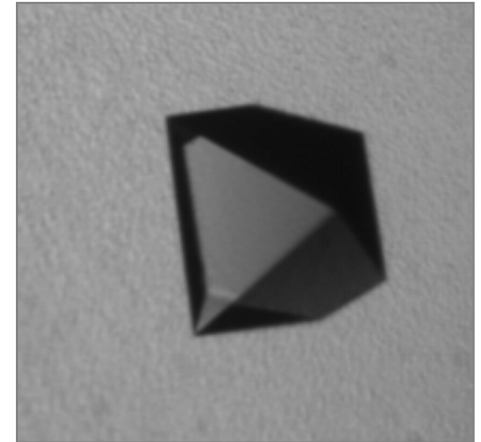


Calculating Dose (*RADDOSE-3D*)

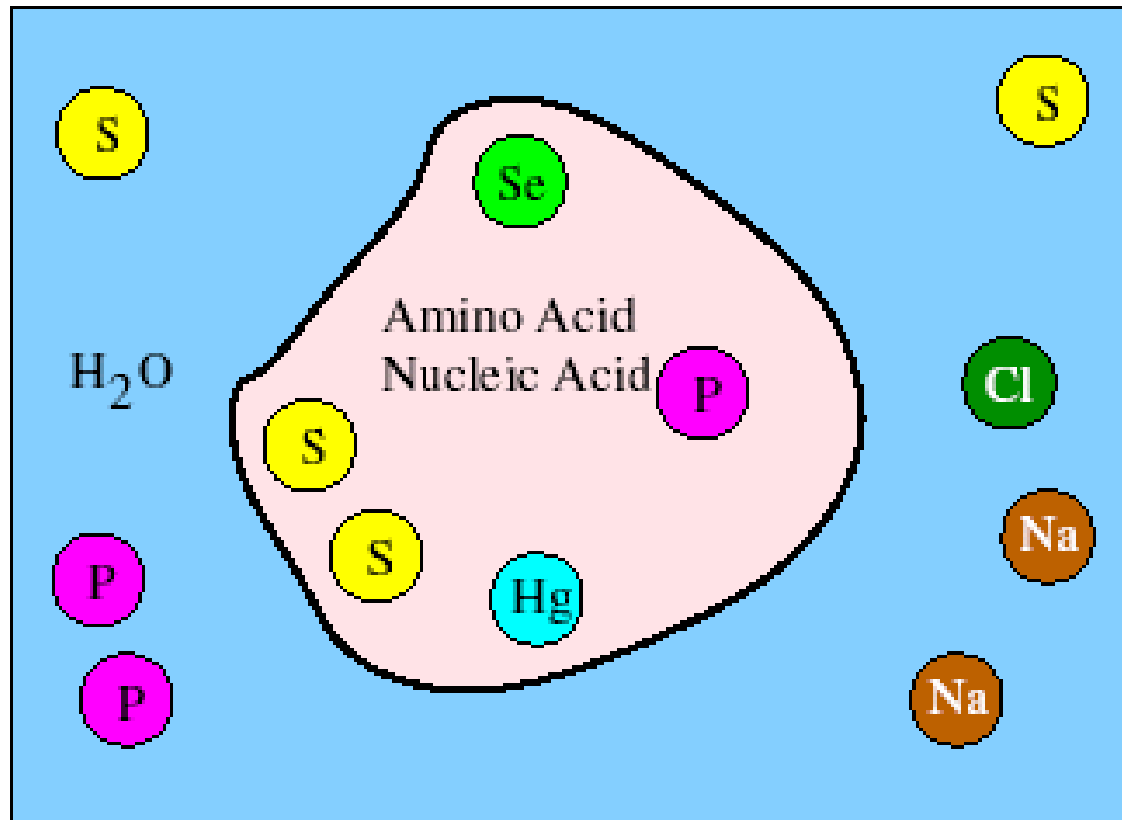
Crystal Characteristics



absorption coefficients
e.g. apoferritin: 0.406mm^{-1}
holoferritin: 1.133mm^{-1}



200 μm

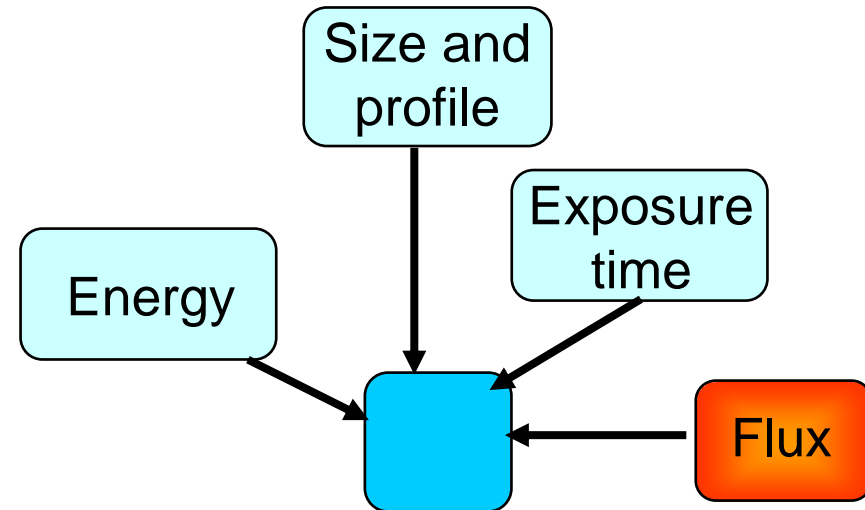
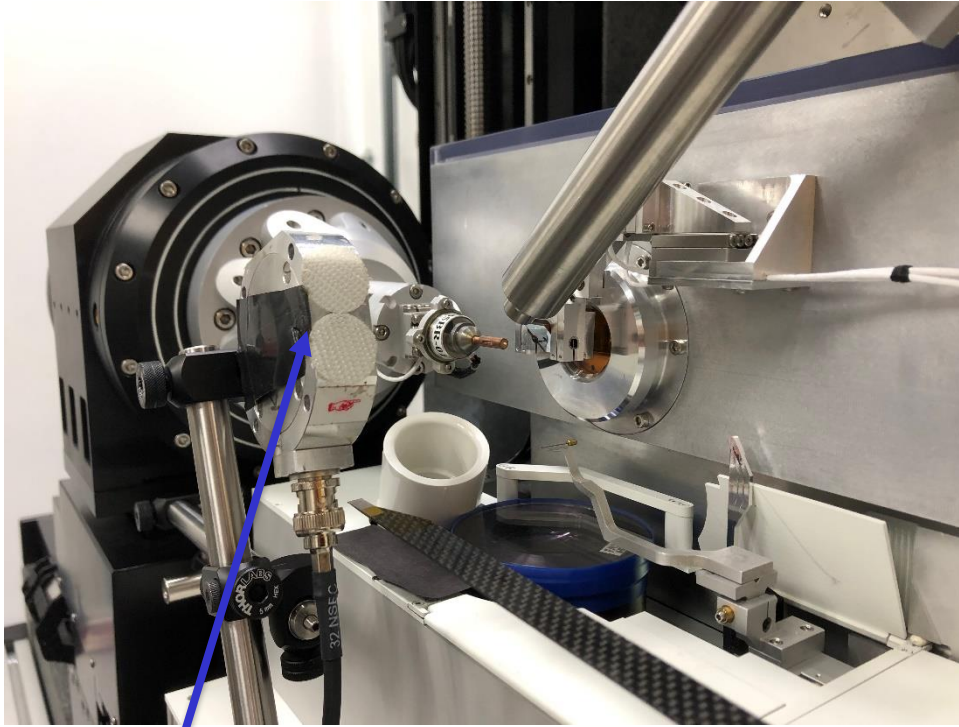


Number of Amino Acids

‘HA’ atoms per monomer, e.g. S, Se, Hg

Solvent - concentrations of components, e.g. Na⁺, Cl⁻

Calculating Dose (*RADDOSE-3D*) Beam Characteristics



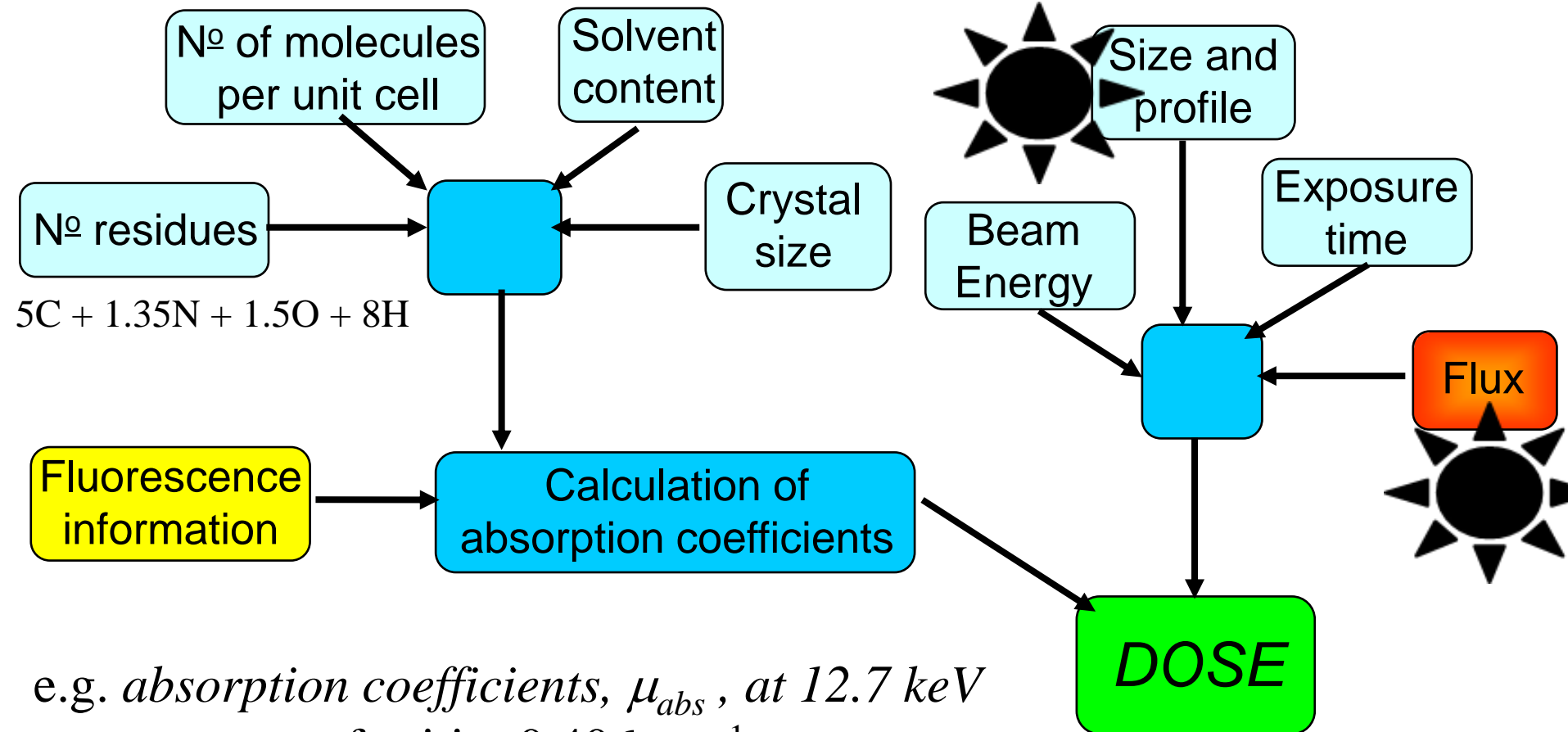
PIN diode to measure flux (ph/s)

Calculating Dose (*RADDOSE*)

[R-v1: Murray, Garman & Ravelli, JAC 2004
R-v2: Paithankar, Owen & Garman, JSR 2009,
R-v3: Paithankar & Garman, Acta D 2010]

Crystal Characteristics

Beam Characteristics



e.g. absorption coefficients, μ_{abs} , at 12.7 keV
apoferritin: 0.406mm^{-1}
holoferritin: 1.133mm^{-1}

MX experimental dose limit (@ 2Å) measurement: Ferritin

N.B. INCIDENT FLUX is the SAME but the absorbed dose is DOUBLE

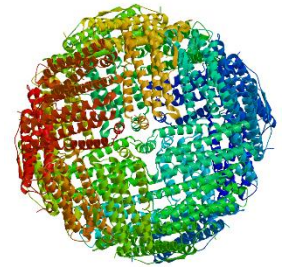
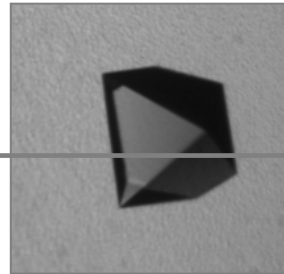
The heavy atom ($z \geq 16$) content of a crystal is not crystallographically defined, but we need it.

Apo ferritin

I_0

$0.98 I_0$

absorption coefficient
 0.406mm^{-1}



Holo ferritin

I_0

$0.96 I_0$

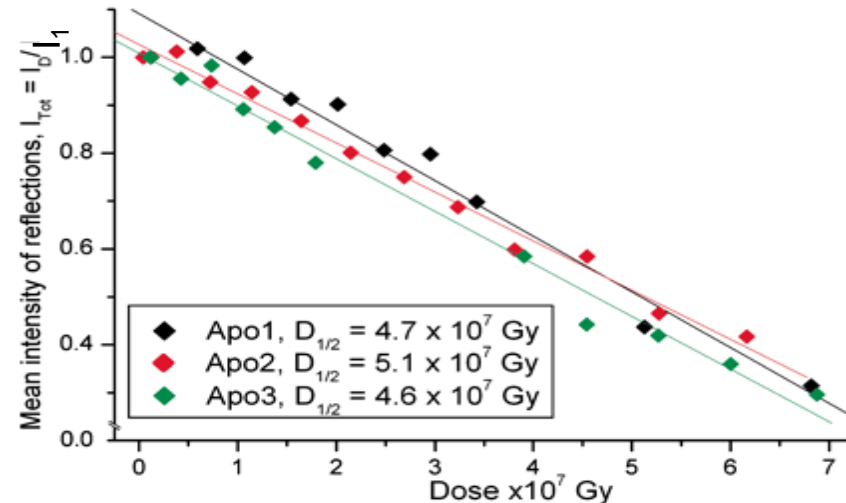
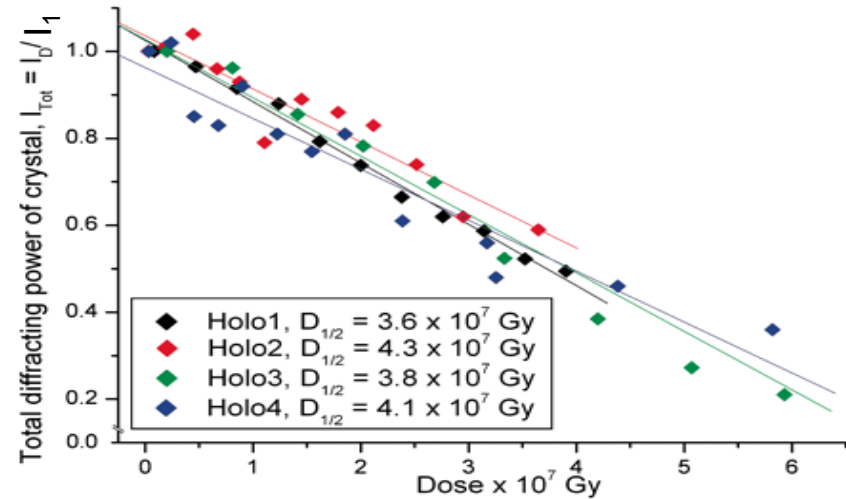
absorption coefficient
 1.133mm^{-1}



$\lambda = 1 \text{\AA}$
100 μm thick xtal

Dose Limit Quantification at 100 K

- Holoferitin & Apoferritin as model: absorption coefficients differ by factor of 2
- Intensity: ~linear dependence on dose
- $D_{1/2}$ is dose to $I_0/2$
- $D_{1/2} = 4.3 (\pm 0.4) \times 10^7$ Gy
= 43 MGy
- **Henderson limit, $D_{1/2} = 20$ MGy**
- Howells et al (2005): resolution dependent limit of 10 MGy/ Å

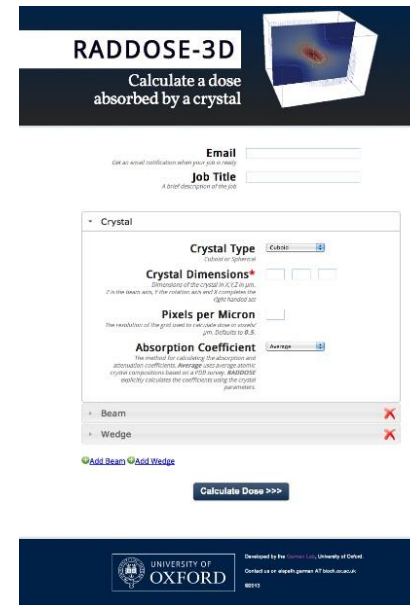
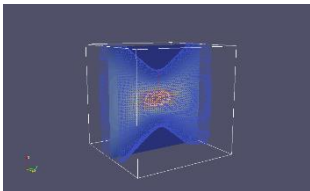


Suggested limit to retain biological 'fidelity'

$$D_{0.7} = 3.0 \times 10^7 \text{ Gy} = \mathbf{30 \text{ MGy}}$$

RADDOSE-3D

- TIME- and SPACE-resolved modeling of dose distributions in MX in Java, replaces RADDOSE:
- Full 3-D simulation of dose absorption by the crystal
- Can deal with multiple wedges of data and different energy beams (e.g. MAD)
- Models beam as Top-Hat or Gaussian **or can use measured experimental profiles**
- Engineered for easy extendibility:
can use any crystal shape
- On server: www.raddo.se (!!)
- On github.com/GarmanGroup/RADDOSE-3D



[Zeldin, Gerstel, Garman *JAC* (2013)

Bury, Brooks-Bartlett, Walsh, Garman, *Protein Science* (2018)]

RADDOSE-3D New GUI

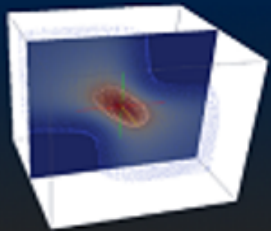
Download from: https://github.com/jdickerson95/qt_RADDDOSE-3D/releases

Versions for a PC (Windows_release.zip) and for Linux (Linux_release.zip).

RADDOSE-3D

File

RADDOSE-3D
Calculate the dose absorbed by a crystal



Subprogram: Standard RADDOSE-3D

Crystal Beam Wedge

Start Angle: 0 End Angle: 90

Exposure time: 50

Angular Resolution: 2

Starting offset X: 0 Y: 0 Z: 0

Translation per degree X: 0 Y: 0 Z: 0

Rotation offset: 0

Manually edit input

Run



Josh
Dickerson

RADDOSE-3D GUI

- Demonstration

RADDOSE-3D (XFEL etc)

TEST our new GUI (Josh Dickerson)!!

To run RADDOSE-3D (or XFEL etc)

Step 1: Download and unzip the RADDOSE-3D GUI from:

https://github.com/jdickerson95/qt_RADDDOSE-3D/releases

There are versions for a PC (Windows_release.zip) and for Linux (Linux_release.zip).

If you have a MAC, there is no new GUI yet, but one will come.

To run the GUI you need to have Java installed which you can get free at

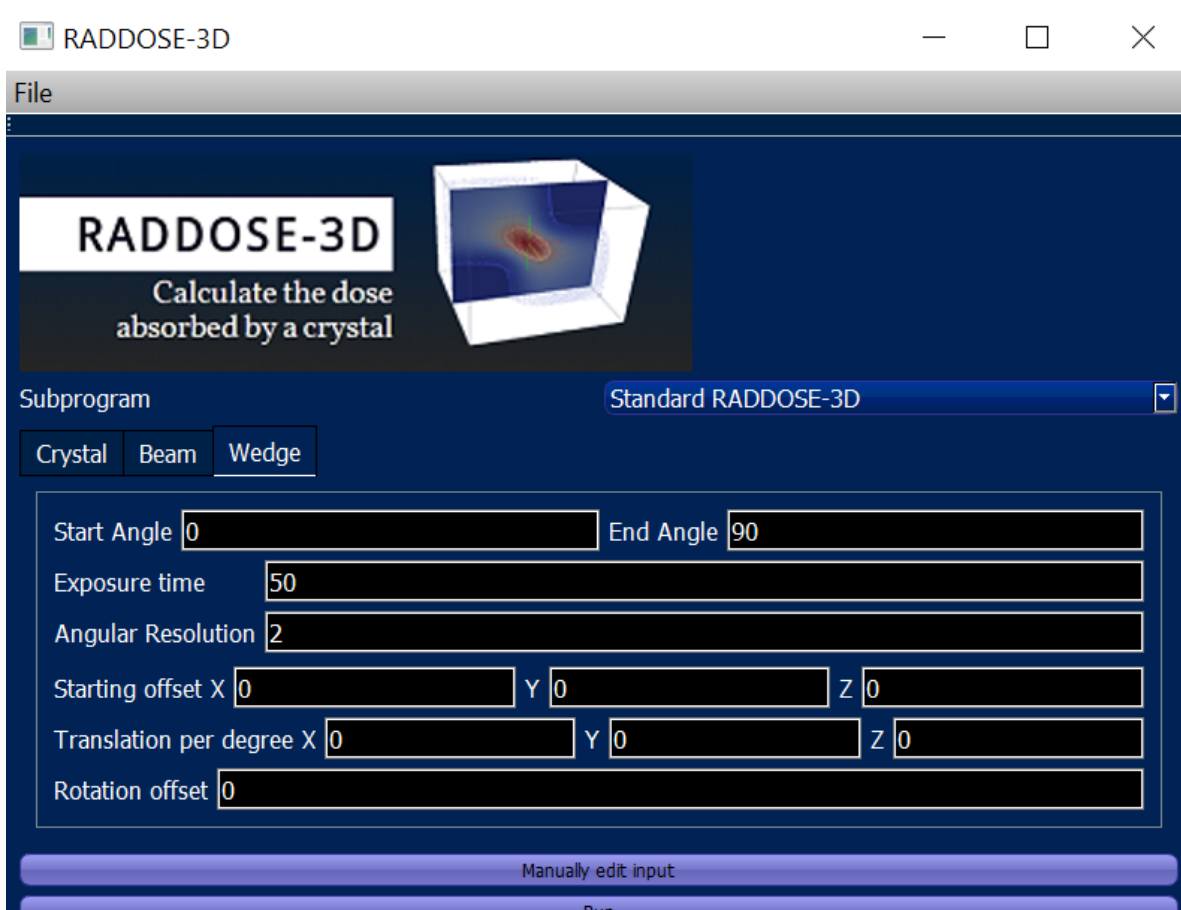
https://www.java.com/download/ie_manual.jsp

Also, if you have R (<https://www.r-project.org/>) installed, from the RADDOSE-XFEL output you will be able to produce 3D representations of the dose distribution in your sample.

Step 2: Unzip the file you have downloaded. On Windows machines I recommend the best place to install the executable (e.g. Documents versus Program Files) is in Documents as I tend to avoid putting programs in Program Files unless I am using an installer, to avoid the need for admin privileges and to keep things self-contained.

Step 3: Find the file RD3D_GUI.EXE and if on a PC click on it. For Linux run it however you usually run executable files. The GUI should open, and you can enter input on 3 tabs: crystal, beam and wedge.

Under ‘Standard RADDOSE-3D, select ‘XFEL’



Wedge 1:

Collecting data for a total of 50.0s from $\phi = 0.0$ to 90.0 deg.

Crystal coefficients calculated with RADDOSE-3D.

Photoelectric Coefficient: $3.21\text{e-}04$ / μm .

Inelastic Coefficient: $1.86\text{e-}05$ / μm .

Elastic Coefficient: $2.10\text{e-}05$ / μm .

Attenuation Coefficient: $3.61\text{e-}04$ / μm .

Density: 1.14 g/ml.

Average Diffraction Weighted Dose : 6.238509 MGy

Last Diffraction Weighted Dose : 10.508146 MGy

Elastic Yield : $2.27\text{e+}11$ photons

Diffraction Efficiency (Elastic Yield/DWD): $3.64\text{e+}10$ photons/MGy

Average Dose (Whole Crystal) : 5.921152 MGy

Average Dose (Exposed Region) : 5.921152 MGy

Max Dose : 37.291488 MGy

Average Dose (95.0 % of total absorbed energy threshold (2.85 MGy)): 9.501849 MGy

Dose Contrast (Max/Threshold Av.) : 3.92

Used Volume : 100.0%

Absorbed Energy (this Wedge) : $6.88\text{e-}03$ J.

Dose Inefficiency (Max Dose/mJ Absorbed) : 5.4 1/g

Dose Inefficiency PE (Max Dose/mJ Deposited): 5.5 1/g

Final Dose Histogram:

Bin 1, 0.0 to 0.1 MGy: 16.6%

Bin 2, 0.1 to 3.4 MGy: 26.1%

Bin 3, 3.4 to 6.7 MGy: 21.0%

Bin 4, 6.7 to 10.1 MGy: 16.0%

Bin 5, 10.1 to 13.4 MGy: 9.4%

Bin 6, 13.4 to 16.7 MGy: 5.0%

Bin 7, 16.7 to 20.0 MGy: 1.8%

Bin 8, 20.0 to 23.4 MGy: 1.6%

Bin 9, 23.4 to 26.7 MGy: 1.0%

Bin 10, 26.7 to 30.0 MGy: 1.2%

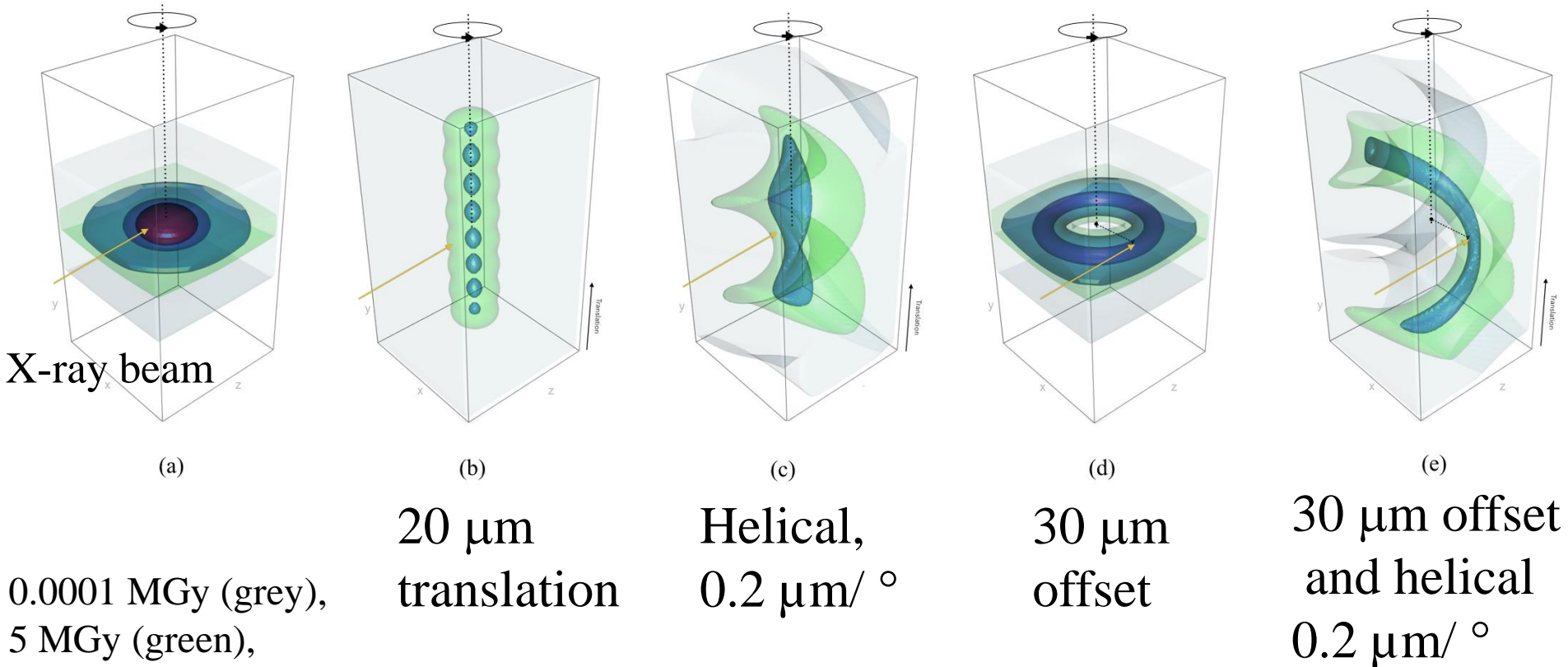
Bin 11, 30.0 MGy upwards: 0.3%

RADDOSE-3D terminated after 0.9 seconds

Plot dose histogram

Back

Dose distribution vs exposure strategy with RADDOSE-3D



0.0001 MGy (grey),
5 MGy (green),
10 MGy (light blue),
20 MGy (dark blue),
30 MGy (red),
360° rotation

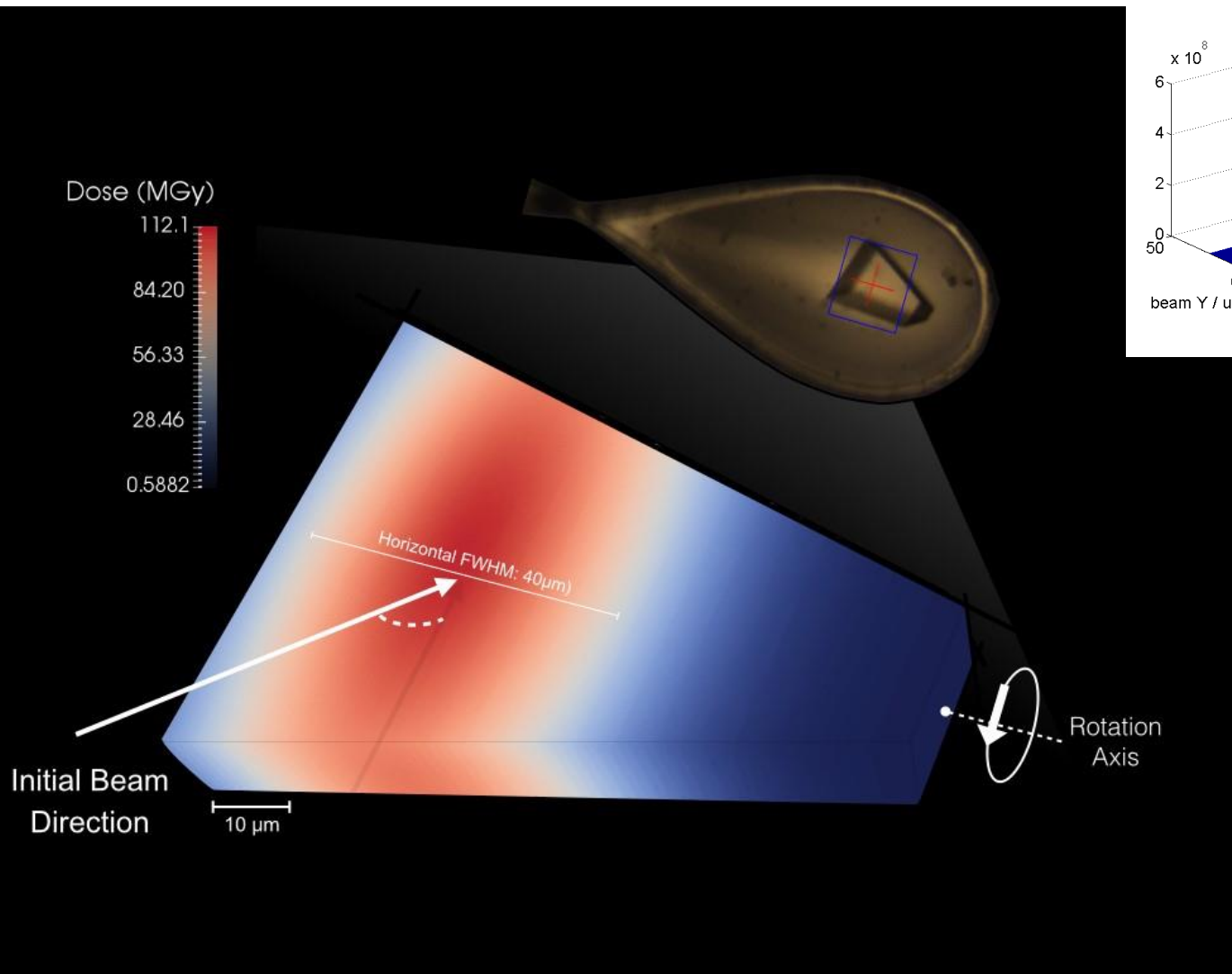
100 μm , 200 μm , 100 μm .

Gaussian beam (FWHM: 20 μm x 20 μm), 12.4 keV, 5×10^{11} ph/s,
1 mm x 1 mm rectangular collimation: full crystal bathed in beam

Helical strategy improvement:
Flot *et al* (2010) *JSR* **17**, 107

Zeldin *et al.* *JSR* (2013), Bury *et al.* *Protein Science* (2018)

Gaussian Beam Profile

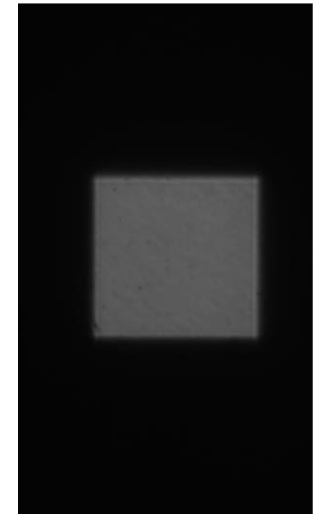
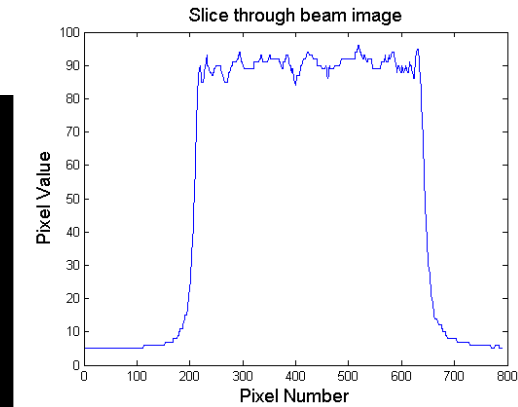
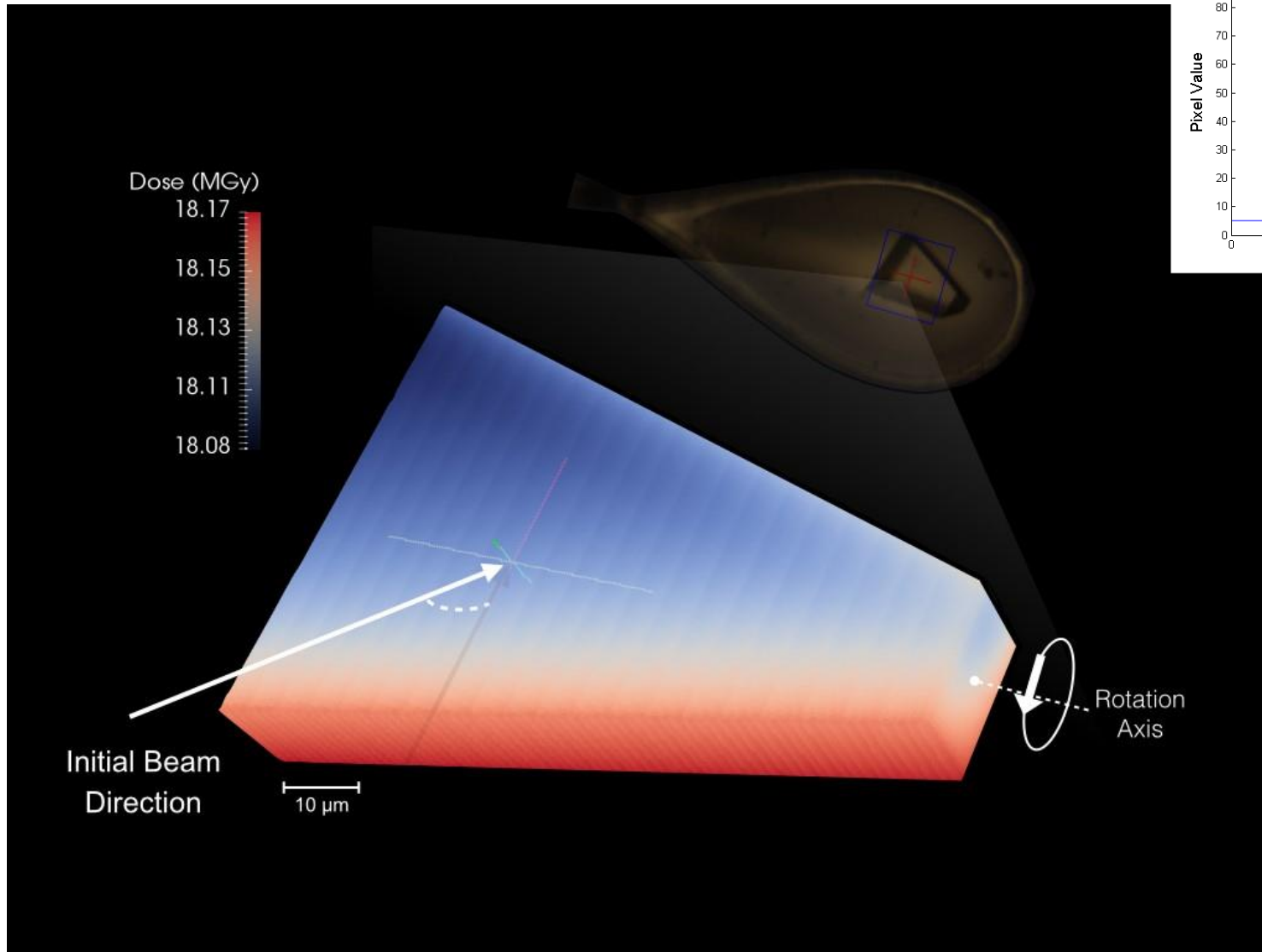


Differential irradiation may lead to differential damage:
get data which merge poorly and are a population of substates

13.2 keV, 60(v)×40μm²(h) FWHM, 100(h)×160μm²(v) coll., 5×10¹¹ ph/s

Garman & Weik *Meth. Mol. Biol* 1607 (2017)

Top-Hat profile

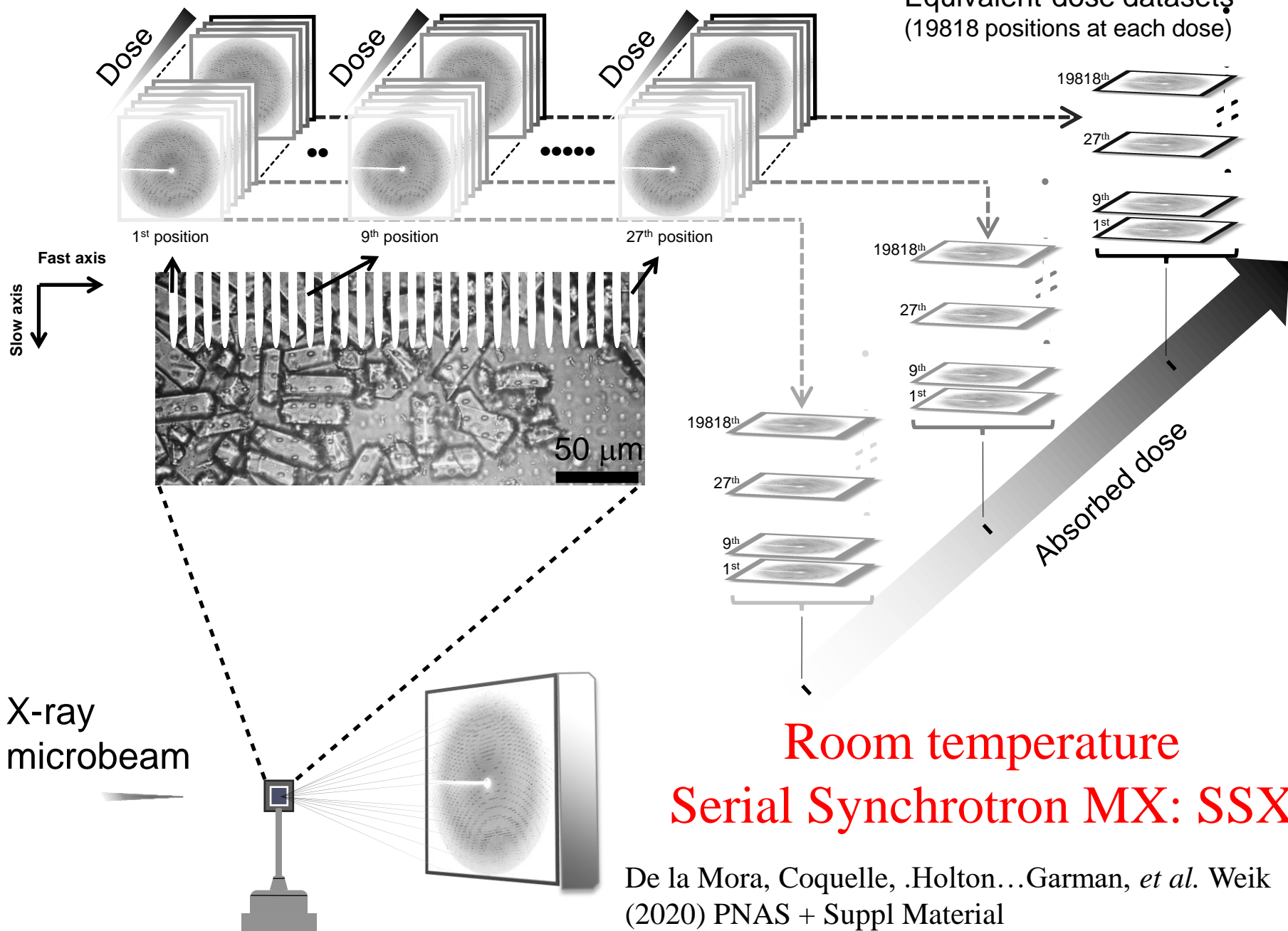


Imaged beam
PETRA III,
Bourenkov,
Schneider

13.2 keV, 100(h) x $160\ \mu\text{m}^2$ (v) coll., $5e11\ \text{ph/s}$

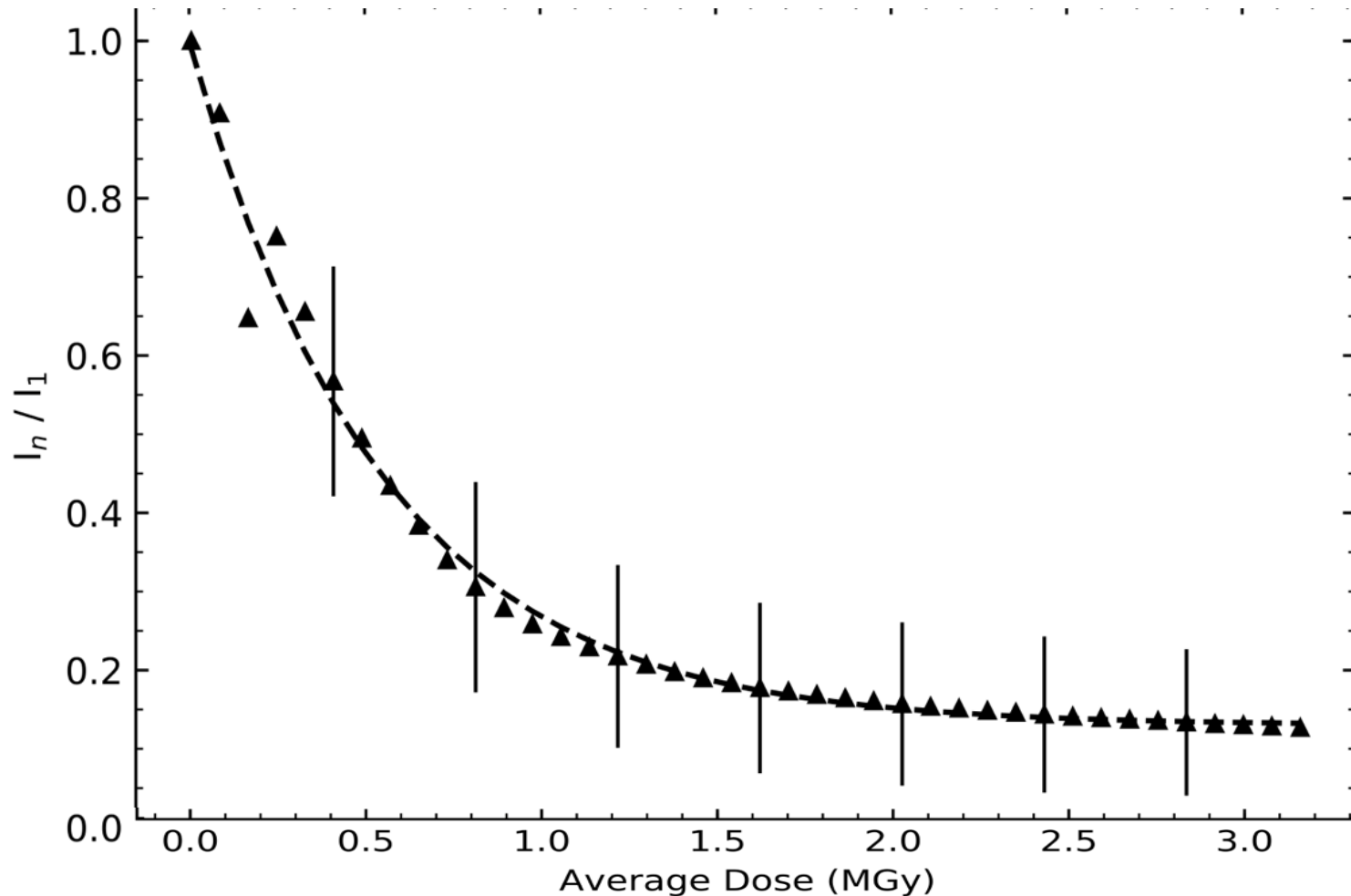
40 consecutive frames per position

Equivalent-dose datasets
(19818 positions at each dose)



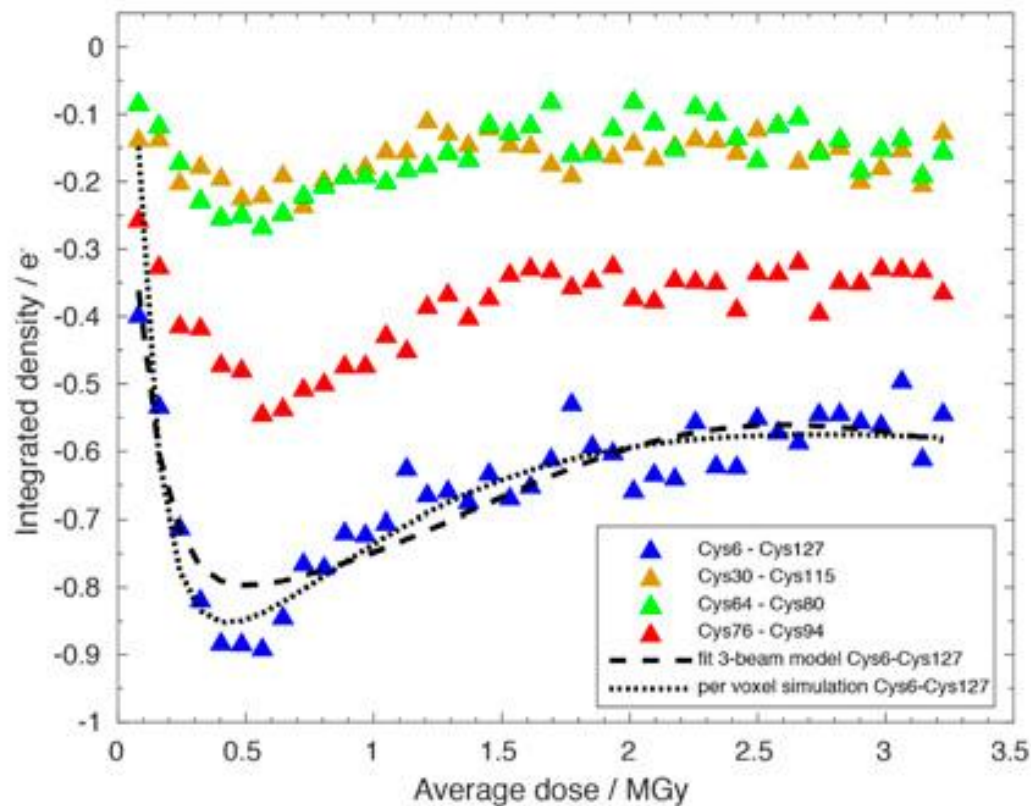
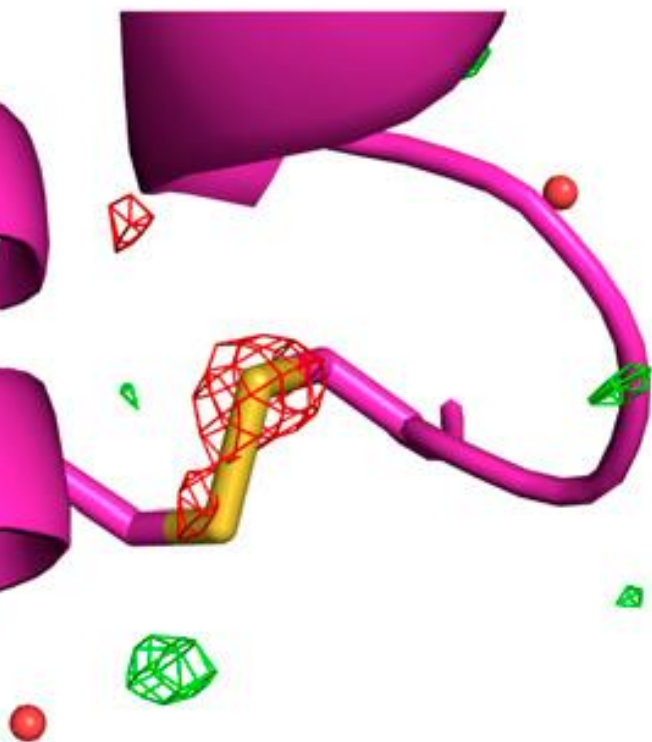
De la Mora, Coquelle, .Holton...Garman, *et al.* Weik
(2020) PNAS + Suppl Material

RadDam signatures in reciprocal space (RT MX)

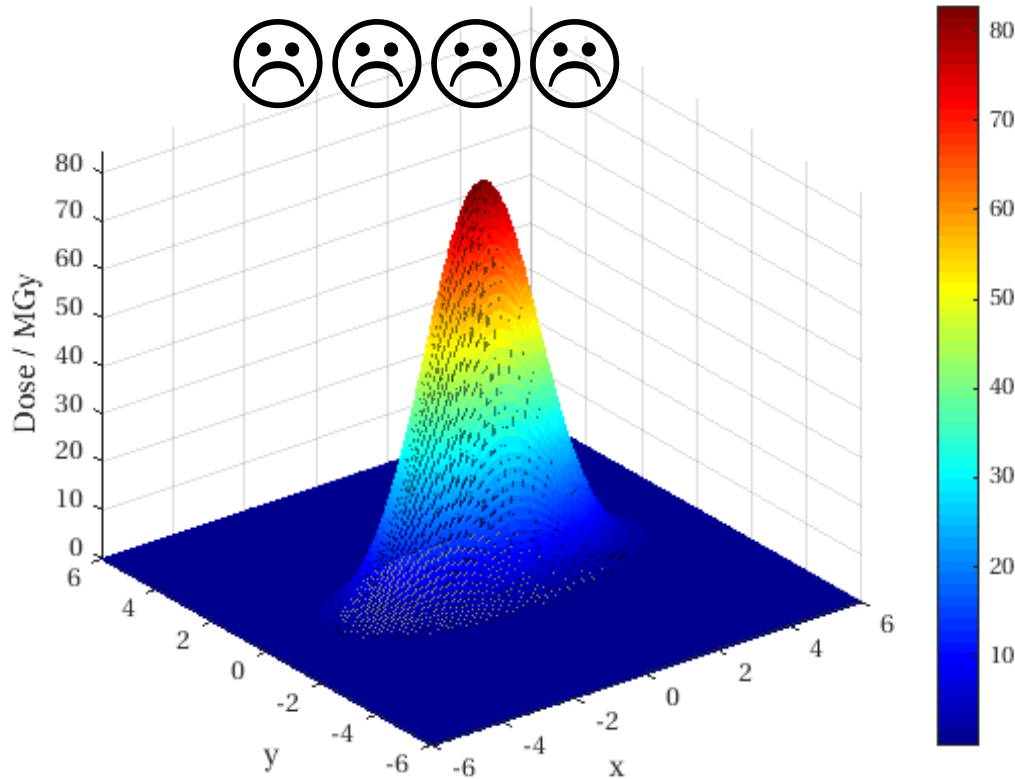


Radiation damage and dose limits in serial synchrotron crystallography at cryo- and room temperatures. De la Mora, Coquelle *et al.* (2020) PNAS

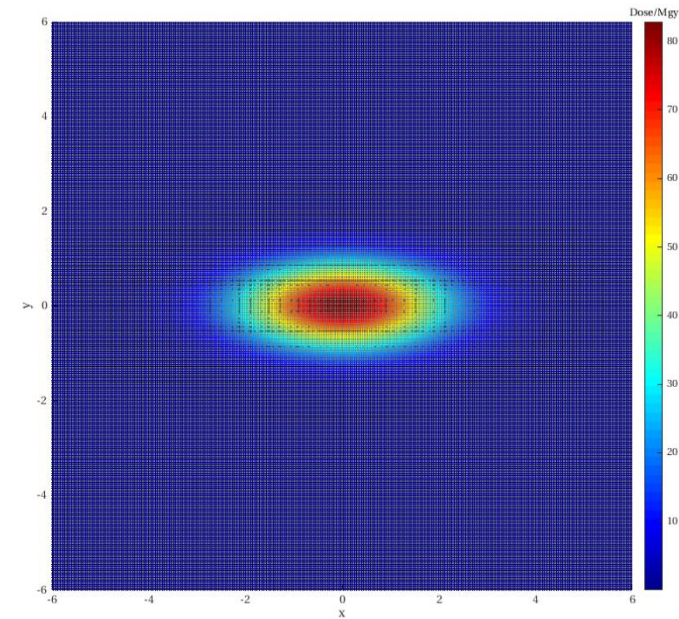
RT high-dose rate ‘recovered’ electron density with dose



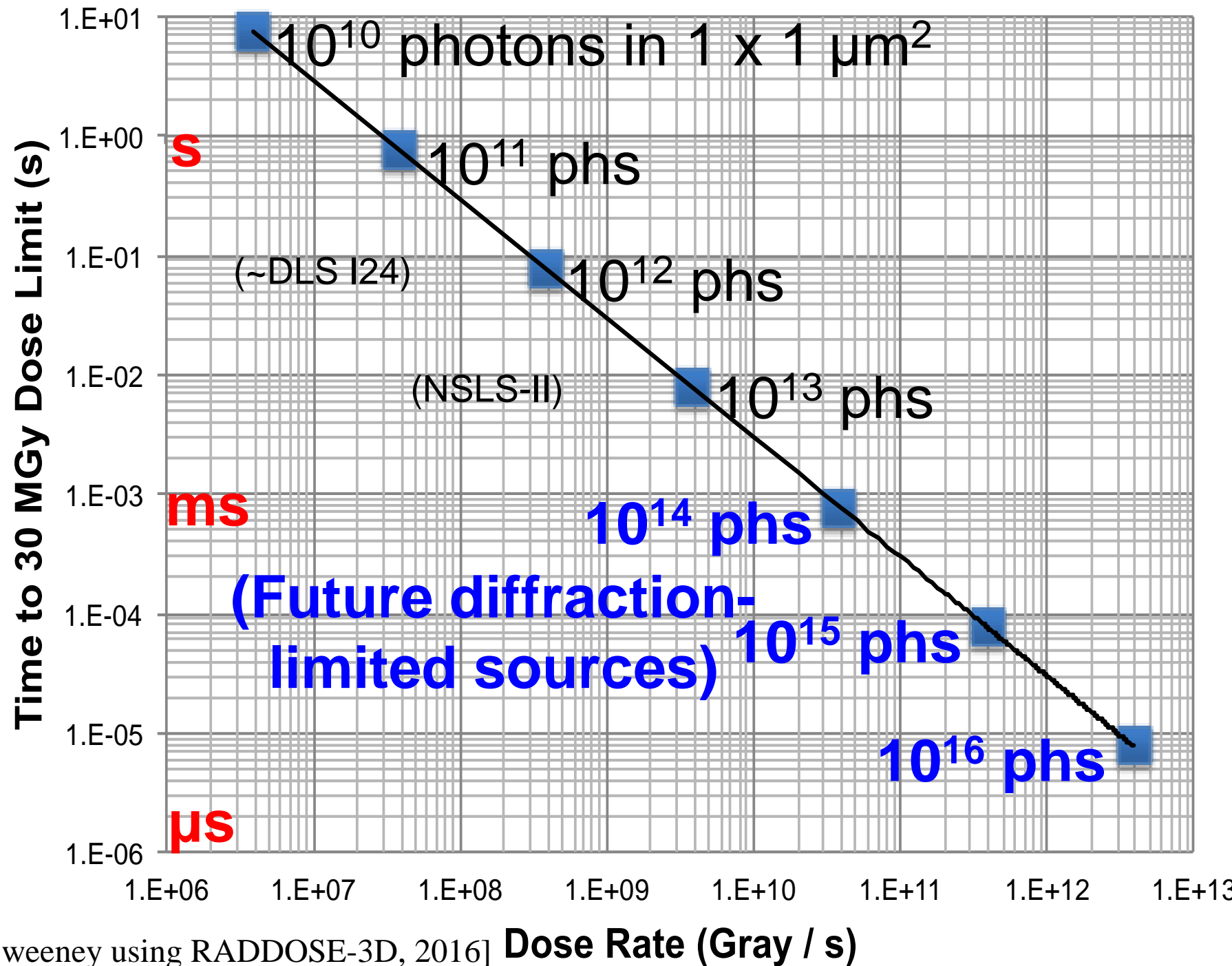
Beam shapes



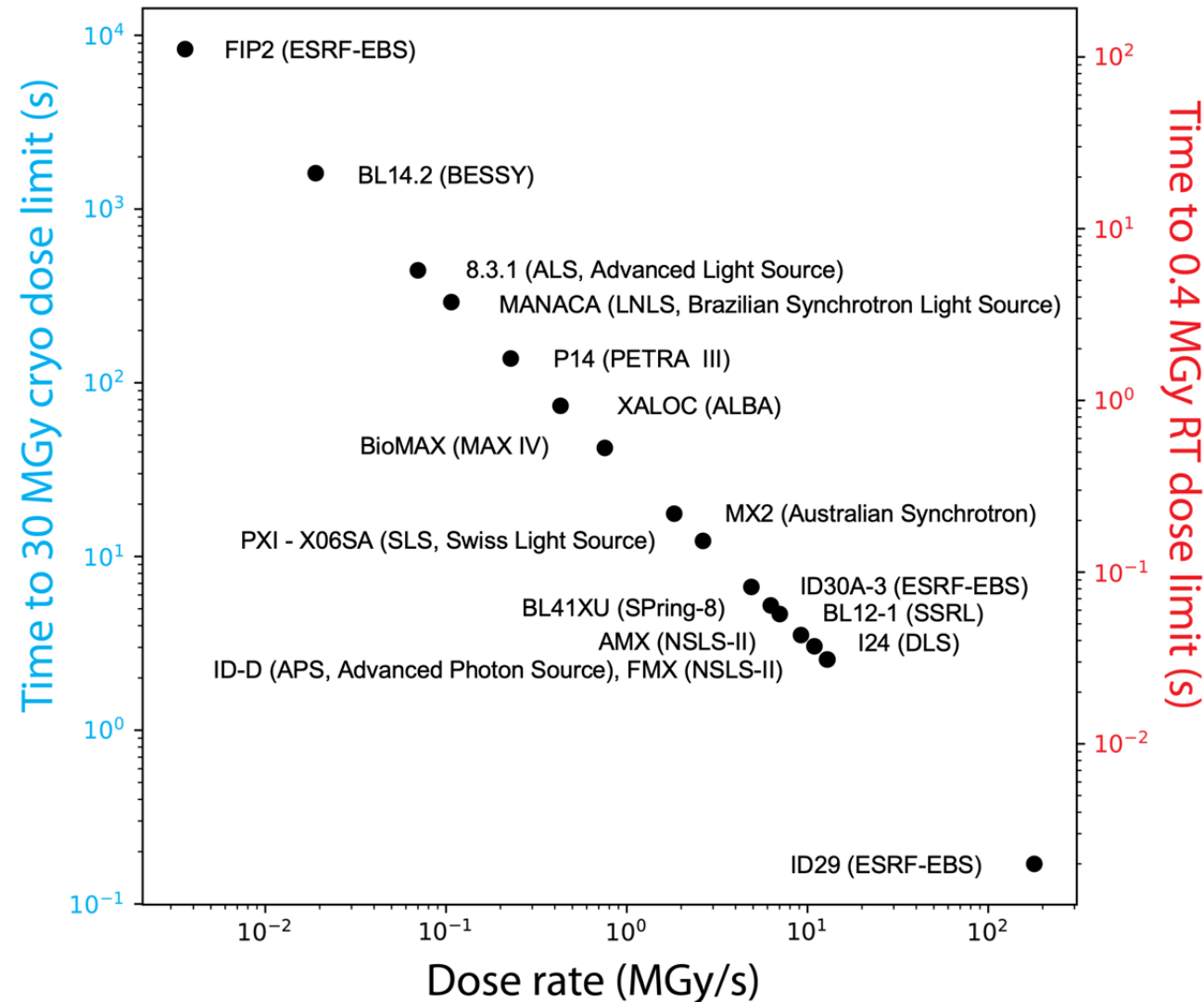
Gaussian/Lorentzian
Pseudo-Voigt



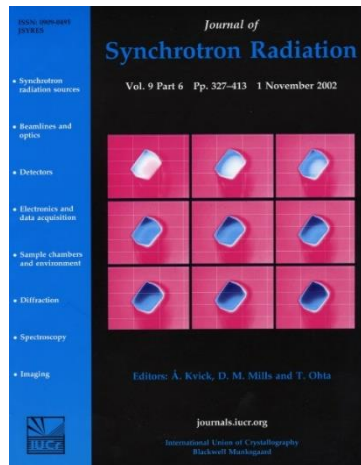
Partition beam
into 3 contributions:
Hot beam
Cold beam
Pedestal beam



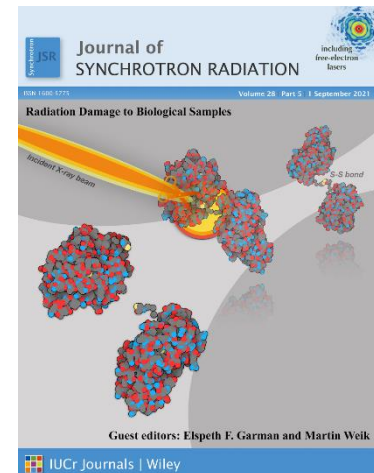
Time to dose limits at current synchrotrons



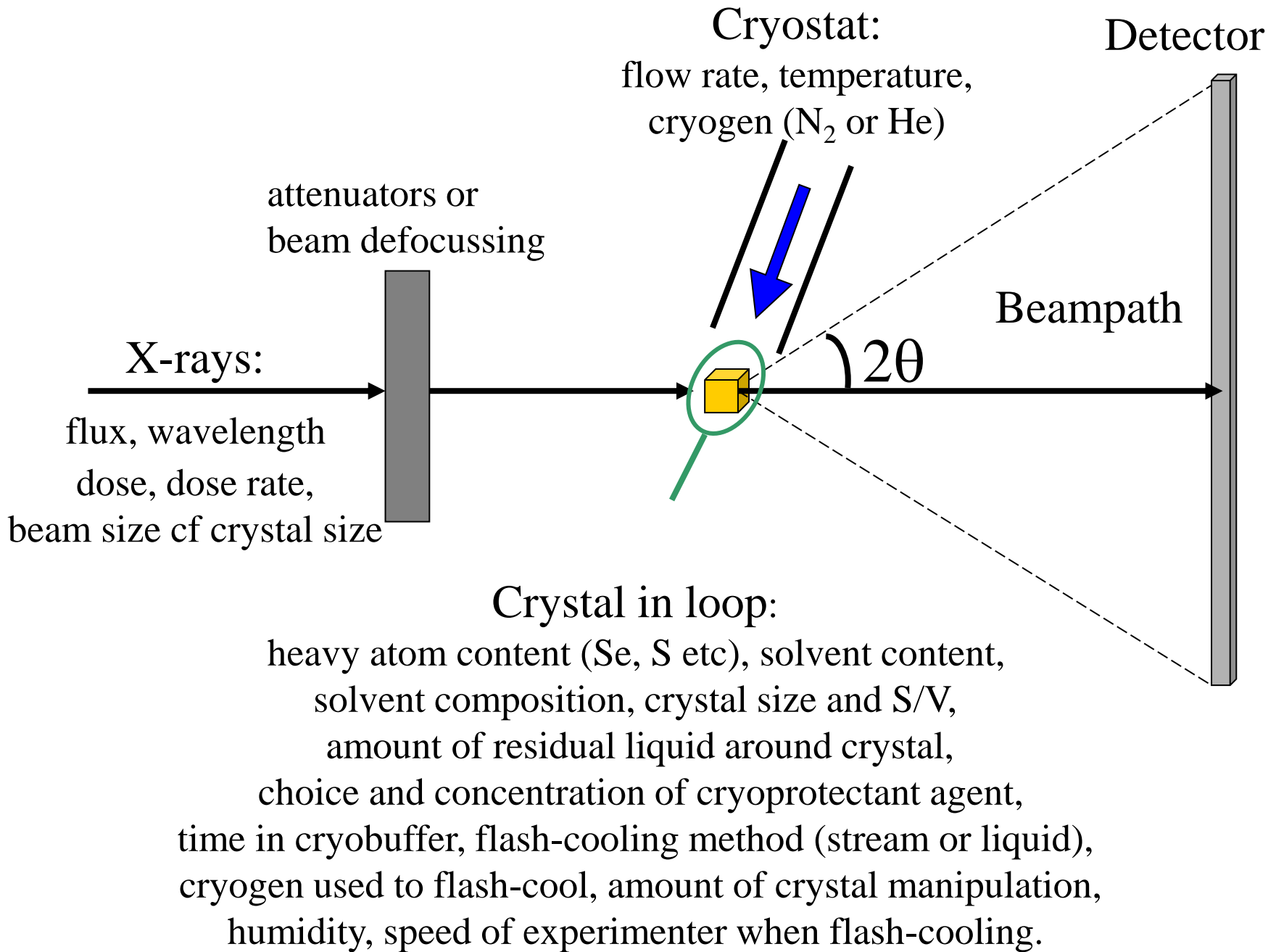
Average diffraction weighted doses (DWD) were calculated for a $(50 \mu\text{m})^3$ lysozyme crystal (grown in 100 mM NaAc and 1M NaCl, solvent fraction 38%) rotated 360° in the flux within the FWHMs of the various beams.



Radiation damage: The Plan:



- What are the symptoms?
- Why do we care? Effect on MAD/SAD.
- What is it?
- How do we calculate the Dose?
- **What do we know/would like to know?**
- A new RD metric



PROBLEM: how do we know that we are making any difference?

- In order to investigate the effects of various parameters on the radiation damage process, we need a robust radiation damage METRIC which is preferably ON-LINE during the diffraction experiment.

No unanimous metric currently/ results from different metrics do not agree.

- Structural changes occur before degradation of diffraction quality is obvious.

Work so far / ongoing:

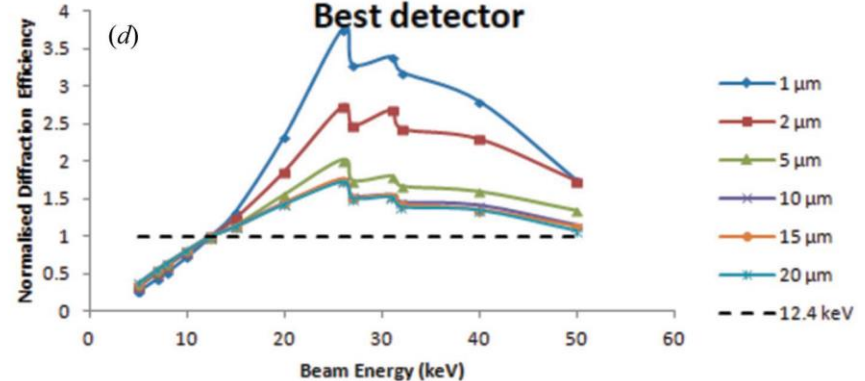
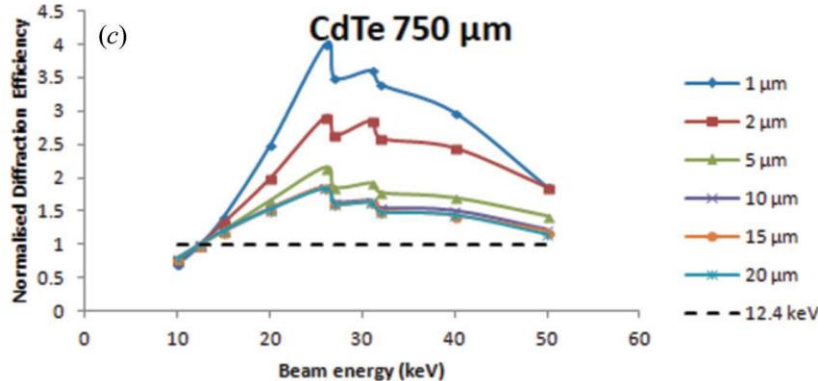
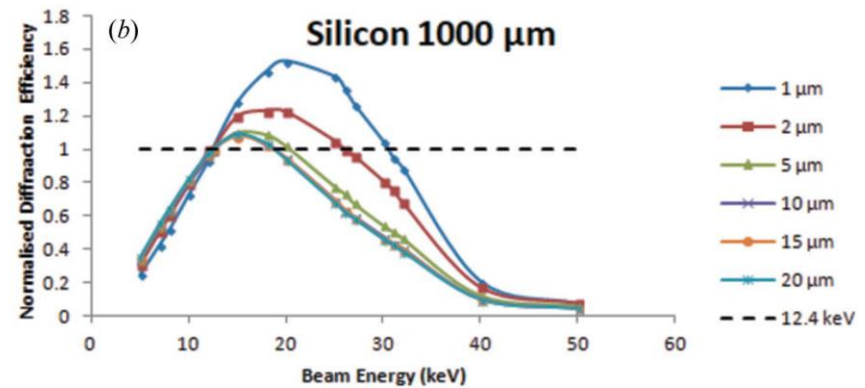
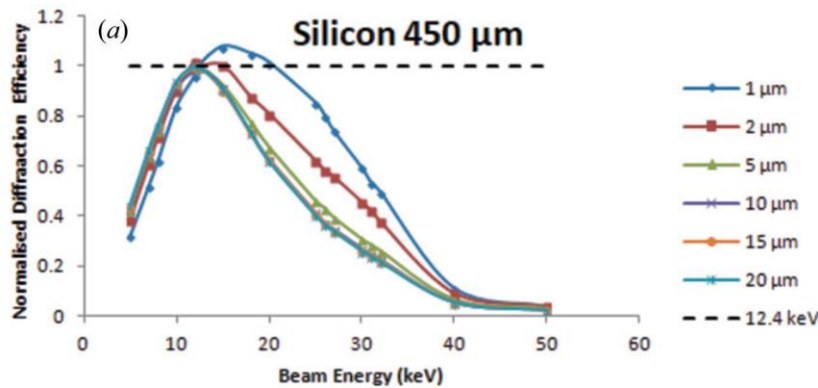
- Lower the cryogen temperature? 40K? 50K? 16K? 140K (no!). **Less than a factor of 2 improvement.**
- Lower the wavelength/increase X-ray energy)? Lots of anecdote + now some systematic results: **no effect on damage rate except for small crystals at higher energies (>20 keV) where photoelectron can escape (needs another talk!**

Varying results. Shimizu et al (2007) 14, 4-10, Liebschner et al, Acta Cryst D71(2015), Storm et al (2021) IUCr 8

- Unit cell expansion as a metric? **No!**
- Change/ regulate the dose/dose rate regime? **No, not at current!**
- Effect on MAD/SAD? Order of data collection?
- Minimum crystal size? Several papers (see Holton 2009)
- Beam heating. **Not a big factor at current flux densities at cryotemperatures (Owen et al JSR 2019).**

Optimum incident energy?

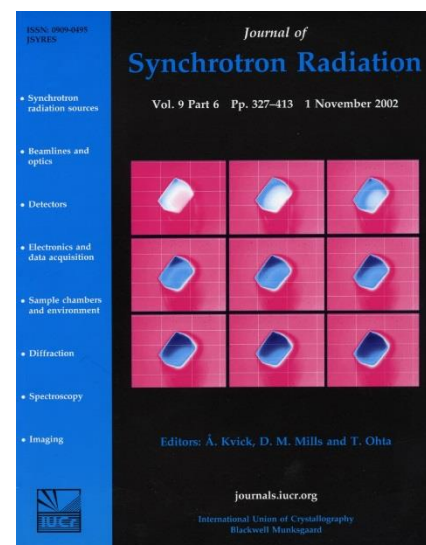
Simulations: CdTe detector, 26 keV,
diffraction efficiency = diffraction/dose



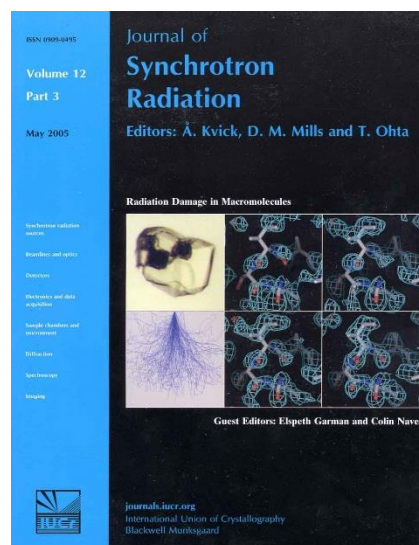
Work so far / ongoing:

- X-ray absorption – **important parameters defined.**
- Remove oxygen? **Nothing yet.**
- Radiation damage Induced Phasing (RIP)
- Software developments – **big progress.**
- Add radical scavengers: **results disagree.**
- Biological implications/applications to mechanistic studies. **Now many.**
- Room temperature studies: dose rate effects?
Results disagree.
- N.B. Need for **systematic statistically significant** experiments.
- **1999-2021 Series of Radiation Damage Workshops**

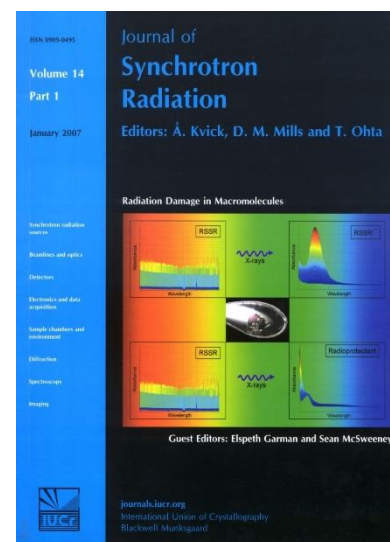
RD2: Dec 2001 RD3: Nov 2003 RD4: March 2006 RD5: March 2008



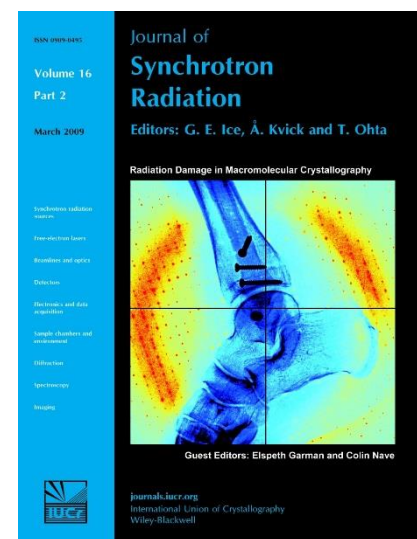
JSR, Nov 2002 (8)



JSR, May 2005 (9)



JSR, Jan 2007 (14)



JSR, March 2009 (8)

RD6: Mar 2010

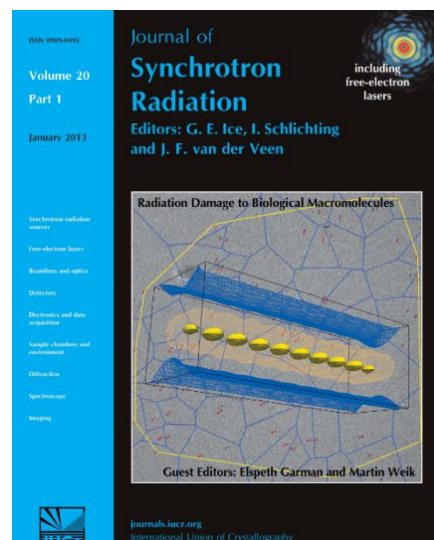
RD7: Mar 2012

RD8: Apr 2014

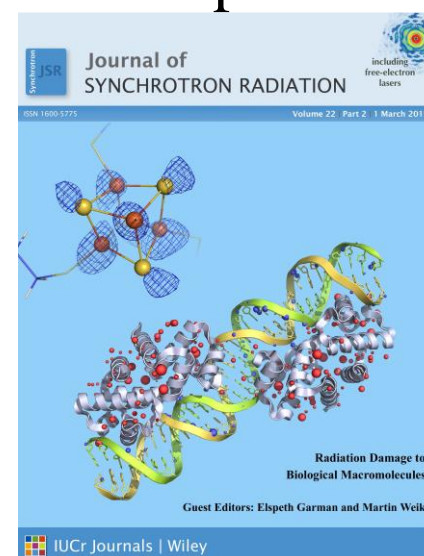
RD9: Mar 2016



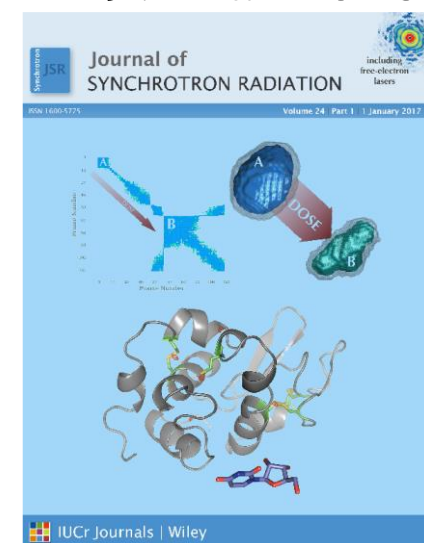
JSR, May 2011 (10)



JSR, Jan 2013 (6)



JSR, March 2015 (8)



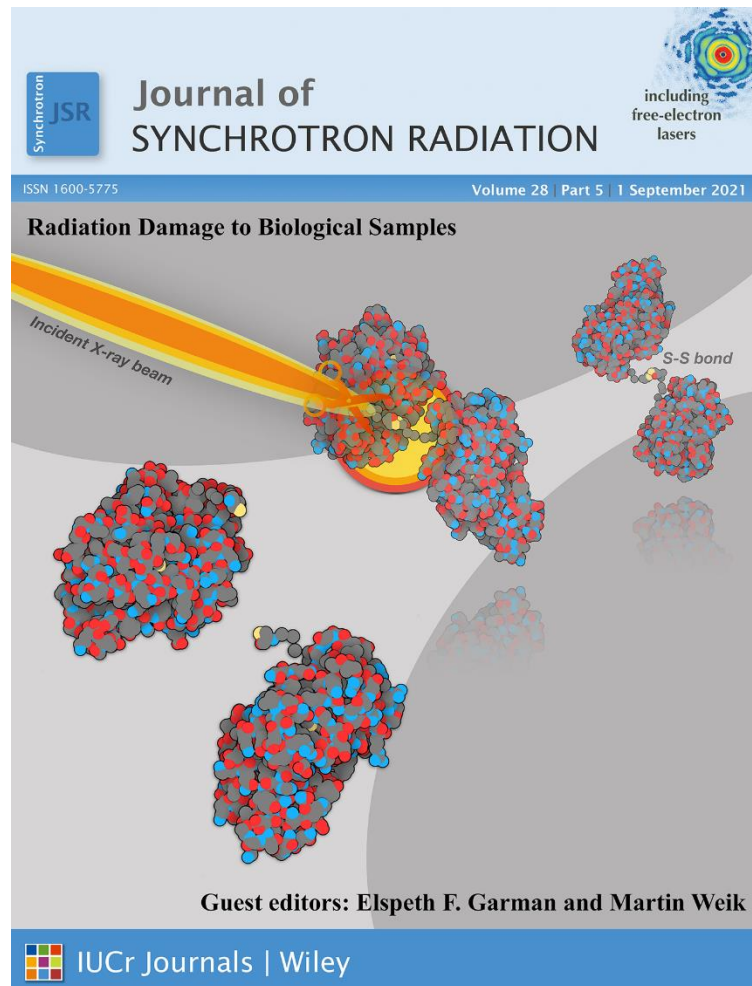
JSR Jan 2017 (8)

RD10: Sep 2018

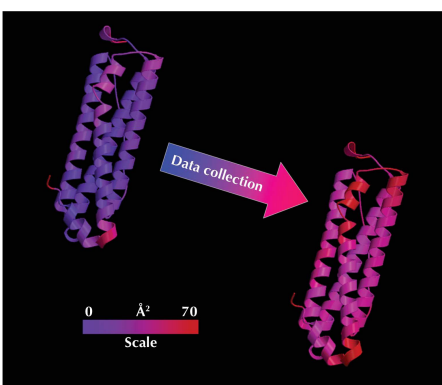


JSR, July 2019 (9)

RD11: Sep 2021

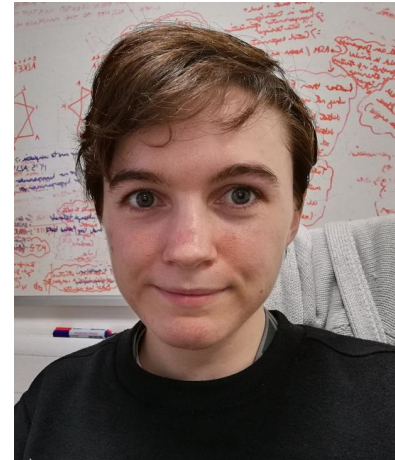


RD2 to 11:
Total of 94 papers published in
JSR Special Issues so far



Radiation damage:

The Plan:



- What are the symptoms?
- Why do we care? Effect on MAD/SAD.
- What is it?
- How do we calculate the Dose?
- What do we know/would like to know?
- **A new RD metric**

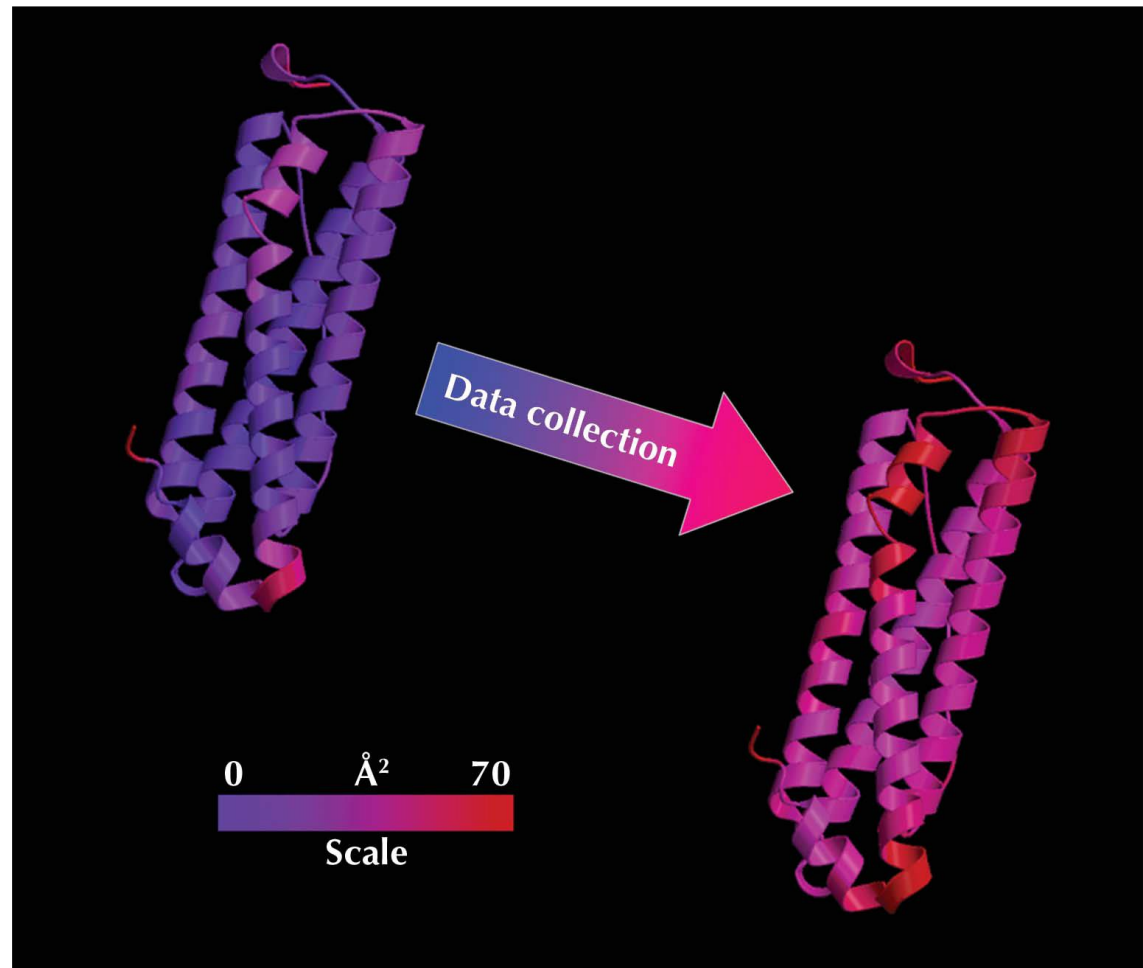
Question:

There are <30 ‘radiation damage series’ in the PDB.

Can we give an isolated deposited PDB file a
‘radiation damage index’?

In the PDB file:

*x , y , z , B ,
occupancy*



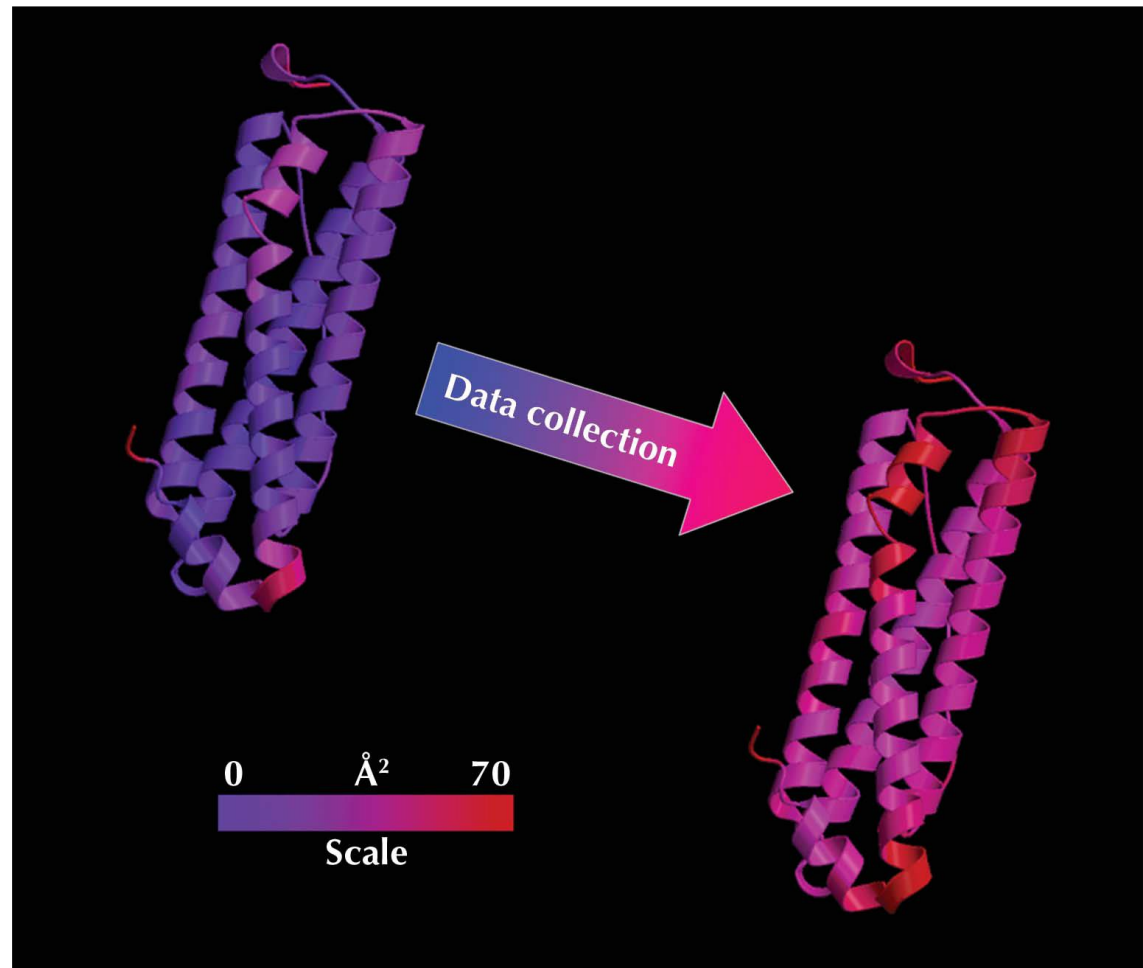
Question:

There are <30 ‘radiation damage series’ in the PDB.

Can we give an isolated deposited PDB file a **YES!!**
‘radiation damage index’?

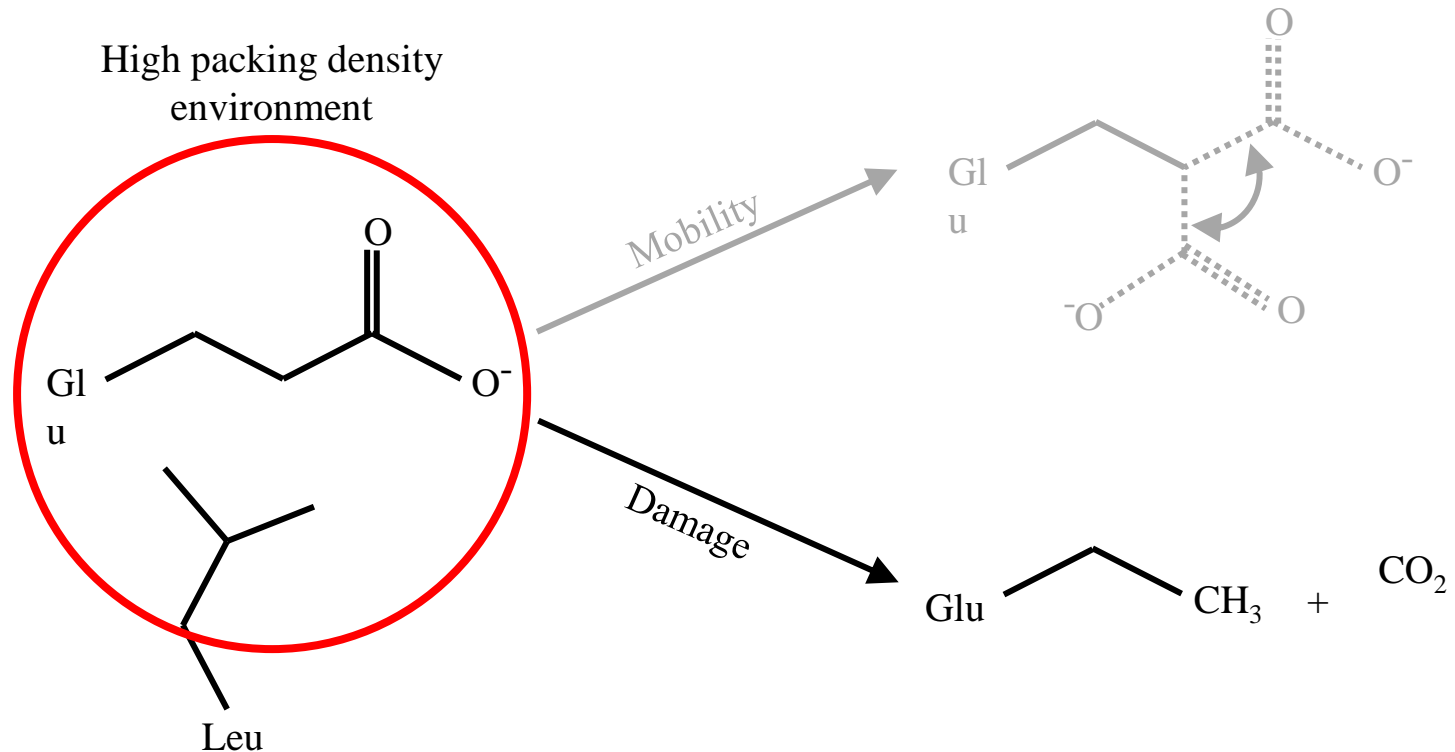


Kathryn Shelley



The B_{Damage} metric

- There is a strong correlation between mobility and packing density
- Correcting B -factor for packing density enables the distinction of damage from mobility



The B_{Damage} metric

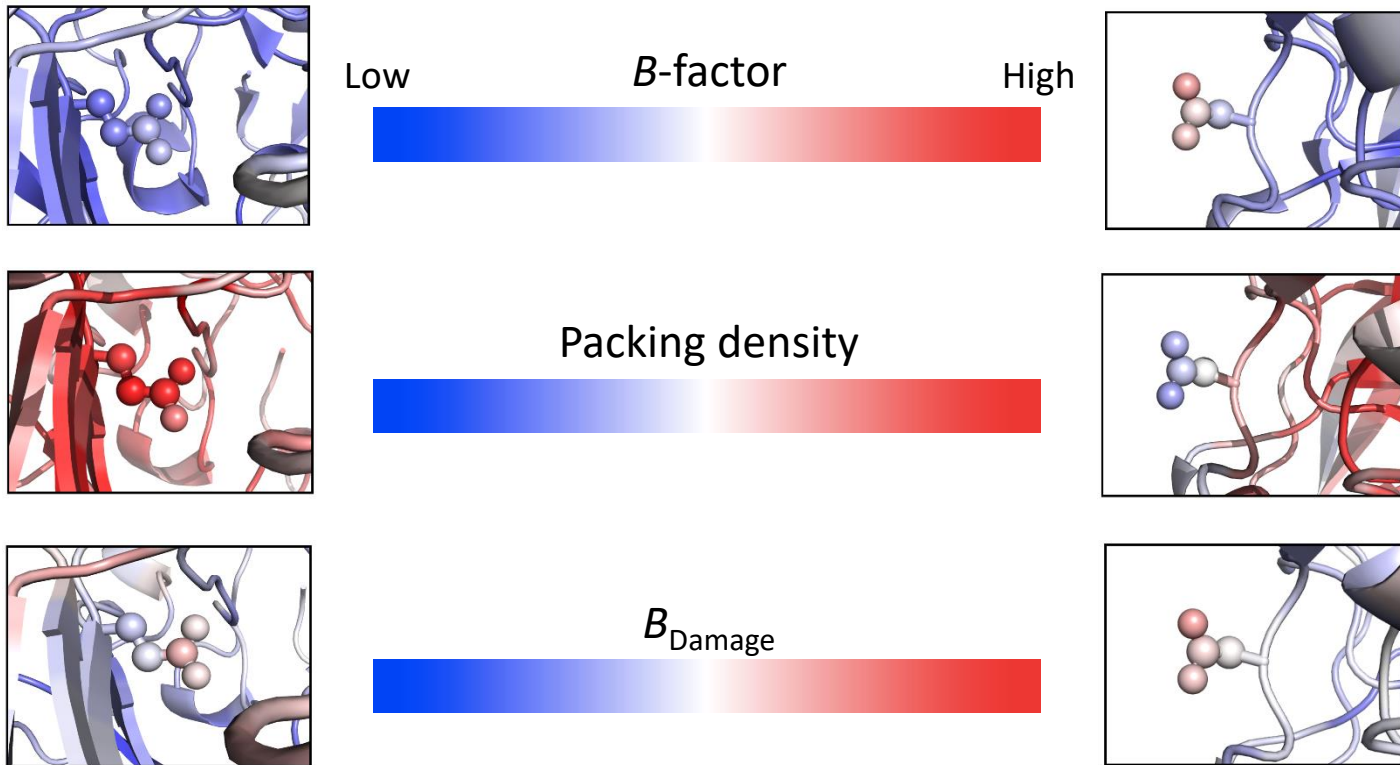
- $B_{\text{Damage}}^{[1]}$ is B -factor corrected for packing density

$$B_{\text{Damage } j} = \frac{B\text{-factor } j}{\frac{1}{n} \sum_{i=1}^n B\text{-factor } i}$$

[1] Gerstel, Deane, Garman (2015) *J Synchrotron Radiat* **22**, 201–212.

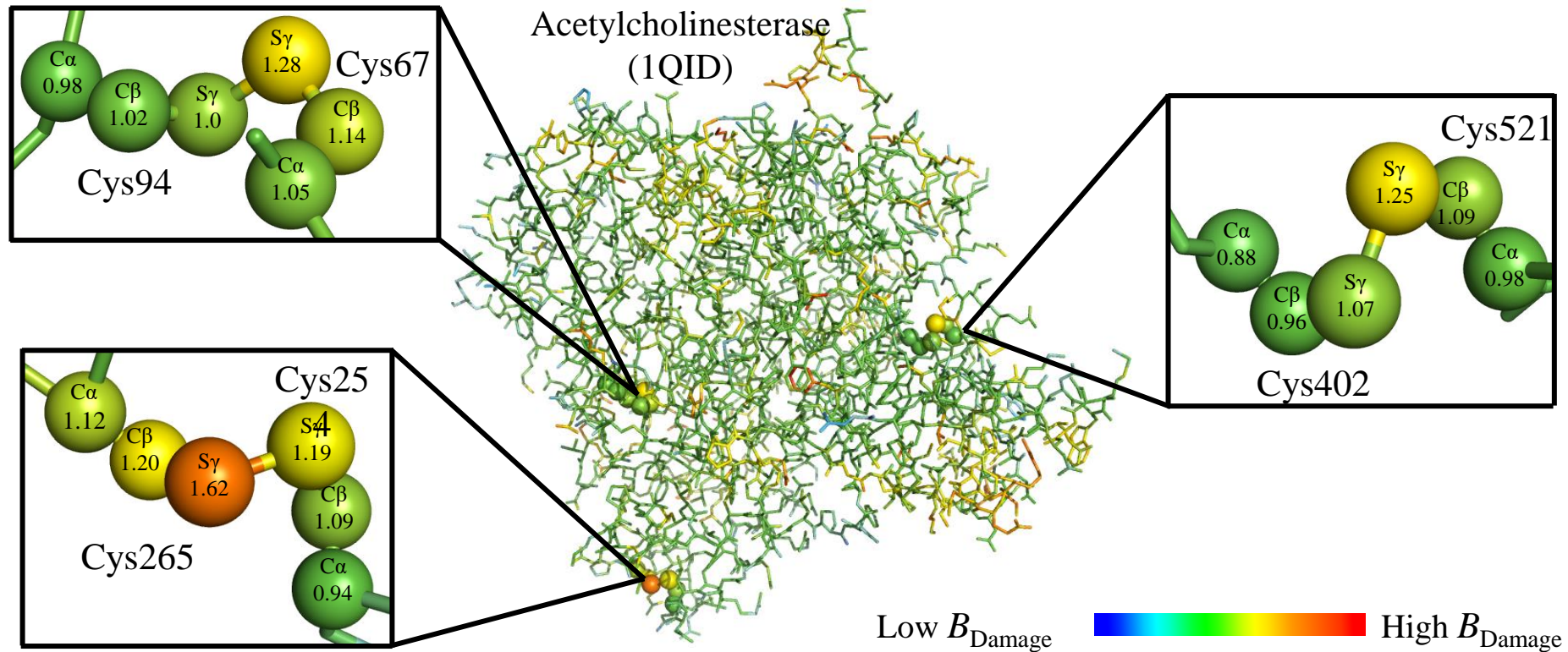
The B_{Damage} metric

- B_{Damage} is B -factor corrected for packing density



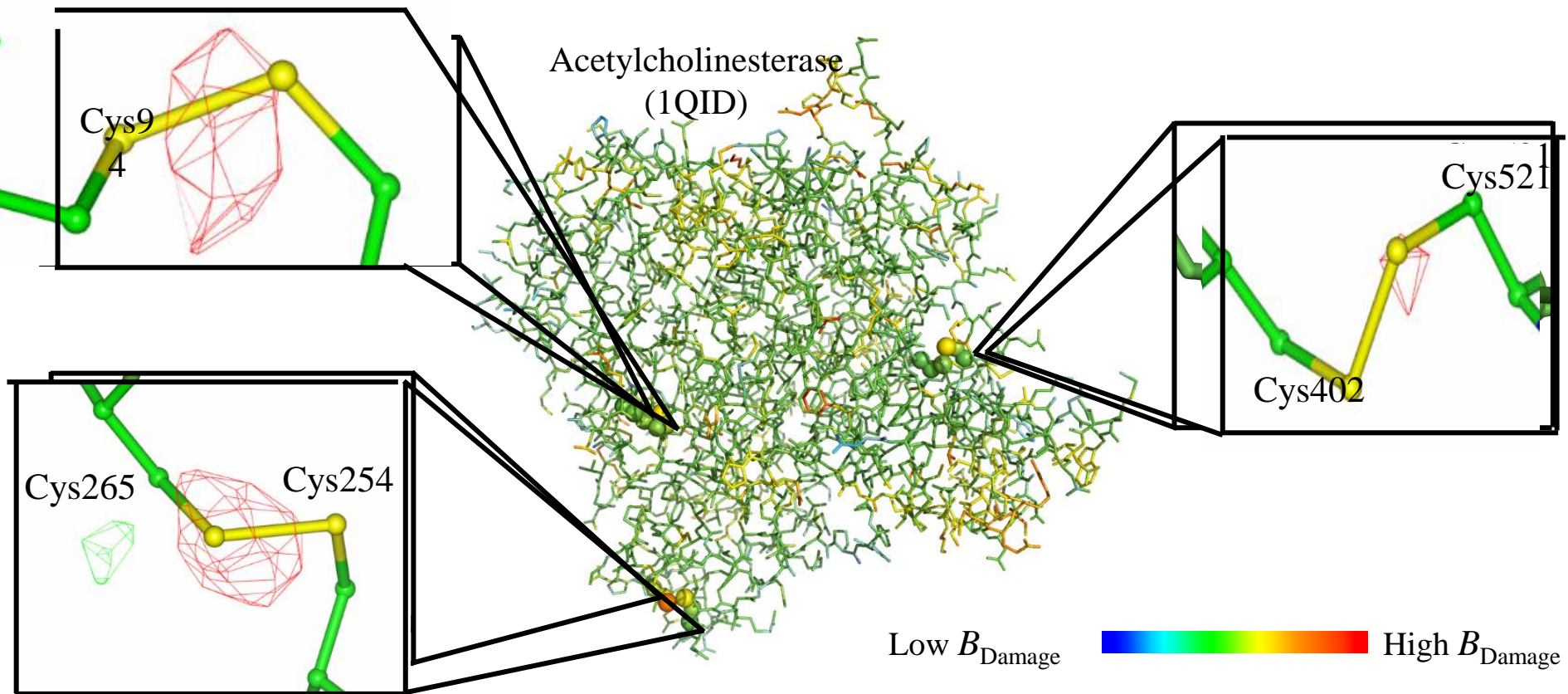
RABDAM (now in the CCP4 computing suite)

- RABDAM calculates B_{Damage} for all selected atoms in any standard format PDB file
- B_{Damage} highlights expected sites of specific radiation damage
- RABDAM provides several useful outputs to aid radiation damage evaluation



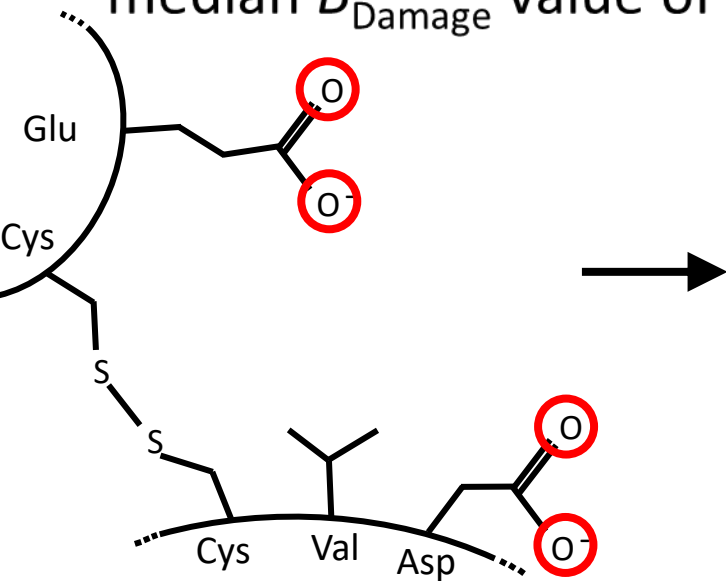
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- B_{Damage} highlights expected sites of specific radiation damage
- RABDAM provides several useful outputs to aid radiation damage evaluation

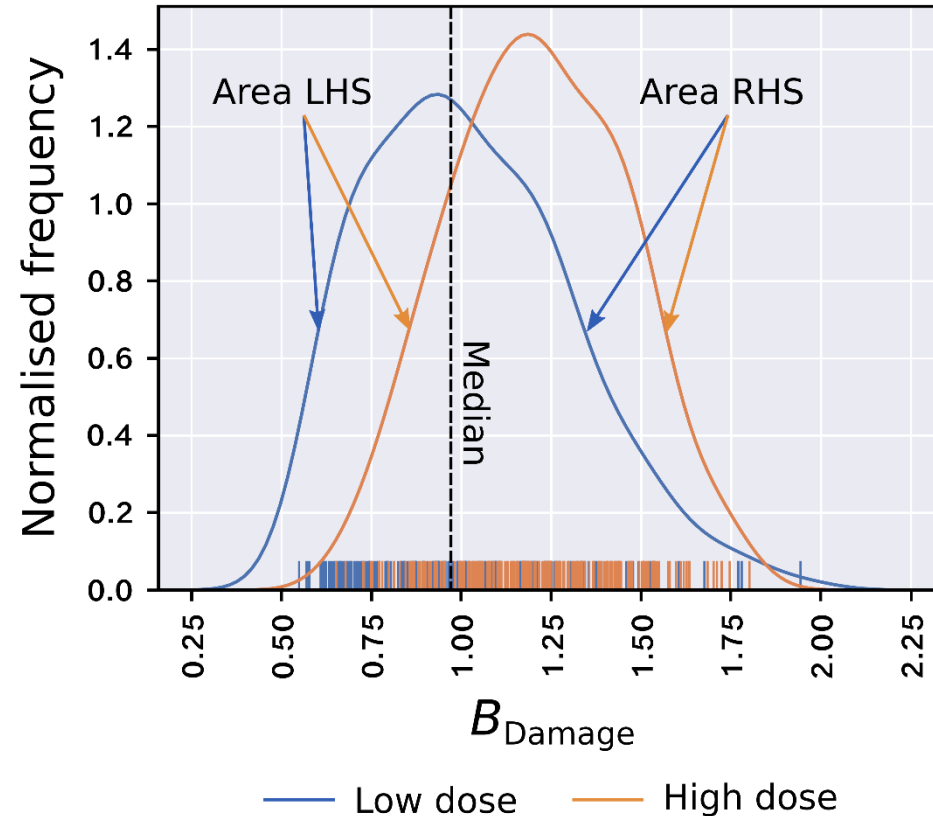


The B_{net} metric

- B_{net} is calculated from the distribution of the B_{Damage} values of Asp $\text{O}\delta$ and Glu $\text{O}\epsilon$ atoms
- Equal to the ratio of the area under the curve either side of the median B_{Damage} value of all protein atoms

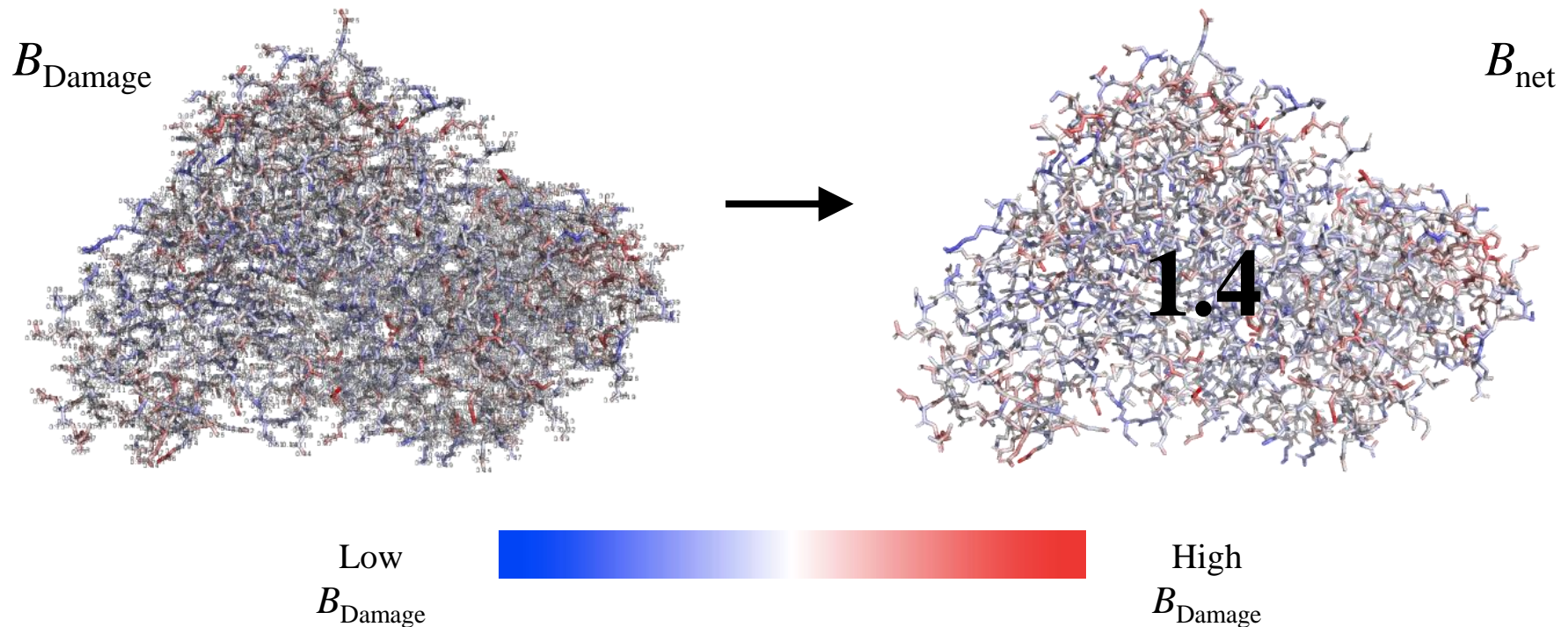


$$B_{\text{net}} = \frac{\text{Area RHS}}{\text{Area LHS}}$$

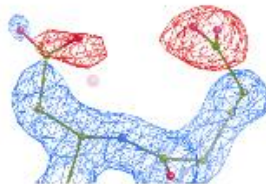
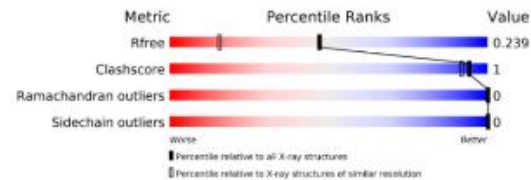


The B_{net} metric

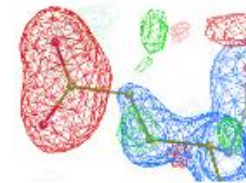
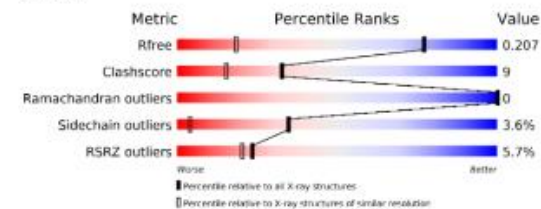
- B_{Damage} is a per-atom metric
- The B_{net} metric is a derivative of B_{Damage} that summarises the total extent of specific radiation damage suffered by a PX structure in a single value



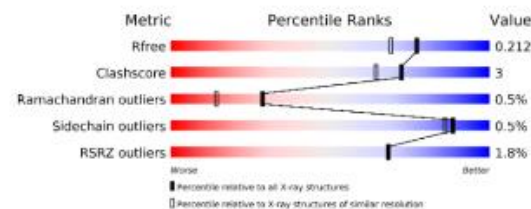
5WUC



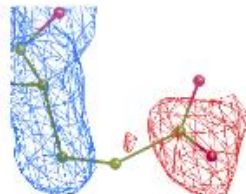
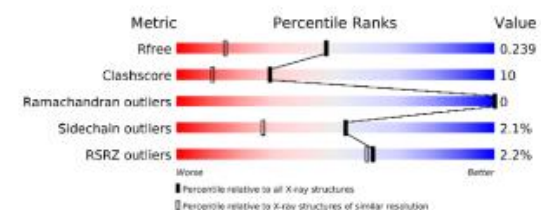
1V70



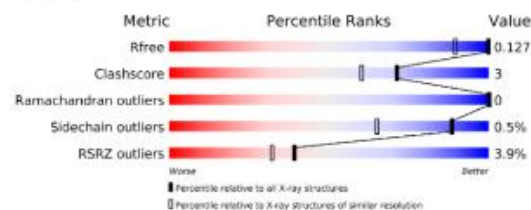
5FXL



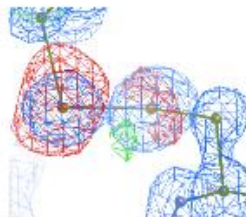
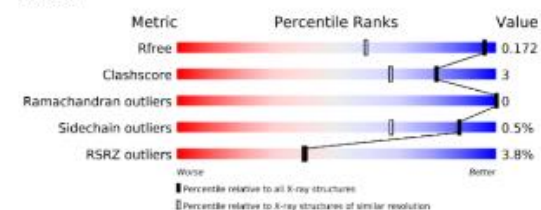
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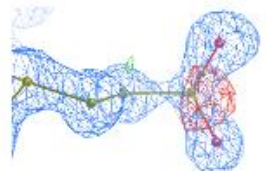
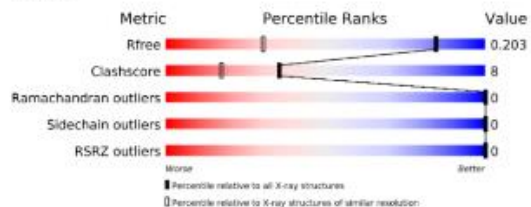
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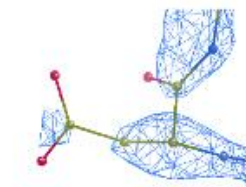
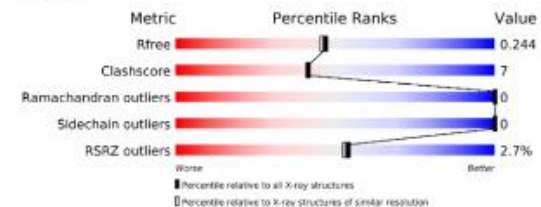
3A07



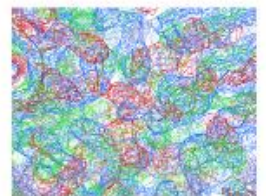
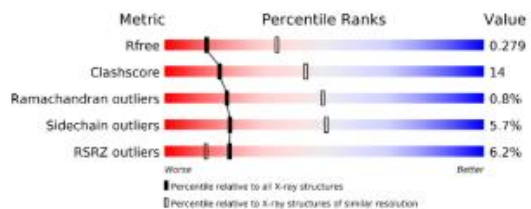
3S8S



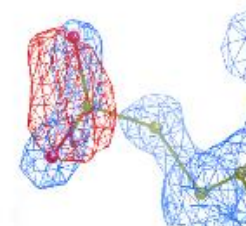
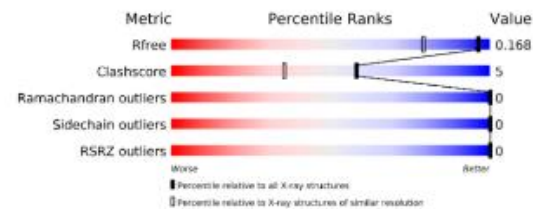
6BKL



3UX1

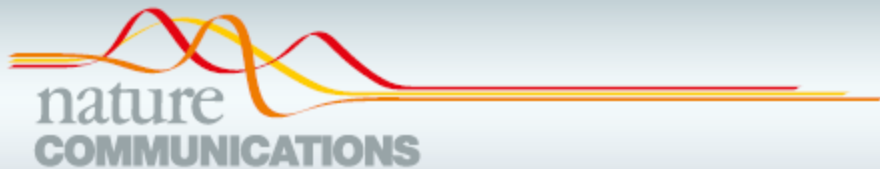


2XMK



B_{net} Summary

- B_{Damage} can detect potential sites of specific radiation damage in individual PX structures.
- B_{net} summarises the extent of damage suffered by a PX structure in a single value. Unlike B_{Damage} values, B_{net} values can be fairly compared between different structures.
- Both B_{Damage} and B_{net} values can be calculated with the CCP4 program *RABDAM*.
- *RABDAM* has the capacity to open up the entire PDB for large-scale statistical analysis of specific radiation damage.
- Currently radiation damage is largely overlooked when assessing the quality of structures in the PDB – we hope our metrics will help to change this.



14th March 2022

ARTICLE

<https://doi.org/10.1038/s41467-022-28934-0>

OPEN



Quantifying and comparing radiation damage in Protein Data Bank

Kathryn L. Shelley^{1,2} & Elspeth F. Garman¹

Summary 1: what can YOU, the experimenter do?

- Do not be afraid to merge data taken from different isomorphous crystals which all had lower doses.
- Back soak non-specifically bound heavier atoms out of your crystals.
- Be ‘absorption aware’ of the contents of your crystal (e.g. Se and buffer) and if possible, avoid cacodylate buffer (arsenic mass=75).
- Match beam size to crystal size

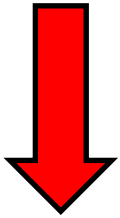
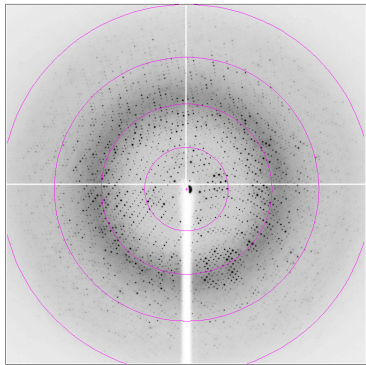
Summary 2: what can YOU, the experimenter do?

- Scavengers: try electron scavengers at 100 K (nitrate/ascorbate/benzoquinone).
- Dose ‘spreading’: use a tophat profile beam if possible. Consider Helical/Translational data collection.
- So you can estimate the dose, ASK at the beamline:
 - What is the flux today at this energy and with this slit size (‘flux density’)?
 - What is the beam profile today at this beam energy? FWHM in x and y ?

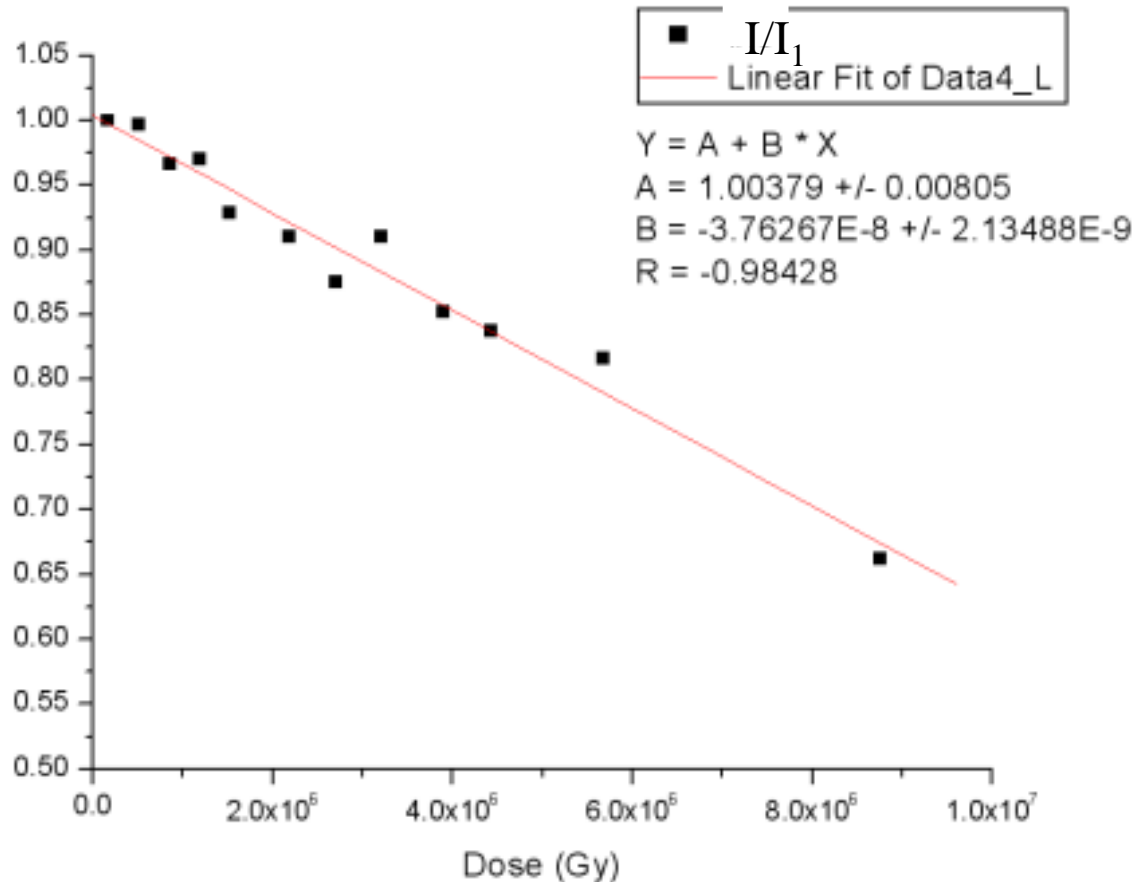
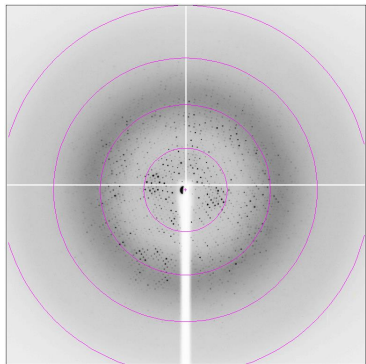
Sacrificial crystal to characterise damage rate



Diffraction fading



I/I_1



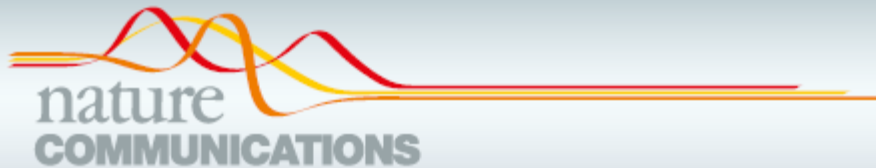
Dose = absorbed energy (J) / mass (kg)

Current status: radiation damage in protein crystals

- Understand a lot more than 20 years ago, but still not nearly enough...
- Understand how to do experiments better.
- Research has prompted some very exciting new approaches.
- Many complementary methods now being used on the problem in concert with crystallography
- Experiments must involve more than one sample (!) to get statistically significant results: labour intensive and time consuming. Also MUST know incident FLUX density...
- **Radiation damage has DEFINITELY become a mainstream concern**



FURTHER READING:

- ‘Beginner’s guide to Radiation Damage’ Holton, (2009)
JSR **16**,133-142
- General summary in: Garman, *Acta D* (2010) **66**, 339-355
- Garman and Weik, Chapter 20 in ‘*Protein Crystallography: Methods in Molecular Biology*’ (2017) **1607**, 477-489



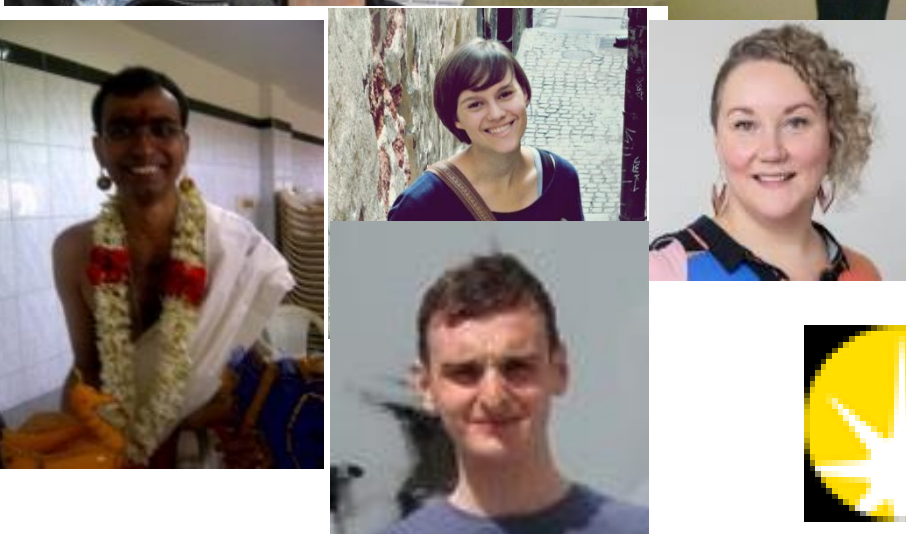
14th March 2022

Quantifying and comparing radiation damage in
Protein Data Bank

Kathryn L. Shelley ^{1,2} & Elspeth F. Garman ¹

- Garman & Weik *Curr. Opin. Struct. Biol.* (2023) **82**:102662

I thank my past and present group and our collaborators, and
acknowledge their huge contribution to the work



Some Garman Group PhD students
and Postdocs 2000-2018:
next generation Crystallographers...



Graduate students

James Murray (Imperial College)

Robin Owen (DLS)

Eugenio de la Mora Lugo (IBS, Grenoble)

Oliver Zeldin (Facebook)

Markus Gerstel (Cybersecurity)

Helen Ginn (U of Hamburg)

Jonathan Brooks-Bartlett (Spotify)

Charlie Bury (Exscientia)

Postdocs

Karthik Paithankar (U. Frankfurt)

Undergraduate Project students

Kathryn Shelley (RABDAM)

Josh Dickerson (RADDPOSE-3D)

Collaborators:

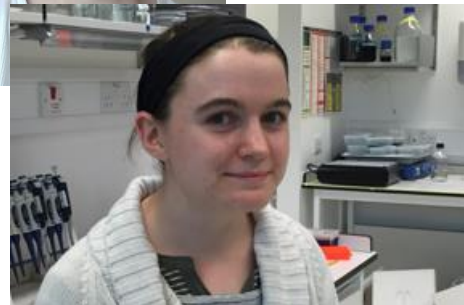
Raimond Ravelli, Maastricht.

Martin Weik, IBS

Ian Carmichael, NDRL, USA

John McGeehan, Portsmouth

James Holton, UCSF



This was a lot of material delivered very fast so...



questions expected and very welcome

elspeth.garman@bioch.ox.ac.uk

The Crystallographer's DILEMMA:



Rate of damage
versus diffraction
intensity

RADDOSE-3D

TEST our new GUI!!

To run RADDOSE-3D for MX, SMX or SAXS (which ever you like!)

Step 1: Download and unzip the RADDOSE-3D GUI from:

https://github.com/jdickerson95/qt_RADDDOSE-3D/releases

There are versions for a PC (Windows_release.zip) and for Linux (Linux_release.zip).

If you have a MAC, there is no new GUI yet, but you can run a limited capability RADDOSE-3D from the WWW site:

raddo.se

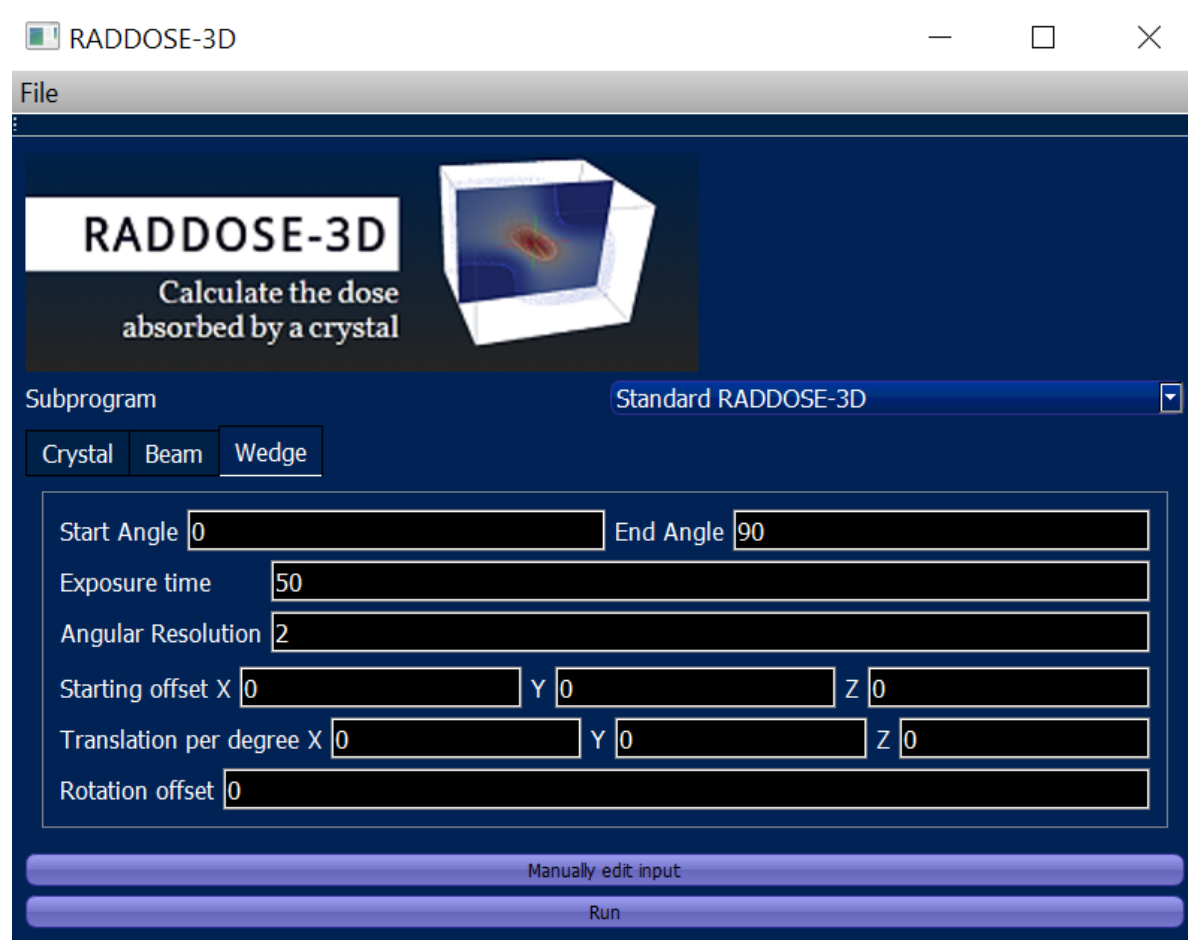
(click on ‘manual interface’ and run the test example first. Then edit the input for a case you would like to try)

To run the GUI you need to have Java installed which you can get free at

https://www.java.com/download/ie_manual.jsp

Also, if you have R (<https://www.r-project.org/>) installed, from the RADDOSE-3D output you will be able to produce 3D representations of the dose distribution in your sample.

Step 3: Find the file RD3D_GUI.EXE and if on a PC click on it. For Linux run it however you usually run executable files. The GUI should open, and you can enter input on 3 tabs: crystal, beam and wedge.

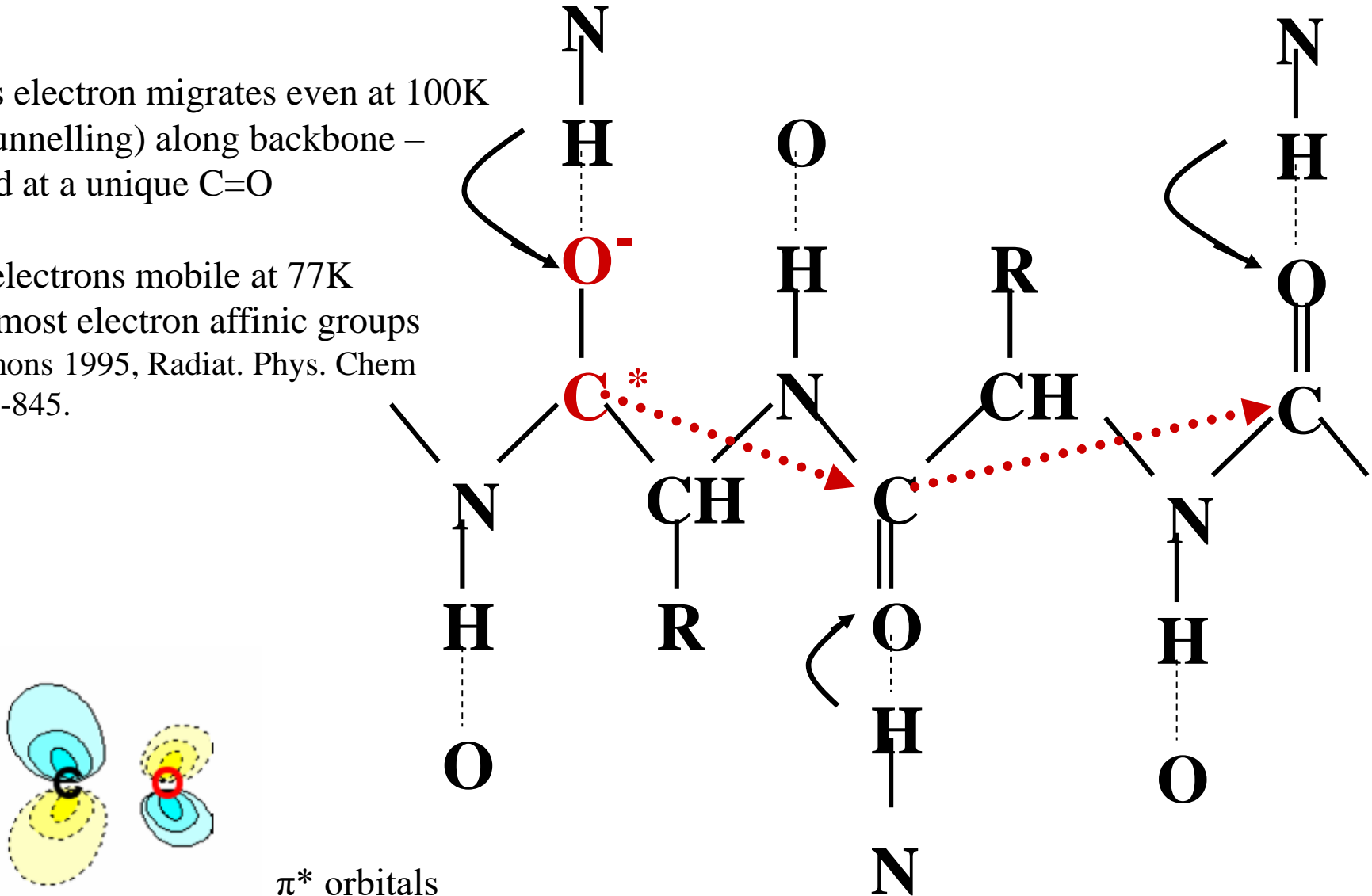


2) DIRECT RADIATION DAMAGE. Protein Redox

a) electron migration and trapping.

Excess electron migrates even at 100K
(q.m.tunnelling) along backbone –
trapped at a unique C=O

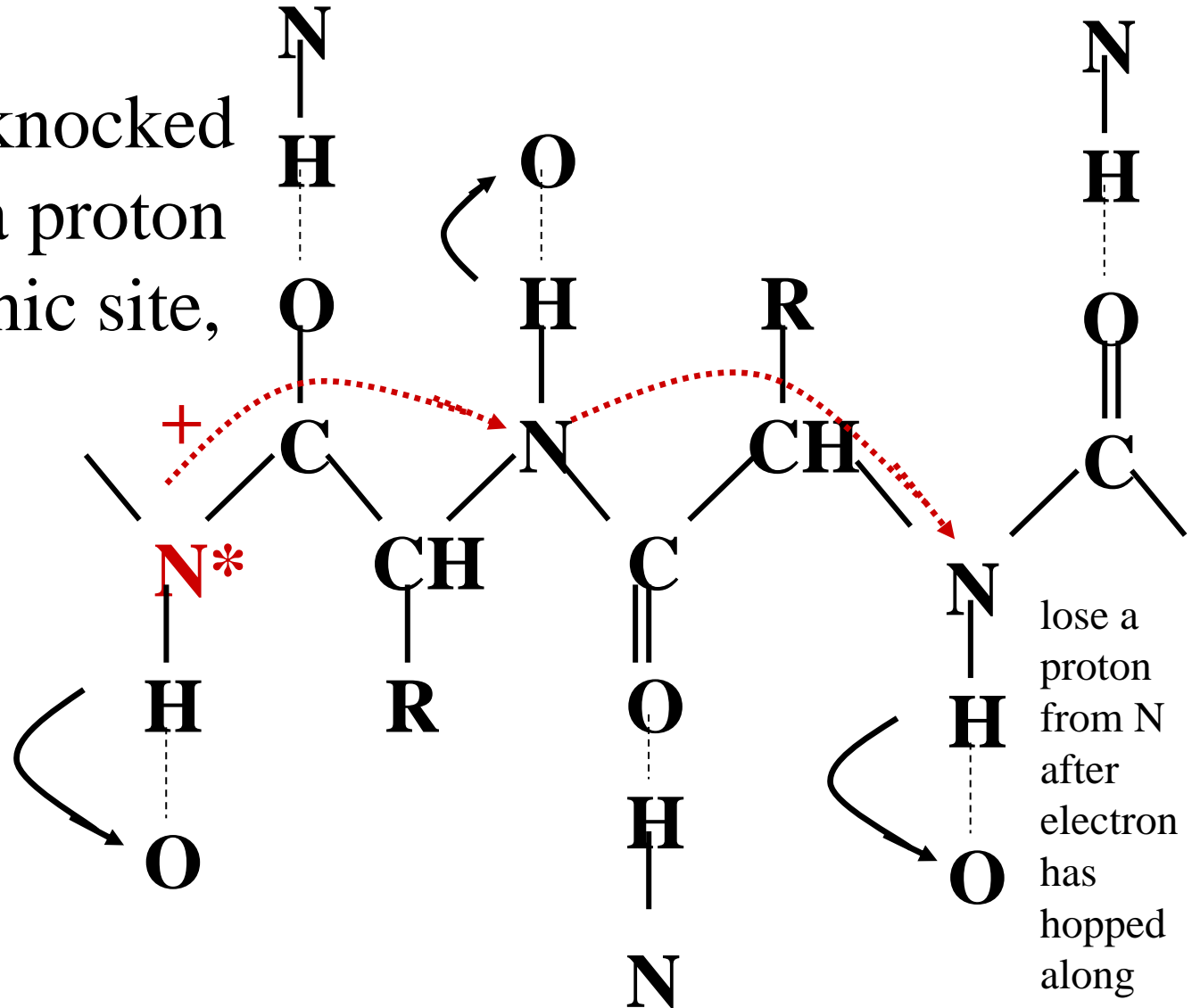
ESR: electrons mobile at 77K
Go to most electron affinic groups
M. Symons 1995, Radiat. Phys. Chem
45, 837-845.



2) DIRECT RADIATION DAMAGE. Protein Redox- b) proton hole migration.

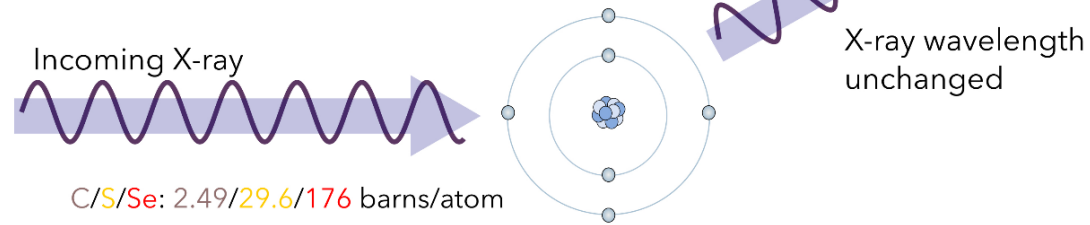
Electron gets knocked off, then lose a proton from the cationic site, get a radical.

lose an electron from N

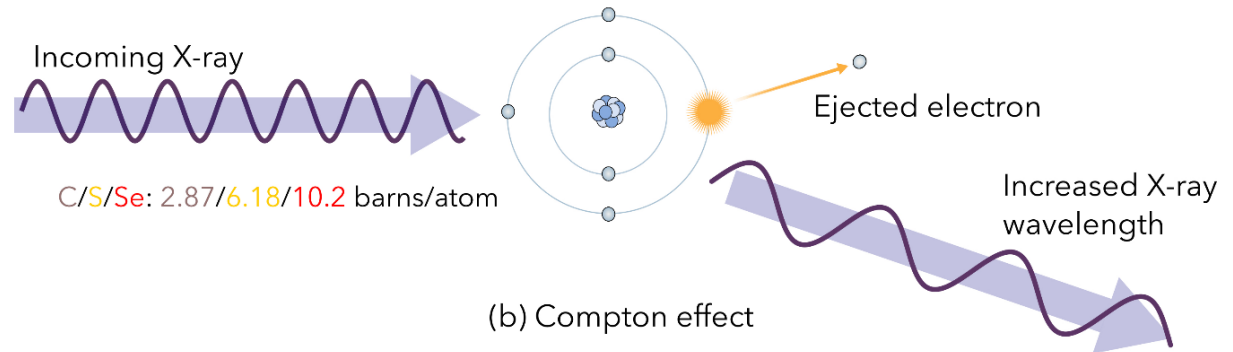


Interactions of X-rays with atoms in a crystal

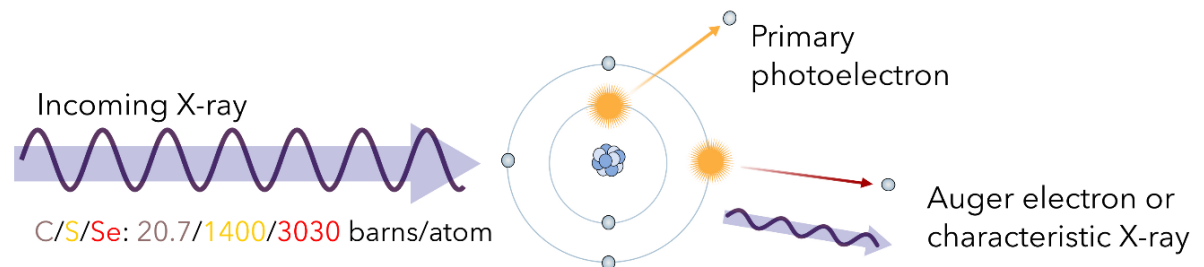
Cross sections at E_{inc} 12.4 keV (1 Å)



(a) Elastic scattering



(b) Compton effect



(c) Photoelectric effect