

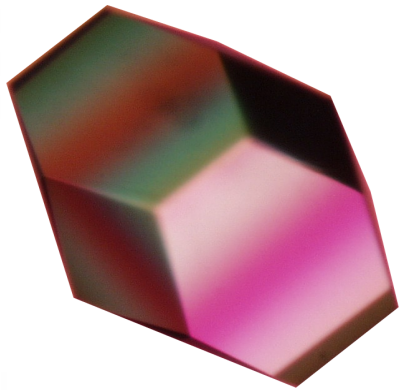
BLEND

Complete data sets from multiple crystals

James Foadi, Pierre Aller, Gwyndaf Evans

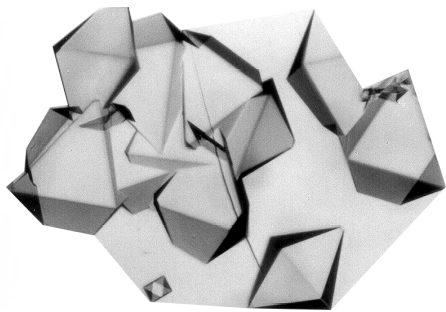
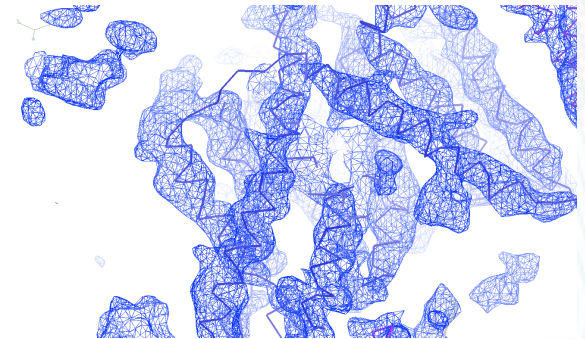
Membrane Protein Laboratory (MPL)
Imperial College London
Diamond Light Source Ltd

This talk in a nutshell



Single Crystal

Established Methods and Software

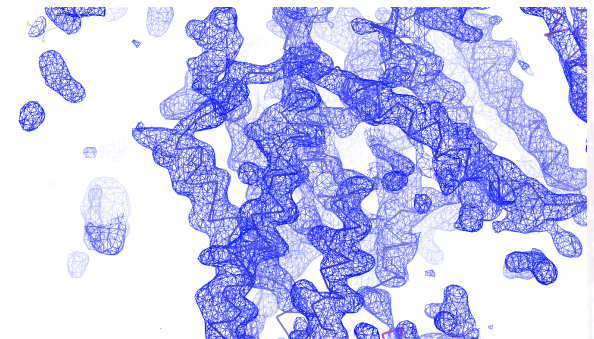


Multiple Crystals

New Methods and Software



BLEND

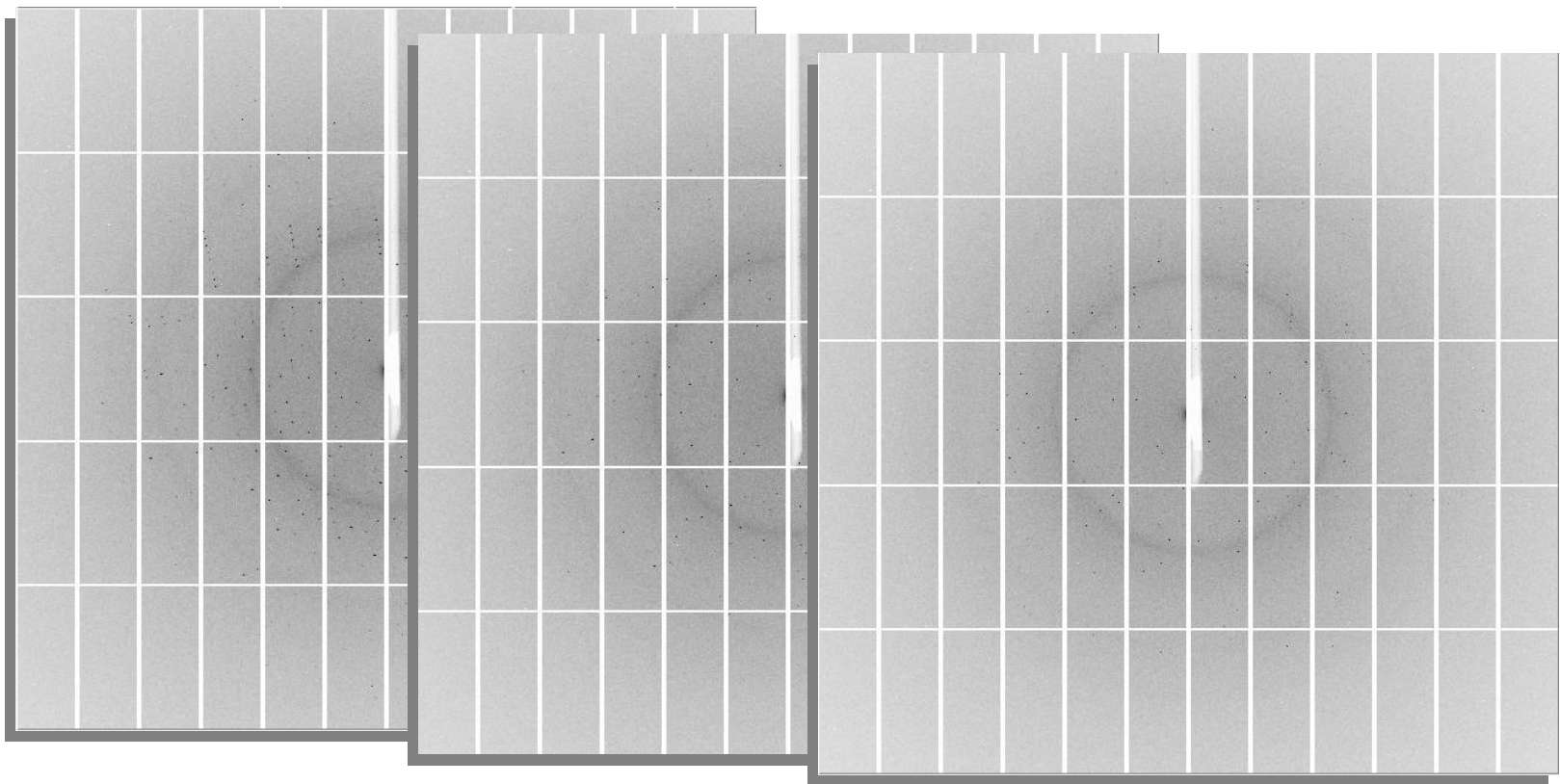


Outline

- (some) Reasons to use multiple crystals
- BLEND
- Established results
- Applications
- Slide on BLEND tutorials

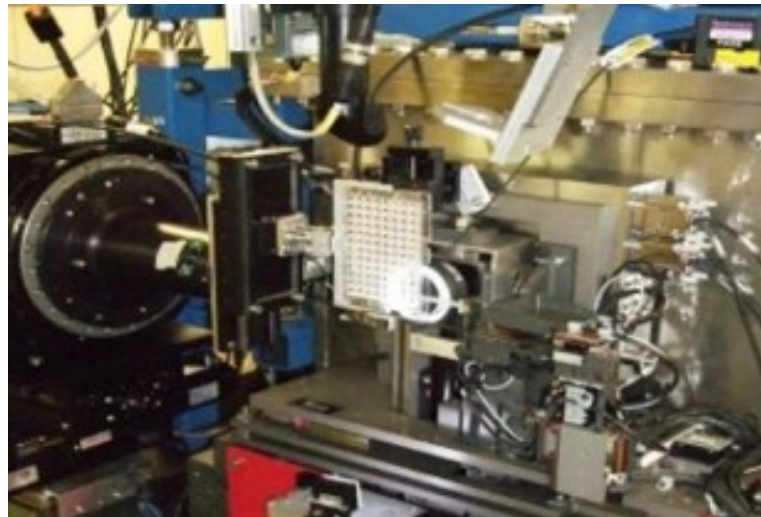
(some) Reasons to use multiple crystals

To reduce radiation damage



(some) Reasons to use multiple crystals

Constraints in data collection (*in situ*)

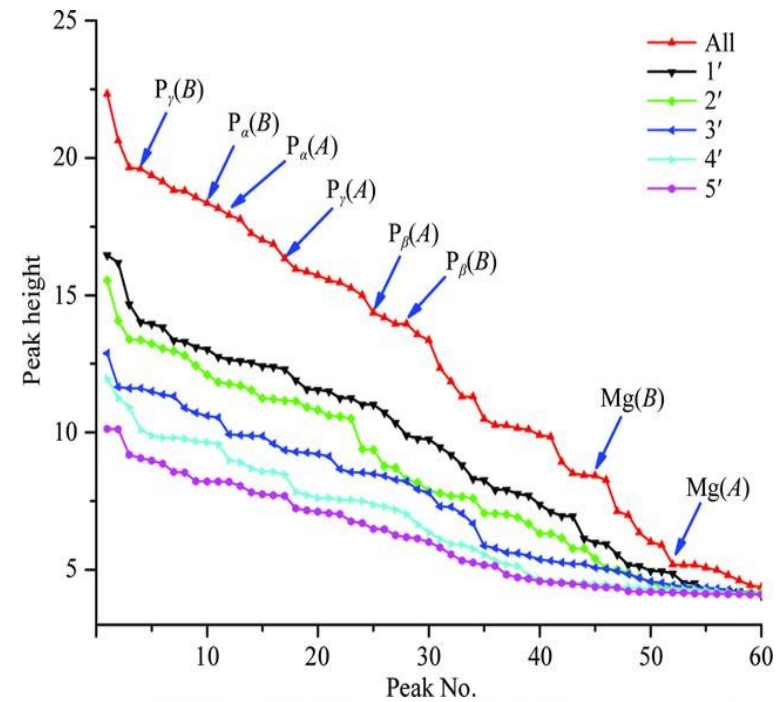
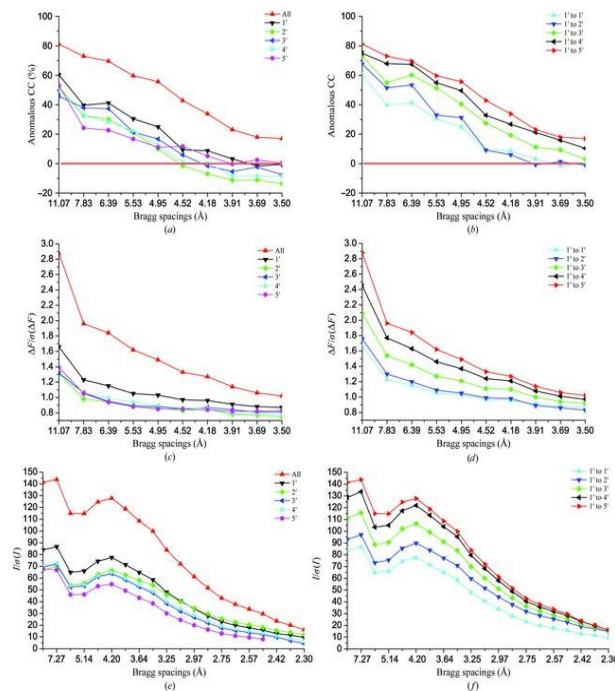


Diamond
I24 beamline

(some) Reasons to use multiple crystals

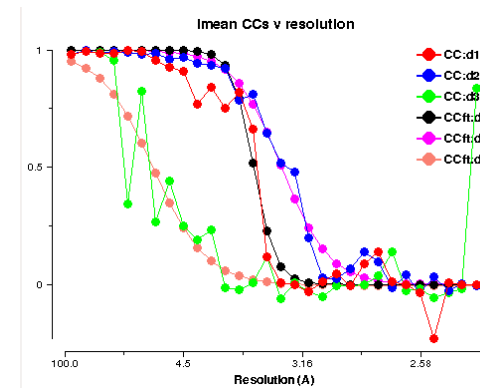
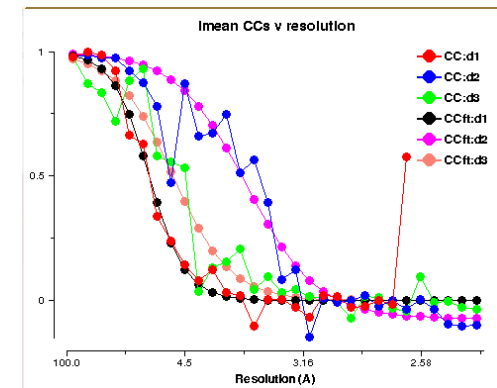
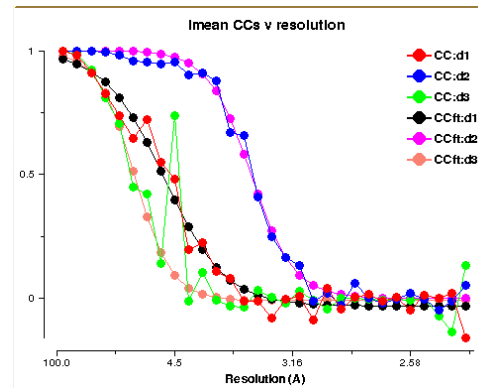
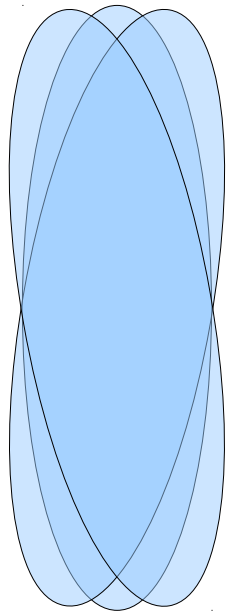
To increase the anomalous signal

Q. Liu, Q. Liu and W. A. Hendrickson (2013), Acta Cryst. D69, 1314-1332



(some) Reasons to use multiple crystals

Slight improvement of anisotropic data



BLEND

Main reference:

J. Foadi, P. Aller, Y. Alguel, A. Cameron, D. Axford,
R. L. Owen, W. Armour, D. G. Waterman, S. Iwata and G. Evans
*Clustering procedures for the optimal selection of data sets
from multiple crystals in macromolecular crystallography*
Acta Cryst. (2013), **D69**, 1617-1632

See also:

Slides from various presentations
(web-search them)

CCP4 html documentation

<http://www.ccp4.ac.uk/dist/html/blend.html>

→ also on personal web site:

http://www.jfoadi.me.uk/BLEND_documentation/CCP4_blend.html
Tutorials

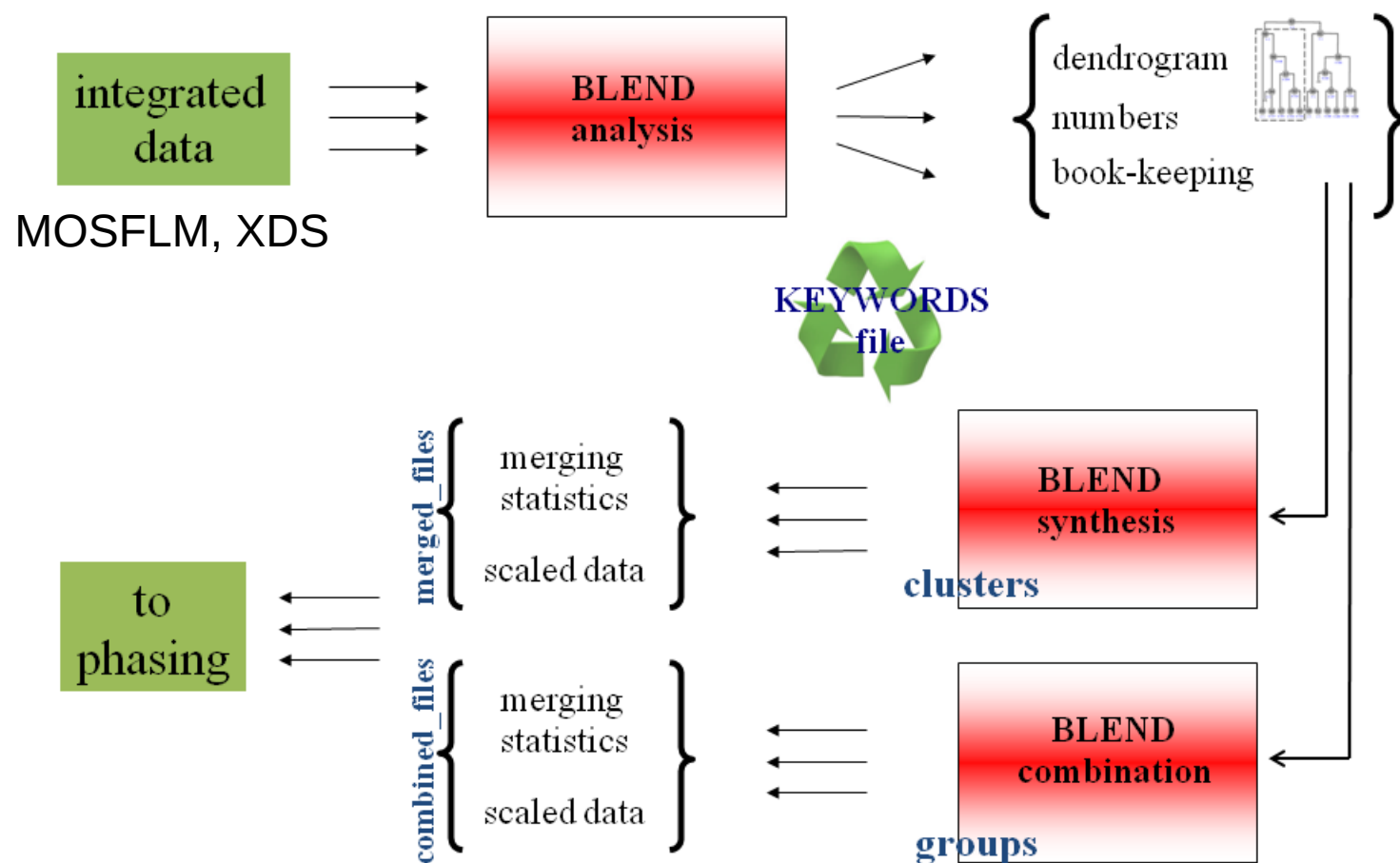
On CCP4 web site

[http://www.ccp4.ac.uk/tutorials/tutorial_files/blend_tutorial/
BLEND_tutorial.html](http://www.ccp4.ac.uk/tutorials/tutorial_files/blend_tutorial/BLEND_tutorial.html)

→ also on personal web site:

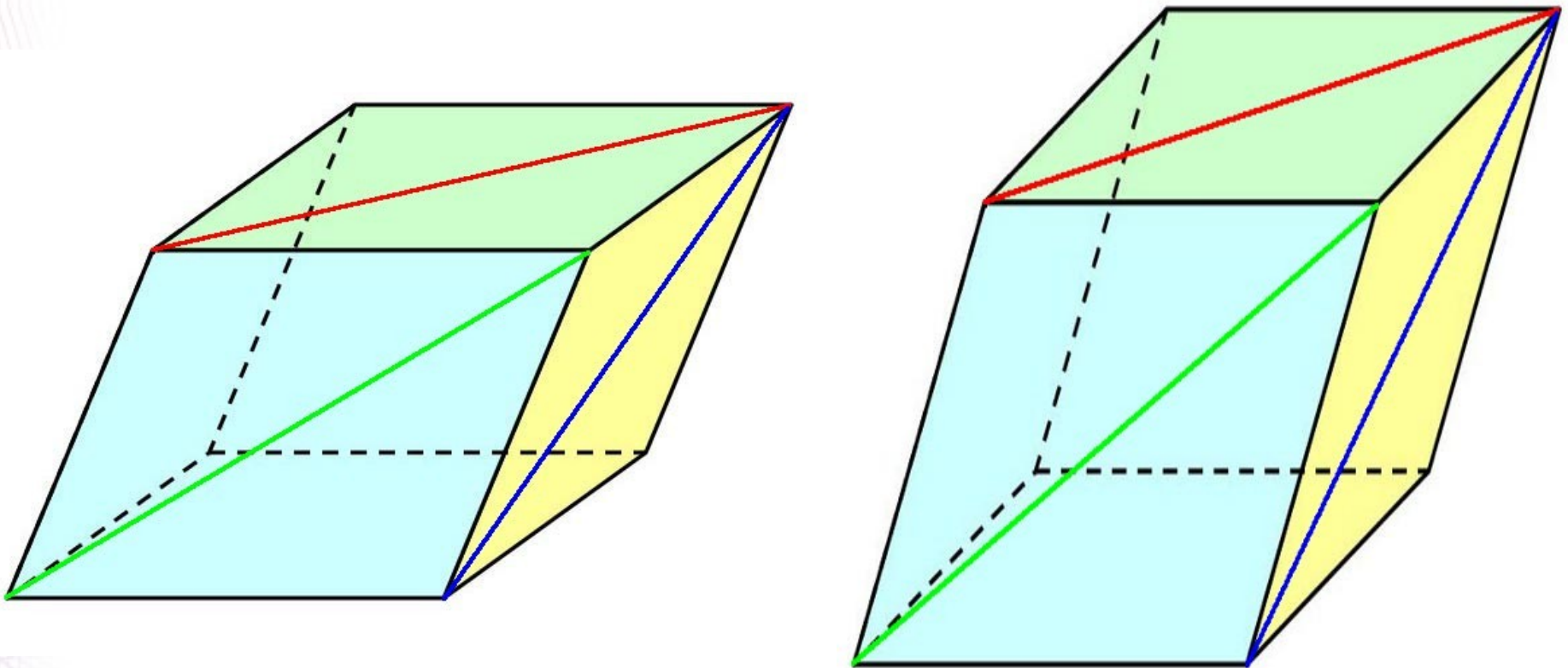
http://www.jfoadi.me.uk/BLEND_documentation/BLEND_tutorial.html

BLEND

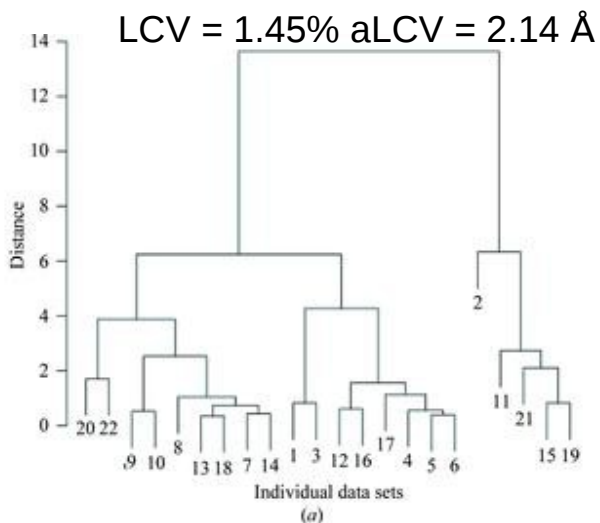


BLEND

Meaning of LCV and aLCV

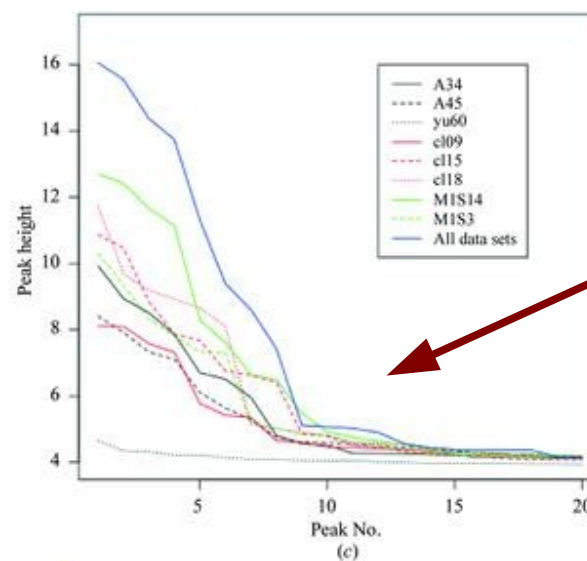
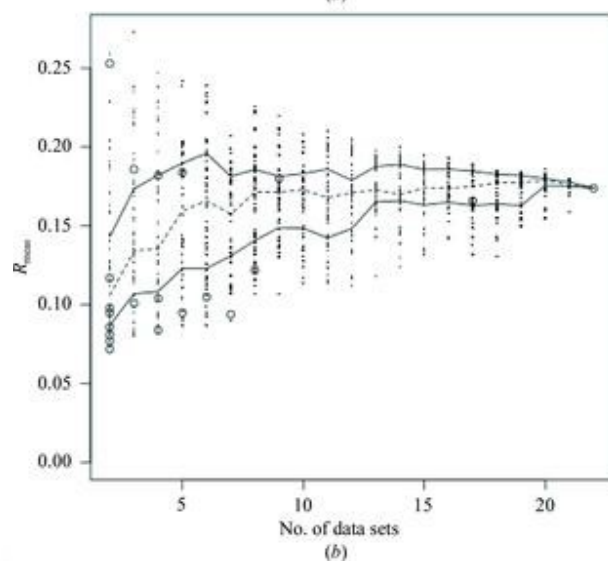


Established Results



Cryo-cooled memPROT
22 data sets, 6 crystals – 4.21 Å

(Yilmaz Alguel, Alexander Cameron, So Iwata et al)



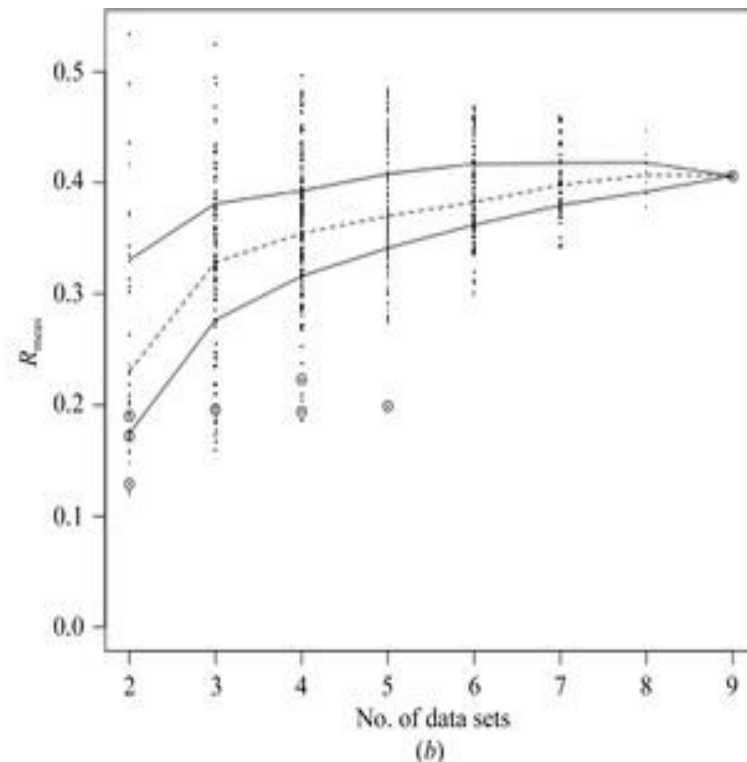
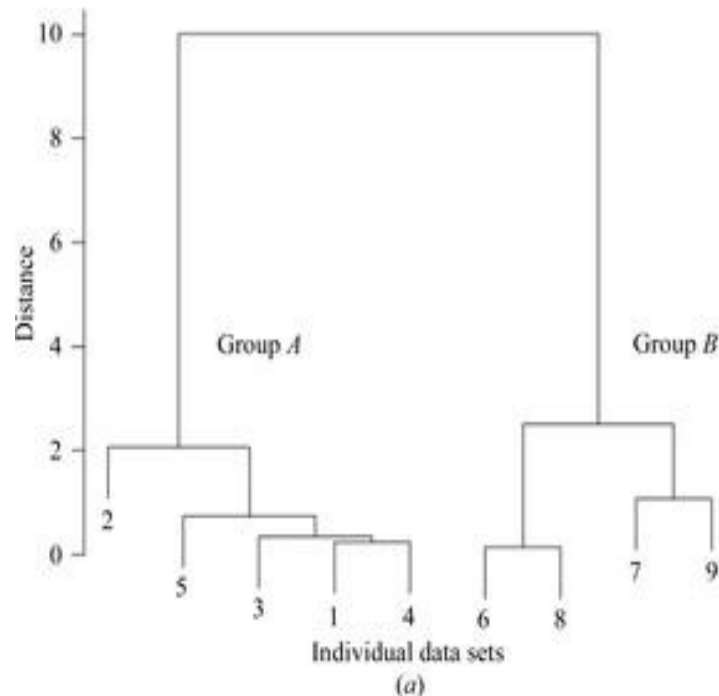
Confirms
Hendrickson's
findings

Established Results

Cryo-cooled ultralente insulin (9 crystals)

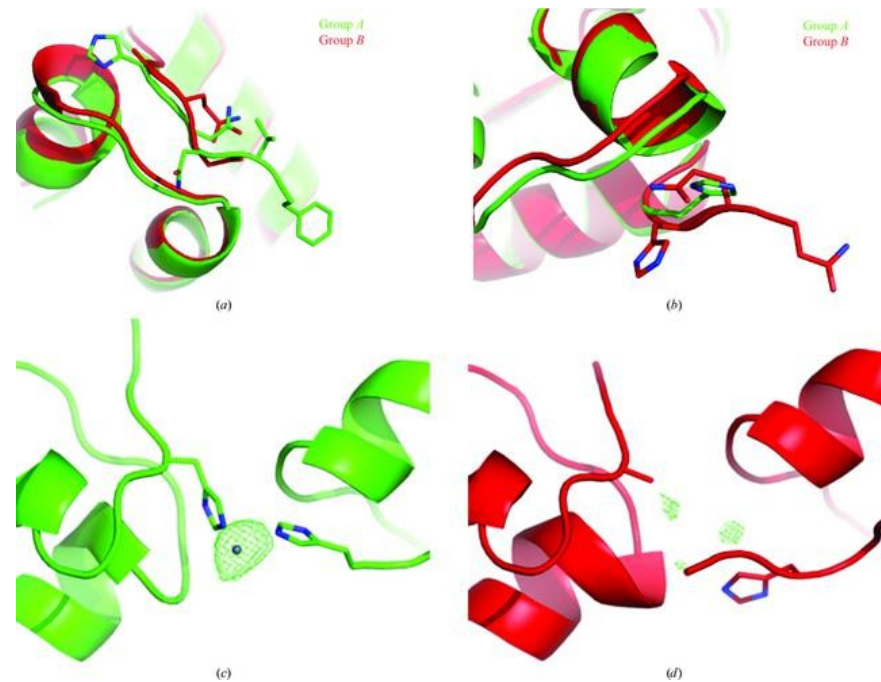
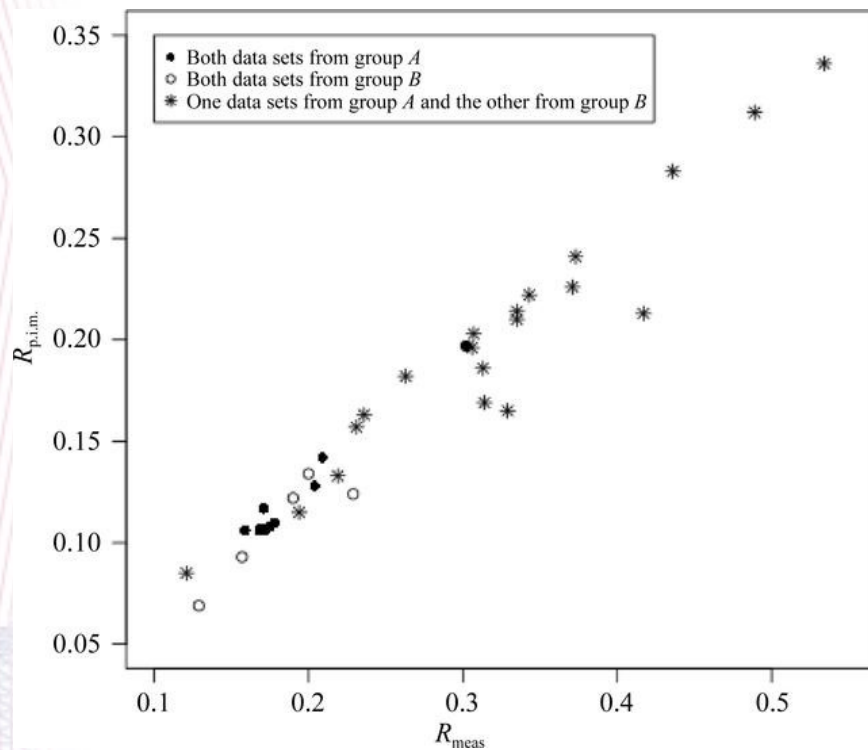
(courtesy of Armin Wagner)

LCV = 1.96% aLCV = 1.56 Å



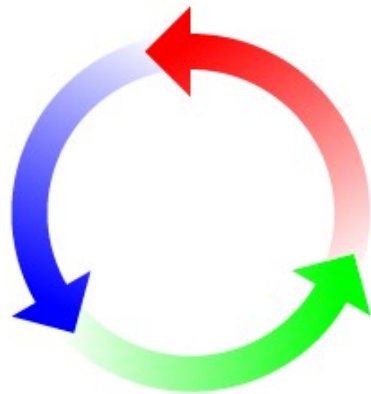
Established Results

Cryo-cooled ultralente insulin (9 crystals)



Applications

- 1) in situ data collection of a tellurite resistance protein homolog – **tehA**
67 data sets, 56 crystals (2.3 Å)
- 2) SAD data collection on crystals of lysozyme soaked with bromine
12 crystals (1.6 Å)



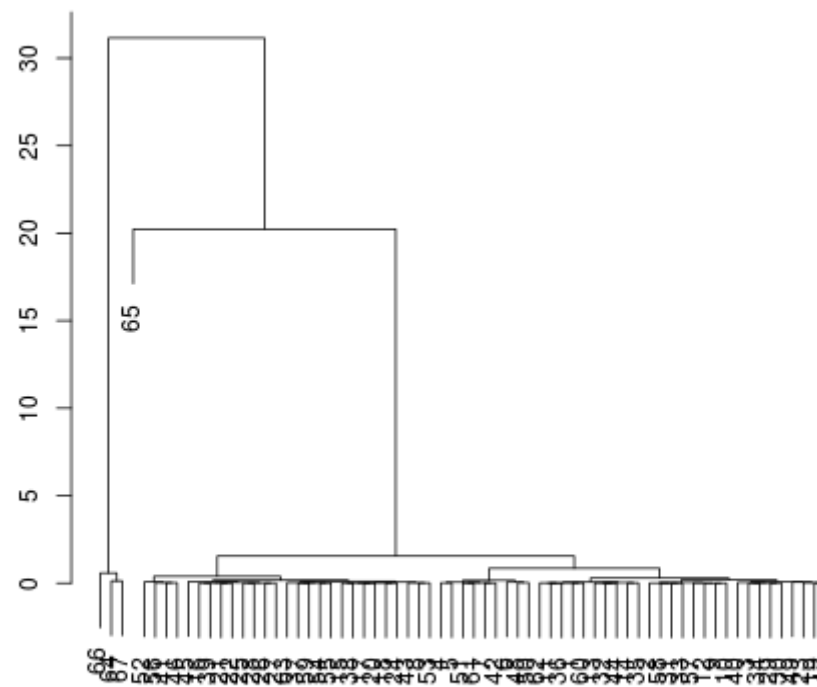
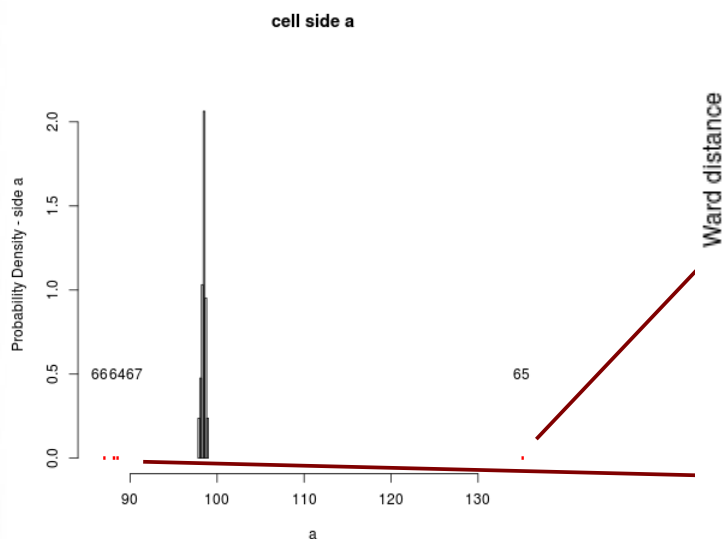
selection
combination
filtering

Applications

in situ Tellurite resistance protein homolog

huge value! ←

LCV: 57.38% (absolute LCV: 48.10 Å)



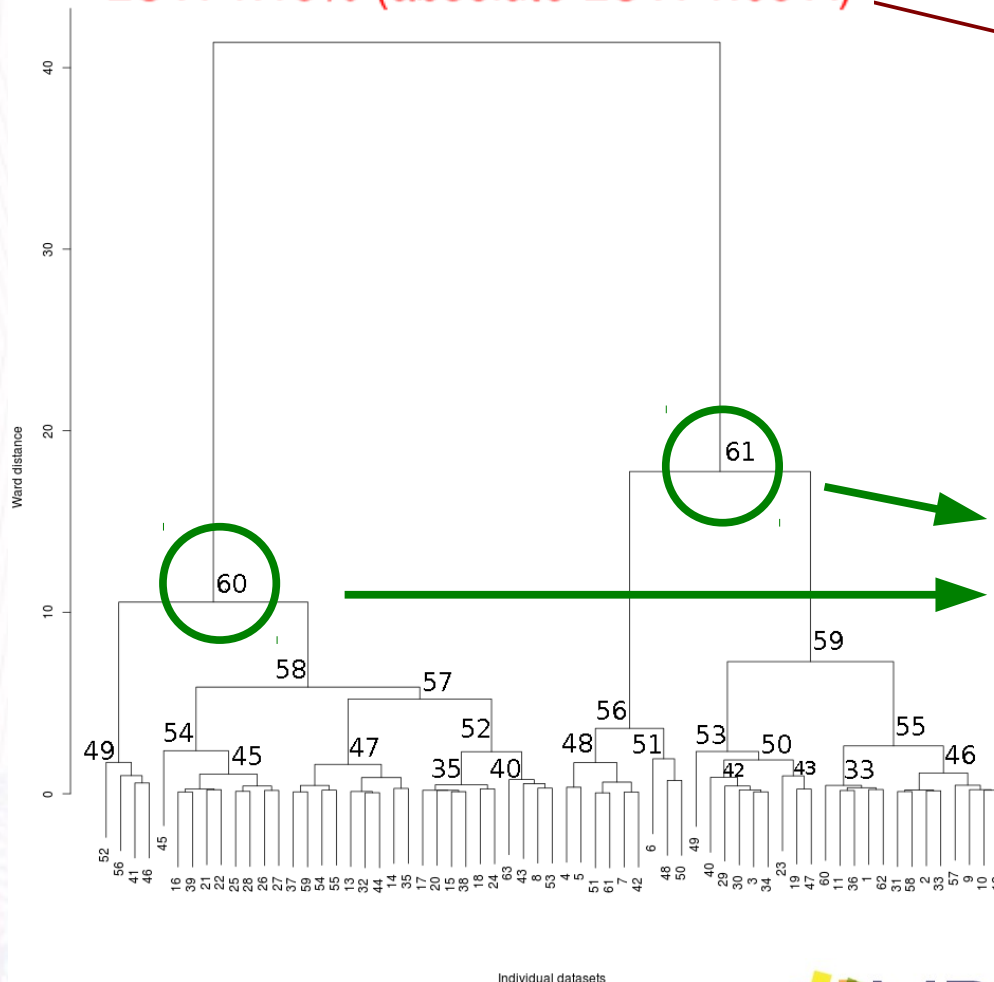
➡ 64, 65, 66 and 67 can be filtered out

Applications

in situ Tellurite resistance protein homolog

LCV: 1.18% (absolute LCV: 1.08 Å)

LCV = 1.18%
a reasonable value



The two different branches might point to some mild non-isomorphism.

But ...

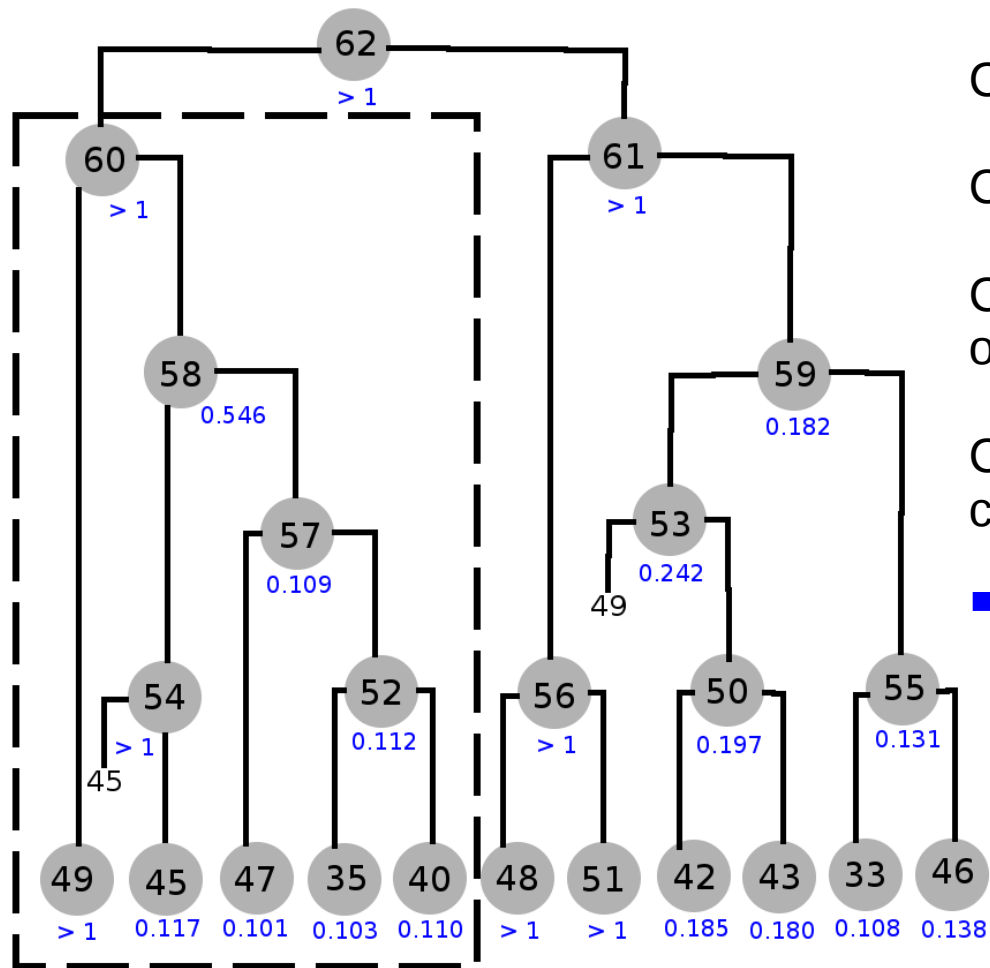
Cluster 60 is **89.7%** complete

Cluster 61 is **70.7%** complete

We select cluster 60

Applications

in situ Tellurite resistance protein homolog



Cluster 57 shows a good value of Rmeas.

Cluster 58 shows a bad value of Rmeas.

Cluster 54 is “polluting” the good quality of cluster 57.

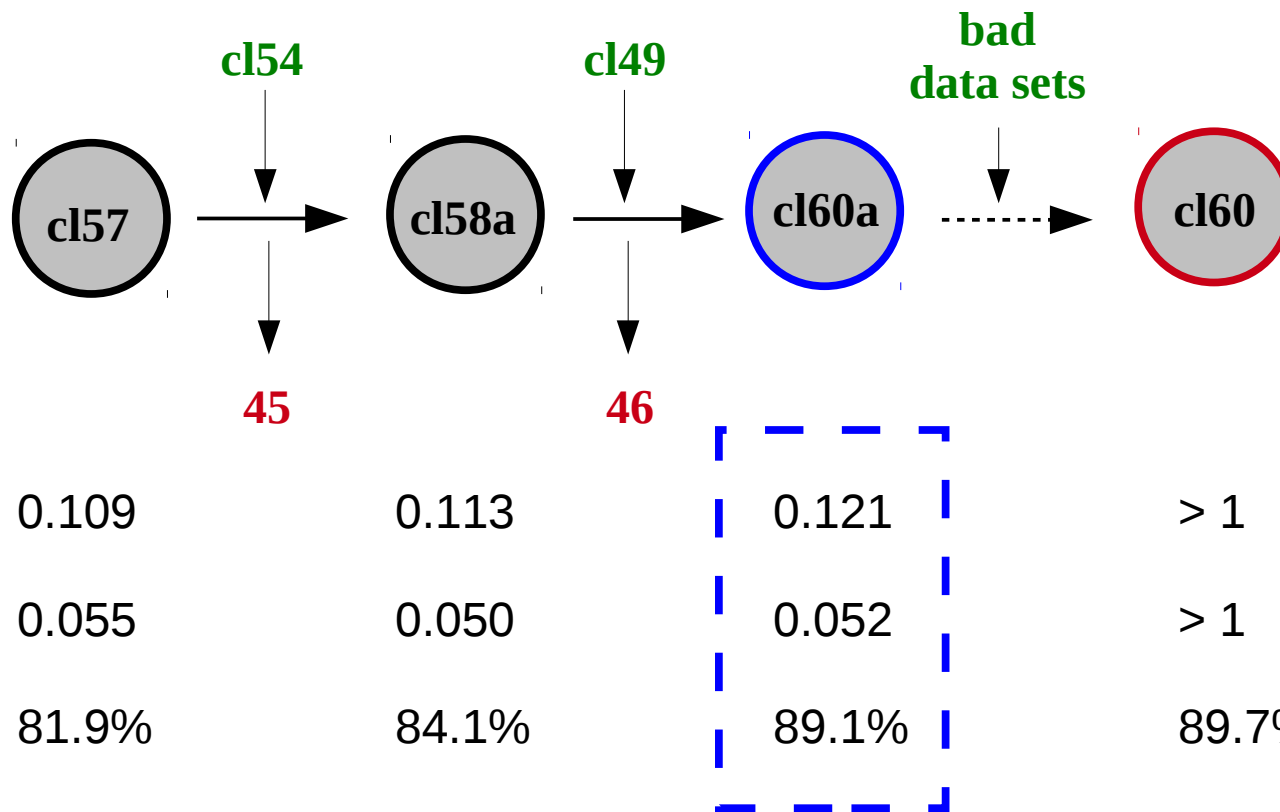
Cluster 60 is “polluted” by cluster 58, but could also be “polluted” by cluster 49.



We need to combine cluster 57 with groups of data sets or individual data sets of cluster 54, in order to form a “cleaned” cluster 58. Then we combine with data sets from cluster 49, in order to form a “cleaned” cluster 60.

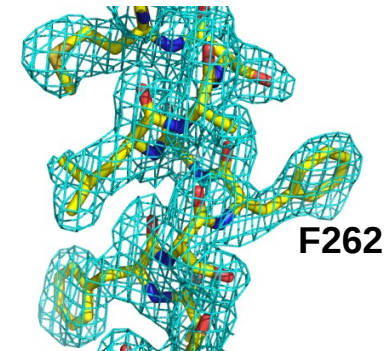
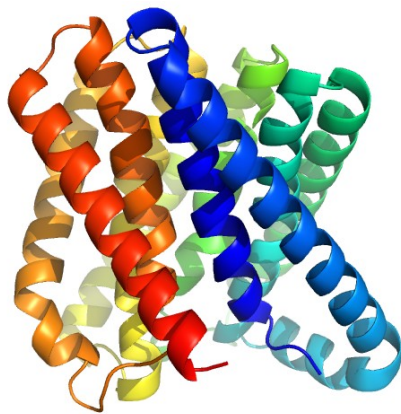
Applications

in situ Tellurite resistance protein homolog

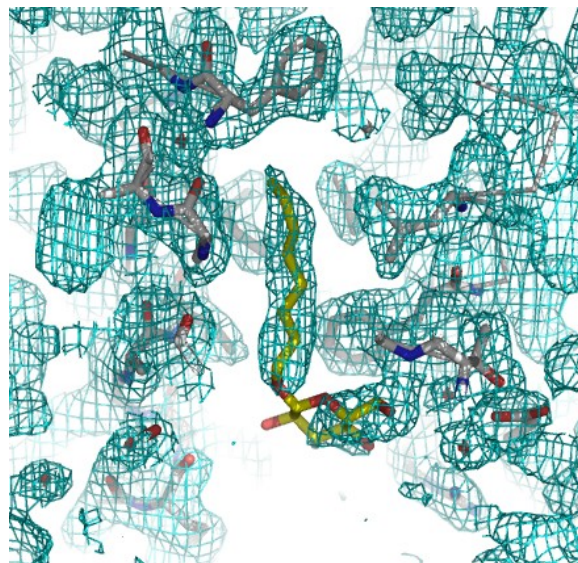


Applications

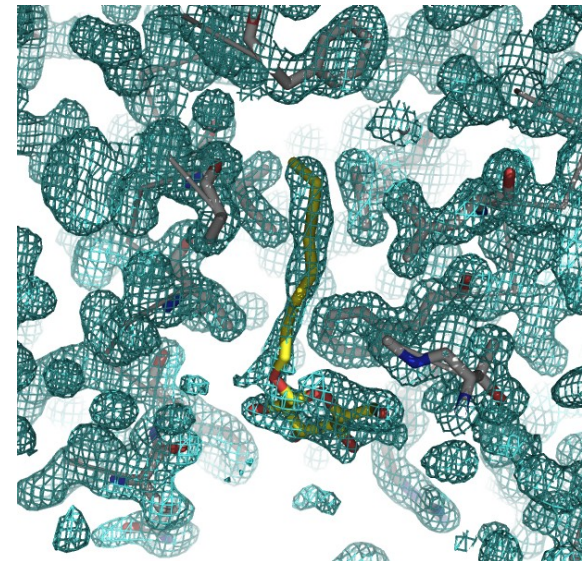
in situ Tellurite resistance protein homolog



2.3 Å



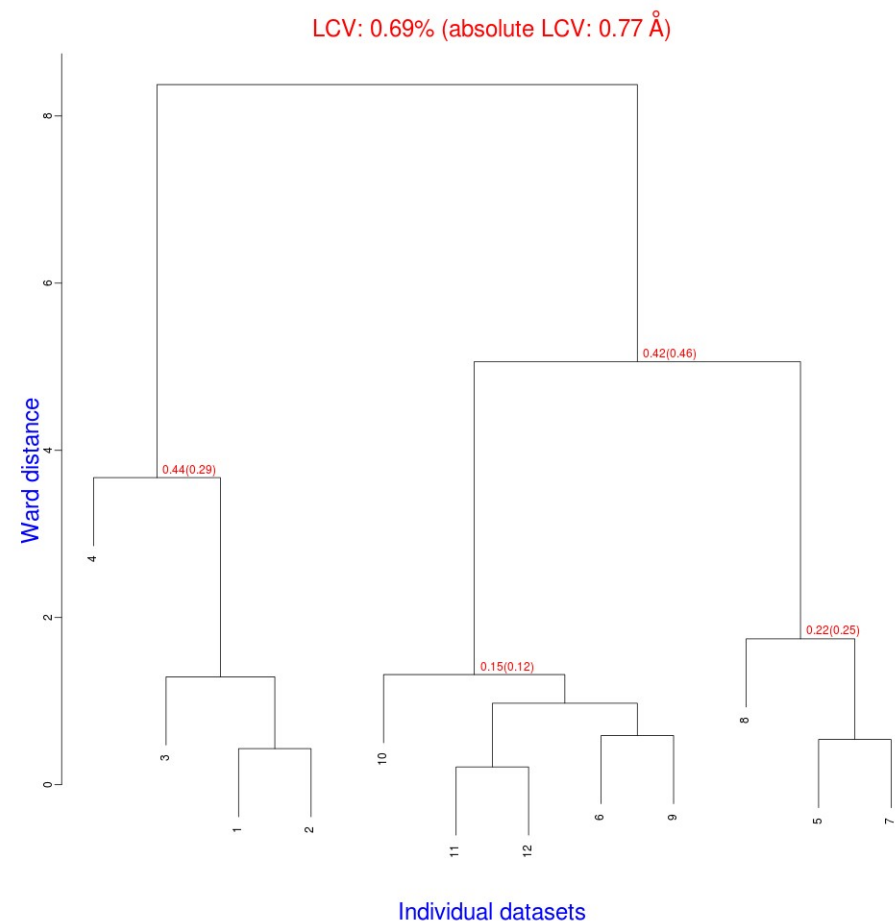
1.5 Å



Applications

SAD lysozyme with bromine

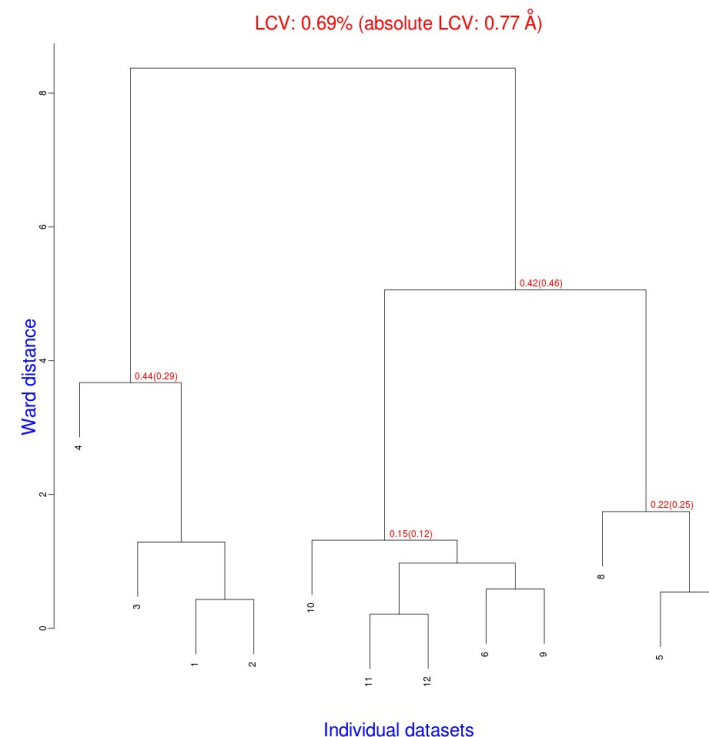
Data Set	Completeness	Anomalous Completeness
1	70.90	30.3
2	73.30	65.3
3	94.70	80.1
4	75.20	36.0
5	84.30	22.2
6	85.50	52.6
7	71.10	51.0
8	60.90	42.3
9	82.30	40.5
10	79.30	20.2
11	81.30	51.0
12	90.40	76.2



Applications

SAD lysozyme with bromine

Cluster	Completeness	Anomalous Completeness
1	88.3	73.7
2	93.6	70.4
3	92.9	70.9
4	96.0	69.5
5	98.0	92.6
6	99.8	98.4
7	98.7	94.8
8	96.3	83.9
9	99.9	98.9
10	99.4	98.2
11	99.9	99.9



19.5

multiplicity

5.8

Applications

SAD lysozyme with bromine

SHELXC → SHELXD

Data set	Sufficient d"/sig ?	Solution ?	CCall	CCweak	Cfom
ind_003	yes (-)	yes (-)	33.68	15.34	49.02
ind_012	yes	yes	43.52	25.45	68.97
005	yes	yes	45.07	24.94	70.01
006	yes	yes	37.17	19.07	56.25
007	yes	yes	44.72	26.03	70.75
009	yes	yes	36.78	18.05	54.83
010	yes	yes	45.68	26.01	71.69
011	yes	yes	44.25	26.60	70.85

Future Work

- Good indicators for anomalous phasing
- Preparation for anomalous phasing (sca files, etc.)
- Mapping data sets reciprocal space completeness (GUI)
- Splitting of discontinuous sweeps (inverse beam)
- Improved procedure for radiation damage

Acknowledgments

Phil Evans

David Waterman

DIAMOND SOFTWARE SUPPORT

Alun Ashton

Karl Levik

MPL SUPPORT

Isabel DeMoraes

Momi Iwata

So Iwata

CCP4 SOFTWARE SUPPORT

Marcin Wojdyr

Ville Uski

Andrey Lebedev

FUNDING

Wellcome Trust

BBSRC

Tutorials

http://www.jfoadi.me.uk/BLEND_documentation/BLEND_tutorial.html

http://www.jfoadi.me.uk/BLEND_documentation/CCP4_blend.html

BLEND: managing, scaling and merging multiple datasets

James Foadi^(a) and Pierre Aller^(b)

(a) Imperial College London (j.foadi@imperial.ac.uk) , Diamond Light Source Ltd (james.foadi@diamond.ac.uk)

(b) Diamond Light Source Ltd (pierre.aller@diamond.ac.uk)

Main aim of this tutorials is to get users acquainted with the program BLEND ([Foadi et al. 2013](#)) and to show how it can be used effectively to obtain complete data sets out of several partial or complete ones. The reader interested in exploring pros and cons of using data from multiple crystals can refer, for instance, to [Liu et al. \(2011\)](#), [Giordano et al. \(2012\)](#), [Axford et al. \(2012\)](#), [Hanson et al. \(2012\)](#).

CONTENTS

[Data sets included with this tutorial](#)

[Download data set 01](#)

[Download data set 02](#)

[Download data set 03](#)

[Tutorial 1](#)

[a\) Using ccp4i](#)

[b\) Without ccp4i](#)

[Tutorial 2](#)

[Tutorial 3](#)

[References](#)

DATA SETS INCLUDED WITH THIS TUTORIALS

For convenience both tutorials can be carried out inside a same directory and specific results can be collected in separate sub-directories. We will assume that "blend_tutorial" is the name of the overall directory and that it has been created in the \$HOME area. Different directory names and locations can be used, but appropriate modifications will have to be applied to the paths displayed in this document. Data downloaded using the above links, when unpacked, yield a directory named "data". For the rest of the tutorials this directory can be moved inside "blend_tutorial". There are three groups of data in the sub-directory "data". One group is formed by crystals of insulin, the second by crystals of lysozyme and the third still by lysozyme soaked with bromine.