



UNIVERSITY OF
LIVERPOOL

Ronan Keegan CCP₄ Group

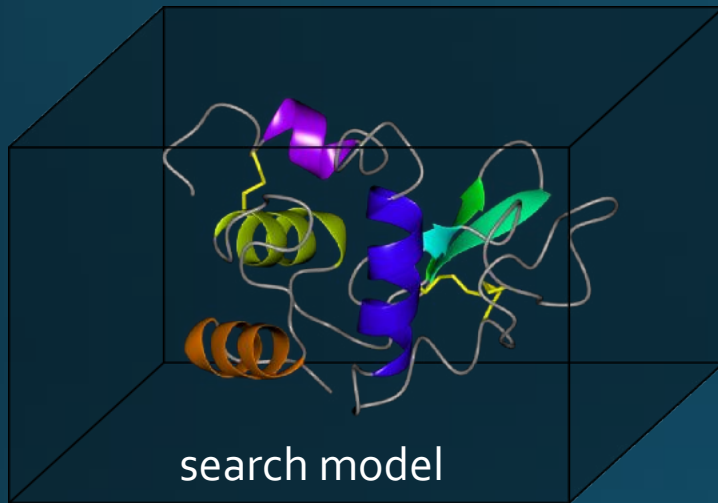
MR model selection, preparation and assessing the solution

DLS-CCP₄ Data Collection and Structure Solution Workshop 2018

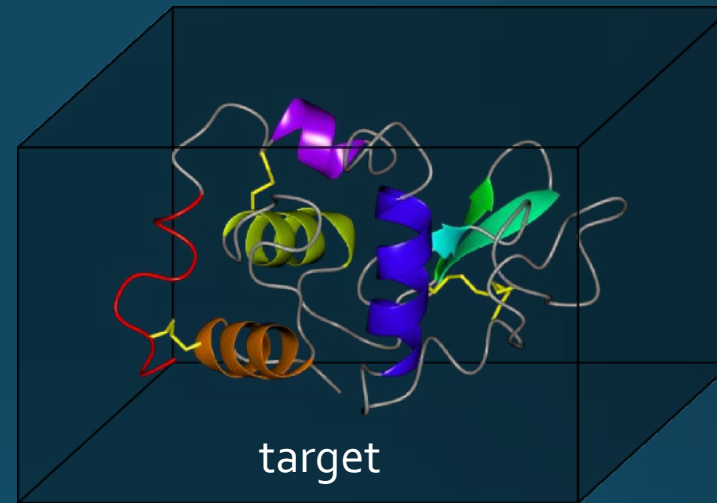
Overview

- Introduction
- Step-by-step guide to performing Molecular Replacement
- Automatic Molecular Replacement in CCP4
- What to do if Molecular Replacement doesn't work
- Acknowledgements

Introduction: *Basics of Molecular Replacement*



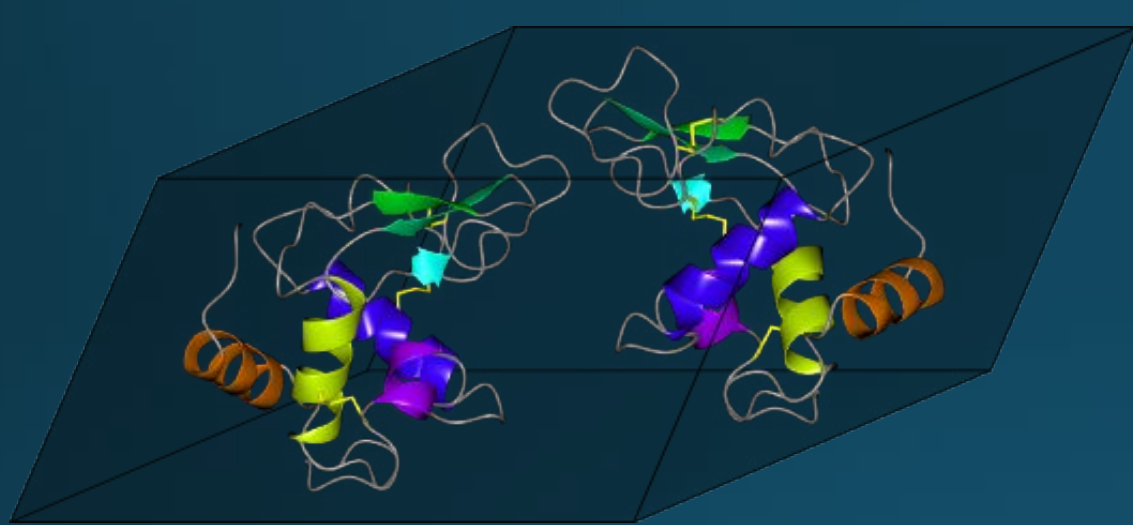
h	k	l	F	ϕ
0	0	1	12.6	123
0	0	2	2.1	12
0	0	3	69.9	287



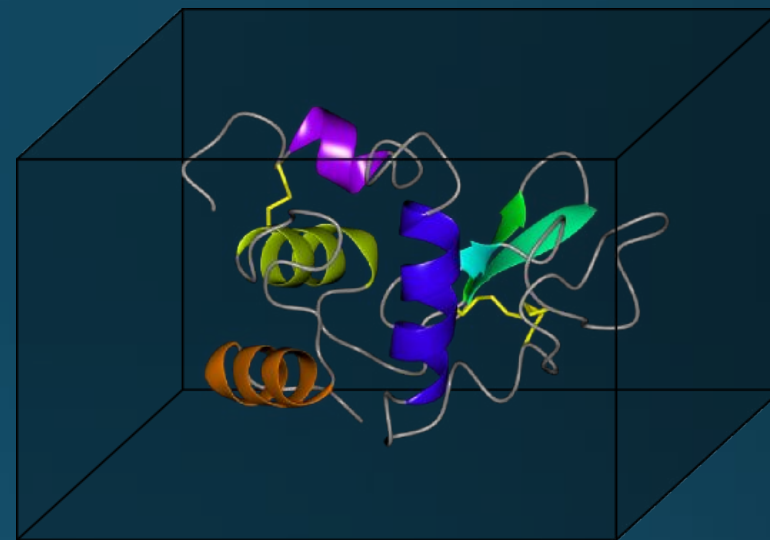
h	k	l	F	ϕ
0	0	1	10.4	113
0	0	2	3.5	18
0	0	3	57.2	265

(slide from Gabor Bunkoczi)

Introduction: *Basics of Molecular Replacement*



Search model crystal form

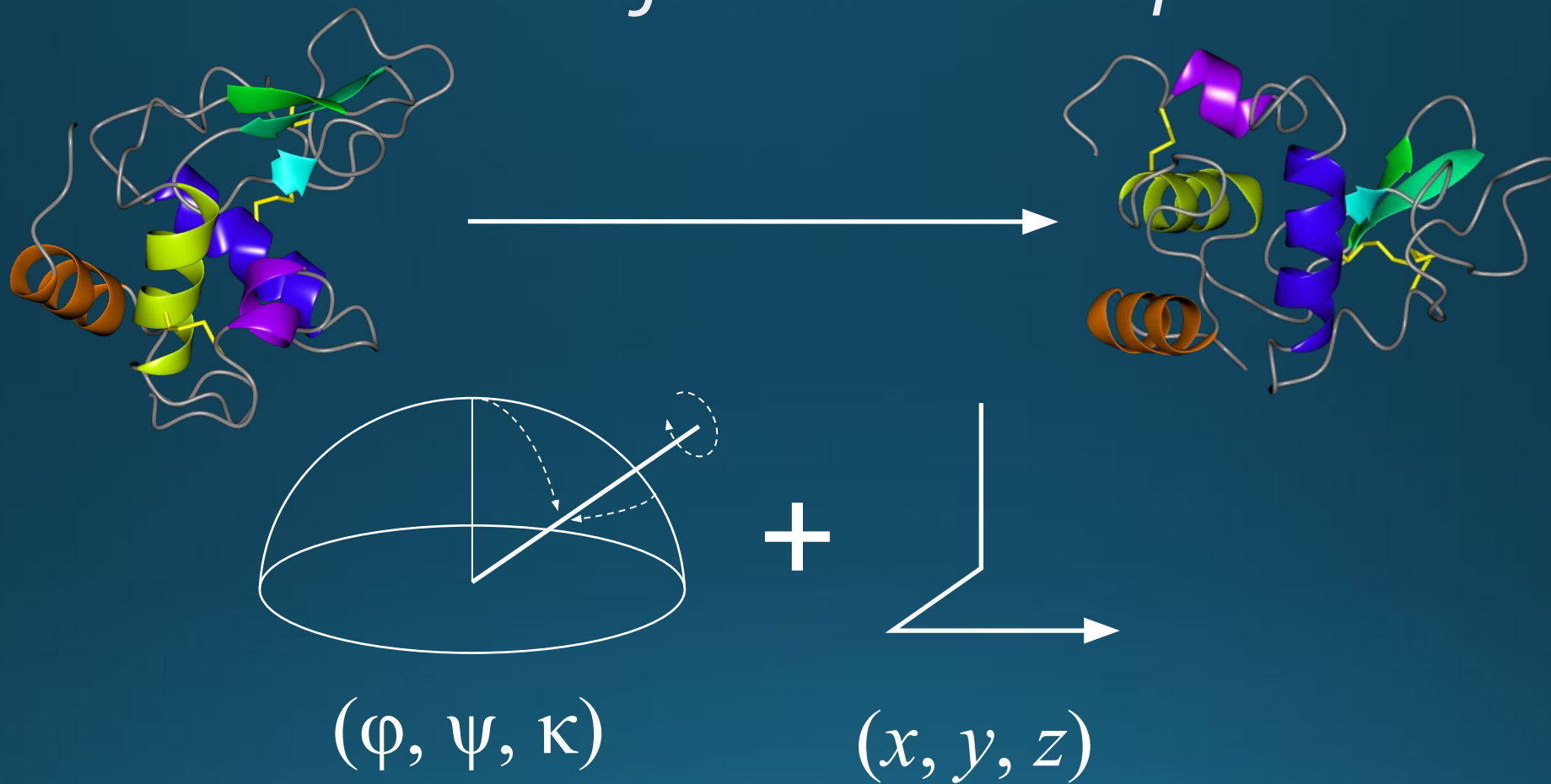


Target crystal form

$$\mathcal{R}\{\varphi, \psi, \kappa, x, y, z\}$$

(slide from Gabor Bunkoczi)

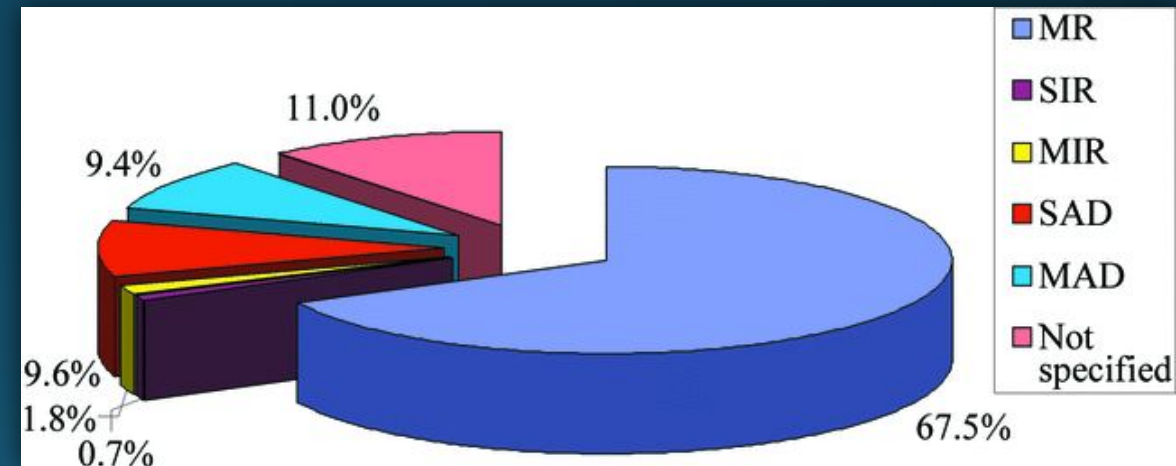
Introduction: *Basics of Molecular Replacement*



(slide from Gabor Bunkoczi)

Introduction: *Molecular Replacement and the PDB*

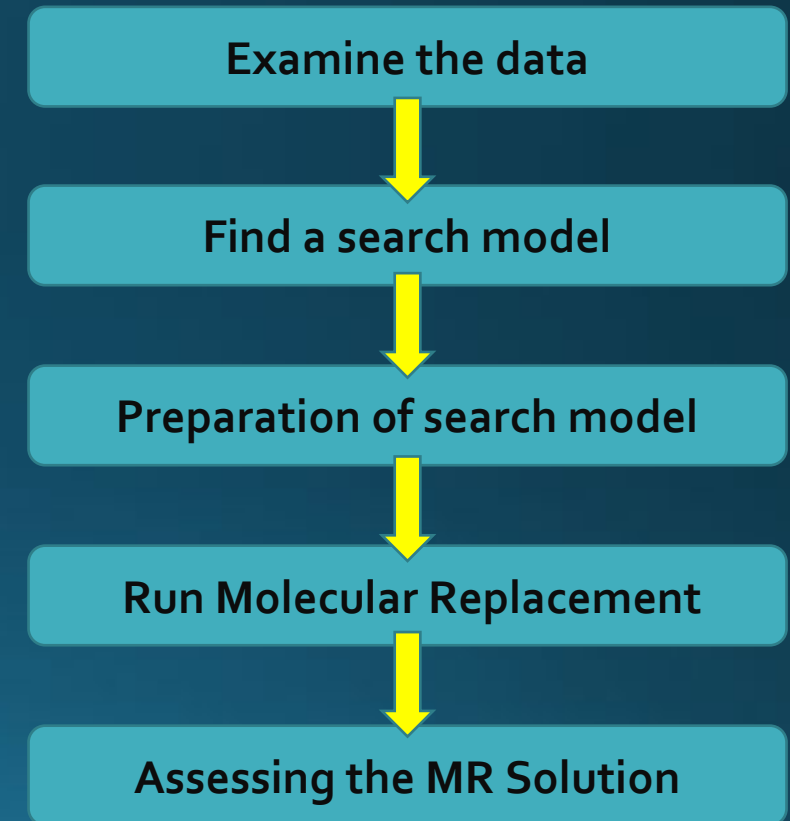
- Today, the vast majority of entries in the PDB have been phased using MR
- Increasing success rate of MR has been due to:
 - Improving methods in software e.g. maximum likelihood in Phaser
 - Increasing availability of suitable search models



(image from Fei Long)

Introduction: *Step-by-step Molecular Replacement*

- Despite this, successfully performing MR depends on attention to detail both before and after running the Molecular Replacement program



Step-by-Step MR: *Examining the data*

- Data issues can have an impact on how well MR will work

Examine the data

Step-by-Step MR: *Examining the data*

- Data issues can have an impact on how well MR will work
- Things to think about:
 - How many copies in the asymmetric unit?
 - What's the resolution of the data?
 - Self-rotation function – signs of NCS?

Examine the data

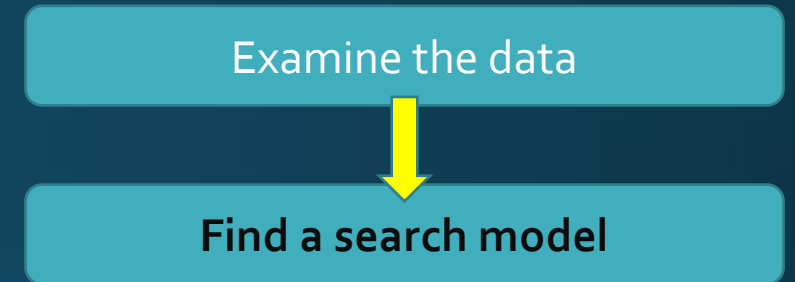
Step-by-Step MR: *Examining the data*

- Data issues can have an impact on how well MR will work
- Things to think about:
 - How many copies in the asymmetric unit?
 - What's the resolution of the data?
 - Self-rotation function – signs of NCS?
- Potential problems:
 - Pseudo translation?
 - Twinned data?
 - Space group assignment correct?

Examine the data

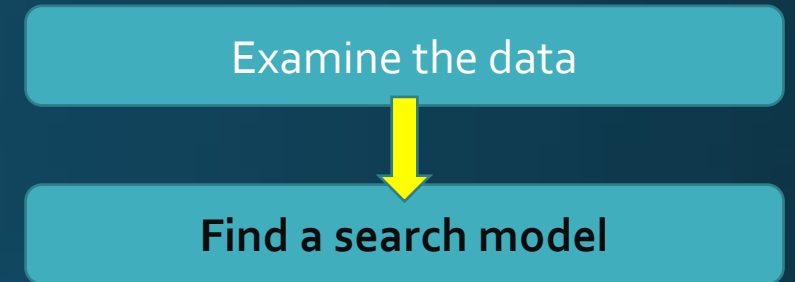
Step-by-Step MR: *Finding a search model*

- A well chosen and optimized search model has two key advantages



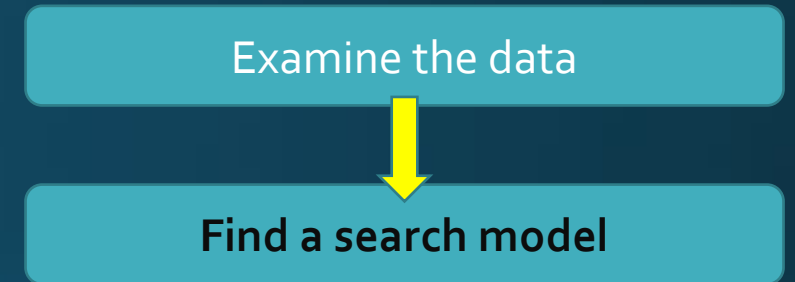
Step-by-Step MR: *Finding a search model*

- A well chosen and optimized search model has two key advantages
 1. Makes it easier for the MR program to find the correct position for that search model and hence sensible starting phases



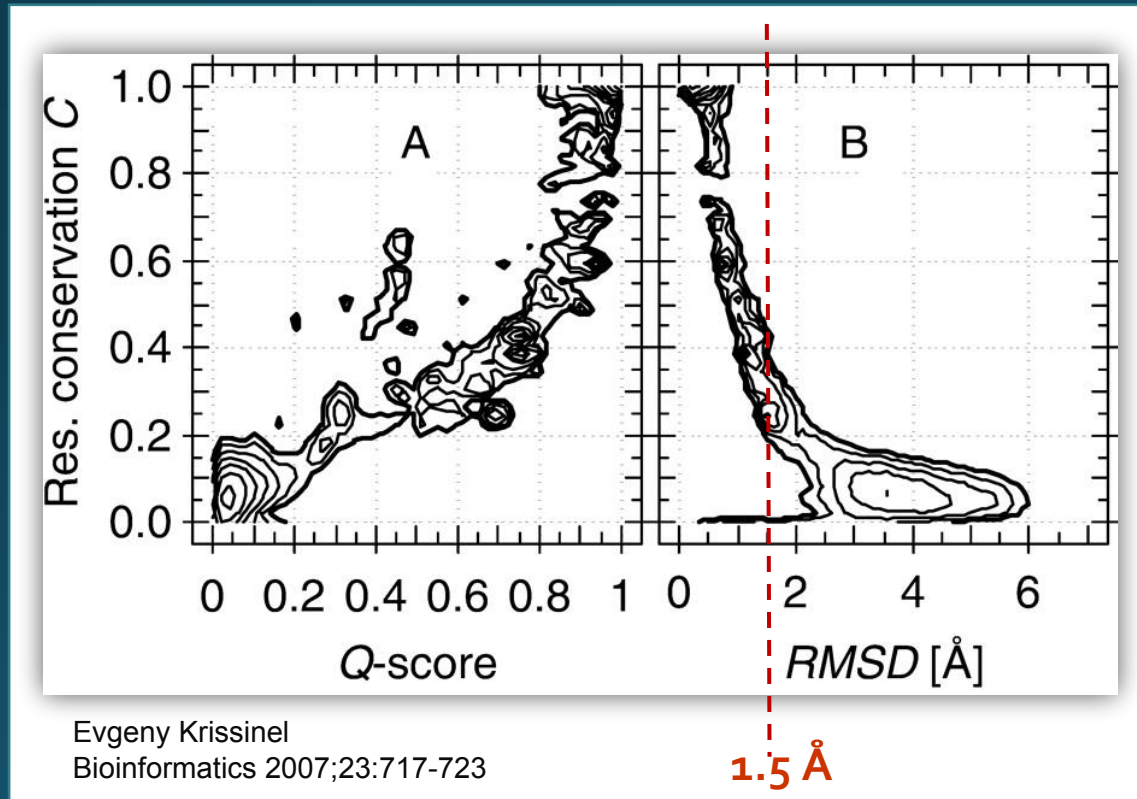
Step-by-Step MR: *Finding a search model*

- A well chosen and optimized search model has two key advantages
 1. Makes it easier for the MR program to find the correct position for that search model and hence sensible starting phases
 2. Facilitates quicker and easier model building and refinement post MR



Step-by-Step MR: *Finding a search model*

- How to find a search model?
 - Amino acid sequence similarity often correlates well with structural similarity



Examine the data



Find a search model

Step-by-Step MR: *Finding a search model*

- Considerations

- Target Size:

- The bigger the structure the more likely there will be sizeable conformational changes

- Data Resolution :

- Lower than 2.5 Angstroms – search model should represent large fraction of target molecule
 - Between 1.0 and 2.5 Angstroms – search model can be small fragment of target – DM and model building can improve phases
 - Better than 1.0 Angstrom – search model can be single atom (McCoy et al. 2017, PNAS)

- Homologue Sequence identity < 30%

- Suitable search models may exist – sequence identity calculation is difficult

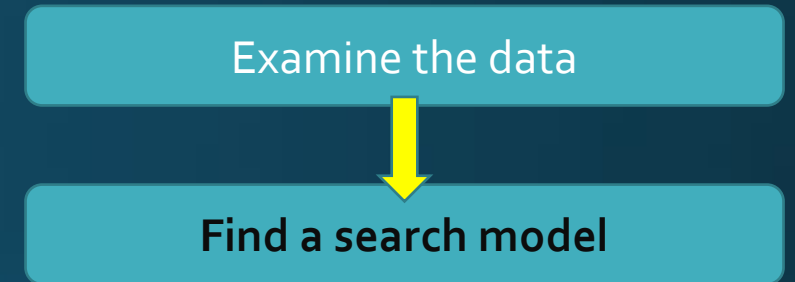
Examine the data



Find a search model

Step-by-Step MR: *Finding a search model*

- The field of Bioinformatics has given us a wealth of sophisticated and sensitive sequence-based search tools for finding potential homologues from which we can derive search models for MR



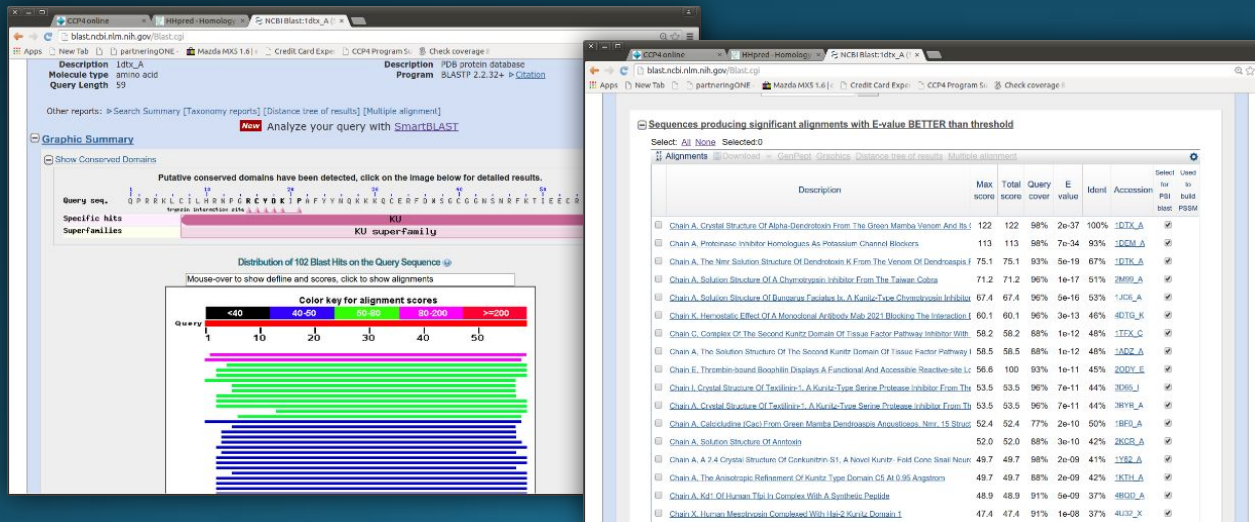
Step-by-Step MR: *Finding a search model*

- PSI-Blast
 - Profile-based searching
 - Online server, fast
 - Works well at finding suitable homologues down to sequence identities of 30%

Examine the data



Find a search model



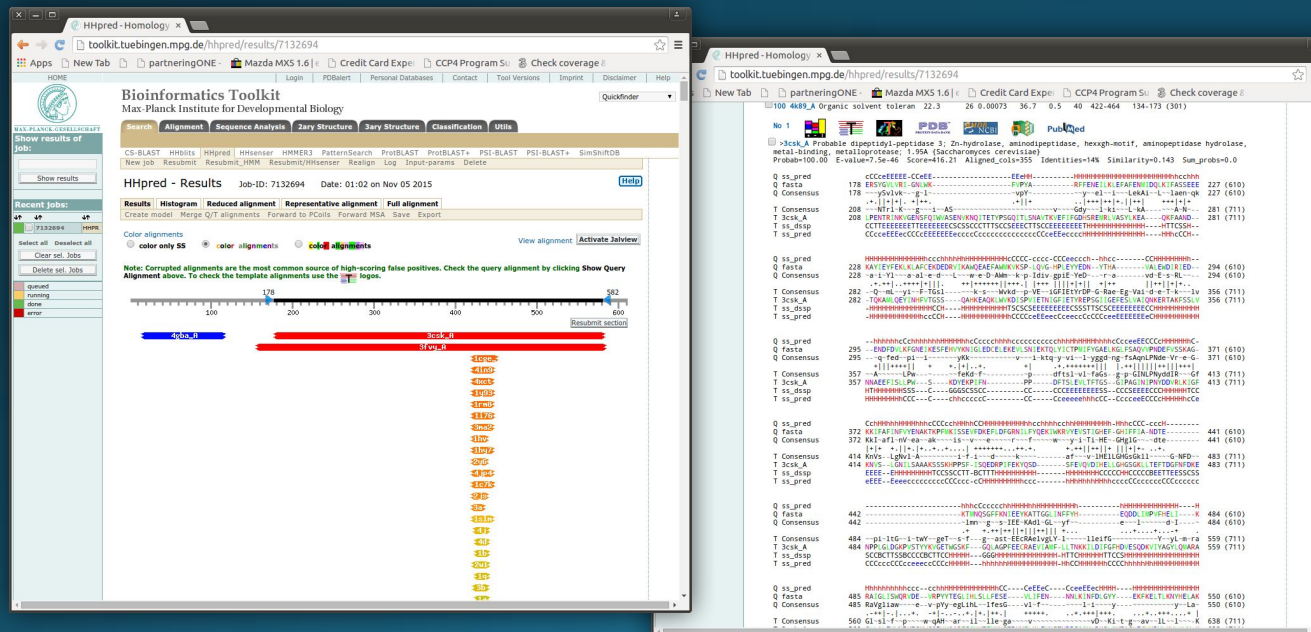
Step-by-Step MR: *Finding a search model*

- HHpred
 - Hidden Markov Model approach
 - Suitable for more difficult cases where no obvious homologue is available

Examine the data

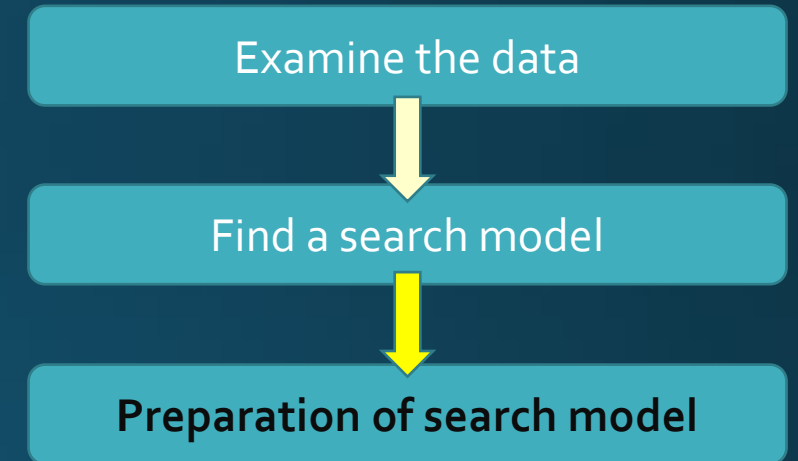


Find a search model



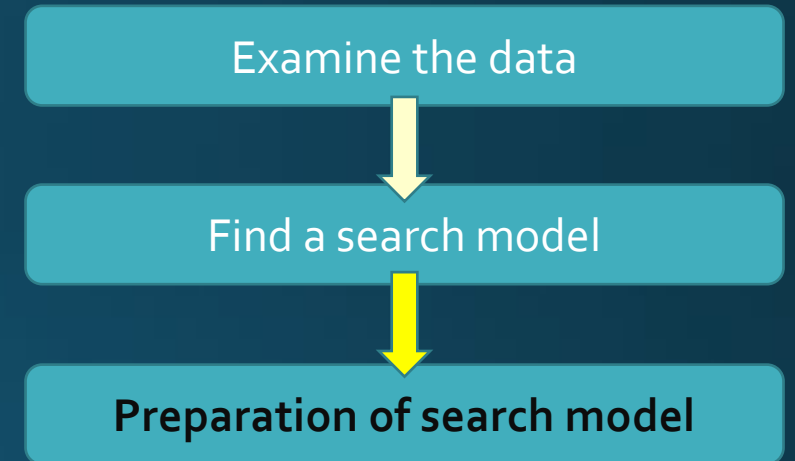
Step-by-Step MR: *Preparation of search model*

- Once a suitable homologue or set of homologues have been found they need to be prepared for use as search models



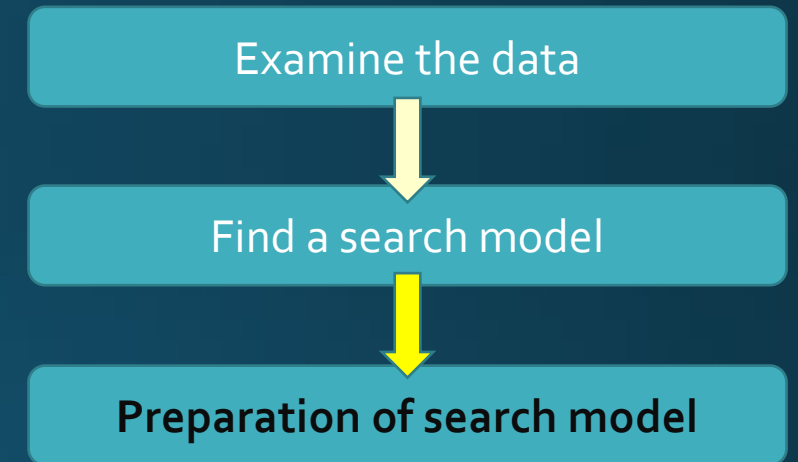
Step-by-Step MR: *Preparation of search model*

- Once a suitable homologue or set of homologues have been found they need to be prepared for use as search models
- The closer we can make the homologue to the target in terms of structural similarity the more likely it is to be successfully positioned in Molecular Replacement



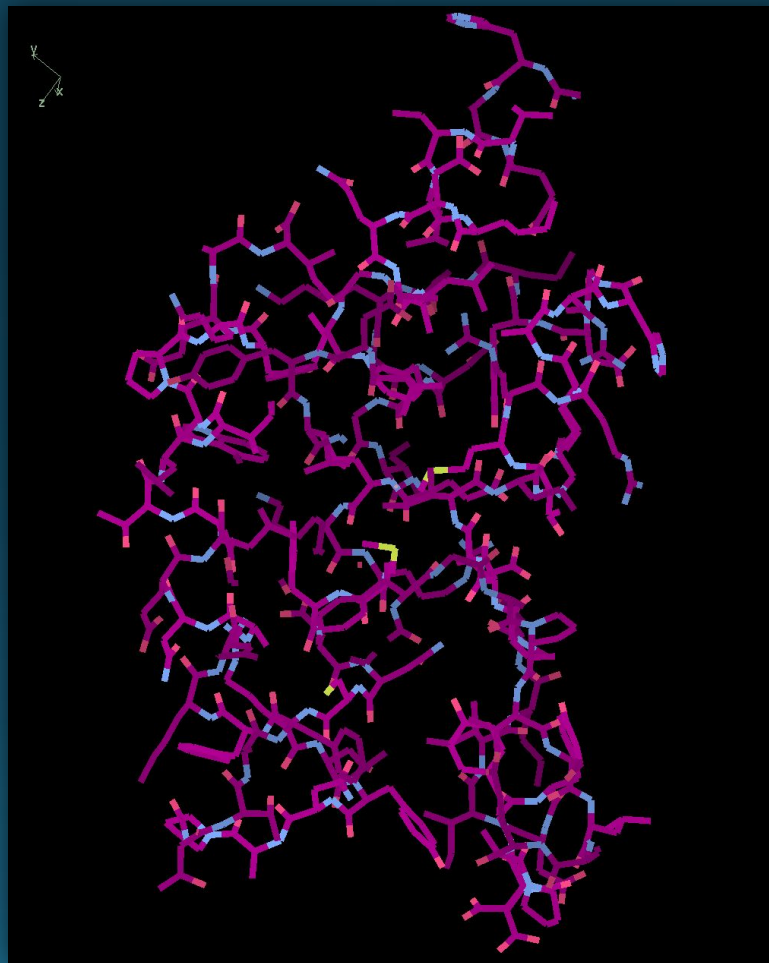
Step-by-Step MR: *Preparation of search model*

- Once a suitable homologue or set of homologues have been found they need to be prepared for use as search models
- The closer we can make the homologue to the target in terms of structural similarity the more likely it is to be successfully positioned in Molecular Replacement
- The sequence alignment generated in the search step can be used as a guide for the pruning of the homologue



Step-by-Step MR: *Preparation of search model*

- Example target:
 - 5TPX - Bromodomain from Plasmodium Faciparum Gcn5



Examine the data



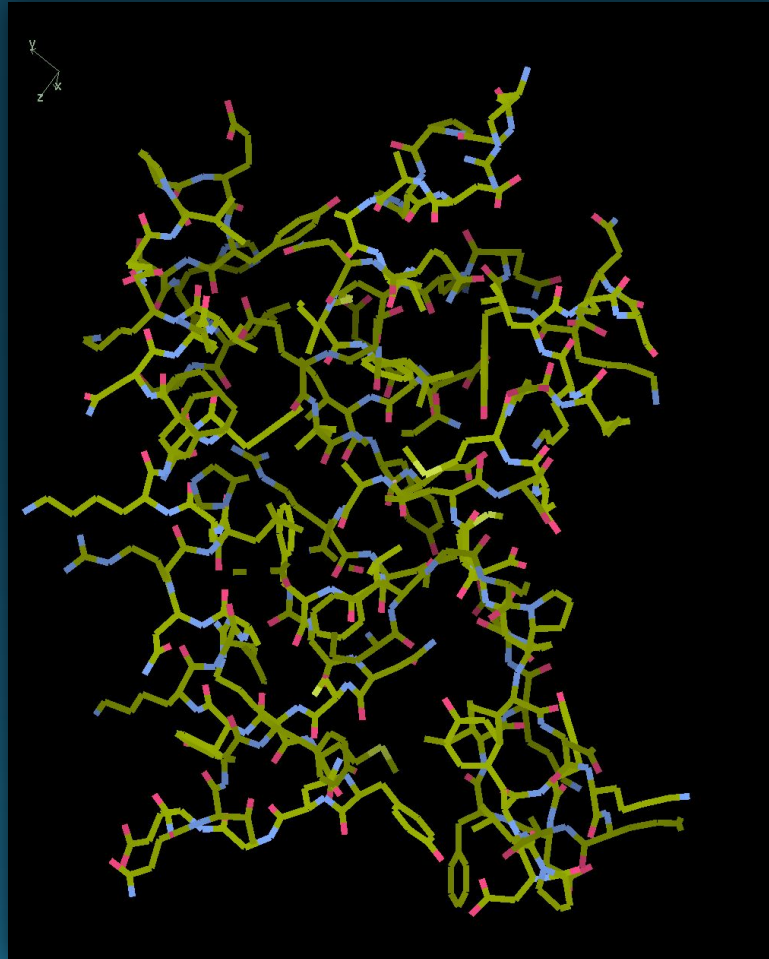
Find a search model



Preparation of search model

Step-by-Step MR: *Preparation of search model*

- Example target:
 - 5TPX - Bromodomain from Plasmodium Faciparum Gcn5
- Homologue:
 - 1E6I – chain A
 - 45% sequence identity over 82% coverage of target



Examine the data



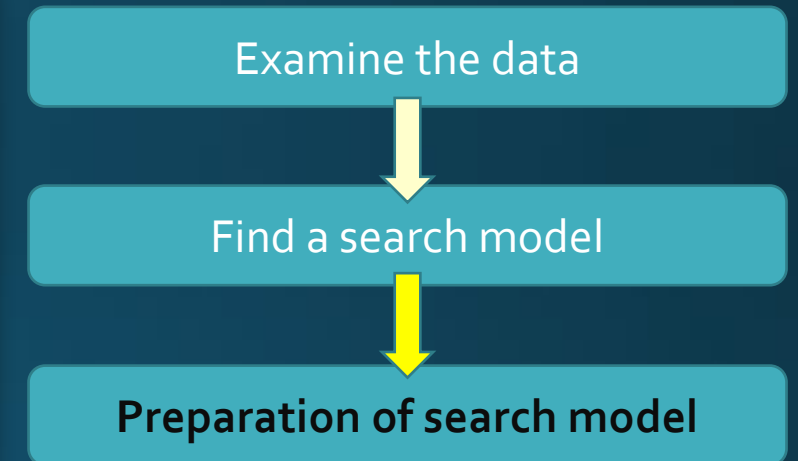
Find a search model



Preparation of search model

Step-by-Step MR: *Preparation of search model*

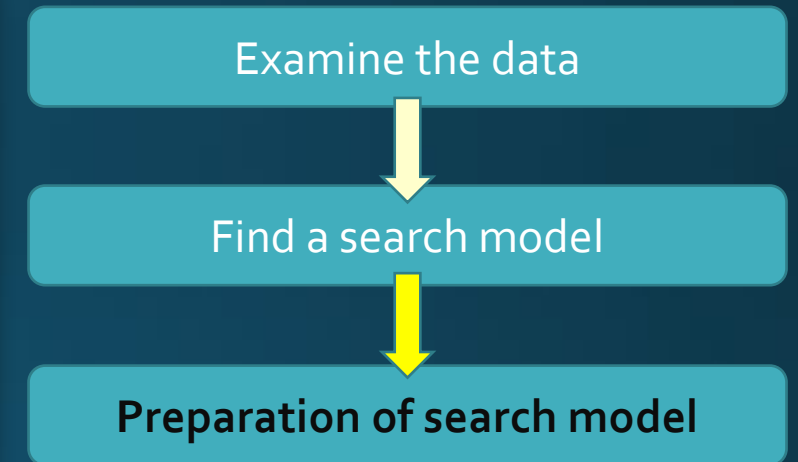
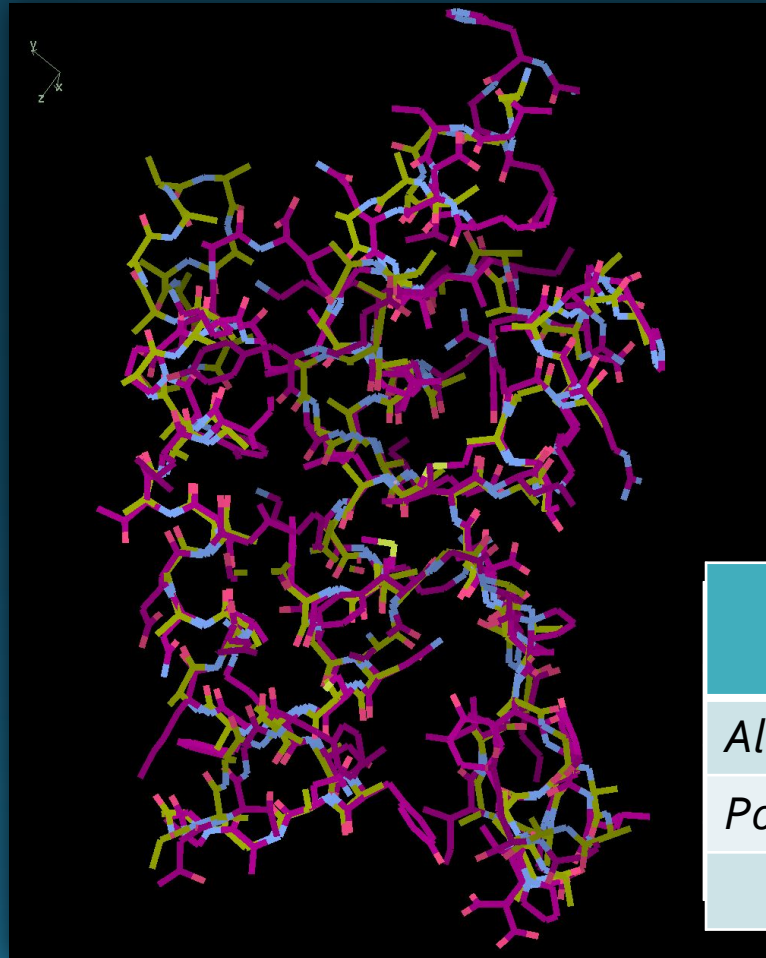
- Search Model preparation and MR:
 - All atoms?



	<i>Molrep</i> TF/sigma	<i>Refmac</i> Rfactor	<i>Refmac</i> Rfree
<i>All atoms</i>	6.82	0.418	0.481

Step-by-Step MR: *Preparation of search model*

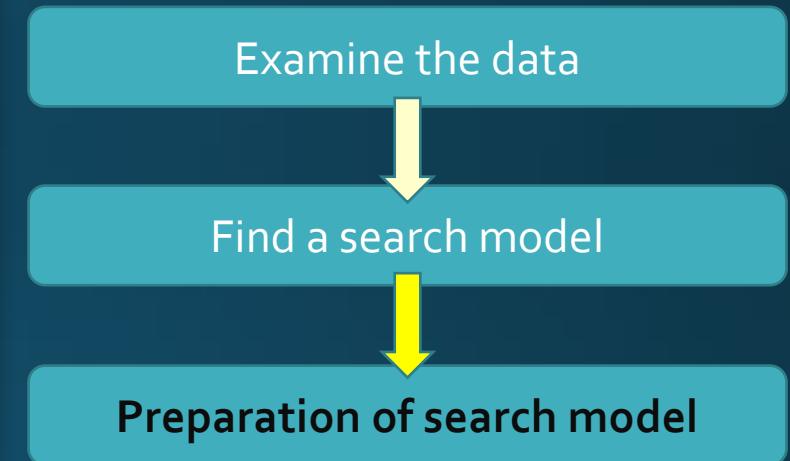
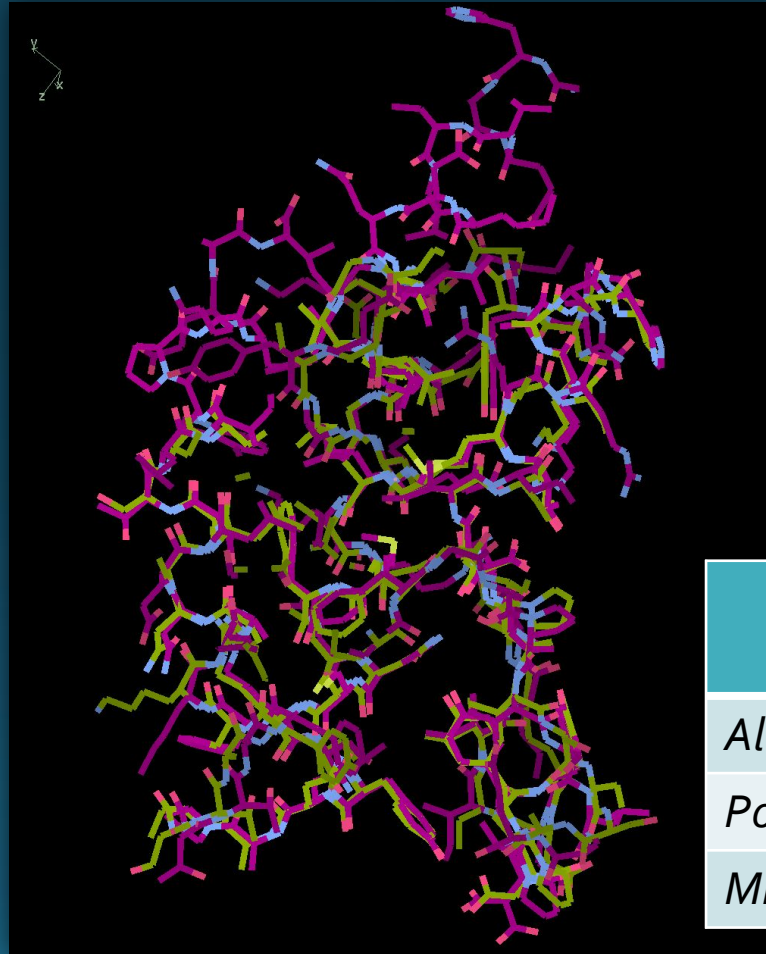
- Search Model preparation and MR:
 - All atoms?
 - Polyalanine?



	<i>Molrep</i> TF/sigma	<i>Refmac</i> Rfactor	<i>Refmac</i> Rfree
<i>All atoms</i>	6.82	0.418	0.481
<i>Poly</i>	4.98	0.46	0.482

Step-by-Step MR: *Preparation of search model*

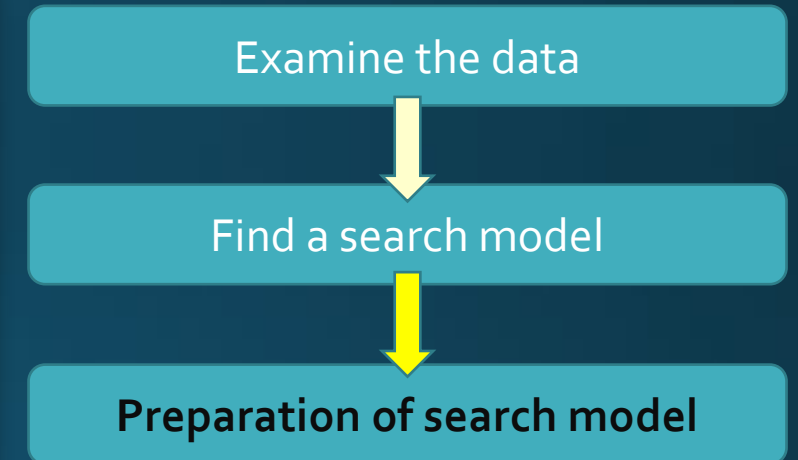
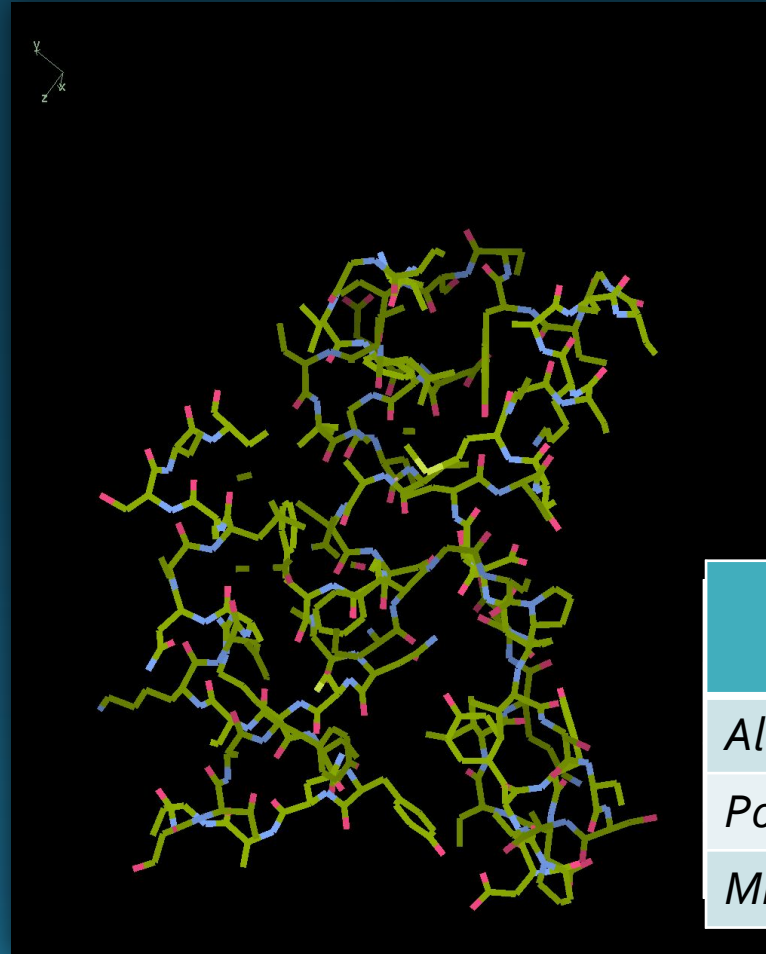
- Search Model preparation and MR:
 - All atoms?
 - Polyalanine?
 - Mixed model based on sequence alignment
 - Keep common residues
 - Truncate to common atoms on aligned residues



	<i>Molrep</i> TF/sigma	<i>Refmac</i> Rfactor	<i>Refmac</i> Rfree
<i>All atoms</i>	6.82	0.418	0.481
<i>Poly</i>	4.98	0.46	0.482
<i>Mixed</i>	10.27	0.379	0.418

Step-by-Step MR: *Preparation of search model*

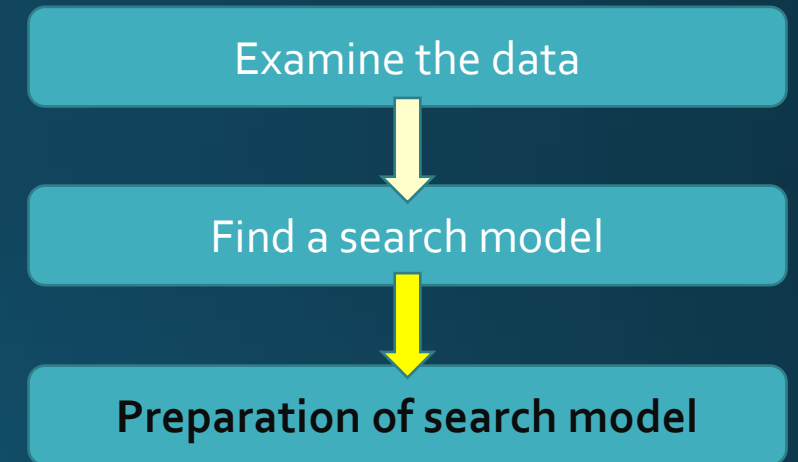
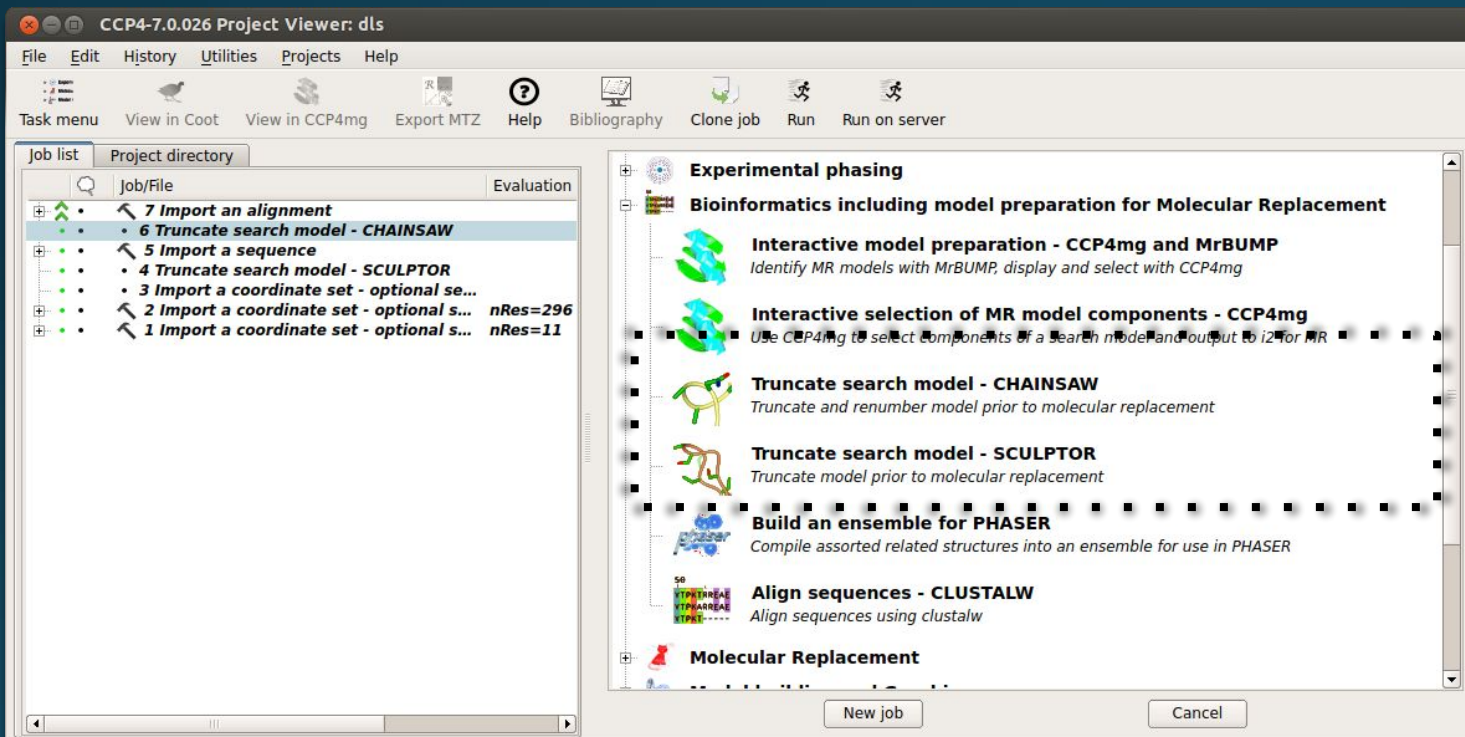
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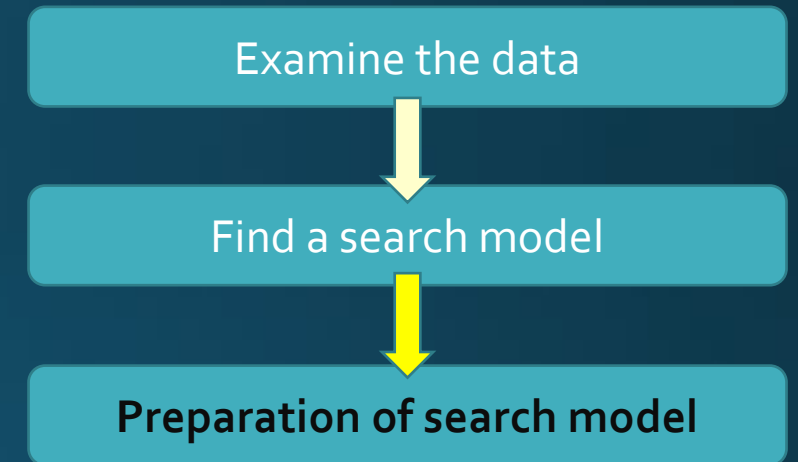
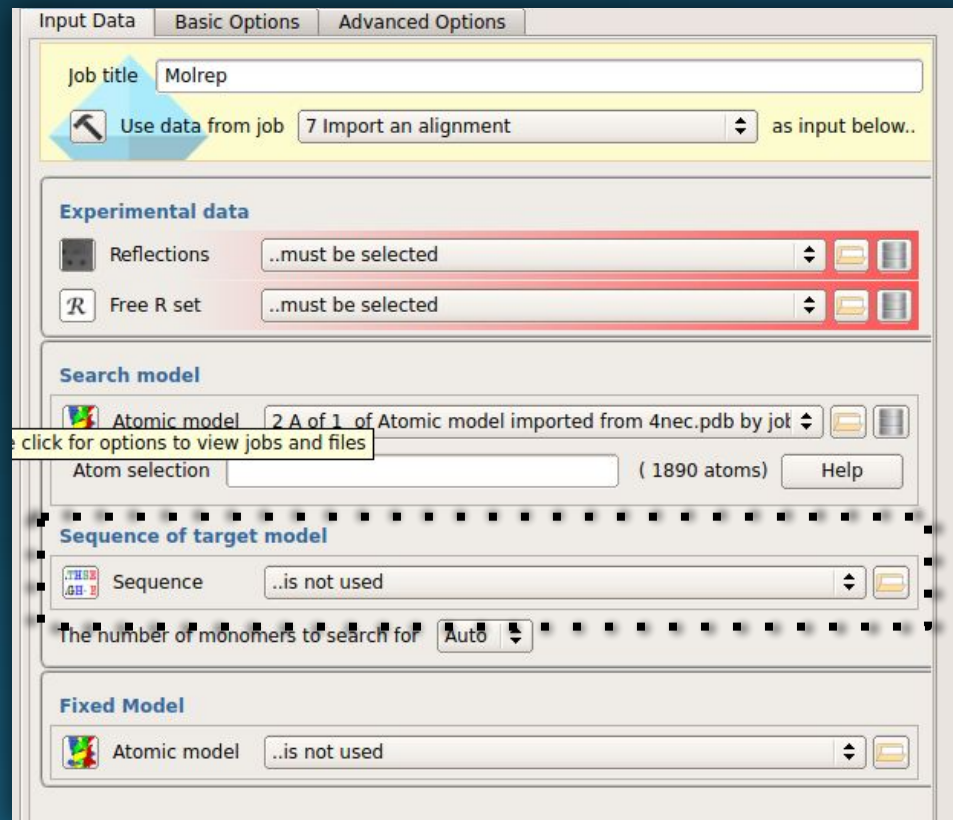
Step-by-Step MR: *Preparation of search model*

- Mixed-model generation in CCP4
- Chainsaw & Sculptor



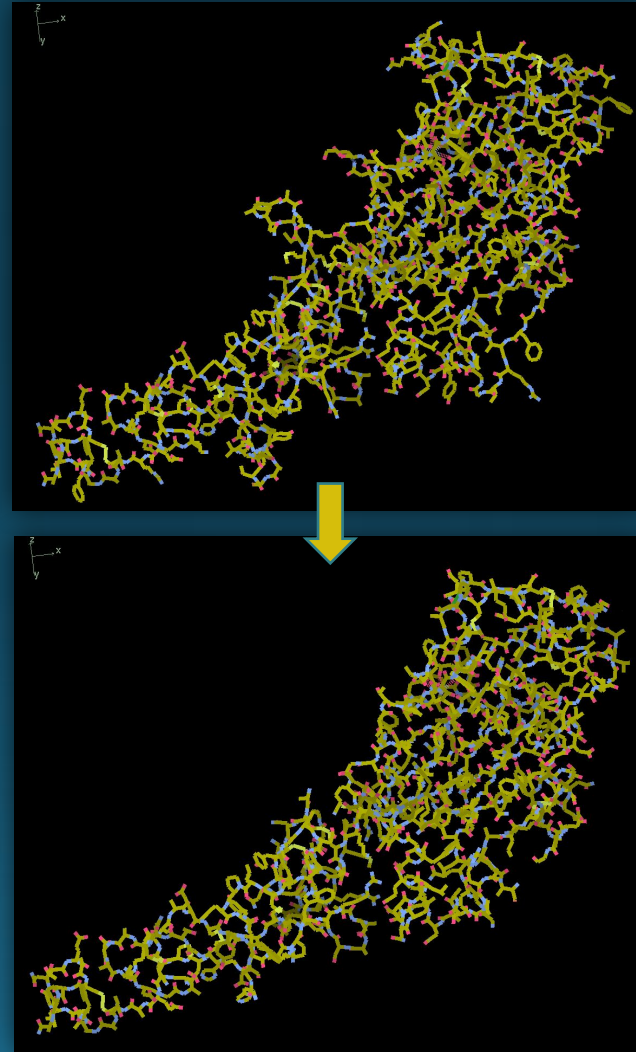
Step-by-Step MR: *Preparation of search model*

- Mixed-model generation in CCP4
 - Molrep



Step-by-Step MR: *Preparation of search model*

- Manual pruning in Coot
- Remove anything that is likely to be flexible:
 - External loops
 - Longer side chains on the surface of the molecule (e.g. LYS)
 - Domains if there is evidence of “hinge motion”



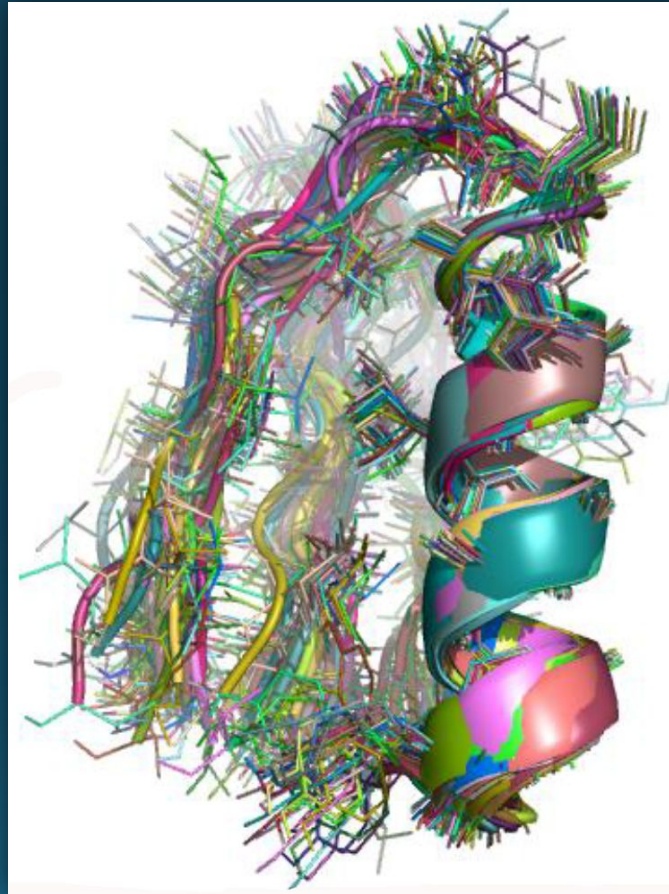
Examine the data

Find a search model

Preparation of search model

Step-by-Step MR: *Preparation of search model*

- Ensembles: alignment of search models



Examine the data



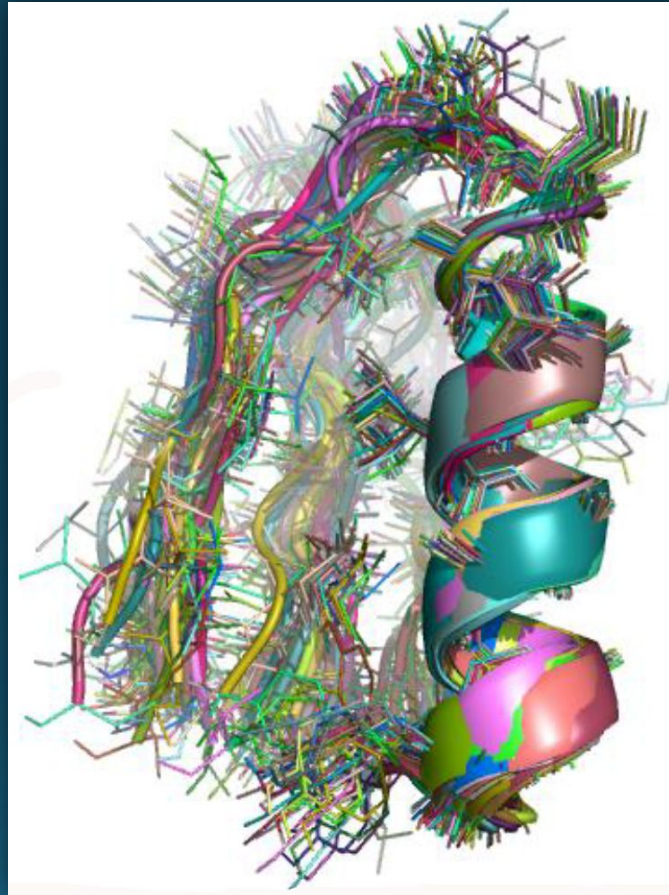
Find a search model



Preparation of search model

Step-by-Step MR: *Preparation of search model*

- Ensembles: alignment of search models
 1. Variance across members can guide experimental data weighting in Phaser



Examine the data



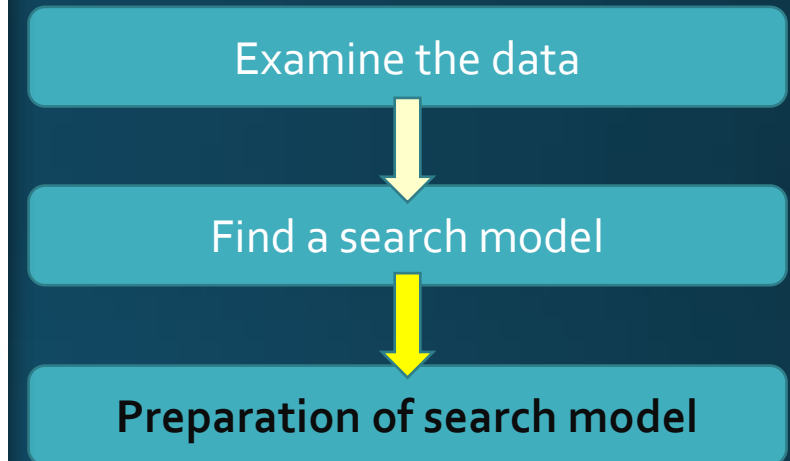
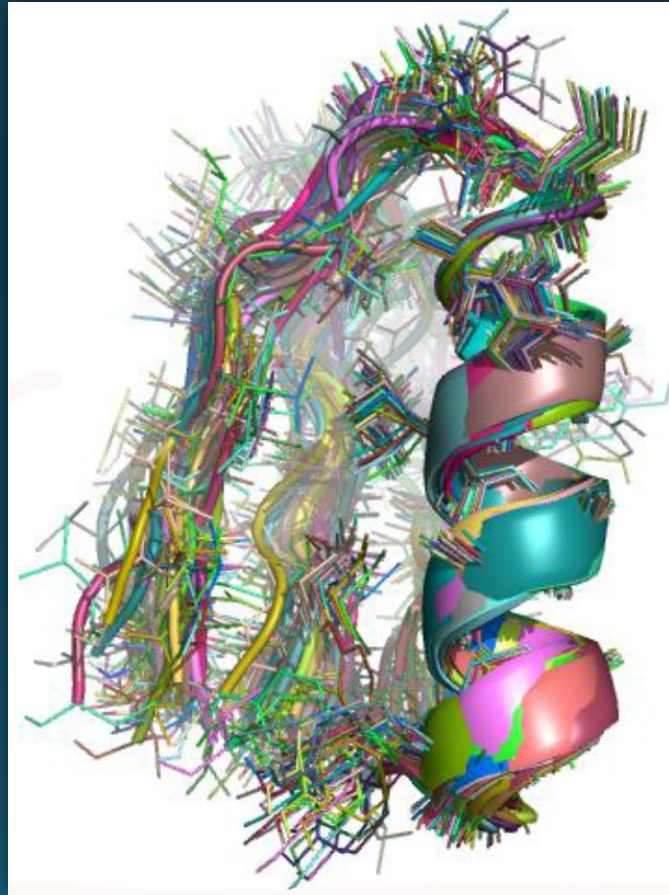
Find a search model



Preparation of search model

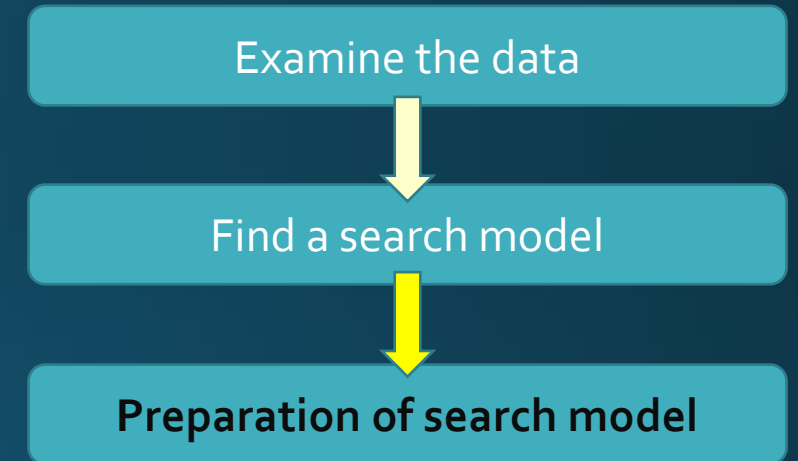
Step-by-Step MR: *Preparation of search model*

- Ensembles: alignment of search models
 1. Variance across members can guide experimental data weighting in Phaser
 2. Truncation based on alignment variance can identify conserved regions or core



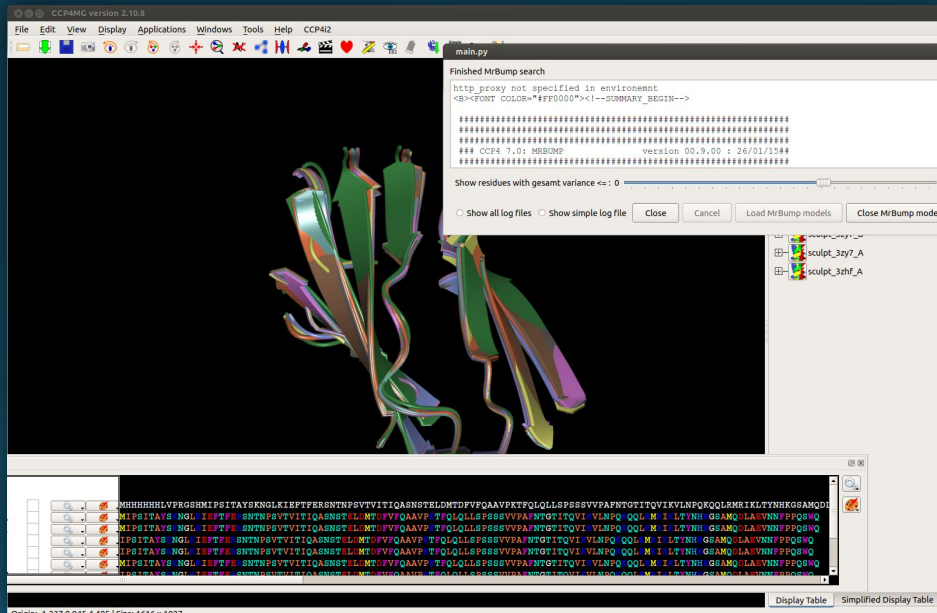
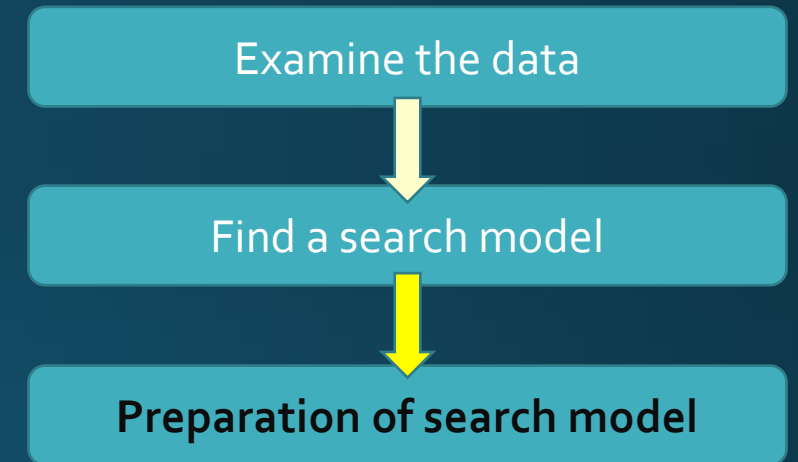
Step-by-Step MR: *Preparation of search model*

- Ensembles: Generating ensembles in CCP4
 - Ensembler:
 - Align models and truncate based on a variance threshold
 - General alignment tools:
 - Gesamt
 - Superpose
 - Can also be run through CCP4mg and Coot graphical interfaces



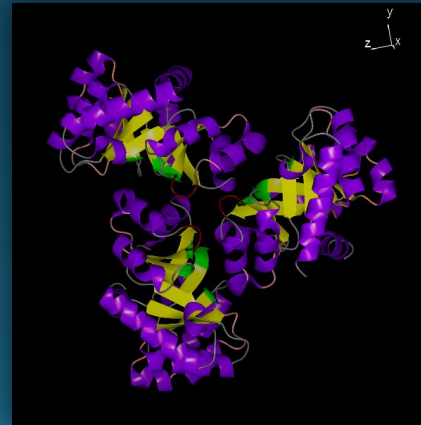
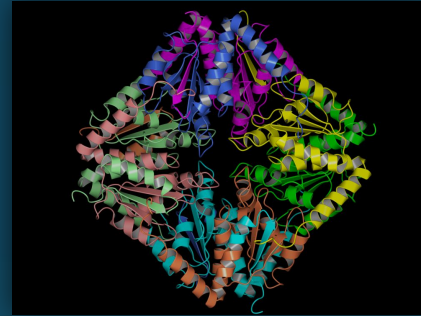
Step-by-Step MR: *Preparation of search model*

- Ensembles: CCP4mg & MrBUMP
 - Finds homologues using Phmmer
 - Runs Chainsaw to prune and create search models
 - Aligns models using Gesamt
 - Truncation tool to adjust variance threshold



Step-by-Step MR: *Preparation of search model*

- Multimers as search models
 - A single chain search model can be too small if the target has crystallised in multimeric form
 - The signal for the correct position is too weak against the background noise of incorrect positions
 - Particularly a problem at lower resolutions and crystals with high symmetry



Examine the data



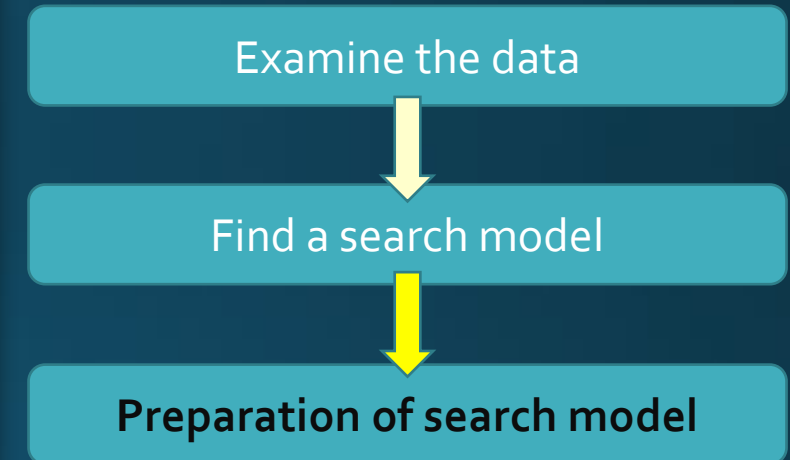
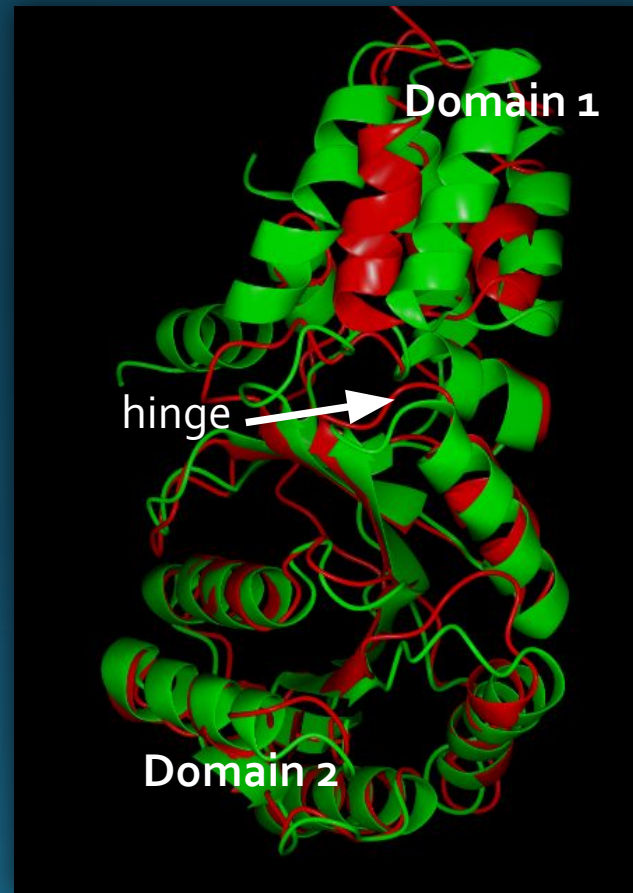
Find a search model



Preparation of search model

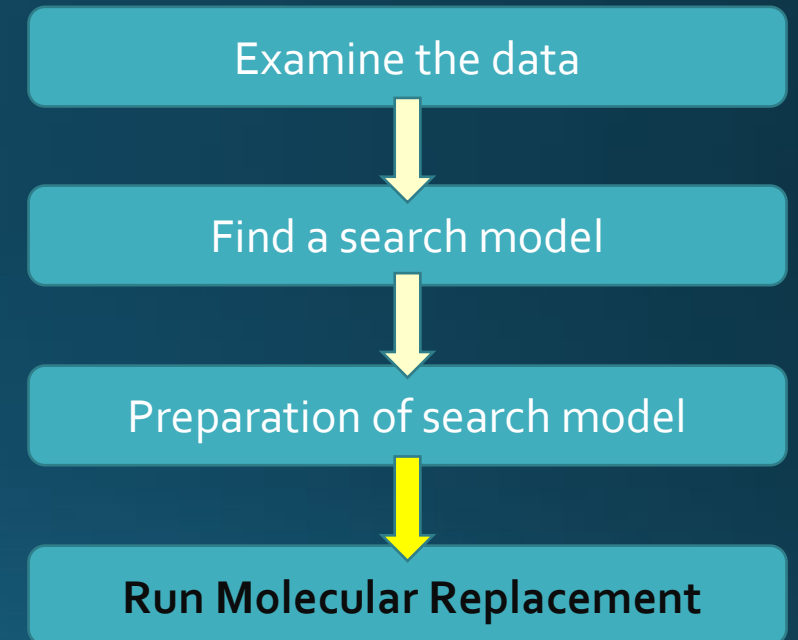
Step-by-Step MR: *Preparation of search model*

- Domains as search models
 - A hinge motion may alter the relative orientation of domains within a chain
- Domain models should be isolated from the parent search model and used separately in MR



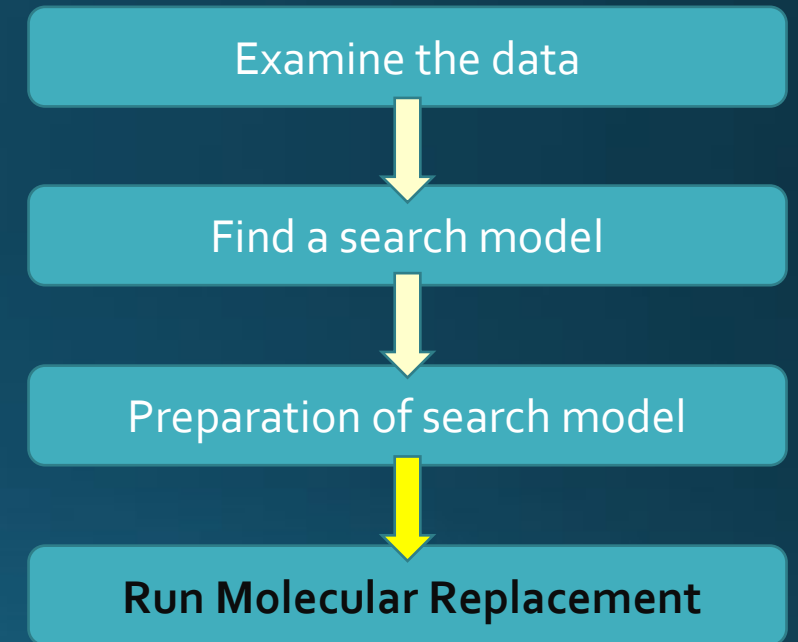
Step-by-Step MR: *Run Molecular Replacement*

- CCP4 has several programs for doing Molecular Replacement
 - Amore
 - Manual steps but very fast
 - Molrep
 - Automated MR
 - Several useful features e.g. searching a map
 - Phaser
 - Maximum likelihood approach
 - Accounts for potential model errors
 - Best for difficult cases and for correctly positioning fragment search models



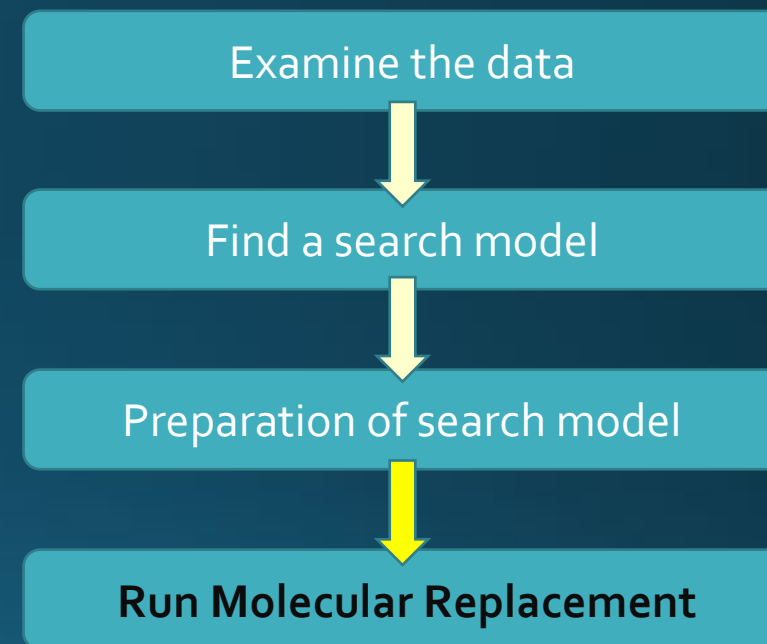
Step-by-Step MR: *Run Molecular Replacement*

- Important points on using Phaser



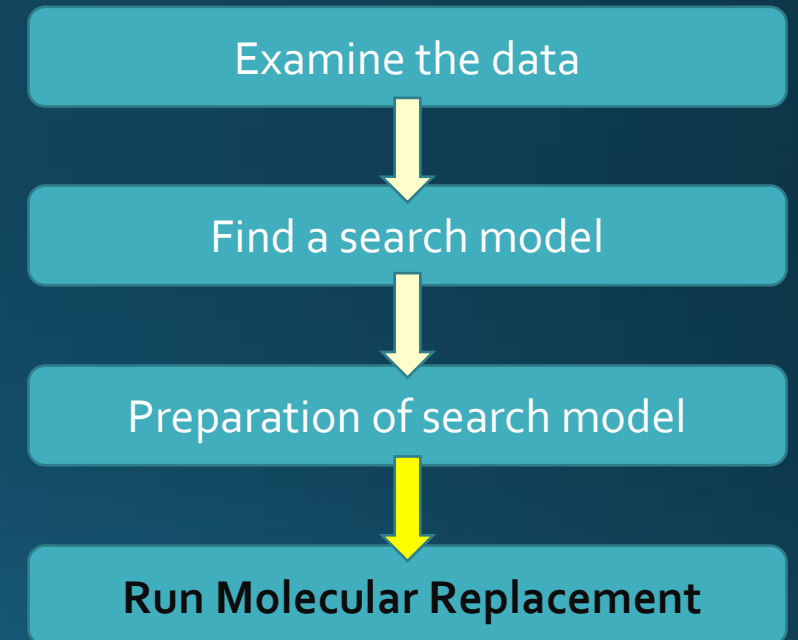
Step-by-Step MR: *Run Molecular Replacement*

- Important points on using Phaser
 - Phaser accounts for errors in:



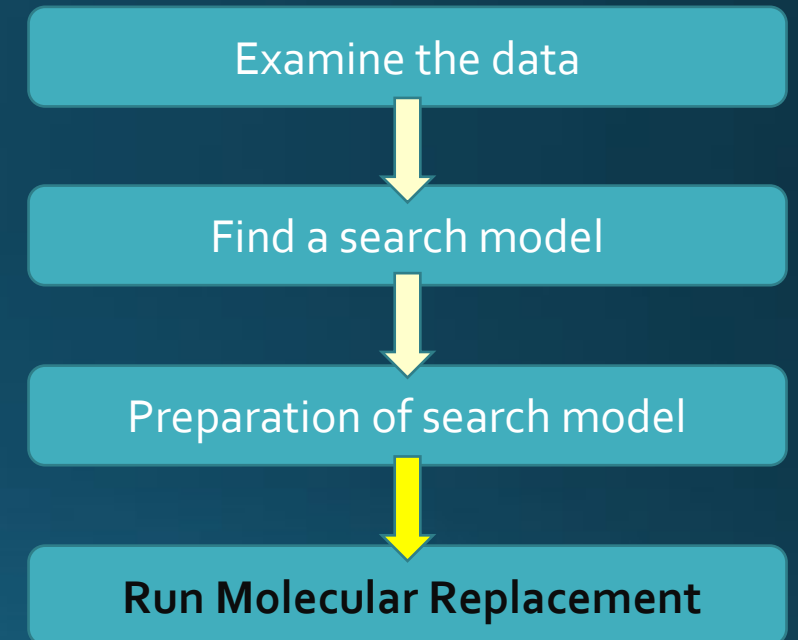
Step-by-Step MR: *Run Molecular Replacement*

- Important points on using Phaser
 - Phaser accounts for errors in:
 1. Model
 - Provide accurate details of AU composition
 - Use RMS value rather than sequence identity and try different values if first attempt doesn't work



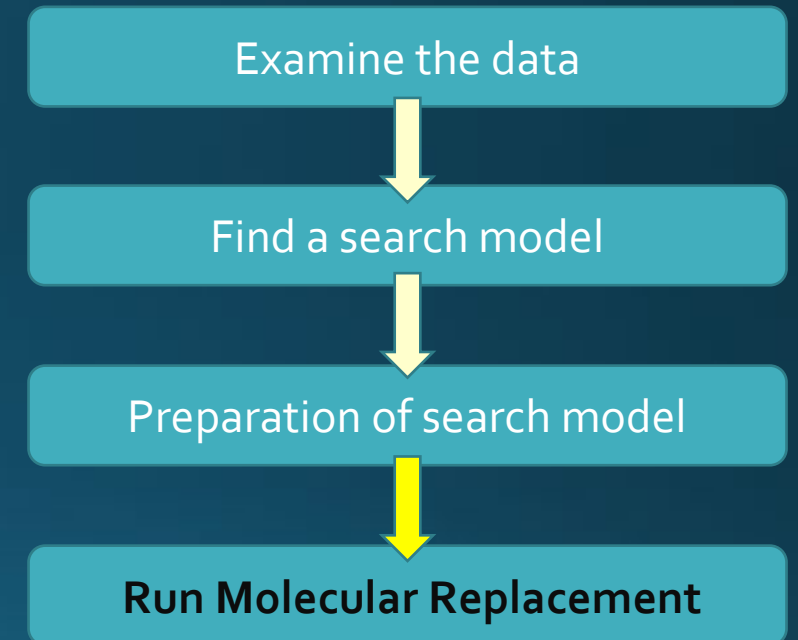
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 2. Data
 - Provide intensities – internally works out amplitudes accounting for experimental errors

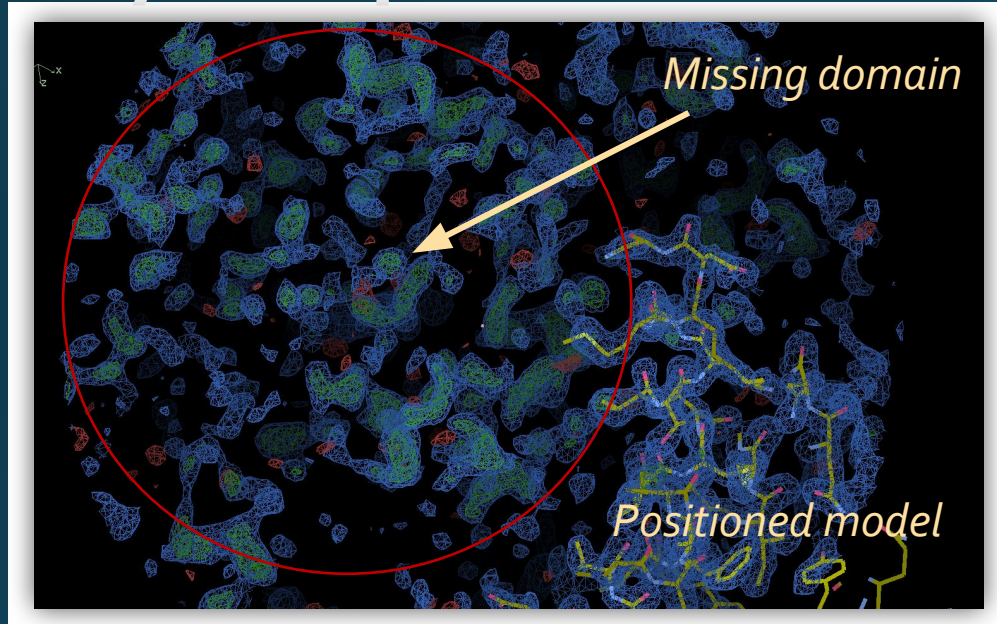


Step-by-Step MR: *Run Molecular Replacement*

- Important points on using Phaser
 - Phaser accounts for errors in:
 1. Model
 - Provide accurate details of AU composition
 - Use RMS value rather than sequence identity and try different values if first attempt doesn't work
 2. Data
 - Provide intensities – internally works out amplitudes accounting for experimental errors
 - Phaser performs clever decision making for automation
 - Provide minimal details and let Phaser make it's own decisions e.g. search order, search all possible space groups
 - If it doesn't work take step-by-step approach – 1 copy at a time

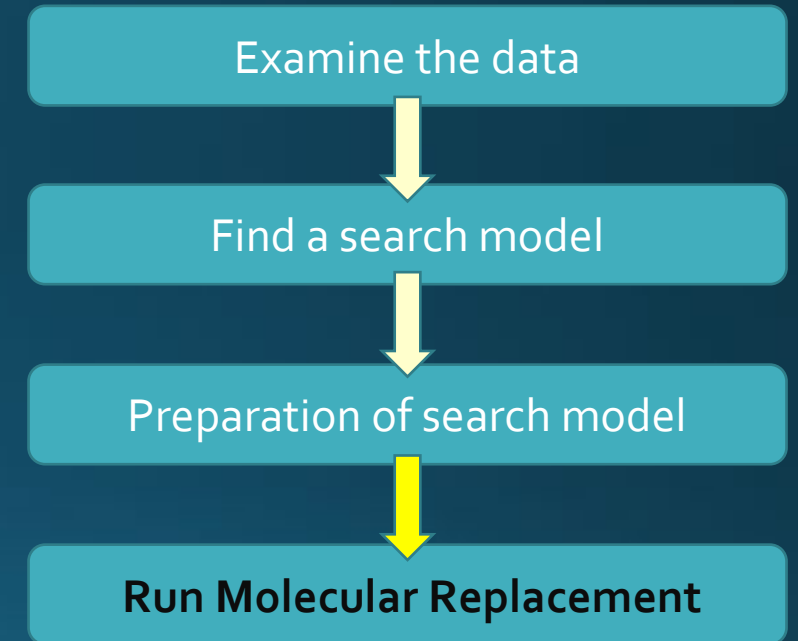


Step-by-Step MR: *Run Molecular Replacement*



- Phased translation search

- Used when searching for several copies or dealing with a complex
- Available through MOLREP (3 protocols) and Phaser
- Often more successful than standard MR search approach particularly when looking for small domains



Step-by-Step MR: *Run Molecular Replacement*

- Phased translation search
 - From MOLREP (CCP4i):
 - 3 Protocols for phased search - Try all!

The screenshot shows the 'Molrep Initial parameters' dialog box. The title bar indicates it's from a file in the CCP4 database. The 'Job title' is 'fit model to 4p9g'. The 'Do' button is set to 'MR Using Phases'. Under 'Use MAP files for', both 'observed data' and 'search model' are unchecked. The 'Data' section shows the full path to '4p9g.mtz'. The 'Model' section shows 'trun_7_molrep1.pdb'. The 'Sequence' and 'Fixed' sections are empty. The 'Solution' section shows 'trun_7_molrep4.pdb'. The 'Search Options' section shows 'SAPTF + Local Phased RF + Phased TF' as the search protocol, with 'Number of copies to find' set to 1. The 'Pseudo-Translation' is set to 'Auto'. The 'Experimental Data', 'Model', and 'Infrequently used options' sections are collapsed. At the bottom are 'Run', 'Save or Restore', and 'Close' buttons.

Examine the data

Find a search model

Preparation of search model

Run Molecular Replacement

Step-by-Step MR: *Run Molecular Replacement*

- Phased translation search
 - From Phaser (CCP4i):

The screenshot shows the 'Maximum Likelihood Molecular Replacement Initial parameters' dialog box. The 'Define ensembles (models)' section is active, showing two ensembles. Ensemble #1 is named 'ensemble1' and uses 'pdb file(s)' with the path '/home/rmk65/Projects/ample/concord/concord_13_merge'. Ensemble #2 is named 'ensemble2' and uses 'pdb file(s)' with the path '/home/rmk65/Projects/ample/concord/concord_13_merge'. The 'Search parameters' section shows 'Perform search using' set to 'ensemble2' and 'Number of copies to search for' set to '1'. The 'Additional Search parameters' section shows 'Allow search with alternative ensembles (models) for a single component of ASU' set to 'off', 'Define any known partial structure' set to 'Fix input coordinates', and 'Translation function search target' set to 'Phased'. The 'User parameters' section is also visible.

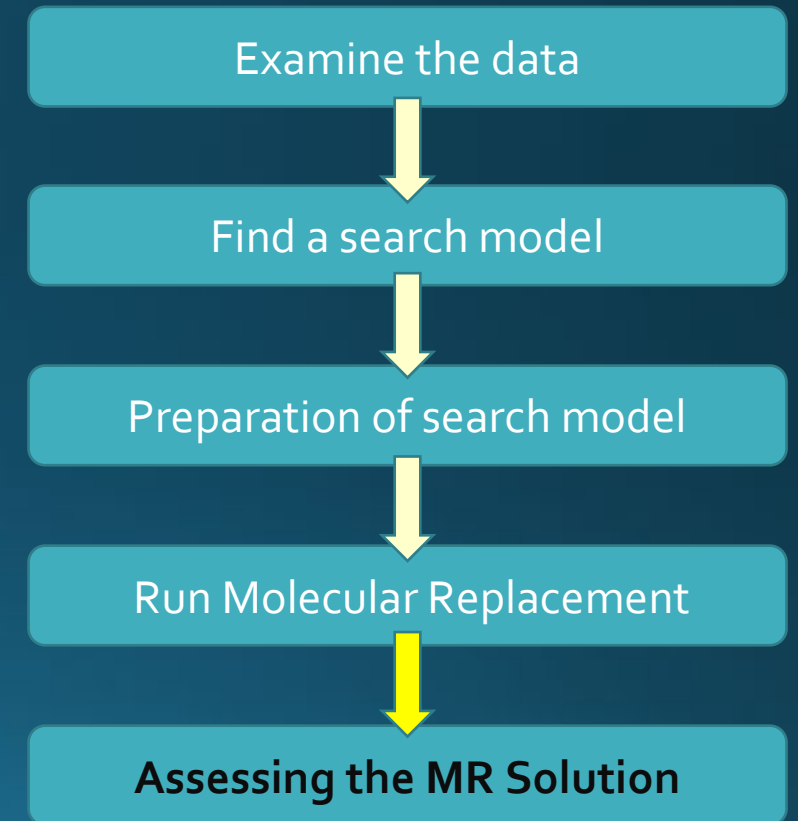
Examine the data

Find a search model

Preparation of search model

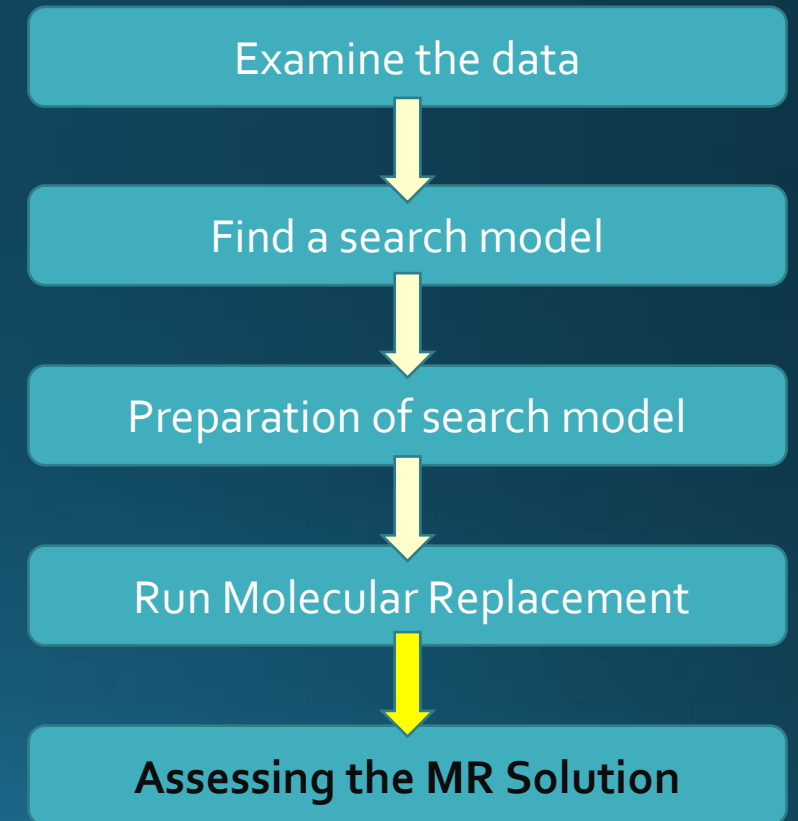
Run Molecular Replacement

Step-by-Step MR: *Assessing the MR solution*



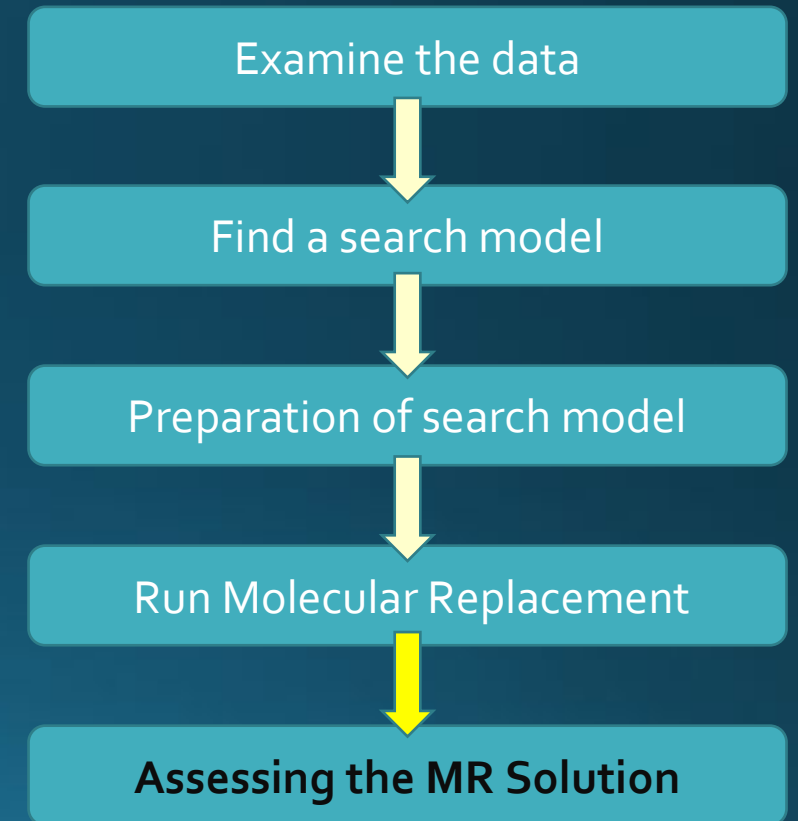
Step-by-Step MR: *Assessing the MR solution*

- How to know if the search model is correctly positioned in the target unit cell



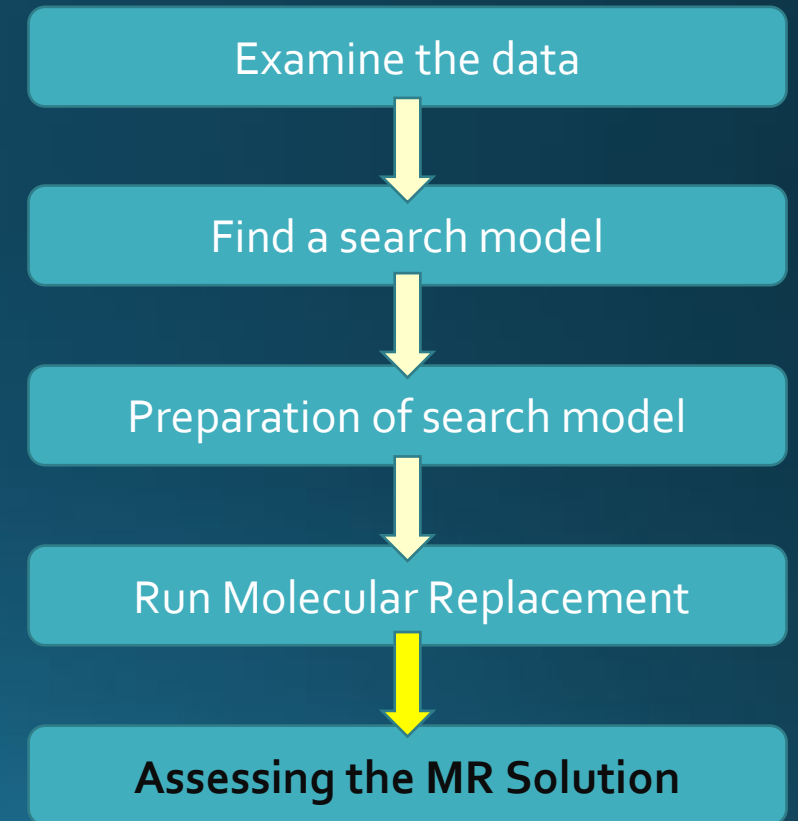
Step-by-Step MR: *Assessing the MR solution*

- How to know if the search model is correctly positioned in the target unit cell
- In difficult cases the position may be correct but getting from the MR solution to a complete model may not be straight forward



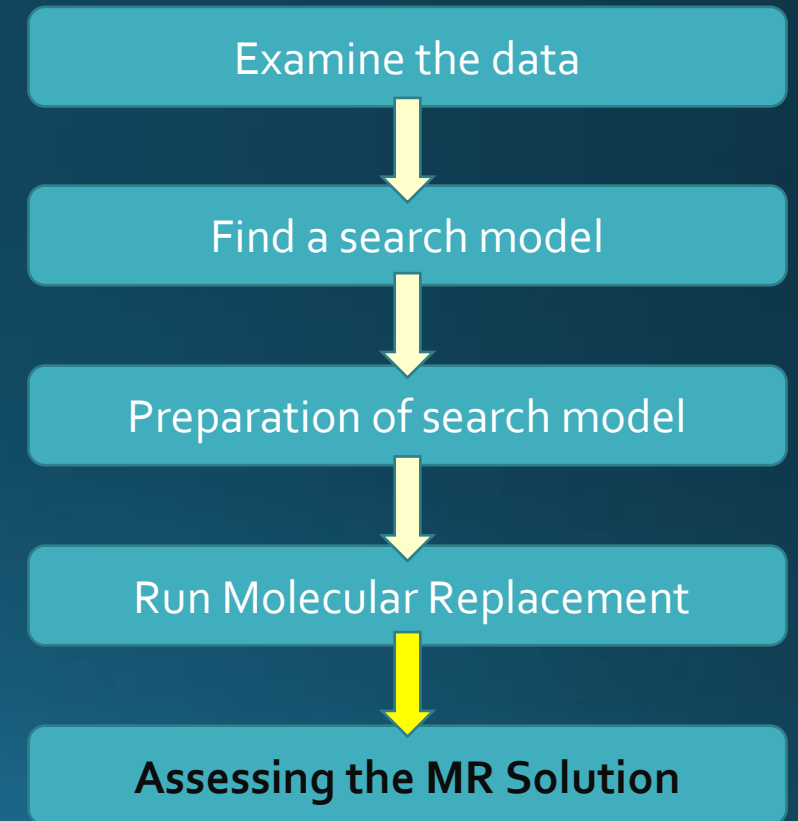
Step-by-Step MR: *Assessing the MR solution*

- Rough guide to MR program scoring



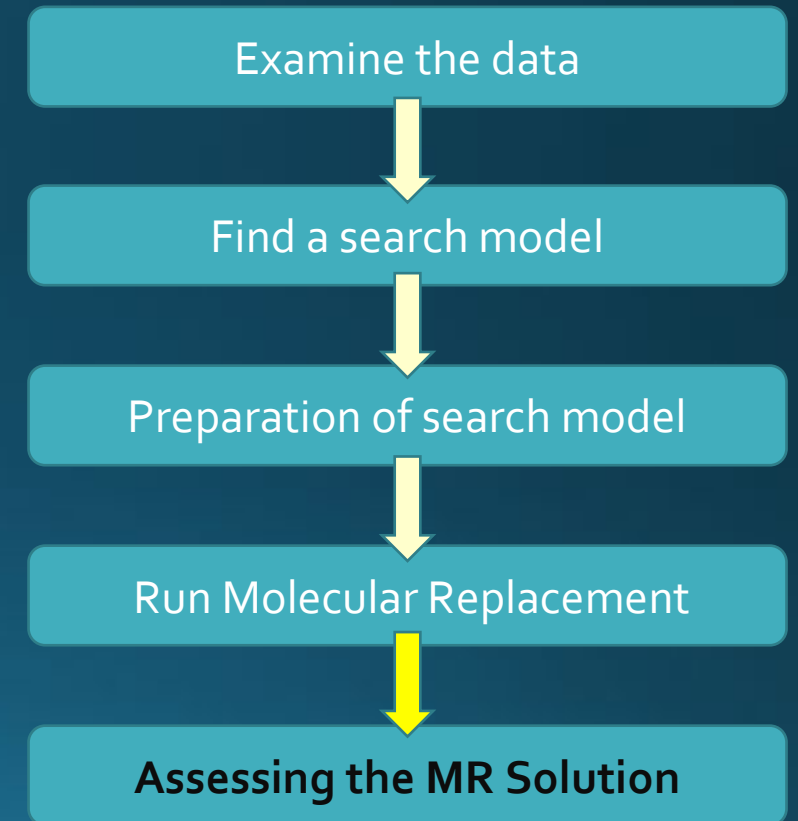
Step-by-Step MR: *Assessing the MR solution*

- Rough guide to MR program scoring
 - Phaser scores
 - LLG scores – has it increased by 60 or more after the placement of a new molecule?
(resolution and space group dependent)
 - TFZ – greater than 8?
 - Few or single solution almost always indicative of success



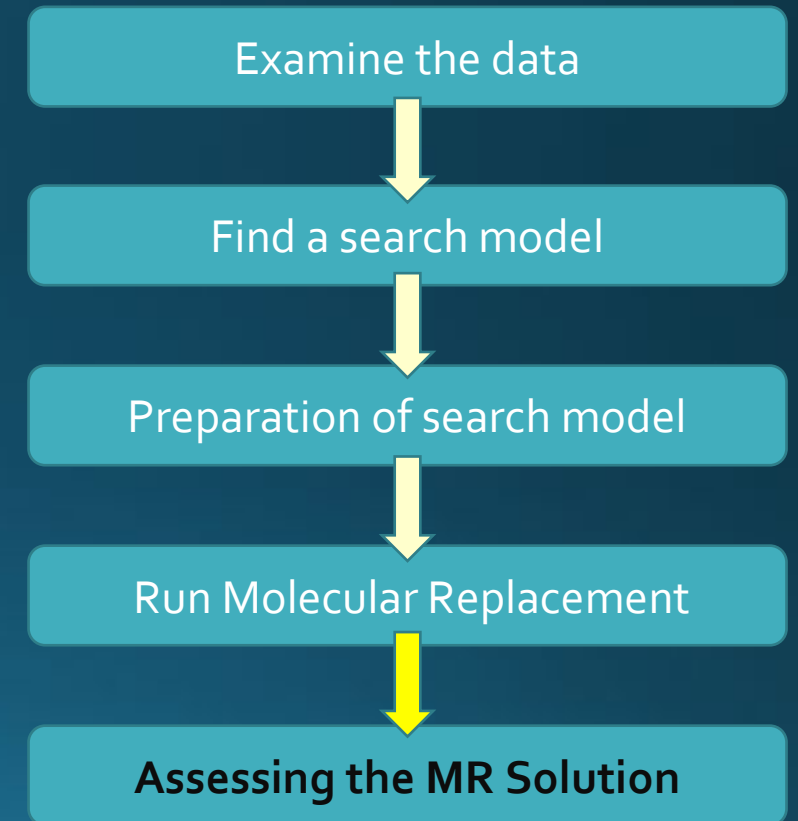
Step-by-Step MR: *Assessing the MR solution*

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 - Phaser scores
 - LLG scores – has it increased by 60 or more after the placement of a new molecule?
(resolution and space group dependent)
 - TFZ – greater than 8?
 - Few or single solution almost always indicative of success
 - Molrep scores
 - RFZ – rotation search score greater than 5 – is there a clear peak?
 - TFZ – translation search score – is there a clear peak?



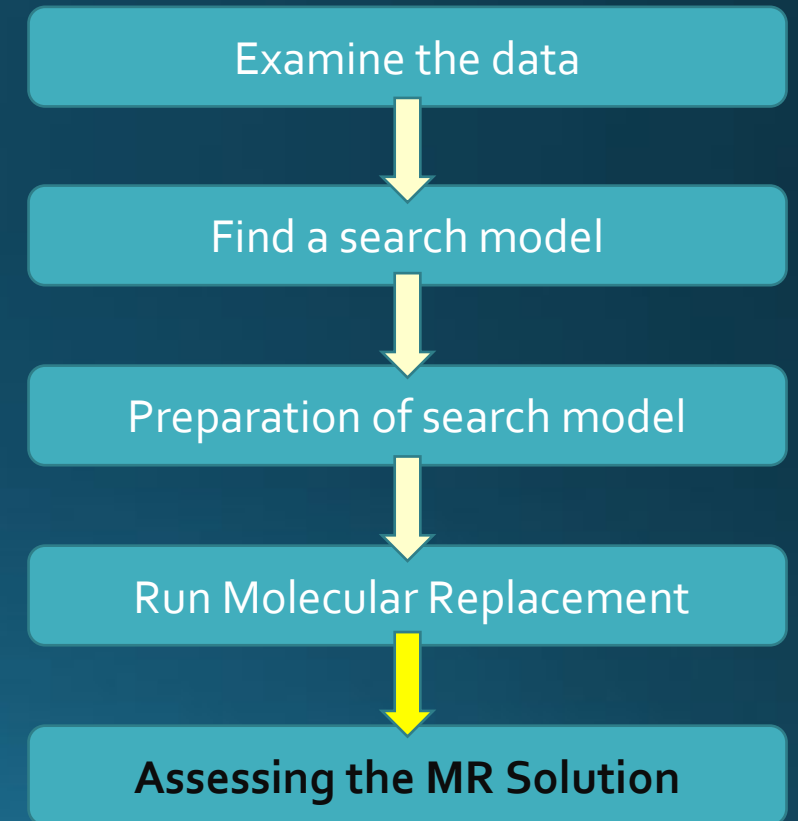
Step-by-Step MR: *Assessing the MR solution*

- Refinement



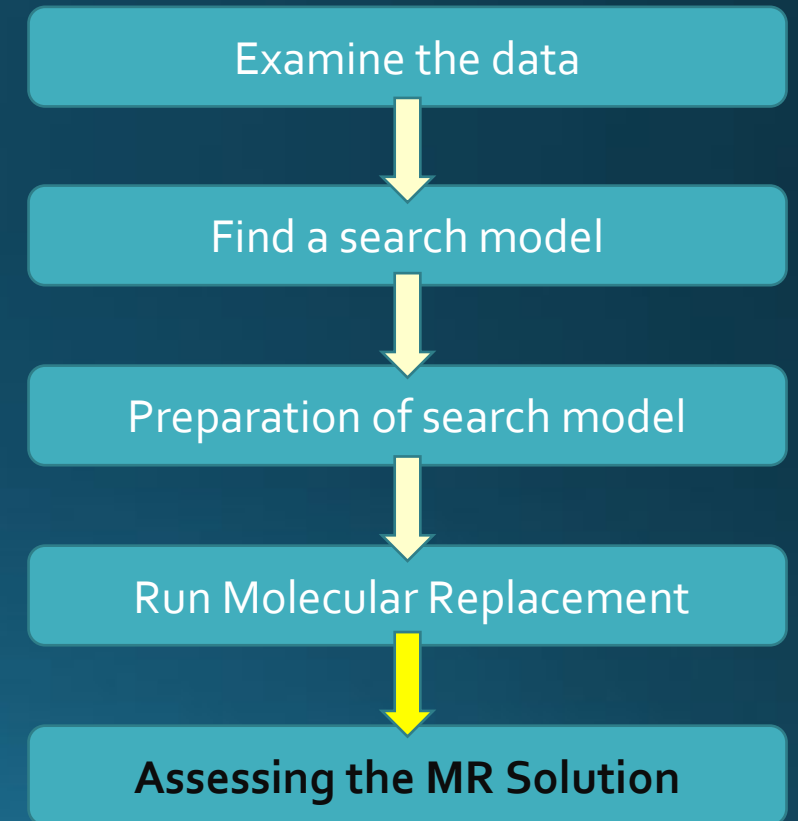
Step-by-Step MR: *Assessing the MR solution*

- Refinement
 - Look at Rfactor/Rfree
 - are they falling? Is Rfree below 0.5?



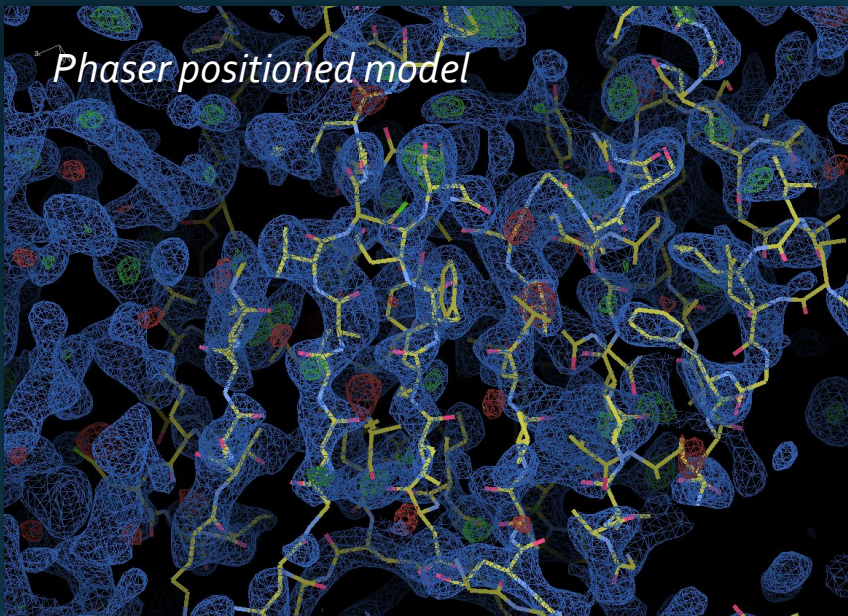
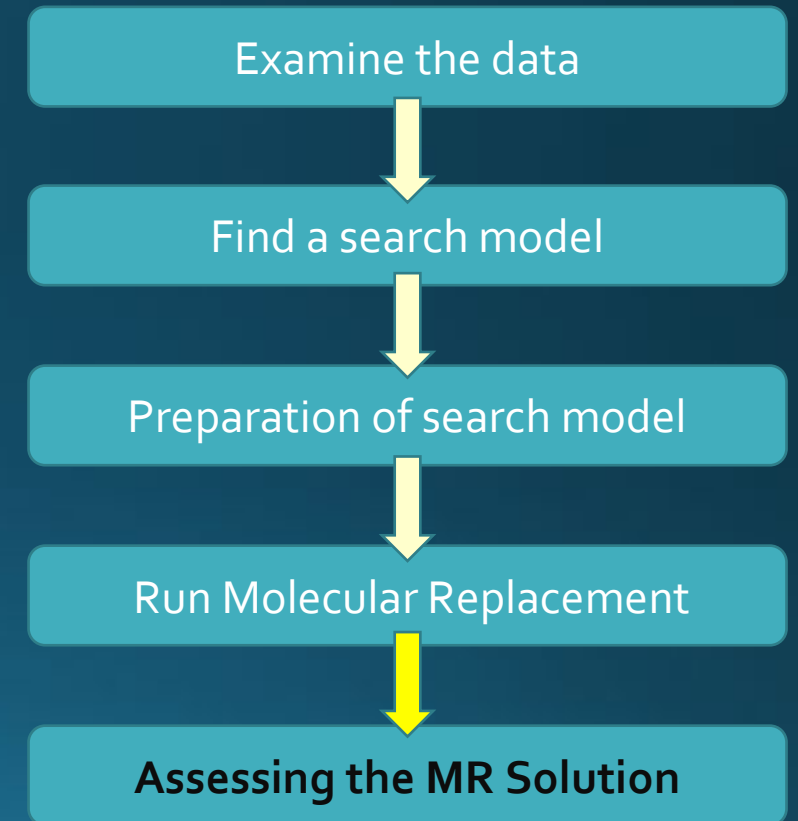
Step-by-Step MR: *Assessing the MR solution*

- Refinement
 - Look at Rfactor/Rfree
 - are they falling? Is Rfree below 0.5?
 - Use 200 cycles of jelly body refinement option in Refmac post MR



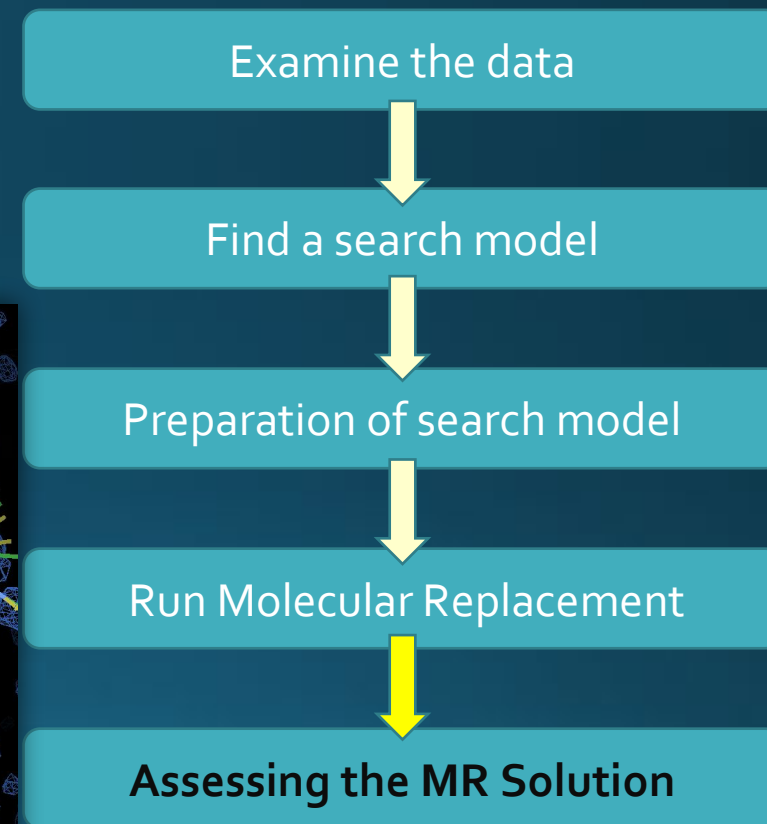
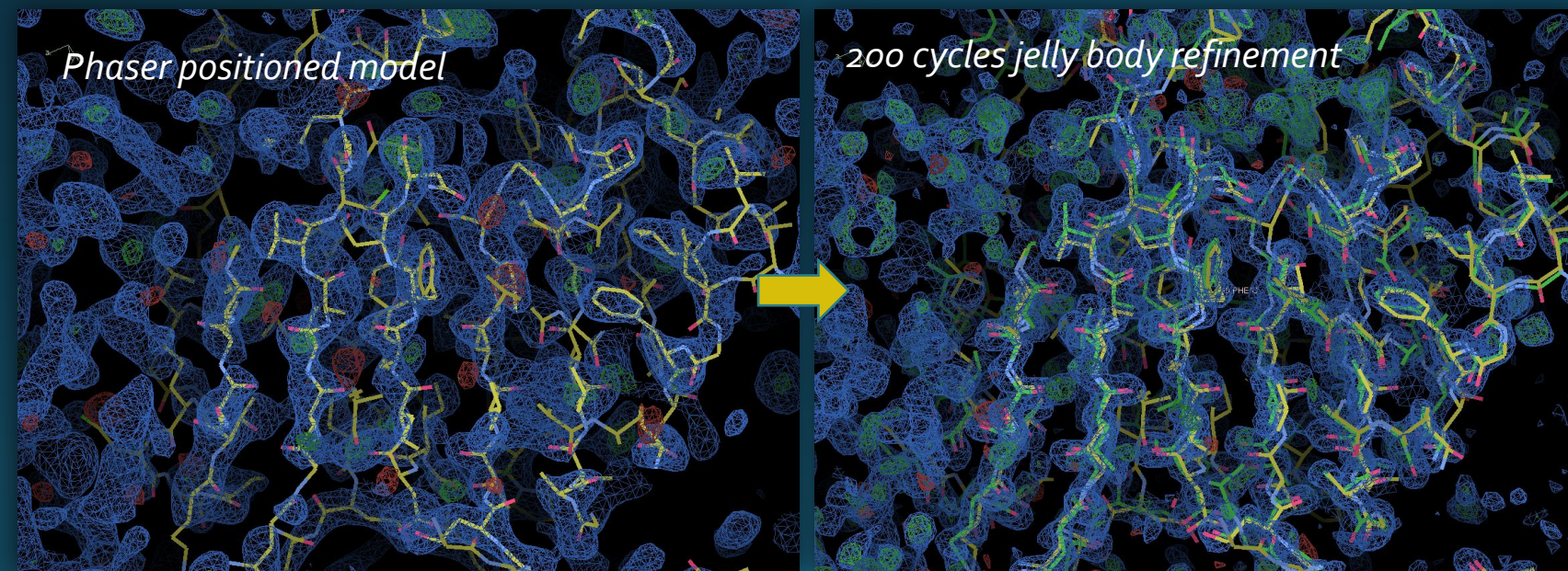
Step-by-Step MR: *Assessing the MR solution*

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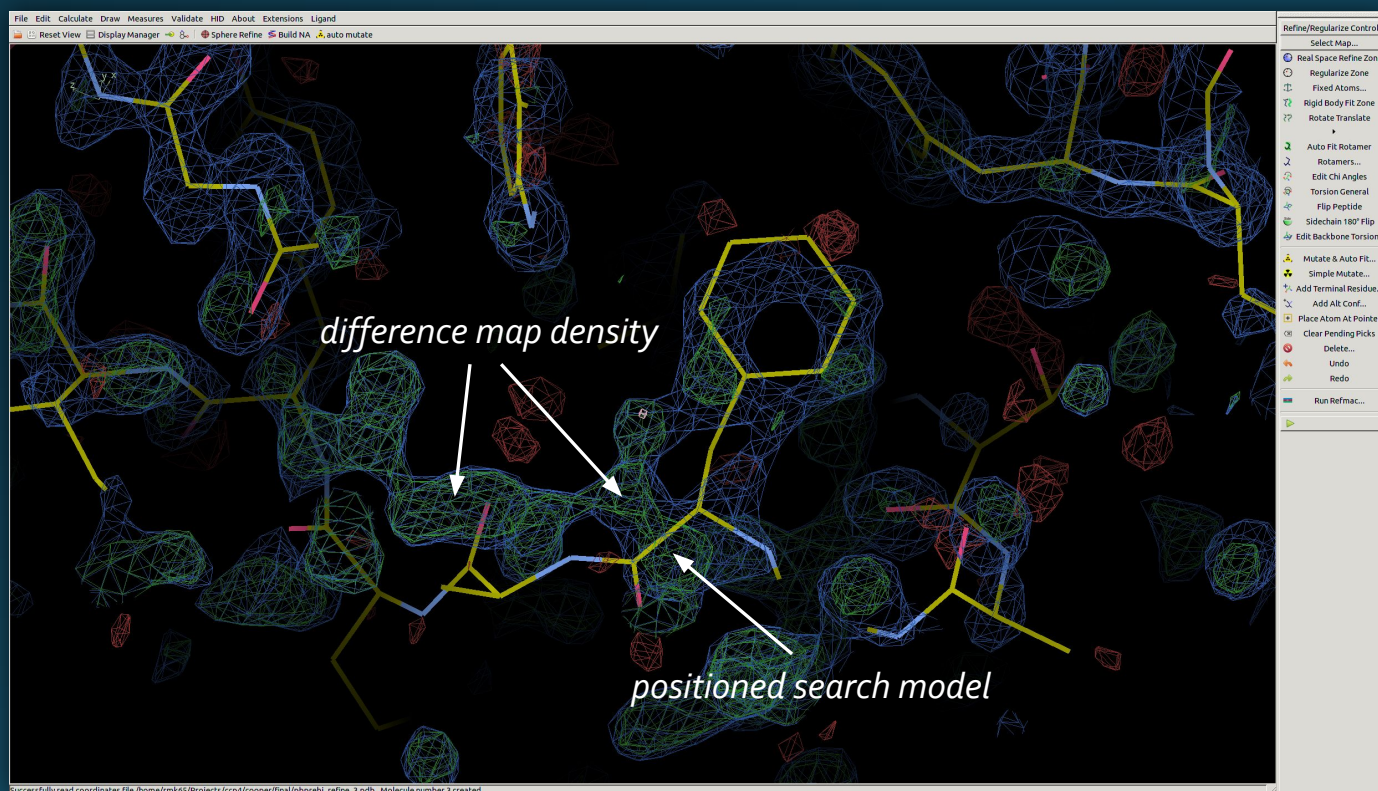
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- Refinement
 - Look at Rfactor/Rfree
 - are they falling? Is Rfree below 0.5?
 - Use 200 cycles of jelly body refinement option in Refmac post MR



Step-by-Step MR: *Assessing the MR solution*

- Examine solution by eye
 - Use Coot to examine positioned models & maps



Examine the data

Find a search model

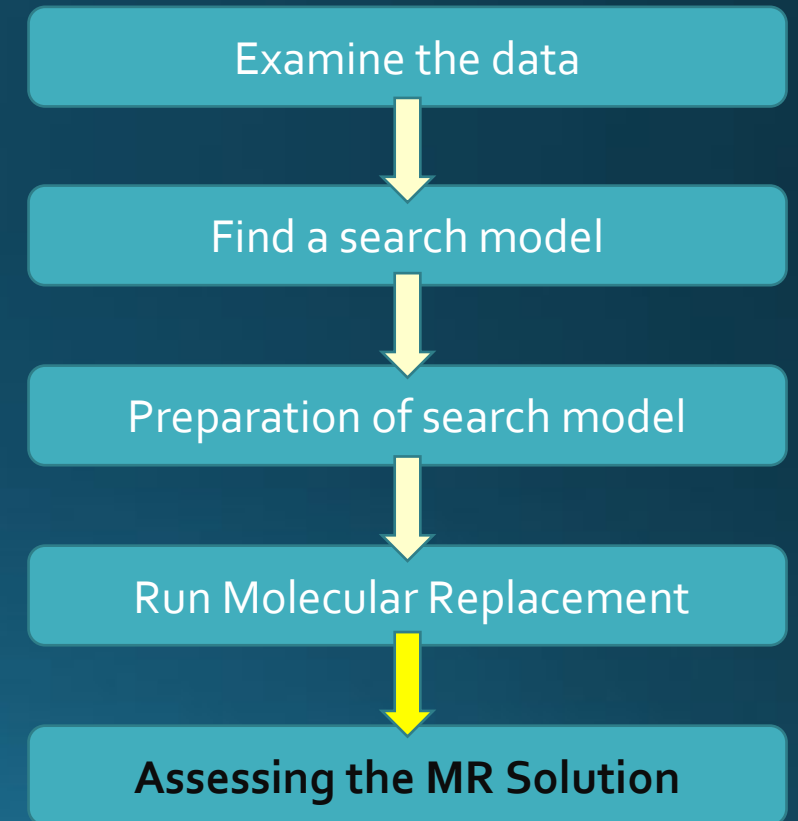
Preparation of search model

Run Molecular Replacement

Assessing the MR Solution

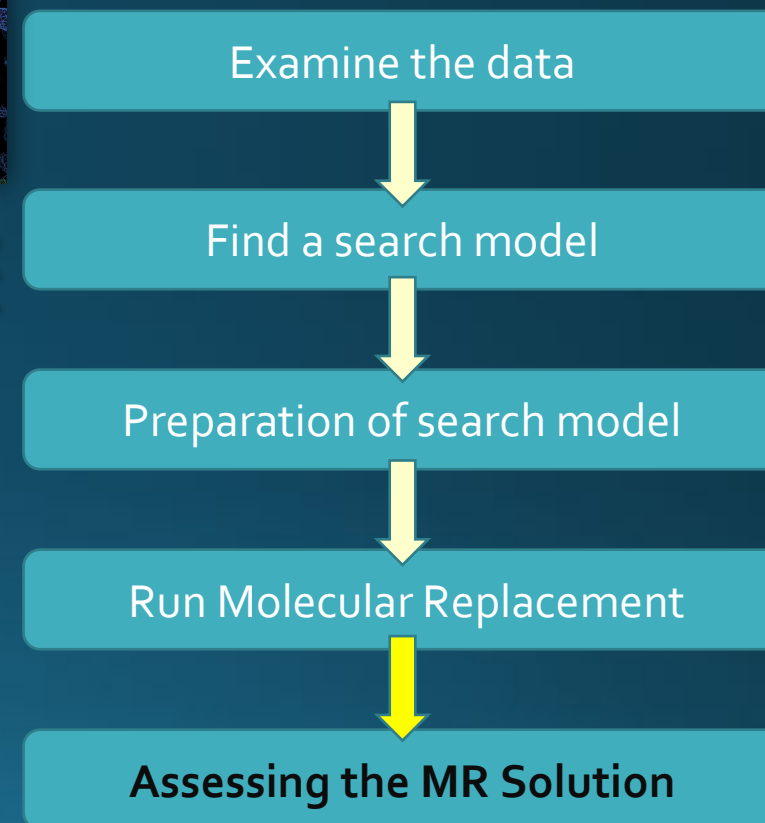
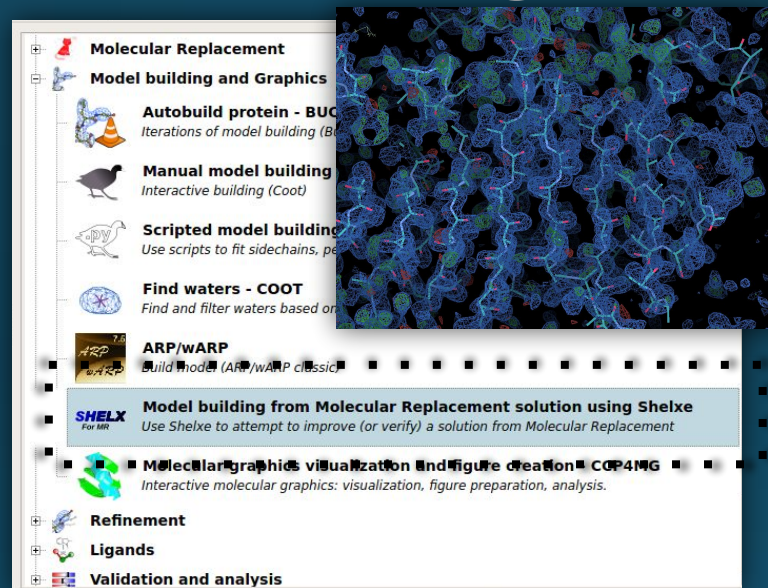
Step-by-Step MR: *Assessing the MR solution*

- Phase Improvement & C-alpha tracing



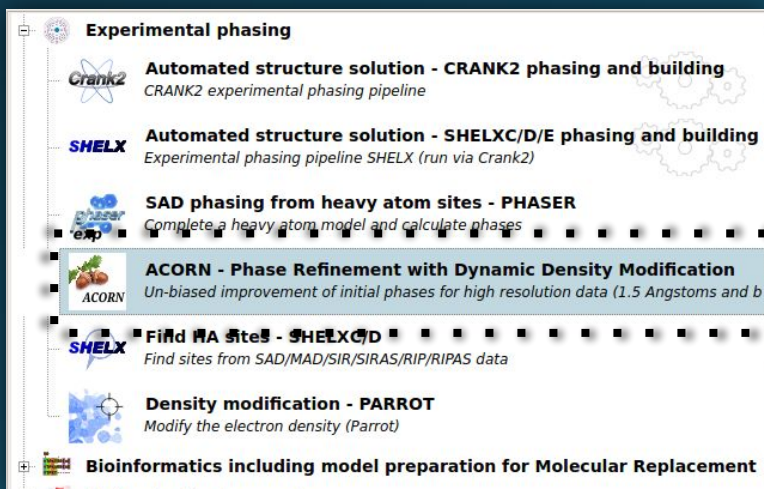
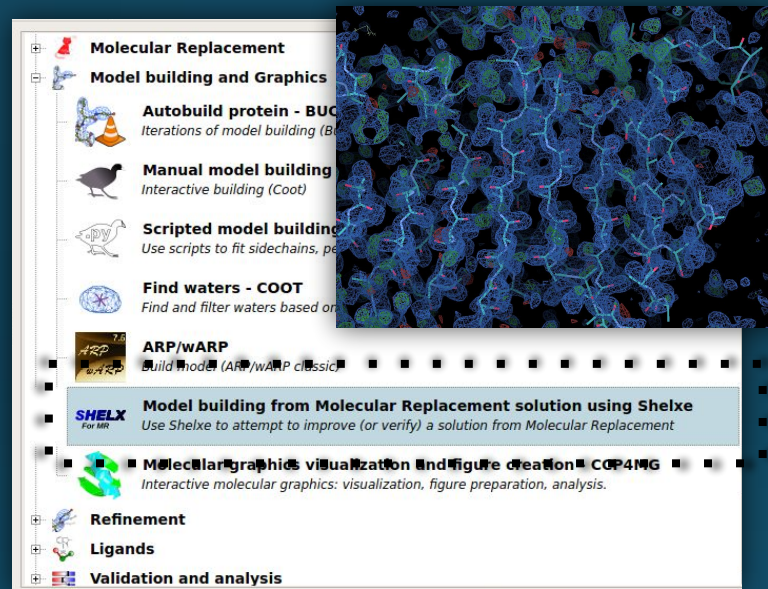
Step-by-Step MR: *Assessing the MR solution*

- Phase Improvement & C-alpha tracing
- SHELXE for MR
(*resolution better than 2.5Å*)

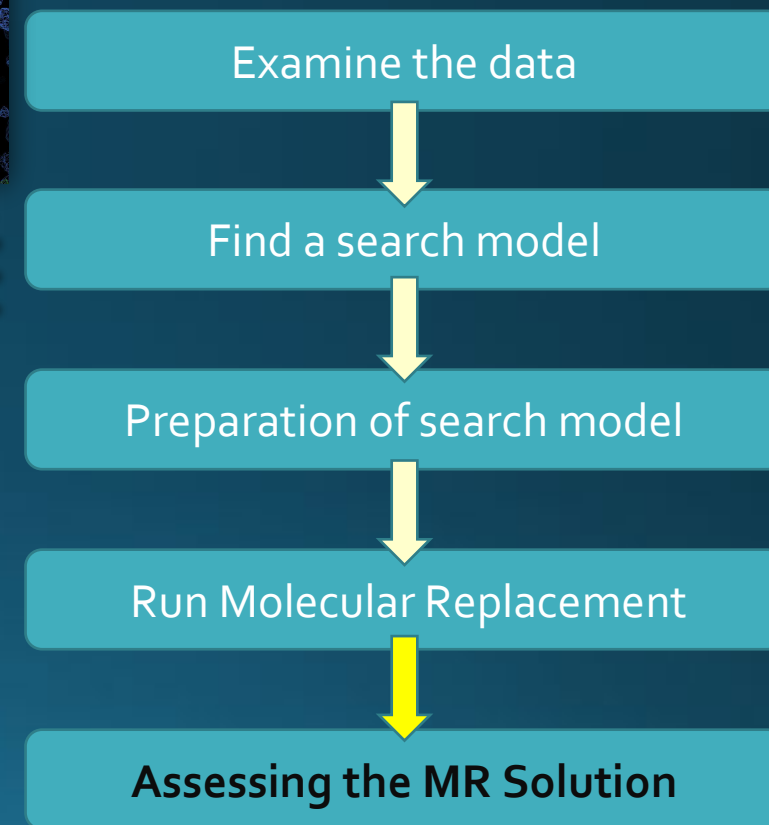


Step-by-Step MR: *Assessing the MR solution*

- Phase Improvement & C-alpha tracing
- SHELXE for MR (*resolution better than 2.5Å*)



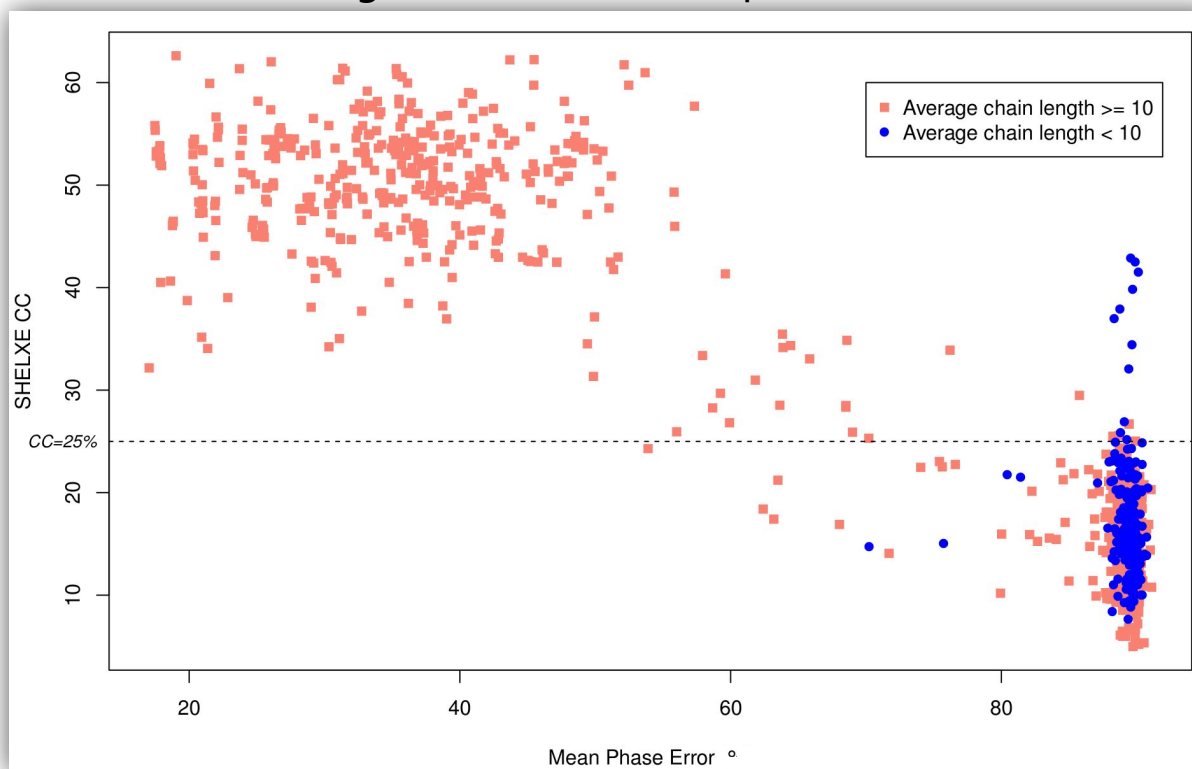
- ACORN (*resolution better than 1.7Å*)



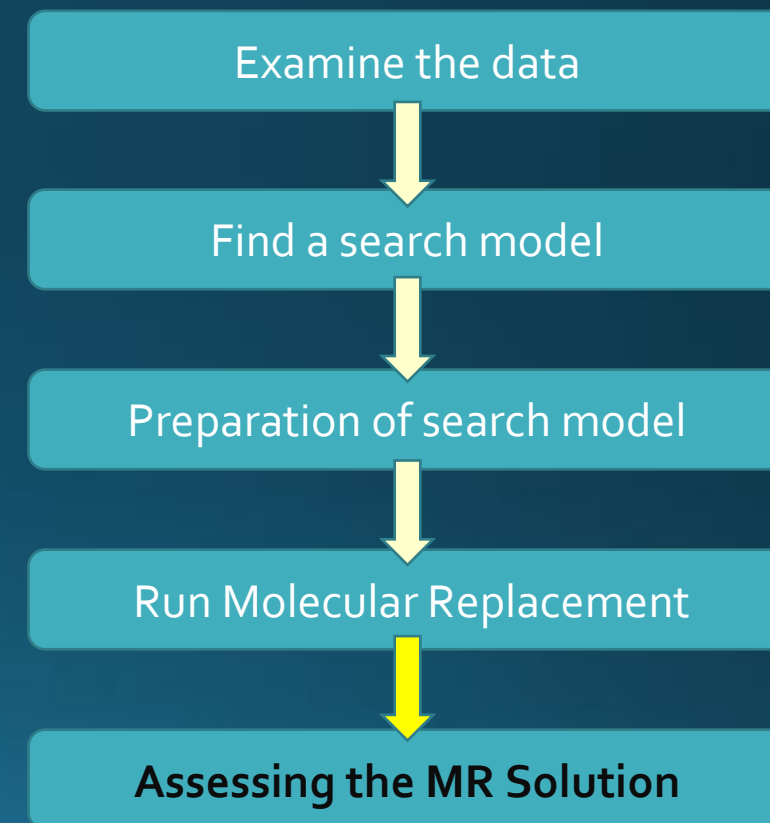
Step-by-Step MR: *Assessing the MR solution*

- SHELXE scoring: $CC \geq 25\%$
(average c-alpha fragment length ≥ 10)

SHELXE scoring tested for 1000 separate MR solutions

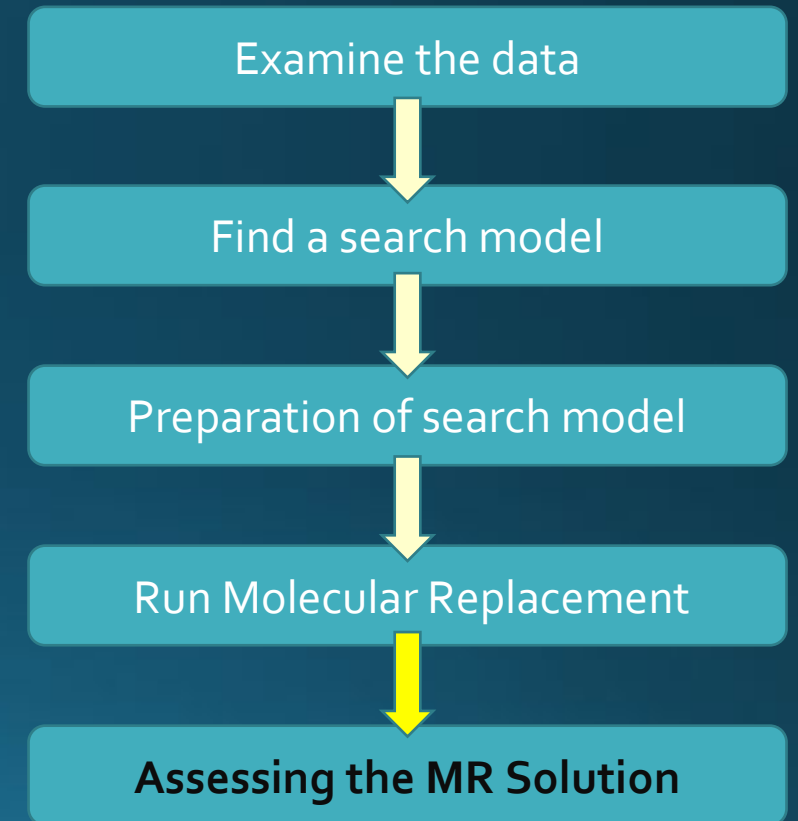


(2015) *Acta Crystallogr D Struct Biol* **71**: 338-343



Step-by-Step MR: *Assessing the MR solution*

- Automatic model building: Buccaneer & ARP/wARP
 - Can be used post-MR for generation of better model and phases for the target
 - Rebuilding parts that may not be present in search model
 - Useful for assessing whether or not your positioned MR model is true – eliminates bias

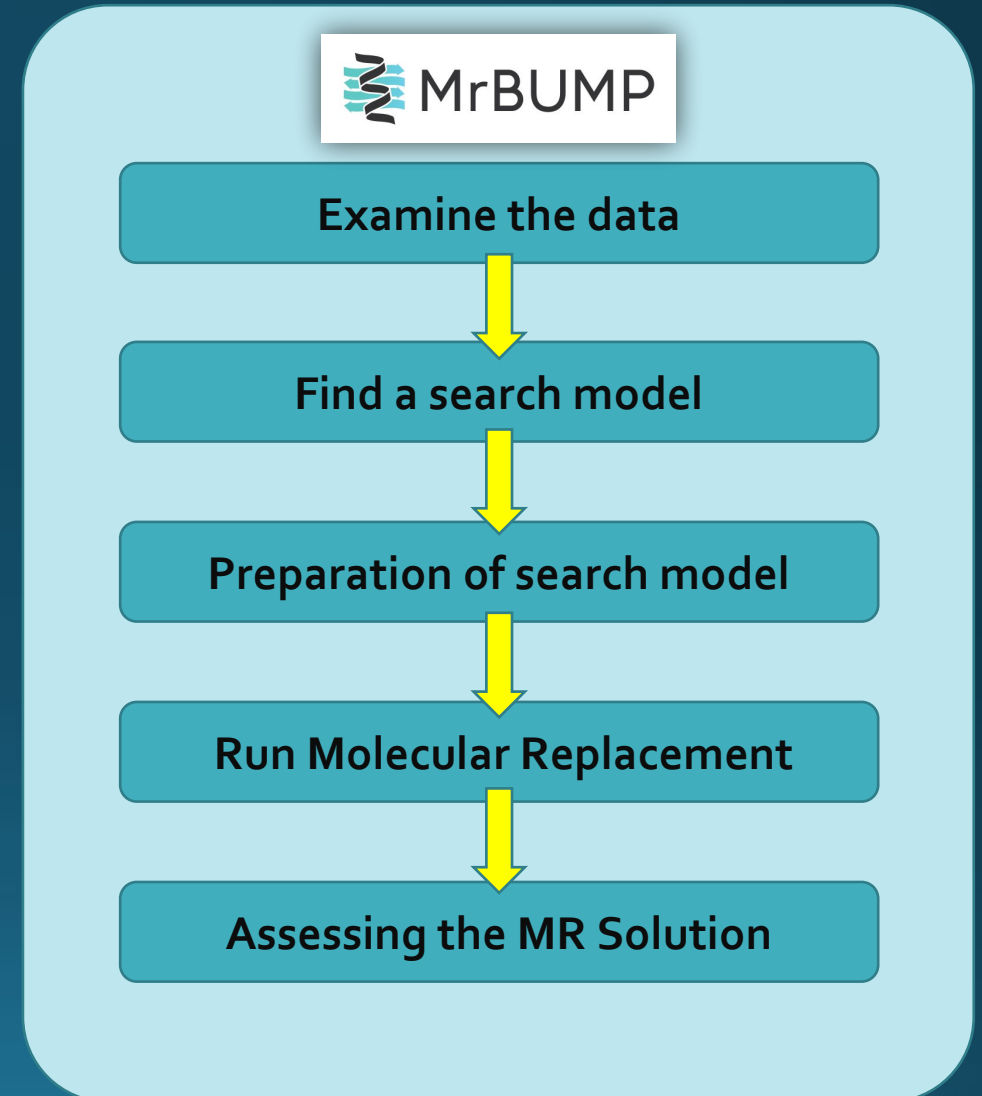


Automated Molecular Replacement in CCP4

- Several automation pipelines for MR in CCP4:
 - *MrBUMP* – model search, preparation, MR and refinement
 - *BALBES* – model search in custom version of PDB database
 - *MoRDa* – similar to BALBES

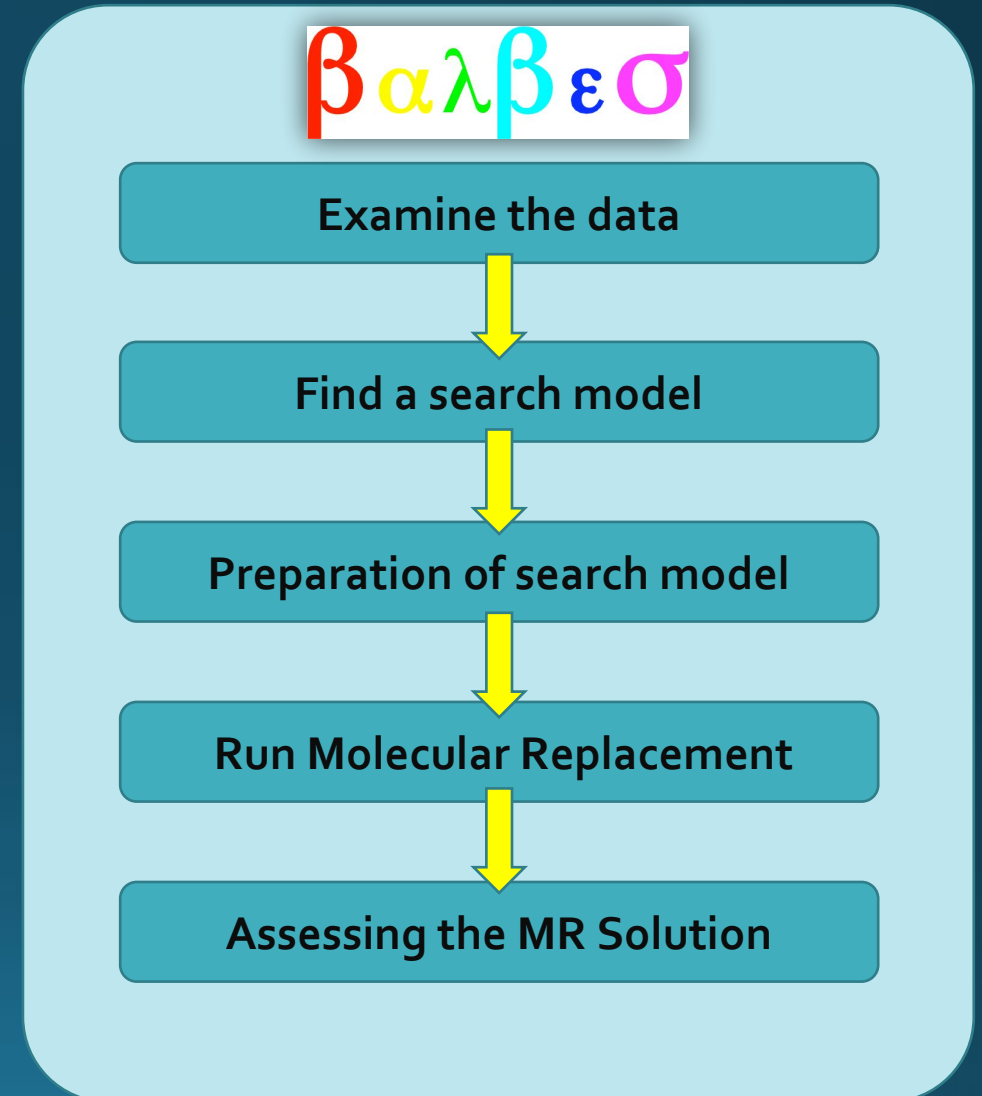
MrBUMP

- Covers all stages from data examination through to initial model building
- Two modes:
 1. Model generation only
 2. Model generation and Molecular Replacement through to model building
- Can be run from:
 - CCP4i2, CCP4i & CCP4 Online
 - CCP4mg – model generation step with graphical view

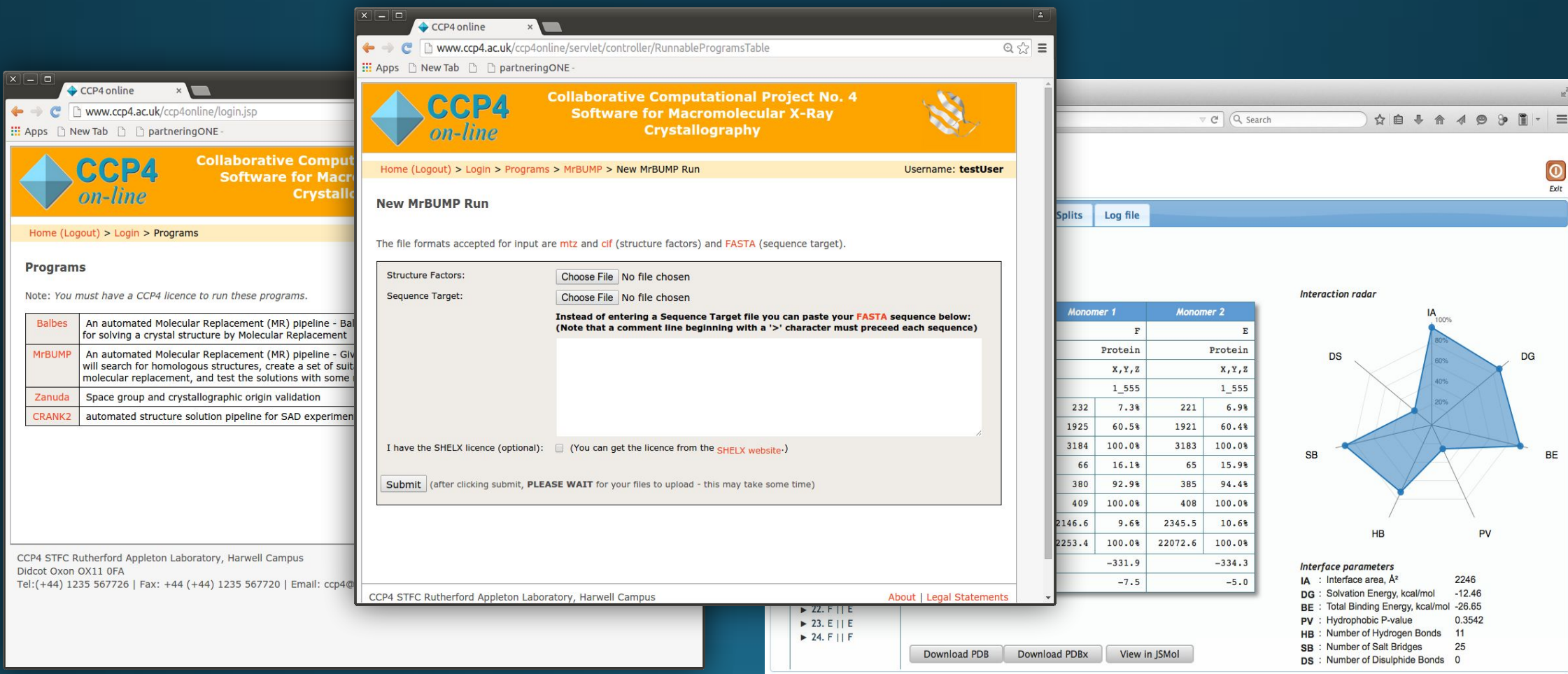


BALBES/MoRDa

- Covers all stages from data examination through to refinement
- Custom database:
 1. Non-redundant version of PDB
 2. Basic entry is a domain with definitions for constructing full chains, multimers and ensembles
- Other features:
 - Search all spacegroups
 - Uses Molrep for MR and Refmac for refinement
 - Available in CCP4i & on CCP4 Online



CCP4 Online: *MrBUMP*, *BALBES*, *MoRDa* & more



CCP4 on-line Collaborative Computational Project No. 4 Software for Macromolecular X-Ray Crystallography

Home (Logout) > Login > Programs > MrBUMP > New MrBUMP Run Username: testUser

New MrBUMP Run

The file formats accepted for input are **mtz** and **cif** (structure factors) and **FASTA** (sequence target).

Structure Factors: No file chosen

Sequence Target: No file chosen

Instead of entering a Sequence Target file you can paste your **FASTA** sequence below:
(Note that a comment line beginning with a '>' character must precede each sequence)

I have the SHELX licence (optional): ☐ (You can get the licence from the [SHELX website](#).)

(after clicking submit, **PLEASE WAIT** for your files to upload - this may take some time)

Monomer 1	Monomer 2
F	E
Protein	Protein
X, Y, Z	X, Y, Z
1_555	1_555
232	221
7.3%	6.9%
1925	1921
60.5%	60.4%
3184	3183
100.0%	100.0%
66	65
16.1%	15.9%
380	385
92.9%	94.4%
409	408
100.0%	100.0%
2146.6	2345.5
9.6%	10.6%
2253.4	22072.6
100.0%	100.0%
-331.9	-334.3
-7.5	-5.0

Interaction radar

Interface parameters

Parameter	Value
IA : Interface area, Å²	2246
DG : Solvation Energy, kcal/mol	-12.46
BE : Total Binding Energy, kcal/mol	-26.65
PV : Hydrophobic P-value	0.3542
HB : Number of Hydrogen Bonds	11
SB : Number of Salt Bridges	25
DS : Number of Disulphide Bonds	0

CCP4 STFC Rutherford Appleton Laboratory, Harwell Campus

Download PDB Download PDBx View in JSMol

<http://www.ccp4.ac.uk/ccp4online>

What to do if Molecular Replacement doesn't work

What to do if Molecular Replacement doesn't work

- Try different search models or different preparation methods for the homologues

What to do if Molecular Replacement doesn't work

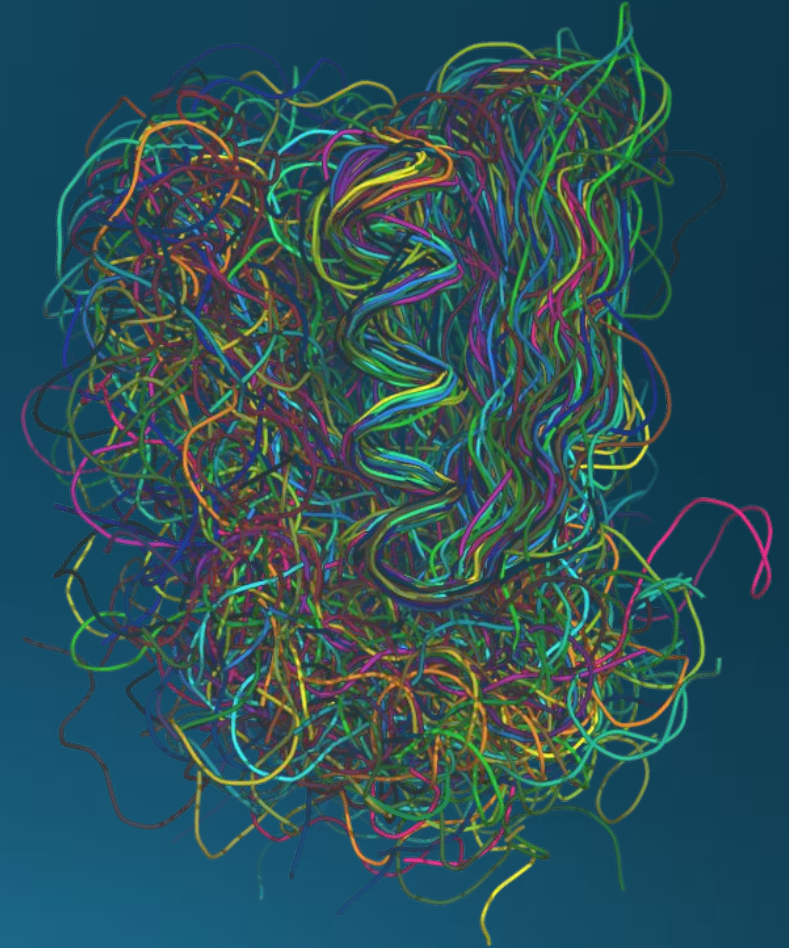
- Try different search models or different preparation methods for the homologues
- Try more distant homologues
 - Sequence similarity does not always imply structural similarity e.g. S100 family of proteins
 - Distant homologues can be structurally similar e.g. globular enzymes

What to do if Molecular Replacement doesn't work

- Try different search models or different preparation methods for the homologues
- Try more distant homologues
 - Sequence similarity does not always imply structural similarity e.g. S100 family of proteins
 - Distant homologues can be structurally similar e.g. globular enzymes
- Experimental Phasing
 - MR-SAD – Phaser, Crank2 or SHELX
 - Sulphur SAD

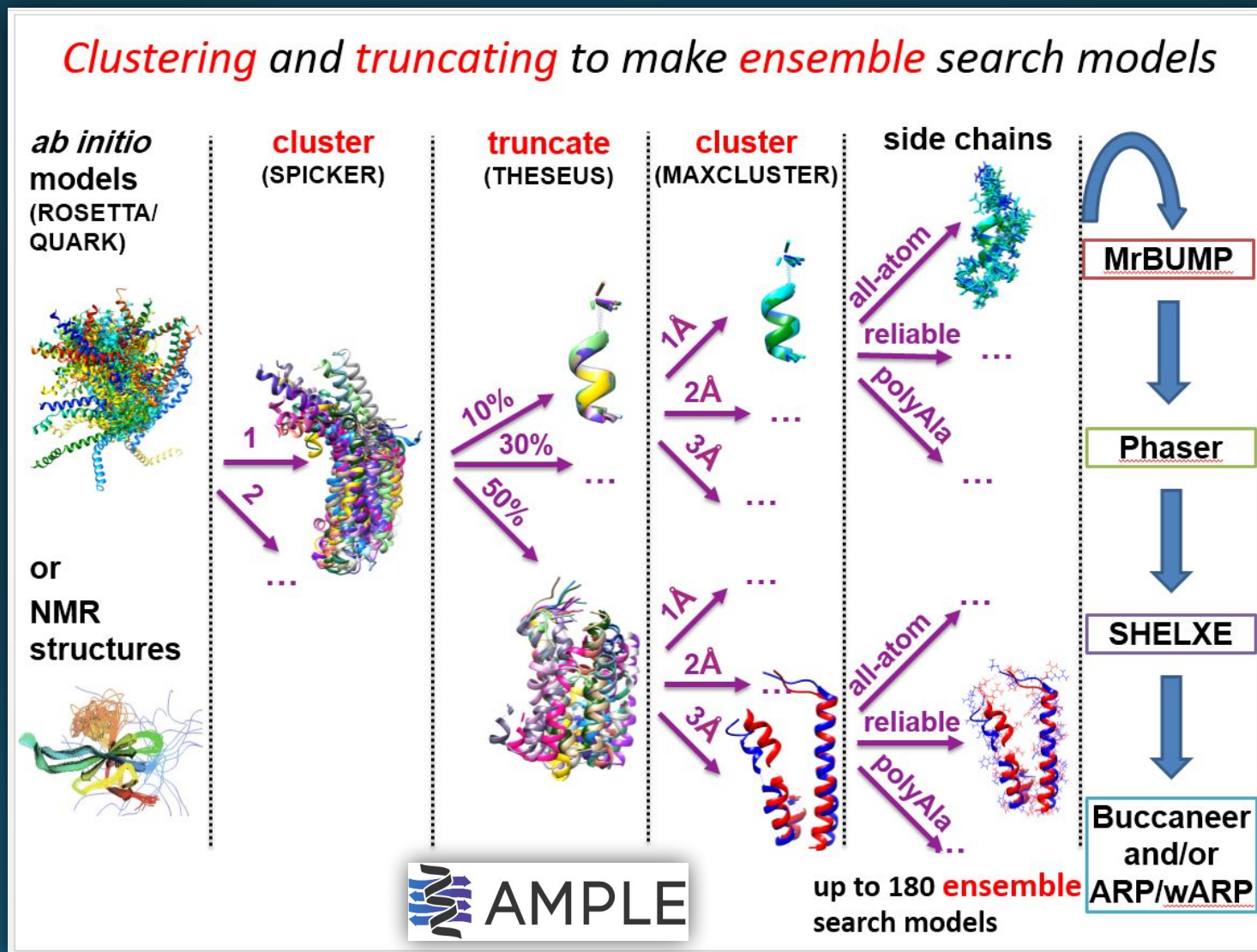
What to do if Molecular Replacement doesn't work

- **AMPLE**: *Ab initio* modelling to generate models for molecular replacement
- Uses programs like *Rosetta* and *Quark* to generate search models from the target sequence
- Can exploit sequence alignment derived residue contact predictions



What to do if Molecular Replacement doesn't work

AMPLE Pipeline



(image from
Jens Thomas)

What to do if Molecular Replacement doesn't work

- *ARCIMBOLDO* (Uson group Barcelona)
- Lite
 - Makes use of simple secondary structure elements such as helices in MR
 - Attempts to position fragments and build up the rest of the c-alpha backbone using SHELXE
- Borges
 - Similar to Lite but draws on a library of common motifs from the PDB as search models e.g. Zinc fingers or common beta sheet fragments
- Shredder
 - Cuts distant homologues into fragments and will use them as search models in ARCIMBOLDO
- <http://chango.ibmb.csic.es/ARCIMBOLDO/>

What to do if Molecular Replacement doesn't work

•Crystal Contaminants

- Always check that you don't have a contaminant present
- Perform mass spectroscopy on your solution and crystals if possible
- Test your data immediately after data collection using SIMBAD or Contamminer

METHODS AND APPLICATIONS

Protein purification and crystallization artifacts: The tale usually not told

Ewa Niedzialkowska,^{1,2,3} Olga Gasiorowska,^{1,3} Katarzyna B. Handing,^{1,3}
Karolina A. Majorek,^{1,3,4} Przemysław J. Porebski,^{1,3} Ivan G. Shabalin,^{1,3,4,5}
Ewelina Zasadzinska,⁶ Marcin Cymborowski,^{1,3} and Wladek Minor^{1,3,4,5*}

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²Jerzy Haber Institute of Catalysis and Surface Chemistry, Polish Academy of Sciences, Niezapominajek 8, Krakow, 30-239, Poland

³Midwest Center for Structural Genomics (MCSG), Argonne, Illinois, 60439

⁴Center for Structural Genomics of Infectious Diseases (CSGID), Chicago, Illinois, 60611

⁵New York Structural Genomics Research Consortium (NYSGR), Bronx, New York, 10461

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Abstract: The misidentification of a protein sample, or contamination of a sample with the wrong protein, may be a potential reason for the non-reproducibility of experiments. This problem may occur in the process of heterologous overexpression and purification of recombinant proteins, as well as purification of proteins from natural sources. If the contaminated or misidentified sample is used for crystallization, in many cases the problem may not be detected until structures are deter-

Abbreviations: CSGID, Center for Structural Genomics of Infectious Diseases; GNAT, Gcn5-related N-acetyltransferase; IMAC, immobilized metal affinity chromatography; MAD, multi-wavelength anomalous diffraction; MR, molecular replacement; MCSG, Midwest Center for Structural Genomics; NYSGR, New York Structural Genomics Research Consortium; PDB, Protein Data Bank; RMSD, root mean square deviation; SAD, single-wavelength anomalous dispersion; SEC, size exclusion chromatography; TEV, tobacco etch virus.

Additional Supporting Information may be found in the online version of this article.

Ewa Niedzialkowska and Olga Gasiorowska have contributed equally to this work.

The authors declare that there is no conflict of interest.

Description of Supporting Information material: Summary of data collection and refinement statistics for the deposited structures and the list of deposits used to identify crystallization artifacts by MR. Filename: Supplementary Materials.

Structural data are available in PDB database under accession numbers 4TNN, 4YYC, and 4ZNZ.

This work focuses on a particular difficulty that may occur as a result of accidental purification or contamination of the sample with a protein different than the protein of interest. Examples where the incorrect protein species were purified and/or crystallized are pro-

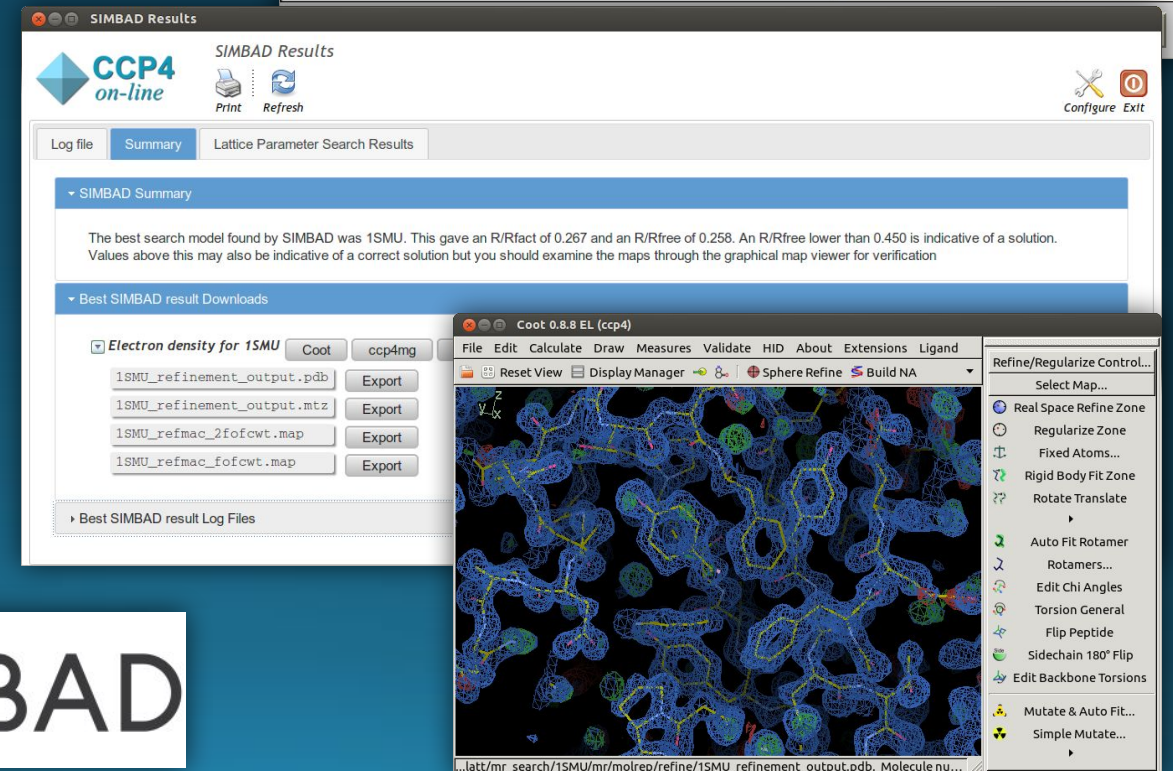
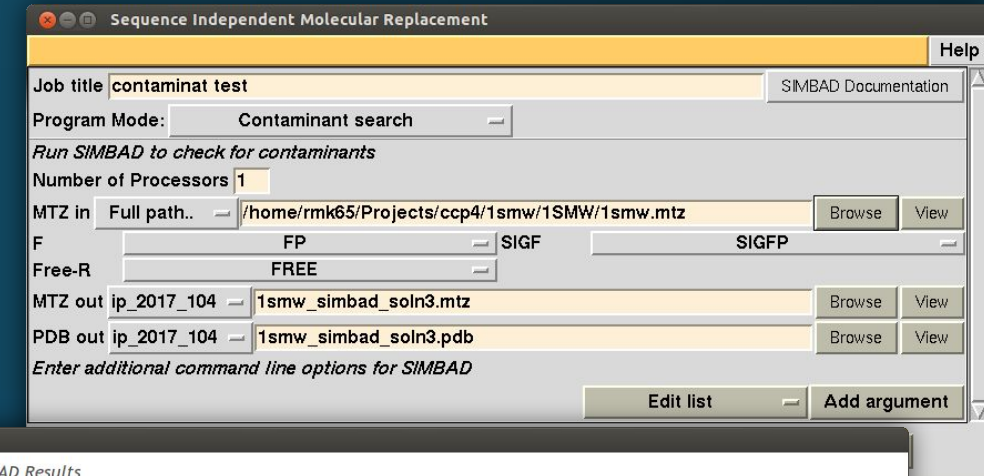
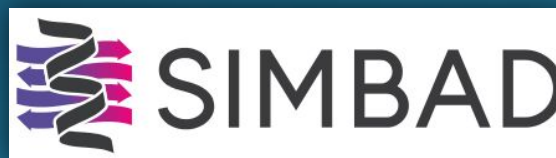
What to do if Molecular Replacement doesn't work

- **SIMBAD: *Sequence-less MR***

1. nearest cell (lattice)
2. contaminants
3. entire non-redundant domain database (85000 models)

- Accessible from *Molecular Replacement* menu

- CCP4 Online
- CCP4i2



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- Daniel Rigden, Jens Thomas, Felix Simkovic, Adam Simpkin, University of Liverpool
- Stuart McNicholas, Keith Wilson, Christian Roth, University of York
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- Andrea Thorn, Isabel Uson, Tim Gruene & George Sheldrick (SHELX)
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- Refmac: Garib Murshudov, LMB-MRC Cambridge
- Phaser: Randy Read, Airlie McCoy & Gabor Bunkozci
- Molrep: Alexei Vagin & Andrey Lebedev
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- Support from the Research Complex at Harwell