

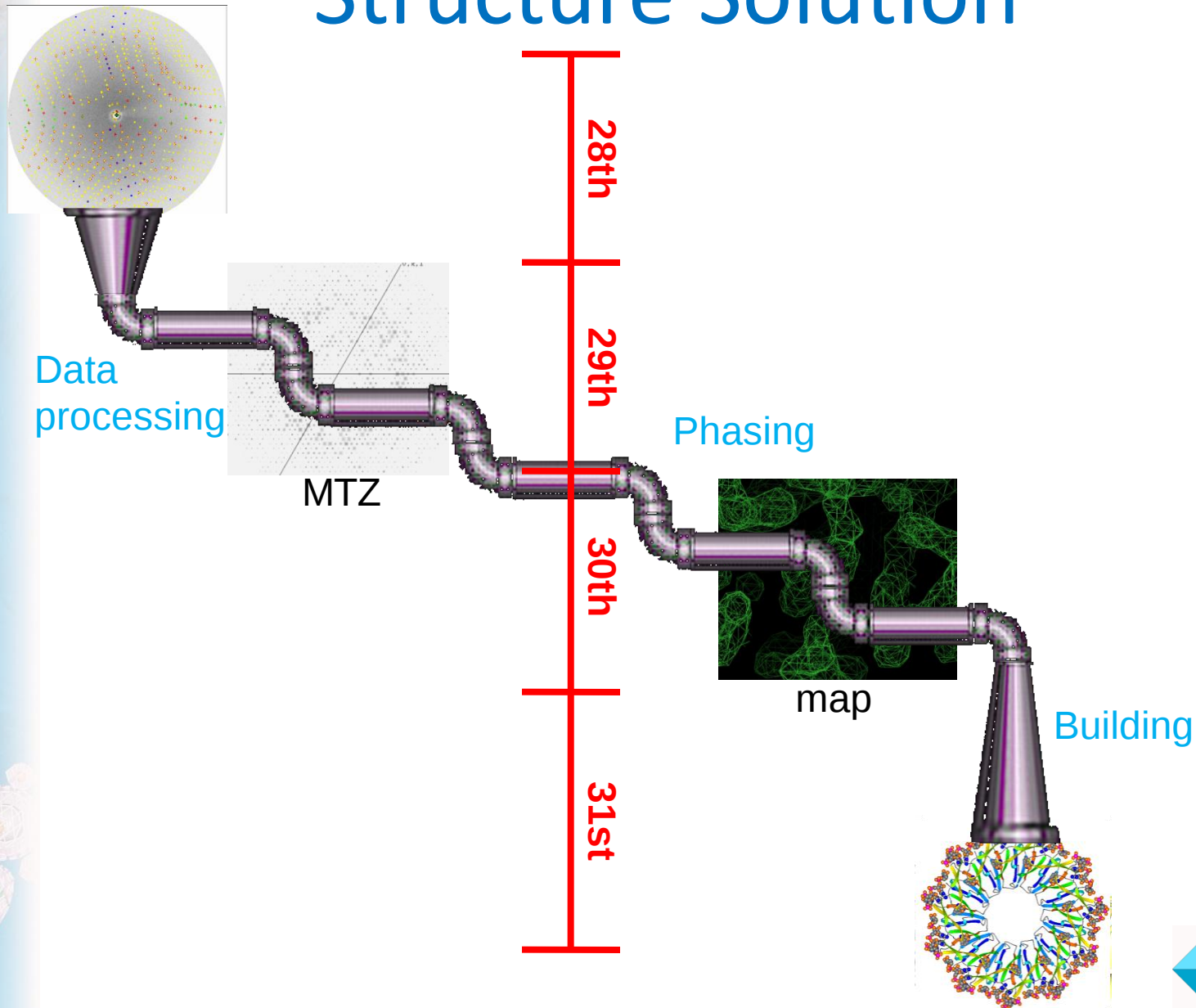


Introduction to Molecular Replacement, Dimple and MrBUMP

Martyn Winn, Daresbury Laboratory

Ronan Keegan, Rutherford Laboratory

Structure Solution





Simplest case: known structure

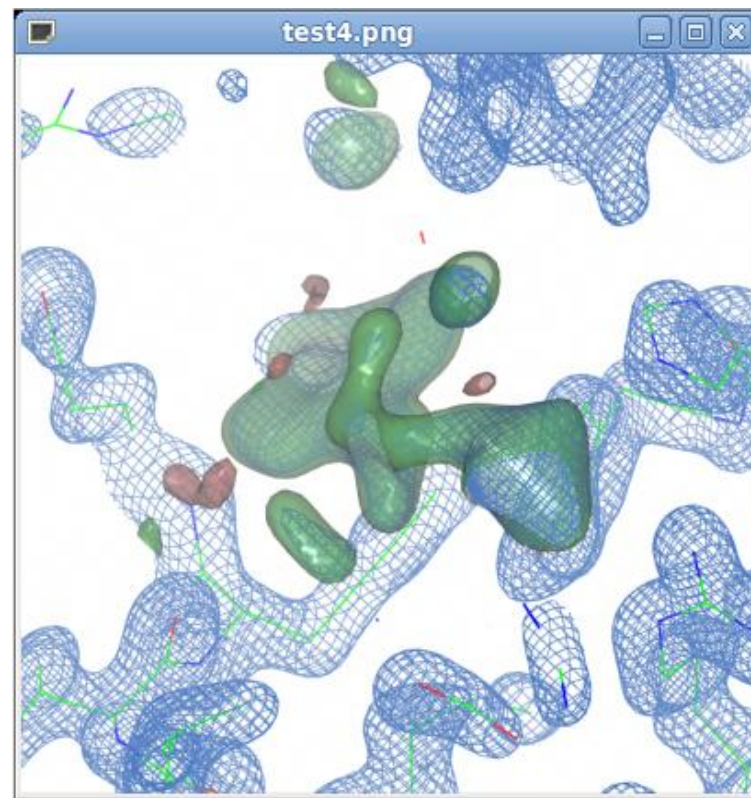
e.g. protein-ligand complex, mutant structure

- Known structure (.pdb file)
 - Data collected and processed (.mtz file)
 - Target structure with same symmetry and cell
- ⇒ phase problem already solved
move directly to refinement

DIMPLE

Need to refine known structure against new data. Shows minor changes, e.g. new ligand

DIMPLE is simple CCP4 pipeline (to be released soon!)

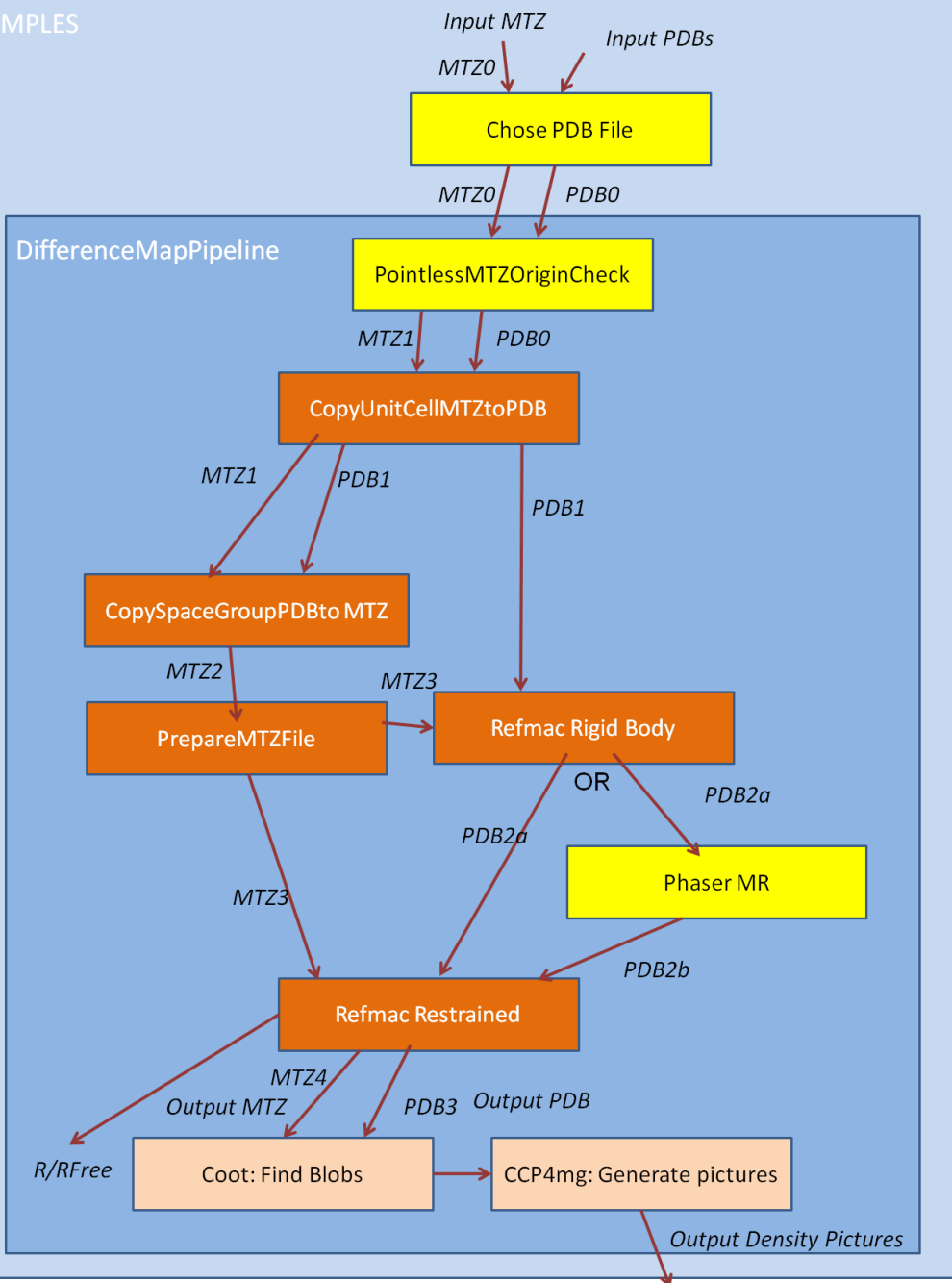


An output map from the DIMPLE pipeline. The blue mesh represents all electron density. The green solid surface indicates an area of positive difference density, i.e. where there is density that is not accounted for by the structure model of the target protein.

⇒ possible location of the bound drug candidate.



DIMPLES





Molecular Replacement

1. What if structure not known?

Similar one?

2. What if target cell/symmetry different?

Position model in new cell?

Suitable search model

Similar to the target structure (this is assumed to be indicated by sequence similarity)

To represent a large fraction of the target structure (or of specific domains)

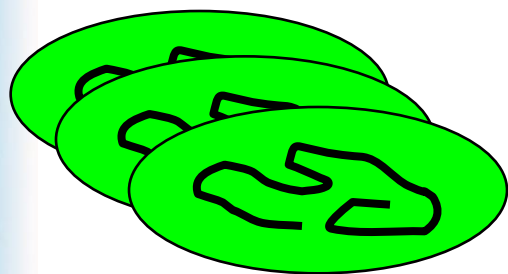
Experimental data

Complete at low resolution to help MR

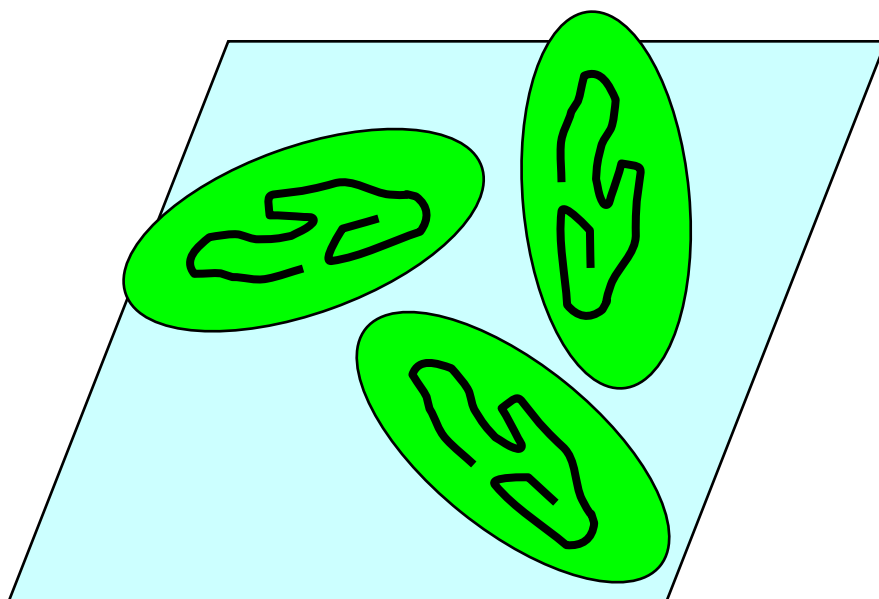
Extend to high resolution to help refinement

MR: What are we trying to do?

Use experimental data to
position protein search
models in the crystal
asymmetric unit



approximate
model of protein



Gives initial phases for model
correction and refinement



Molecular Replacement in CCP4

- MR Programs:
 - *Molrep*
 - automated molecular replacement given an existing template search model
 - *Phaser (MR)*
 - automated molecular replacement based on using likelihood methods to determine the best solution
- Helper programs:
 - Matthews_coef
 - Sfcheck
 - Polarrfn
 - Chainsaw

MR Pipelines in CCP4

- *MrBUMP*
 - automated template model search and preparation through to MR and refinement
 - Brute force approach – emphasis put on generating many search models
 - Can use both Phaser and Molrep for MR
- *BALBES* - Garib talk tomorrow 
 - automated molecular replacement pipeline using a customised version of the PDB database to provide search models for the MR process using Molrep
 - Support for complexes and searching across all related spacegroups



What do we know from the diffraction data?

- Some information on the **spacegroup**:
 - Cannot distinguish between enantiomorphic spacegroups from diffraction data alone, e.g. P43 vs P41
 - May be unsure about some screw axes (*are 00l, l odd really absent??*)
- The **quality** of the experimental intensities.
 - Complete? Saturated at low resolution? Anisotropic?
 - Are the intensity statistics reasonable? Could the crystal be twinned?
- Volume of the asymmetric unit, size of the target protein(s) $\Rightarrow$ likely **number of molecules** it contains (Matthews coefficient).
- Internal arrangement of molecules in asymmetric unit ...
Non-Crystallographic Symmetry (NCS)

Rotational NCS

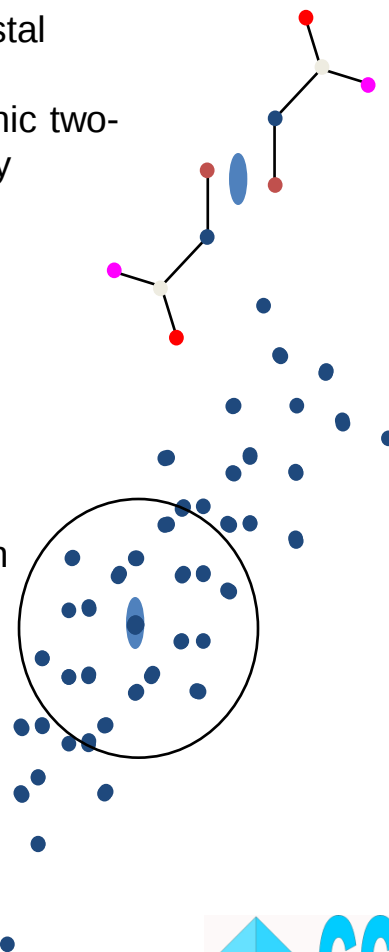
Self rotation function from data alone.

If > 1 molecule in the asymmetric unit, then the **self rotation function** of the Patterson on itself gives a peak at the angle corresponding to the relative rotation between the two.

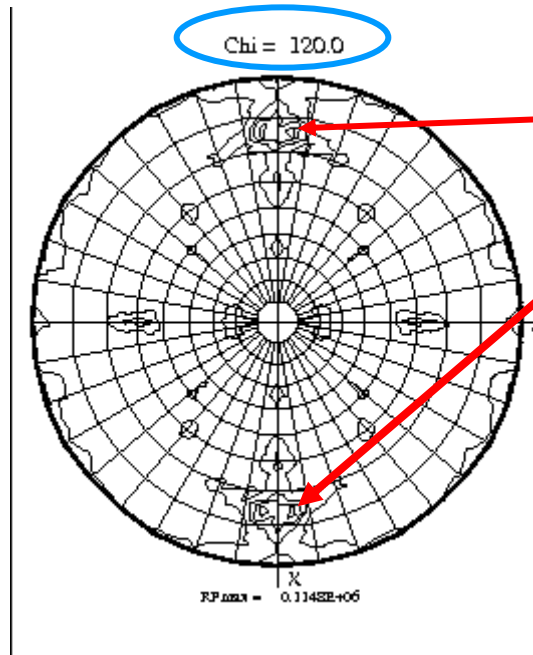
- Confirming how many copies of the structure you have in the asymmetric unit.
- Checking an MR solution
- Used in Locked Rotation Function

Asymmetric unit of unknown crystal with non-crystallographic two-fold symmetry

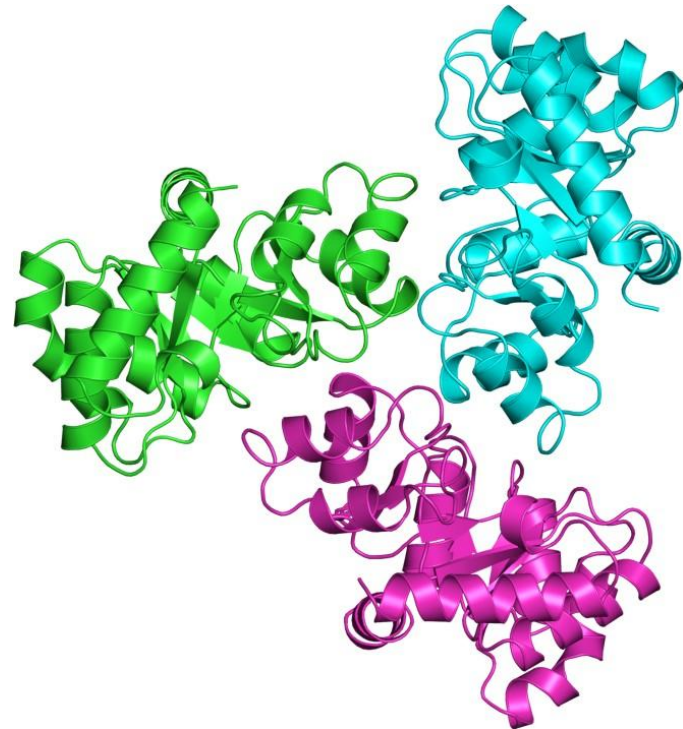
Crystal Patterson has same two-fold symmetry near the origin (intra-molecular peaks only)



Self rotation function for 1vlw



2 symmetry-related 3-folds



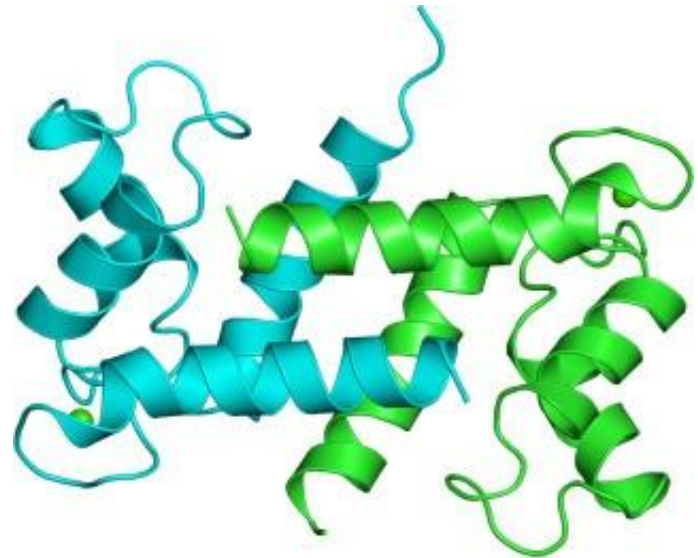
C 2 2 21

Self Rotation Function for S100



H 3

3 symmetry related 2-folds



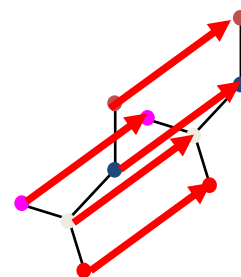
Translational NCS

If the asymmetric unit contains two molecules related by a translation, then the native Patterson will have a large peak at the position representing this translation.

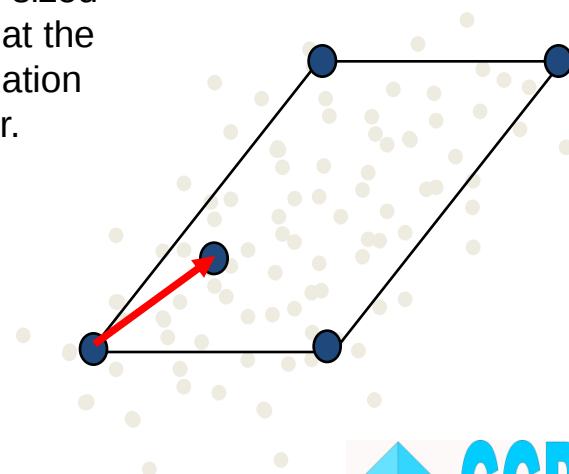
NB Patterson always has peak at origin !

Beware, Patterson peaks can also arise from internal regularities, e.g. helices or DNA

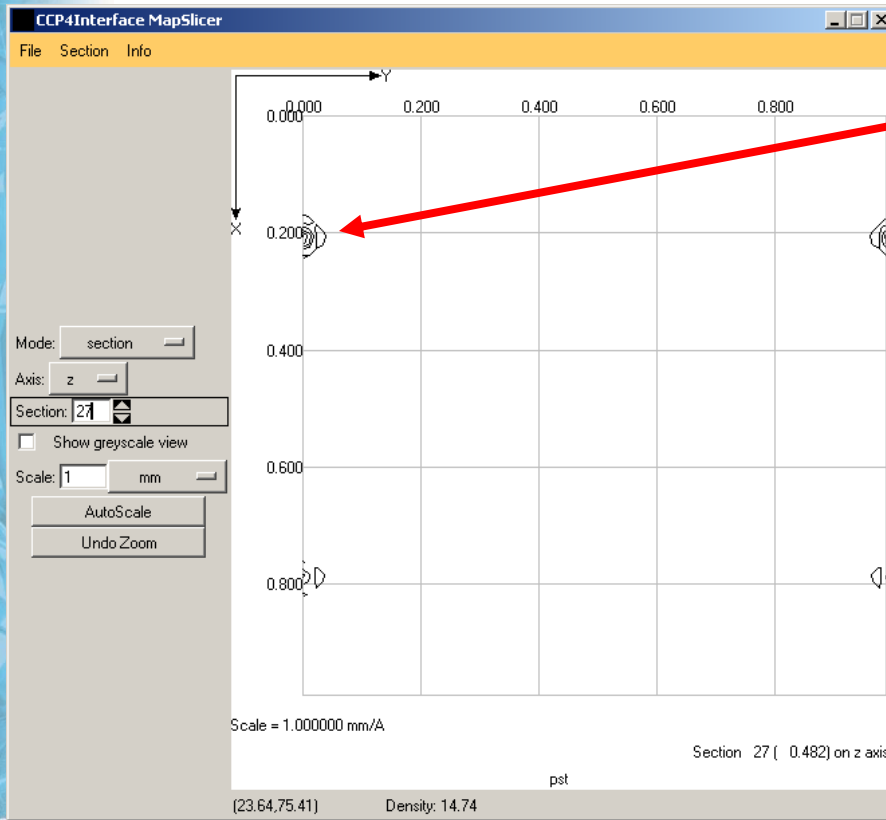
Asymmetric unit of unknown crystal structure with non-crystallographic translation.



Crystal Patterson has origin sized peak at the translation vector.



Native Patterson for pst



Translational NCS vector
(0.209, 0.000, 0.487)

Peak is 0.286 the height of
origin peak.
Molrep looks for peaks higher
than 0.125 of origin peak.



Translational NCS

Non-crystallographic translations introduce awkward structure factor correlations, and can make structures difficult to refine.

Molrep can use NCS translation vector to generate dimer for use in translation search.

Data analysis before MR

Matthews coefficient
Number copies in a.s.u.

Native Patterson
(translational NCS)

Self RF
(rotational NCS)

CCP4 Program Suite 6.1.24 CCP4Interface 2.0.7 running on

Molecular Replacement	
Analysis	☒
Cell Content Analysis	
Analyse Data for MR	
Self RF in polars	
Self RF with molrep	
Model Generation	☐
Run Phaser	
Run Molrep - auto MR	
Run MrBUMP	
Run Balbes	
AMoRe Suite	☐
Utilities	☐

72	01	Mar	1
71	01	Mar	1
70	01	Nov	1
69	01	Nov	1
64	18	Jun	C
60	02	Feb	C
54	12	Jun	C
53	12	Jun	C
44	30	Apr	C
43	30	Apr	C
42	09	Apr	C
41	09	Apr	C
40	16	Jan	C
39	16	Jan	C
30	01	Nov	C
29	01	Nov	C
28	01	Nov	C
27	01	Nov	C
16	31	May	C
9	22	May	C



Molecular Replacement

Traditionally split MR search into 2 steps (cf. EPMR, Queen of Spades, etc.):

1. Determine orientation of search model
 - cross rotation function (**CRF**)
2. Determine position of search model for given orientation
 - translation function (**TF**)
 - check for clashes between symmetry related models

Look at this in context of **Molrep** and **Phaser**

Selection of Search Models

- Search PDB for homologous proteins
- Don't always just take the first
- Look at search model first and edit

Discussed more under MrBUMP and tomorrow.



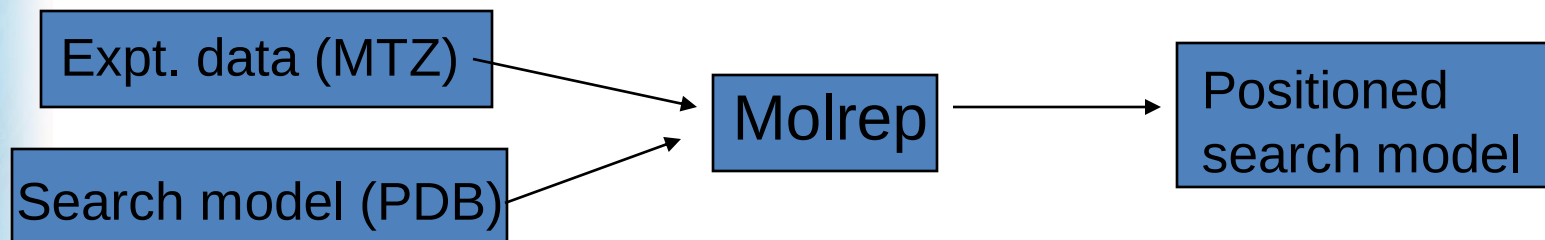
Molrep

Alexei Vagin
University of York

<http://www.ysbl.york.ac.uk/~alexei/molrep.html>

Molrep: overview of functionality

Performs complete MR in single step:



- Individual steps for more difficult cases: CRF, TF, rigid-body
- Self RF, locked CRF
- Multi-copy search: dyad search, multi-monomer
- Phased TF, spherically-averaged phased TF
- Improve search model
- Other search models: electron density map, EM map, NMR models (inc. ensembles of homologous proteins)
- Fit model in electron density map / EM map

MR for straightforward case via GUI

Molrep - Molecular Replacement Initial parameters from /home/mdw/gui_project/CCP4_DATABASE/463_molrep.def

This interface is for version 11.0 of Molrep

Job title 1v3z_B search model

Do molecular replacement performing rotation and translation function

Get input structure factors from MTZ file

☐ Input fixed model

☐ Multi-copy search

☐ Use sequence

MTZ in Full path.. /xtal/ccp4_newCVS/ccp4/examples/mr_tutorial_2006/data/hypF/hypF Browse View

Use ☐ Intensities

FP FP1gxu SIGFP SIGFP1gxu

Model in Full path.. /xtal/ccp4_newCVS/ccp4/examples/mr_tutorial_2006/data/hypF/1v3 Browse View

Coords out testing 1v3z_B_molrep1.pdb Browse View

Experimental Data (Resolution,ANISO,DIFF,BADD,INVER,DSCALE,...) ☐

The Model (SIM,COMPL,SURF,NMR,NCSM,DSCALEM...) ☐

Search Parameters (NMON,NP,NPT,PST,STICK,LOCK,...) ☐

Infrequently Used Parameters (MODE,SAPTF,RAD,PACK,SCORE,LMIN,NOSG) ☐

Run Save or Restore Close

title

mode

MTZ file

MTZ labels

search model

RUN IT!

Other Molrep parameters

SG ALL

Check all compatible spacegroups

High resolution limit

Absolute cut-off (**RESMAX**)

Default estimated

Low resolution cut-off

Molrep uses soft cut-off, B_{off} (**BOFF**)

From minimum resolution (**RESMIN**) or size of search model (**COMPL**)

High resolution cut-off

Molrep uses soft cut-off, B_{add} (**BADD**)

Default from sequence similarity (**SIM**)

$$|F|_{\text{new}} = |F|_{\text{input}} \cdot \exp(-B_{\text{add}} \cdot s^2) \cdot (1 - \exp(-B_{\text{off}} \cdot s^2))$$

Cross Rotation Function

polar angles

R factor

List of top
RF peaks

CCP4I fileviewer 463_molrep.log

Help

--- Peaks of Rotation Function ---

	theta	phi	chi	Rf/sigma
1	168.62	-151.20	33.09	3.94
2	169.00	-163.69	33.53	3.86
3	88.94	76.08	173.73	3.84
4	151.24	-177.80	123.74	3.60
5	151.49	-179.42	124.78	3.59
6	172.08	167.35	76.48	3.52
7	172.41	159.91	77.05	3.51
8	85.29	97.67	175.66	3.50
9	172.63	150.72	77.78	3.44
10	85.26	97.17	172.71	3.42
11	168.57	-166.26	74.62	3.37
12	166.93	-160.55	73.37	3.37
13	73.13	111.42	167.03	3.21
14	168.73	176.49	159.03	3.19
15	158.52	-151.28	60.71	3.15
16	85.15	106.97	179.54	3.11
17	83.05	49.58	174.27	3.10
18	170.42	177.86	43.67	3.03
19	164.11	-177.85	92.09	3.03
20	95.58	-131.50	174.74	2.99
21	67.45	127.63	158.41	2.98

Find Show Log Graphs Show Summary Quit

Translation Function

polar angles

fractional translation

R factor Score

List of top solutions:

--- Summary ---

	RF	TF	theta	phi	chi	tx	ty	tz	TFcnt	wRfac	Score
1	2	1	94.68	-81.60	179.85	0.875	0.718	0.404	5.55	0.559	0.292
2	9	1	85.25	98.00	178.63	0.870	0.717	0.404	5.48	0.566	0.286
3	7	1	85.22	97.75	177.33	0.536	0.050	0.237	4.97	0.569	0.271
4	5	1	85.18	97.57	175.91	0.869	0.715	0.404	3.94	0.574	0.249
5	6	2	85.14	97.27	174.33	0.536	0.046	0.238	3.03	0.577	0.226
6	1	9	168.18	-151.20	33.43	0.966	0.465	0.398	2.20	0.572	0.206
7	8	2	163.58	-175.79	91.12	0.805	0.735	0.397	2.15	0.589	0.205
8	10	3	84.73	96.38	168.47	0.828	0.768	0.245	3.02	0.596	0.196
9	2	4	88.80	76.15	173.60	0.574	0.066	0.238	2.12	0.588	0.195
10	25	5	85.22	106.84	178.36	0.187	0.386	0.445	1.84	0.590	0.193
11	23	3	82.63	49.43	175.94	0.100	0.771	0.243	3.36	0.585	0.193
48	45	10	96.50	-131.77	178.90	0.491	0.427	0.416	2.66	0.601	0.161

Contrast = 3.90

After stick correction:

Move closer to origin

I_sym_operator : 13

new position(frac): 0.208 0.385 0.071

contrast of solution



Identification of solutions

Packing Function (computed from overlap of electron densities, $PF = 1$ means no overlap) integrated into TF search

⇒ downweights solutions with overlapping molecules.

SCORE = product Correlation Coefficient of intensities and maximal value of Packing Function

CONTRAST = ratio of top score to mean score:

>3.0 - definitely solution

<3.0 and > 2.0 - solution

<2.0 and > 1.5 - maybe solution

<1.5 and > 1.3 - maybe not solution, but program accepts it

<1.3 - probably not solution

Finding more than one copy in the asu

By default, Molrep will estimate number of copies to find.

Override with **NMON** keyword

"Multi-copy search" options

CRF

TF for first copy

Fix first copy

TF for second copy

Fix second copy

TF for third copy

⋮

Solving complexes

- Choose first component (largest, highest similarity)
- Solve for first component (probably need to specify **NMON** explicitly)
- New Molrep job
 - Model in** - second component
 - Fixed in** - positioned first component
- Repeat for all other components

Phaser

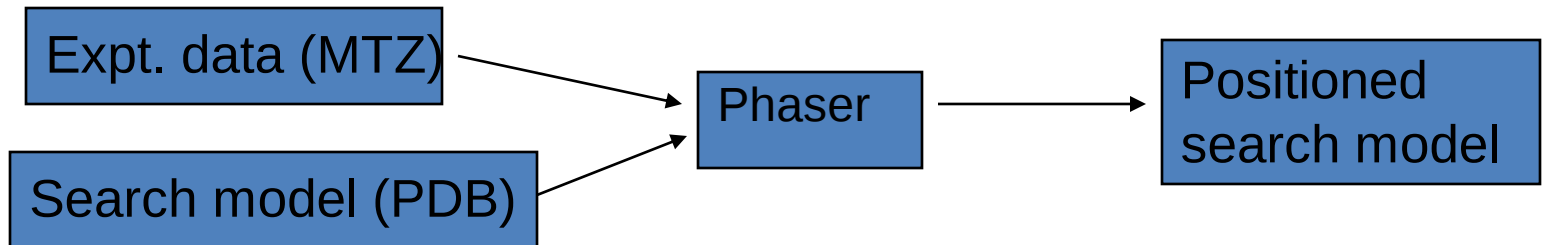


Randy Read, Airlie McCoy, Gabor Bunkoczi
University of Cambridge

<http://www.phaser.cimr.cam.ac.uk/>

Overview

Performs complete MR in single step:



Use “**MODE MR_AUTO**” or “**automated search**” in the GUI

- anisotropy correction
 - fast rotation function
 - fast translation function
 - packing
 - refinement and phasing
-
- loop over models



More functionality ...

- All steps can be run separately
- Search over spacegroups (**SGALTERNATIVE**):
 - MTZ spacegroup and enantiomorph e.g. P41 vs P43
 - All spacegroups in MTZ point-group
 - Selected spacegroups
- Ensemble models (see later)
- Brute RF and TF - slow and accurate
- Normal mode analysis
 - Generates models (.pdb) perturbed along normal modes (e.g. to account for domain movements)

MR for straightforward case

mode

MTZ file

search model

target details

specify search

RUN IT!

Maximum Likelihood Molecular Replacement

Job title 1v3z_B search model

Mode for molecular replacement automated search

Number of processors 2

Define data

MTZ in Full path.. /xtal/ccp4_newCVS/ccp4/examples/mr_tutorial_2006/data/hy Browse View

F Hg SIGF SIGF_Hg

Resolution range 52.129 A to 2.5 A

Space group read from mtz file H 3 2 ; Run Phaser with mtz space group

Define ensembles (models)

Ensemble # 1

Ensemble Name ensemble1 Define ensemble via pdb file(s)

PDB #1 Full path.. /xtal/ccp4_newCVS/ccp4/examples/mr_tutorial_2006/data/ Browse View

Similarity of PDB #1 to the target structure sequence identity 0.38

Edit list Add superimposed PDB file to the ensemble

Edit list Add ensemble

Define composition of the asymmetric unit

Total scattering determined by components in asymmetric unit

Component 1 protein sequence file Number in asymmetric unit 1

SEQ file Full path.. /xtal/ccp4_newCVS/ccp4/examples/mr_tutorial_2006/data/hyp Browse View

Edit list Define another component

Search parameters

Determine ensemble search order automatically on

Allow search with alternative ensembles (models) for a single component of the ASU off

Perform search using ensemble1 Number of copies to search for 1

Edit list Add another search

Additional parameters

Output control

Expert parameters

Run Save or Restore Close

FRF

Euler angles (CCP4)

Top LLG and Z-scores for FRF

CCP4I fileviewer 20_phaser_MR.log

Help

Top Peaks With Clustering From Rescoring of FRF #1

Rank of the peak after clustering search points
 (#) Rank of the peak before clustering search points
 FSS Fast Search Score
 Z-Score Number of standard deviations of FFS above the mean
 Split Distance (in degrees or Angstroms) to top peak
 #Group Number of points clustered in peak
 raw/top FSS of raw (unclustered) peak and top peak in cluster

You requested peaks over 75% of top (i.e. $0.75 * (\text{top} - \text{mean}) + \text{mean}$)
 There were 6 sites over 75% of top
 The sites over 75% are:

#	(#)	Euler1	Euler2	Euler3	LLG	Z-score	Split	#Group	raw/top
1	1	30.3	8.5	235.9	+17.29	4.63	0.0	43	100.0/100.0
		150.3	8.5	235.9					
		270.3	8.5	235.9					
		89.7	171.5	55.9					
		209.7	171.5	55.9					
		329.7	171.5	55.9					
2	16	38.0	8.0	244.0	+16.68	4.50	15.7	29	86.6/ 91.1
		158.0	8.0	244.0					
		278.0	8.0	244.0					
		82.0	172.0	64.0					
		202.0	172.0	64.0					
		322.0	172.0	64.0					
3	47	108.4	5.8	223.6	+15.78	4.32	54.3	30	82.5/ 86.0
		228.4	5.8	223.6					
		348.4	5.8	223.6					
		11.6	174.2	43.6					
		131.6	174.2	43.6					
		251.6	174.2	43.6					
4	20	59.5	7.3	333.2	+15.68	4.30	13.3	62	85.9/ 87.1
		179.5	7.3	333.2					
		299.5	7.3	333.2					
		60.5	172.7	153.2					
		180.5	172.7	153.2					
		300.5	172.7	153.2					
5	73	39.8	12.4	343.3	+13.40	3.84	17.5	1	78.9/ 78.9
		159.8	12.4	343.3					
		279.8	12.4	343.3					
		80.2	167.6	163.3					
		200.2	167.6	163.3					
		320.2	167.6	163.3					
6	209	22.6	46.6	198.9	+11.82	3.53	57.8	1	67.5/ 67.5
		142.6	46.6	198.9					
		262.6	46.6	198.9					
		97.4	133.4	18.9					
		217.4	133.4	18.9					
		337.4	133.4	18.9					

Fast Rotation Function Table

Top	(Z)	Second	(Z)	Third	(Z)
17.29	(4.63)	16.68	(4.50)	15.78	(4.32)

Find Show Summary Quit

FTF

fractional translation

CCP4I fileviewer 20_phaser_MR.log

Help

Top Peaks With Clustering From Rescoring of SET #1 TRIAL #1

Rank of the peak after clustering search points
 (#) Rank of the peak before clustering search points
 FSS Fast Search Score
 Z-Score Number of standard deviations of FFS above the mean
 Split Distance (in degrees or Angstroms) to top peak
 #Group Number of points clustered in peak
 raw/top FSS of raw (unclustered) peak and top peak in cluster

You requested peaks over 75% of top (i.e. $0.75 \times (\text{top} - \text{mean}) + \text{mean}$)
 There were 57 sites over 75% of top
 The sites over 75% are:

#	(#)	Frac X	Frac Y	Frac Z	LLG	Z-score	Split	#Group	raw/top
1	1	0.692	0.691	0.206	+25.09	4.62	0	2	56.60/ 56.60
2	14	0.509	0.989	0.037	+24.74	4.58	29	3	53.10/ 54.87
3	3	0.502	0.981	0.771	+23.07	4.37	12	2	55.89/ 56.07
4	9	0.695	0.695	0.258	+23.06	4.37	8	1	54.10/ 54.10
5	19	0.475	0.844	0.775	+22.62	4.31	16	2	52.43/ 52.43
6	6	0.571	0.450	0.132	+21.80	4.21	17	3	55.18/ 55.18

Find Show Summary Quit

FRF solution number

CCP4I fileviewer 20_phaser_MR.log

Help

TABLES OF RESULTS

Fast Translation Function Table: Space Group R 3 2

#SET	#TRIAL	Top (Z)	Second (Z)	Third (Z)	Ensemble
1	1	25.09 (4.62)	24.74 (4.58)	23.07 (4.37)	ensemble1
1	2	45.14 (7.13)	-	-	ensemble1
1	3	23.91 (5.00)	21.21 (4.63)	20.14 (4.49)	ensemble1
1	4	23.41 (4.53)	23.25 (4.51)	19.77 (4.08)	ensemble1
1	5	21.86 (4.75)	21.46 (4.70)	20.37 (4.56)	ensemble1
1	6	21.01 (4.78)	19.57 (4.61)	19.52 (4.60)	ensemble1

Find Show Summary Quit

Top LLG and Z-scores for FRF

Packing

Phaser does packing check after FTF

Clashes = C α atoms closer than 3Å

Default number of clashes now a percentage of total C α atoms

PACKING FUNCTION

```
There is 1 solution to pack
Packing analysis
0% 100%
|==| DONE
```

Packing Table: Space Group H 3 2

```
Solutions accepted if number of clashes <= 5% of CA atoms
i.e. total number of clashes <= 4 AND each ensemble <= 5%
#  #Clashes #  Accepted Annotation
1   2      1  YES  RFZ=4.4 TFZ=7.0 PAK=2
```

```
1 accepted of 1 solutions
```

If the model is RNA or DNA, phosphate (P) and carbon atoms (C3* and C4*) in the phosphate backbone, and nitrogen atoms in the bases are taken as the marker atoms for clashes.

Solution Files

.sol file produced at end of job

- Contains summary of all solutions
- Each solution contains rotations and usually translations -

3DIM vs 6DIM

- One line per model located
- **.sol file** can be read back into Phaser in later jobs

Z-score	Have I solved it?
less than 5	no
5 - 6	unlikely
6 - 7	possibly
7 - 8	probably
more than 8	definitely

RFZ = RF Z-score

↔ TFZ = TF Z-score

Ensemble models

Phaser refers to search models as “ensembles”

Often, ensemble contains single model, as in traditional MR

But Phaser can use an ensemble of > 1 models, which may work better than any single model

Models in an ensemble must be superposed prior to use in Phaser

- use e.g. **Superpose** in CCP4

N.B. Phaser will complain if:

- MW of models in ensemble are too different
- RMS between models is too large

(In **Molrep**, construct ensemble as pseudo-NMR PDB file)

Finding more than 1 copy in the asu

Specify > 1 in **Composition of the asymmetric unit**
(keyword COMPOSITION ... NUMBER)

Specify > 1 in **Number of copies to search for**
(keyword SEARCH ... NUMBER)

Phaser will issue warnings if these numbers are wrong.

CRF

TF for first copy

Fix first copy (possibly multiple sets)

CRF for second copy

TF for second copy

Fix second copy (possibly multiple sets)

⋮



Complexes

As before, but:

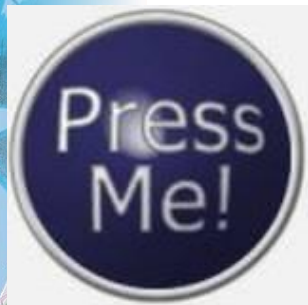
- Define > 1 type of component
 - Define composition of the asymmetric unit**
 - Define another component**
- Define > 1 ensemble
 - Define ensembles (models)**
 - Add ensemble**
- Specify all searches
 - Search parameters**
 - Add another search**

MrBUMP

- An automation framework for Molecular Replacement.
- Particular emphasis on generating a variety of search models.

- Wraps **Phaser** and/or **Molrep**.
- Uses a variety of helper applications (e.g. Chainsaw) and bioinformatics tools (e.g. Fasta, Mafft) to generate search models
- Uses up-to-date on-line databases (e.g. PDB, Scop)

- In favourable cases, gives “one-button” solution
- In Complicated Cases, will suggest likely search models for manual investigation (lead generation)



The Pipeline

Target MTZ
&
Sequence



Target
Details

Template
Search

N templates

Model
Preparation

N x M models

Molecular Replacement
& Refinement

Phase Improvement

*Check scores
and exit or select
the next model*



FAST

— Se

All of t
are ac

These
mode

Other

- S
- C
- FR
- C

- **Sequence based search** using sequence of target structure

These structures are called model **templates**

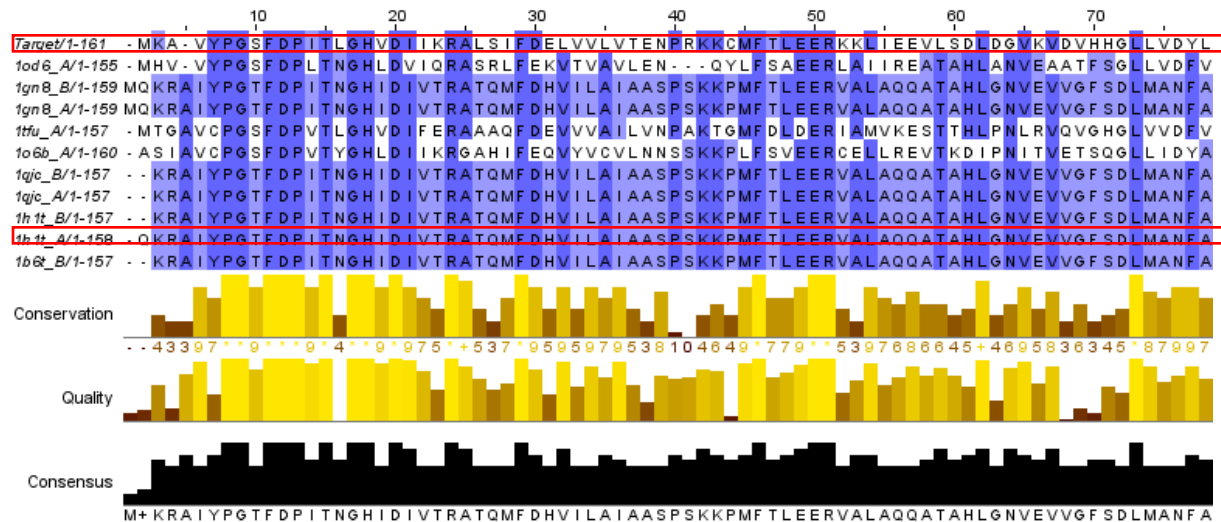
- SSM search using top hit from the FASTA search
- Can add additional PDB id codes to the list, e.g. identified from **FFAS** or **psiBLAST** searches
- Can add local PDB files



Multiple Alignment step

target

model
templates



pairwise
alignment
(used in
Chainsaw)

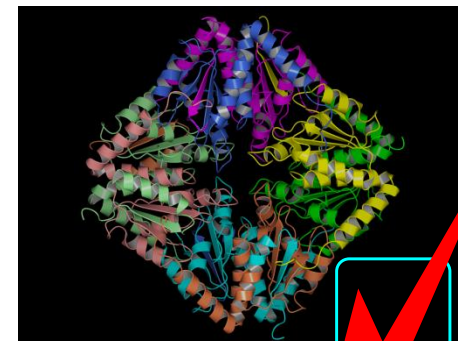
Jalview 2.08.1 Barton group, Dundee

currently support ClustalW, MAFFT, probcons or T-coffee for multiple alignment

Model template scoring: score = sequence identity X alignment quality



Domains
e.g. if relative
domain motion



SCOP

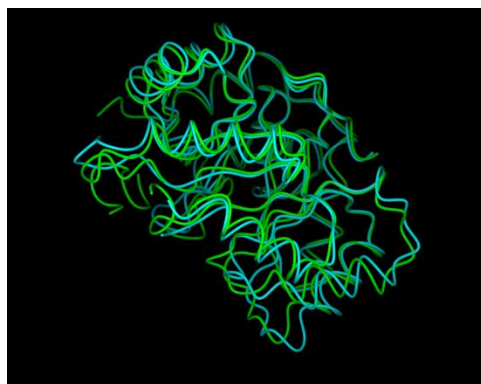
PQS/PISA

template chains

superpose

Multimers

Better signal-to-noise ratio than monomer, *if* assembly is correct for the target.



Ensembles

Create ensembles of search models, for use in additional run of **Phaser**. Need to be similar in MW and rmsd

Search Model Preparation

Search models prepared in four ways:

PDBclip

- original PDB with waters removed, most probable conformations selected and format tidied (e.g. chain ID added)

Molrep

- Molrep model preparation function which aligns the template sequence with the target sequence and prunes the non-conserved side chains accordingly.

Chainsaw

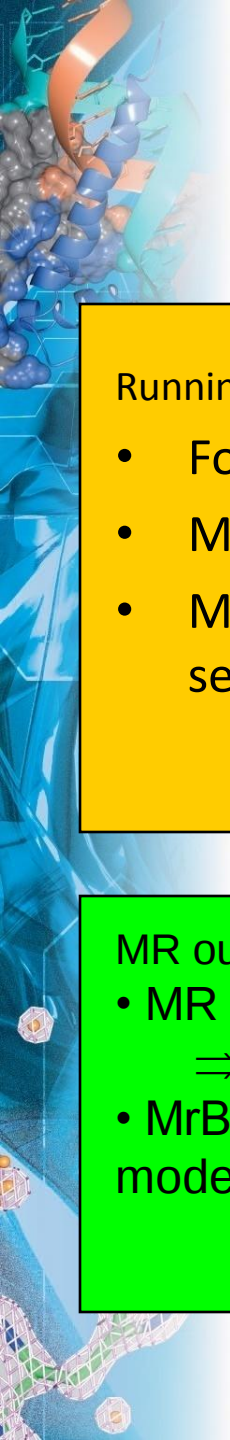
- Can be given any alignment between the target and template sequences. Non-conserved residues are pruned back to the gamma atom.

Polyalanine

more side
chain
truncation

- Created by excluding all of the side chain atoms beyond the CB atom using the Pdbset program

} deal with
deletions



Molecular Replacement step

Running MR

- For each search model, MR done with **Molrep** or **Phaser** or both.
- MR programs run mostly with defaults
- MrBUMP provides LABIN columns, MW of target, sequence identity of search model, number of copies to search for, number of clashes tolerated

MR output

- MR scores and un-refined models available for later inspection
 - ⇒ assess quality of solution, extent of model bias
- MrBUMP doesn't use MR scores, but checks for output file with positioned model, and passes to refinement step

Testing enantiomorphic spacegroups

- 11 pairs of enantiomorphic spacegroups containing screw axes of opposite handedness, e.g. $P4_1$ and $P4_3$)
- usually both need to be tested in MR
- correct spacegroup indicated by TF and packing

Spacegroup from MTZ file: 'P 31 2 1'

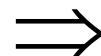
Do MR using enantiomorphic spacegroup as well ☒

- **MrBUMP** can test both in **Molrep** and/or **Phaser**.
- For each search model, best MR results used to fix spacegroup for subsequent steps.
- Discrimination good for good search model + correct MR solution

Restrained Refinement step

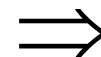
- The resulting models from molecular replacement are passed to **Refmac** for restrained refinement.
- The change in the Rfree value during refinement is used as rough estimate of how good the resulting model is.

final Rfree < 0.35 or
final Rfree < 0.5 and dropped by 20%



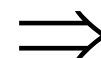
“success”

final Rfree < 0.48 or
final Rfree < 0.55 and dropped by 5%



“marginal”

otherwise



“failure”

conservative

Phase improvement

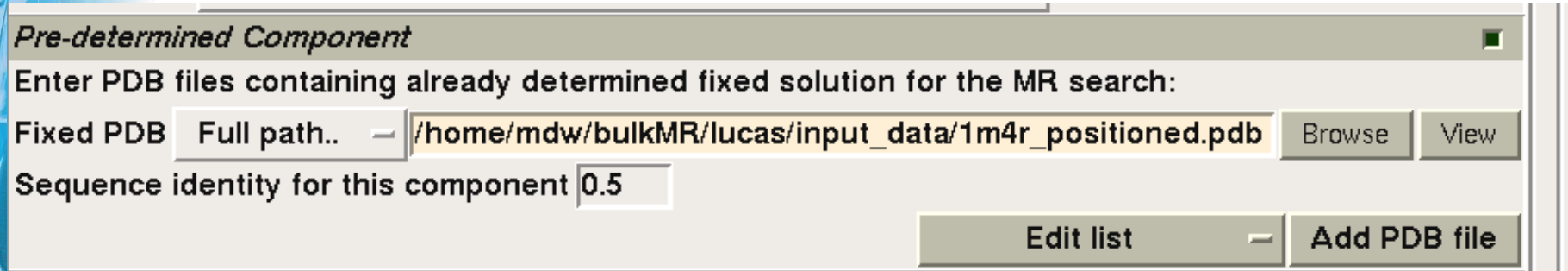
If resolution better than $1.7\text{\AA}$ use Acorn procedure:
initial phase set from refined MR solution
artificial phase extension to $1.0\text{\AA}$
dynamic density modification

Result:

CC for medium Es good indicator of solution
Use E-maps for re-building

Inclusion of fixed models

- MrBUMP will accept one or more positioned models.
- These are included as fixed models in all MR jobs.



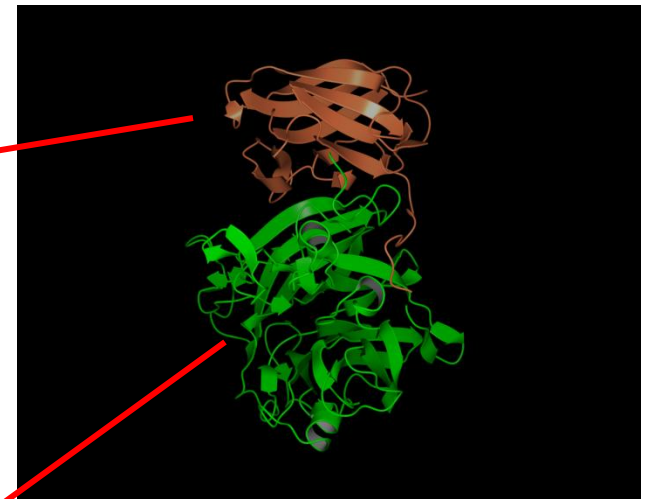
The screenshot shows a software dialog box titled "Pre-determined Component". It contains the instruction "Enter PDB files containing already determined fixed solution for the MR search:". Below this, there is a label "Fixed PDB" followed by a text field labeled "Full path.." containing the file path "/home/mdw/bulkMR/lucas/input_data/1m4r_positioned.pdb". To the right of the text field are "Browse" and "View" buttons. Below the text field is a label "Sequence identity for this component" followed by a text field containing the value "0.5". At the bottom right of the dialog are two buttons: "Edit list" and "Add PDB file".

Thus, solve complexes through consecutive runs of MrBUMP.

Example (thanks to Elien Vandermarliere)

Target is an arabinofuranosidase
Data to 1.55Å in P212121

Small C domain (144 res) solved with 34%
seq ident model
(1w9t_B_MOLREP best out of 4 solutions)



With C domain solution fixed, large N domain (345 res) solved with 28% seq
ident model
(1gyh_C_CHNSAW best out of 7 solutions)

Not yet solved!

Acorn: CC increases from 0.04 to 0.18

ARP/wARP then builds 457/493 residues to R/Rfree 0.185/0.225

MrBUMP output

Log file gives summary of models tried and results of MR

- May get several putative solutions
- *Ease of subsequent model re-building, model completion may depend on choice of solution*
- Worth checking “failed” solutions

Top solution available from ccp4i

Detailed results located in: `<ccp4i project directory>/search_<job number>`

In this directory, there are a number of subdirectories, including:

data

Contains the data files and log files from all jobs run. The directory hierarchy is of the form `<template>/<search model>/<pipeline step>`
e.g. `<ccp4i project directory>/search_55/data/loc0_A/chainsaw/mr`

results

Results from the successful search model are placed into subdirectory "solution". Other results are placed into subdirectory "marginal_solns".